Bioorganic & Medicinal Chemistry Letters 21 (2011) 3676-3681

Contents lists available at ScienceDirect



Bioorganic & Medicinal Chemistry Letters

journal homepage: www.elsevier.com/locate/bmcl



Discovery of XEN907, a spirooxindole blocker of Na_v1.7 for the treatment of pain

Sultan Chowdhury^a, Mikhail Chafeev^a, Shifeng Liu^a, Jianyu Sun^a, Vandna Raina^a, Ray Chui^b, Wendy Young^b, Rainbow Kwan^b, Jianmin Fu^a, Jay A. Cadieux^{a,*}

^a Department of Medicinal Chemistry, Xenon Pharmaceuticals Inc., 3650 Gilmore Way, Burnaby, British Columbia, Canada V5G 4W8 ^b Department of Biological Sciences, Xenon Pharmaceuticals Inc., 3650 Gilmore Way, Burnaby, British Columbia, Canada V5G 4W8

ARTICLE INFO

Article history: Received 17 March 2011 Revised 14 April 2011 Accepted 19 April 2011 Available online 24 April 2011

Keywords: Sodium channel SCN9A Pain Spirooxindole Oxindole

ABSTRACT

Starting from the oxindole **2a** identified through a high-throughput screening campaign, a series of Na_V1.7 blockers were developed. Following the elimination of undesirable structural features, preliminary optimization of the oxindole C-3 and N-1 substituents afforded the simplified analogue **9b**, which demonstrated a 10-fold increase in target potency versus the original HTS hit. A scaffold rigidification strategy then led to the discovery of XEN907, a novel spirooxindole Na_V1.7 blocker. This lead compound, which in turn showed a further 10-fold increase in potency, represents a promising structure for further optimization efforts.

© 2011 Elsevier Ltd. All rights reserved.

Voltage-gated sodium channels have been implicated in a number of neurological pathologies, including epilepsy, migraine, multiple sclerosis, neurodegenerative diseases and neuropathic pain.¹ In particular, the Na_V1.7 subtype (encoded by the *SCN9A* gene), primarily expressed in the peripheral nervous system, has elicited significant attention as a drug target for the treatment of chronic pain. Loss-of-function mutations in *SCN9A* have been linked to a condition known as congenital indifference to pain.² Individuals with such mutations are completely unable to experience pain but otherwise display normal neurological function. On the other hand, corresponding gain-of-function mutations correlate to two distinct pain syndromes: paroxysmal extreme pain disorder and hereditary erythromelalgia.³

In light of this genetic validation of Na_V1.7 as a target for the treatment of chronic pain, a high-throughput screening campaign, predicated on a previously described [¹⁴C]guanidinium influx assay adapted for use in HEK-293 cells expressing human Na_V1.7, was undertaken.^{4,5} From this effort, the 3-hydroxyoxindole analogue **2a** (Scheme 1) was identified as a promising candidate for hit-to-lead activities. The 3-hydroxy-2-oxindole framework is featured in a number of natural products with a broad range of biological activities.⁶ Initial efforts were focused on replacing the β -hydroxyyketone motif present in **2a** in order to obviate potential α , β -unsaturated ketone formation as well as on identifying a suitable

surrogate for the metabolically labile furyl moiety.⁷ Initial substitution of the furyl moiety with a thienyl ring was tolerated (Table 1). Geminal demethylation α to the ketone moiety led to a sharp reduction in potency observed in analogue **4**, whereas truncation of the methylene spacer to afford derivative **7** also led to reduced activity. However, upon further shortening of the spacer to obtain the 3-aryl-3-hydroxyoxindole **3f** (Table 2), modest potency was restored.

The SAR around the 3-aryl substituent was then explored (Table 2). Addition of aryl Grignard reagents or aryllithium species to the N-substituted isatin 1 allowed for the rapid generation of a series of analogues. The 3,4-methylenedioxyphenyl analogue 3a showed the greatest target potency (equipotent with the original HTS hit 2a). The corresponding 3,4-ethylenedioxyphenyl and 3,4-propylenedioxyphenyl analogues 3c and 3f demonstrated a decrease in potency with increasing ring size. The methoxypyridine analogue **3b** and 3-methoxyphenyl derivative **3d** were both potent, suggesting a preference for a H-bond acceptor at the aryl ring's 3-position. In contrast, both the 2-methoxyphenyl and the 4-methoxyphenyl analogues **3h** and **3j** were significantly less potent. The replacement of the 3,4-methylenedioxyphenyl moiety in **3a** with the corresponding monocyclic 3,4-dimethoxyphenyl ring in 3i was poorly tolerated. Geminal difluorination of the dioxolane carbon in **3a** also led to a reduction in potency (**3k**).

The next phase of optimization involved variation of the oxindole N-substituent while preserving the 3-(3,4-methylenedioxyphenyl) moiety present in **3a** (Scheme 2, Table 3). While the N-unsubstituted compound **9r** was poorly active, a wide variety

^{*} Corresponding author. Tel.: +1 604 484 3300x157; fax: +1 604 484 3321. *E-mail address:* jcadieux@xenon-pharma.com (J.A. Cadieux).

⁰⁹⁶⁰⁻⁸⁹⁴X/ $\$ - see front matter \odot 2011 Elsevier Ltd. All rights reserved. doi:10.1016/j.bmcl.2011.04.088



Scheme 1. Reagents and conditions: (a) *p*-chlorobenzyl chloride, NaH, DMF, 1 h; (b) *i*Pr₂NH (cat.), EtOH, reflux, 1 h; (c) ArMgX, 0 °C to rt or ArLi, -78 °C to rt, THF, 2-16 h; (d) (i) *N*-methylaniline, EtMgBr, PhH, rt, 1 h; (ii) **1**, THF, -15 °C to rt, 3 h; (e) HS(CH₂)₃SH, BF₃-Et₂O (cat.), CH₂Cl₂, rt, 24 h; (f) "BuLi, THF, -78 °C then **1**, -78 °C to rt, 16 h; (g) PhI(OCOCF₃)₂, MeCN/H₂O, 0 °C to rt, 2.5 h.

Table 1								
Guanidinium	influx	assay	potencies	of	initial	3-hydroxyoxindole	analogues	and
reference con	pound	s						

Compound	$hNa_V 1.7 \ IC_{50} \ (\mu M)$	c log P
2a	0.30	2.50
2b	0.20	3.87
4	6.08	5.14
7	>10	3.92
Tetrodotoxin	0.006	<-1
Tetracaine	7.51	0.41

of substituted alkyl and benzyl moieties were well tolerated at that position. Of note, potency was invariant with respect to the electronic nature of the benzyl substituent (c.f. compounds **9d**, **9e** and **9o**). Amongst the *N*-alkyl derivatives, branching (e.g., **9c** and **9i**) as well as the inclusion of saturated carbocycles (e.g., **9a**, **9j**, **9m**, and **9n**) were also tolerated. When compared to the starting *p*-chlorobenzyl compound **3a**, several analogues demonstrated improved activity along with reduced lipophilicity. The simple *n*-pentyl derivative **9b** was selected over the equipotent but somewhat less soluble and permeable (2-cyclopropyl)ethyl analogue **9a** as the starting point for investigations aimed at further improvements by replacement of the potentially labile tertiary alcohol.

Derivatization of the tertiary alcohol in **9b** to the corresponding methyl ether **12** was not tolerated (Scheme 3, Table 4). Triethylsilane-mediated dehydroxylation of **9b** afforded the key intermediate **10**.⁸ Treatment of **10** with sodium hydride, followed by alkylation with iodomethane furnished the modestly active 3methyl analogue **13**. A lanthanide triflate-catalyzed aldol condensation between *p*-formaldehyde and oxindole **10** yielded the hydroxymethylene compound **11**, which was ca. 20-fold less potent than **9b**.⁹ Deprotonation of the intermediate **10**, followed by treatment with methyl cyanoformate led to the carboxylic ester



Scheme 2. Reagents and conditions: (a) RX, NaH, DMF, rt, 1-16 h; (b) 3,4-methylenedioxyphenylmagnesium bromide, THF, 0 °C to rt, 16 h.

14, which was subsequently elaborated to the carboxamide derivatives **15** and **16** with loss of target potency. However, conversion of the ester **14** to the corresponding acyl hydrazide, followed by oxidation and Curtius rearrangement, provided the highly potent 3-amino-3-aryloxindole **17**.¹⁰ Finally, alkylation of intermediate **10** with either methyl 3-bromopropionate or methyl bromoacetate led to a series of inactive homologated esters, acid and amides (**18**– **23**).

In light of the results obtained with the 3-methyl derivative **13**, which suggested that substitution of the oxindole 3-position with an aliphatic moiety allowed for some target potency to be retained, a rigidification strategy involving the preparation of a number of spirooxindole derivatives was then explored (Scheme 4). Addition of the Grignard reagent obtained from treating sesamol (3,4-meth-ylenedioxyphenol) with isopropylmagnesium chloride to the substituted isatin **8b** afforded the diol **24**.¹¹ Triethylsilane-mediated dehydroxylation to the phenol derivative **25**, followed by aldol condensation and intramolecular cyclization under Mitsunobu conditions afforded spiroether **29** (XEN907).^{12,13} This analogue was 10-fold more active than the most potent acyclic oxindole analogue **9b** (Table 5). The estimated lipophilicity of XEN907 ($c \log P 3.97$) is comparable to or less than those of some reported sodium channel blockers under investigation for the

Table 2

SAR of 1-(4'-Chlorobenzyl)-3-aryl-3-hydroxyoxindoles



Compound	Ar	hNa _v 1.7 IC ₅₀ (μM)	c log P	Solubility ^a (µg/mL)	Permeability ^b (×10 ⁻⁶ cm/s)
3a	3,4-Methylenedioxyphenyl	0.20	4.19	<5	9.0/10.9
3b	(2-Methoxypyridin)-5-yl	0.27	4.08	<5	ND ^c
3c	3,4-Ethylenedioxphenyl	0.48	3.91	9	1.9/3.5
3d	3-Methoxyphenyl	1.19	4.28	6	ND
3e	Phenyl	1.58	4.41	5	13.1/12.5
3f	3,4-Propylenedioxyphenyl	3.21	4.02	<5	ND
3g	3-Furyl	4.17	3.02	11	ND
3h	2-Methoxyphenyl	6.08	4.28	<5	ND
3i	3,4-Dimethoxyphenyl	6.14	4.15	26	ND
3j	4-Methoxyphenyl	7.37	4.28	<5	ND
3k	2,2'-Difluoromethylenedioxyphenyl	>5	5.59	<5	ND
31	(Benzo[b]furan)-6-yl	>5	4.02	<5	ND

^a Solubility at pH 7.4 in phosphate-buffered saline.

^b Permeability $(a \rightarrow b/b \rightarrow a)$ determined in CaCo-2 cells.

^c ND = not determined.

treatment of pain, such as PPPA¹⁴ ($c \log P 4.25$) or BZP¹⁵ ($c \log P 5.95$). The corresponding five- and six-membered carbocyclic derivatives were prepared from the 3-aryloxindole derivative **10** by treatment with an excess of sodium hydride, followed by alkyl-

ation with a suitable bromoester. Following saponification and acyl halide formation, the poorly active ketones **27** and **28** were obtained via tin (IV) chloride-promoted internal Friedel–Crafts acylation.¹⁶ Reduction of the benzylic ketone with triethylsilane

Table 3

SAR of 1-substituted-3-(3,4-methylenedioxyphenyl)-3-hydroxyoxindoles



Compound	R	hNa _v 1.7 IC ₅₀ (μM)	c log P	Solubility ^a (µg/mL)	Permeability ^b $(a \rightarrow b/b \rightarrow a)$ $(\times 10^{-6} \text{ cm/s})$
9a	(2-Cyclopropyl)ethyl	0.02	2.97	11	19.9/19.8
9b	n-Pentyl	0.03	3.56	37	28.9/25.4
9c	3-Methylpentyl	0.04	3.89	14	10.5/11.5
9d	4-Methoxybenzyl	0.04	3.50	6	6.5/11.9
9e	Benzyl	0.06	3.63	<5	9.3/13.6
9f	(5-Chloro-2-thienyl)methyl	0.06	3.98	<5	ND ^c
9g	3,4-Methylenedioxybenzyl	0.06	3.41	14	7.7/12.1
9h	4,4,4-Trifluorobutyl	0.07	3.43	9	ND
9i	Isopentyl	0.07	3.47	<5	ND
9j	(Cyclopropyl)methyl	0.08	2.62	22	ND
9k	n-Hexyl	0.09	3.97	10	ND
91	4-Fluorobenzyl	0.09	3.79	<5	ND
9m	(Cyclohexyl)methyl	0.11	3.87	<5	ND
9n	(Cyclobutyl)methyl	0.13	3.04	38	21.7/21.1
90	3,4-Difluorobenzyl	0.14	3.95	<5	ND
3a	4-Chlorobenzyl	0.20	4.19	<5	9.0/10.9
9p	4-(Trifluoromethyl)benzyl	0.22	4.55	<5	5.4/7.9
9q	(1-Napthyl)methyl	2.14	4.63	<5	ND
9r	Н	>5	1.66	56	ND

^a Solubility at pH 7.4 in phosphate-buffered saline.

^b Permeability $(a \rightarrow b/b \rightarrow a)$ determined in CaCo-2 cells.



Scheme 3. Reagents and conditions: (a) Et₃SiH, TFA, DCM, rt, 6 h; (b) (i) TMSCl, NEt₃, CH_2Cl_2 , 0 °C, 2 h; (ii) (CH_2O)_n, Yb(OTf)₃ (cat.), THF, rt, 48 h; (c) MeI, NaOMe, THF, 0 °C to rt, 16 h; (d) MeI, NaH, THF, 0 °C, 1 h; (e) NaH, NCCO₂Me, THF, 0 °C to rt, 2 h; (f) MeNH₂ or BnNH₂, MeOH, sealed tube, 70–100 °C, 16 h; (g) hydrazine monohydrate, *n*-PrOH, 100 °C, 2 h; (h) NaNO₂, AcOH, 10 °C, 1 h; (i) *t*-BuOH, PhMe, reflux, 1 h then HBr, HOAc, rt, 1 h; (j) (c) NaH, THF, 0 °C, 0.5 h, then methyl 3-bromoproionate or methyl bromoacetate, rt; (k) LiOH, THF/H₂O, rt, 16 h; (l) (i) (COCl)₂, DMF (cat.), PhMe, rt, 4 h; (ii) amine, NaHCO₃, CH₂Cl₂, rt, 16 h.

Table 4

SAR of 1-pentyl-3-(3,4-methylenedioxyphenyl)-3-substituted oxindoles



Compound	R	hNa _v 1.7 IC ₅₀ (μM)	c log P	Solubility ^a (µg/mL)	Permeability ^b (×10 ⁻⁶ cm/s)
9b 10	ОН	0.03	3.17	37 6	28.9/25.4
10	п	1./4	4.10	0	ND
11	CH ₂ OH	0.63	3.82	<5	ND
12	OMe	>5	3.92	<5	ND
13	Me	1.63	4.89	<5	2.4/3.6
14	CO ₂ Me	1.90	4.22	<5	ND
15	CONHMe	>5	3.54	10	ND
16	CONHBn	1.10	5.27	<5	ND
17	NH ₂	0.03	3.56	34	22.0/22.3
18	CH ₂ CO ₂ Me	>5	4.16	<5	ND
19	$(CH_2)_2CO_2Me$	>5	4.58	<5	ND
20	CH ₂ CO ₂ H	>5	3.90	73	ND

Table 4 (continued)

Compound	R	hNa _V 1.7 IC ₅₀ (μM)	c log P	Solubility ^a (µg/mL)	Permeability ^b (×10 ⁻⁶ cm/s)
21	(CH ₂) ₂ CO ₂ H	>5	4.32	35	ND
22	CH_2CONH_2	>5	3.25	35	ND
23	CH ₂ CONMe	>5	3.48	31	ND
24	CH ₂ CONMe ₂	>5	3.72	<5	ND

^a Solubility at pH 7.4 in phosphate-buffered saline.

^b Permeability $(a \rightarrow b/b \rightarrow a)$ determined in CaCo-2 cells.

^c ND = not determined.

afforded the carbocycles **30** and **31**, which themselves were modestly active against $Na_V 1.7$. Finally, the regioisomeric tetrahydrofuran spirocycle **35** was prepared from the bromoaldehyde **32** by means of a reduction/metal-halogen exchange/carbonyl addition sequence, followed by Mitsunobu cyclization. Notably, this analogue was ca. 400-fold less potent than its regioisomer XEN907.

XEN907 showed no significant activity at 10 μ M against a broad panel of 63 receptors and transporters. Determination of the ADME properties of XEN907 (Table 6) revealed that the compound was not cytotoxic and had favourable hepatocyte metabolic stability for both human and dog, although inhibition of CYP3A4 was observed in a recombinant human enzyme assay.



Scheme 4. Reagents and conditions: (a) sesamol, iPrMgCl, THF, 0 °C to rt, 16 h; (b) Et₃SiH, TFA, CH₂Cl₂, rt, 6 h; (c) (i) (COCl)₂, DMF (cat.), PhMe, rt, 4 h; (ii) SnCl₄, CH₂Cl₂, 0 °C to rt, 16 h; (d) (i) TMSCl, NEt₃, CH₂Cl₂, 0 °C, 2 h; (ii) (CH₂O)_n, Yb(OTf)₃ (cat.), THF, rt, 48 h; (e) PPh₃, DEAD, THF, -10 °C to rt, 16 h; (f) Et₃SiH, TFA, rt, 16 h; (g) NaBH₄, THF, 0 °C to rt, 10 h; (h) ⁿBuLi, THF, -78 °C, 0.5 h then isatin **8b**, -78 °C to rt, 16 h.

Table 5

SAR of spirocyclic analogues

Compound	hNa _V 1.7 IC ₅₀ (μM)	c log P	Solubility ^a (µg/mL)	Permeability ^b (a→b/ b→a) (×10 ⁻⁶ cm/s)
27	3.34	3.52	<5	ND ^c
28	1.81	3.94	<5	ND
29 (XEN907)	0.003	3.97	7.3	5.9/7.2
30	0.44	4.87	<5	1.1/1.6
31	1.48	5.29	<5	ND
35	1.18	3.62	<5	ND

^a Solubility at pH 7.4 in phosphate-buffered saline.

^b Permeability $(a \rightarrow b/b \rightarrow a)$ determined in CaCo-2 cells.

^c ND = not determined.

Table 6

Selected ADME properties of 29 (XEN907)

Property	Value for 29
Plasma protein binding (rat)	97%
Cytotoxicity (HepG2, % viable after 16 h)	>99%
Hepatocyte stability, % remaining after 2 h (rat/human/dog)	21%/34%/46%

Pharmacokinetic analysis in rats of XEN907 (Table 7) demonstrated that, consistent with the compound's ADME parameters,

Table 7

Mean pharmacokinetic parameters of 29 (XEN907)) after administration to male lewis
rats ($n = 3/\text{group}$)	

Parameter	po (10 mg/kg)	iv (3 mg/kg)
$C_{\rm max} (\rm ng/mL)$	35 ± 26	953 ± 96
AUC _{last} (h ng/mL)	143 ± 61	320 ± 52
$t_{1/2}$ (h)	_	2.6 ± 0.1
$T_{\rm max}$ (h)	0.33	-
V _{ss} (L/kg)	_	35.0 ± 4.5
Cl (L/h/kg)	_	9.4 ± 1.7
F (%)	13	-

the compound was modestly bioavailable. Following an initial rapid absorption phase (oral $T_{\rm max}$ = 20 min), XEN907 was extensively distributed ($V_{\rm ss} \sim 600$ -fold higher than the plasma volume in rats) and rapidly cleared.

In summary, progressive elaboration of the HTS hit **2a** via structural simplification to the 3-aryl-3-hydroxyoxindole **9b**, preliminary optimization of the N1- and C3-substituents and rigidification to a spirooxindole framework led to the identification of the highly potent ($IC_{50} = 3 \text{ nM}$) lead XEN907. Further optimization efforts in this structural class aimed at defining analogues with improved physico-chemical and ADME properties will be described in due course.

Acknowledgments

The authors are grateful to Audrey Wang, Xing Cheng, Jing Zhong and Ivana Rajlic (for technical assistance) as well as Henry Verschoof (for helpful discussions during the preparation of this Letter).

References and notes

- (a) Mantegazza, M.; Curia, G.; Ragsdale, D. S.; Avoli, M. Lancet Neurol. 2010, 9, 413; (b) Termin, A.; Martinborough, E.; Wilson, D. Ann. Rept. Med. Chem. 2008, 43, 43; (c) Catterall, W. A.; Goldin, A. L.; Waxman, S. G. Pharmacol. Rev. 2005, 57, 397; (d) England, S. Expert Opin. Invest. Drugs 2008, 17, 1849; (e) Dib-Hajj, S. D.; Cummins, T. R.; Black, J. A.; Waxman, S. G. Ann. Rev. Neurosci. 2010, 33, 325.
- (a) Goldberg, Y. P.; MacFarlane, J.; MacDonald, M. L.; Thompson, J.; Dube, M. P.; Mattice, M.; Fraser, R.; Young, C.; Hossain, S.; Pape, T.; Payne, B.; Radomski, C.; Donaldson, G.; Ives, E.; Cox, J.; Younghusband, H. B.; Green, R.; Duff, A.; Boltshauser, E.; Grinspan, G. A.; Dimon, J. H.; Sibley, B. G.; Andria, G.; Toscano, E.; Kerdraon, J.; Bowsher, D.; Pimstone, S. N.; Samuels, M. E.; Sherrington, R.; Hayden, M. R. *Clin. Genet.* **2007**, *71*, 311; (b) Cox, J. J.; Reimann, F.; Nicholas, A. K.; Thornton, G.; Roberts, E.; Springell, K.; Karbani, G.; Jafri, H.; Mannan, J.; Raashid, Y.; Al-Gazali, L.; Hamamy, H.; Valente, E. M.; Gorman, S.; Williams, R.; McHale, D. P.; Wood, I. N.; Gribble, F. M.; Woods, C. G. *Nature* **2006**, *444*, 894.
- (a) Yang, Y.; Wang, Y.; Li, S.; Xu, Z.; Li, H.; Ma, L.; Fan, J.; Bu, D.; Lui, B.; Fan, Z.; Wu, G.; Jin, J.; Ding, B.; Zhu, X.; Shen, Y. J. Med. Genet. 2004, 41, 171; (b) Cummins, T. R.; Dib-Hajj, S. D.; Waxman, S. G. J. Neurosci. 2004, 24, 8232; (c) Fertleman, C. R.; Baker, M. D.; Parker, K. A.; Moffat, S.; Elmslie, F. V.; Abrahamsen, B.; Ostman, J.; Klugbauer, N.; Wood, J. N.; Gardiner, R. M.; Rees, M. Neuron 2006, 52, 767.

- Reddy, N. L.; Fan, W.; Magar, S. S.; Perlman, M. E.; Yost, E.; Zhang, L.; Berlove, D.; Fischer, J. B.; Burke-Howie, K.; Wolcott, T.; Durant, G. J. *J. Med. Chem.* **1998**, *41*, 3298.
- 5. All $\ensuremath{\mathsf{IC}}_{50}$ data reported herein are means of no fewer than two replicate experiments.
- 6. For a review, see: Peddibhotla, S. Curr. Bioact. Compd. 2009, 5, 20.
- (a) Peterson, L. A.; Cummings, M. E.; Vu, C. C.; Matter, B. A. Drug Metab. Dispos. 2005, 33, 1453; (b) Peterson, L. A. Drug Metab. Rev. 2006, 38, 615.
- (a) Melch, W. M.; Williams, M. T. J. Labelled Compd. Radiopharm. 1993, 33, 119;
 (b) Brueckner, C.; Holzinger, H.; Reissig, H.-U. J. Org. Chem. 1988, 52, 2450.
- (a) Kobayashi, S.; Hachiya, I. Tetrahedron Lett. **1992**, 33, 1625; (b) Kobayashi, S.; Hamada, T.; Nagayama, S.; Manabe, K. Org. Lett. **2001**, 3, 165.
- 10. Koza, G.; Oezcan, S.; Sahin, E.; Balci, M. Tetrahedron 2009, 65, 5973.
- 11. Kunihiro, M.; Yokomizo, K.; Uyeda, M. Chem. Pharm. Bull. 2002, 50, 298.
- (a) Krohn, K.; Ahmed, I.; Markus, J. Synthesis 2009, 779; (b) Wilkinson, M. C.; Bell, R.; Landon, R.; Nikiforov, P. O. Synlett 2006, 2151.
- 13. All compounds described herein were characterized by ¹H NMR, ¹³C NMR and ESI-MS and had purities ≥98% a/a by HPLC. Characterization data for **29** (XEN907): mp 89–90 °C; ¹H NMR (300 MHz, CDCl₃) δ 7.34–7.29 (m, 1H), 7.19–7.15 (m, 1H), 7.07–7.01 (m, 1H), 6.92–6.87 (m, 1H), 6.51 (s, 1H), 6.12 (s, 1H), 5.88–5.84 (m, 2H), 4.91, 4.66 (ABq, J_{AB} = 8.9 Hz, 2H), 3.87–3.64 (m, 2H), 1.77–1.65 (m, 2H), 1.41–1.32 (m, 4H), 0.91 (t, *J* = 7.0 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 177.4, 156.0, 148.9, 142.5, 142.4, 132.6, 129.0, 124.0, 132.2, 119.7, 108.7, 103.1, 101.6, 93.7, 80.6, 58.3, 40.5, 29.1, 27.2, 22.5, 14.1; MS (ESI+) *m*/*z* 352.3 (M+1); purity (HPLC, UV at 254 nm) 99.2% a/a.
- Ilyin, V. I.; Pomonis, J. D.; Whiteside, G. T.; Harrison, J. E.; Pearson, M. S.; Mark, L.; Turchin, P. I.; Gottshall, S.; Carter, R. B.; Nguyen, P.; Hogenkamp, D. J.; Olanrewaju, S.; Benjamin, E.; Woodward, R. M. J. Pharmacol. Exp. Ther. 2006, 318, 1083.
- McGowan, E.; Hoyt, S. B.; Li, X.; Lyons, K. A.; Abbadie, C. Anesth. Analg. 2009, 109, 951.
- (a) Starflinger, W.; Kresze, G.; Huss, K. J. Org. Chem. 1986, 51, 37; (b) Han, D. I.; Oh, D. Y. Synth. Commun. 1989, 19, 2213.