Liebigs Ann. Chem. 1983, 1278-1291

Synthesis of Reserpine-type Alkaloids, II¹⁾

Synthesis of 18-Hydroxyyohimbine Stereoisomers and Raunescine Analogues with Normal Skeletons

Csaba Szántay *^{a,b}, Katalin Honty^a, Gábor Blaskó^b, Eszter Baitz-Gács^b, and Pál Kolonits^a

Department of Organic Chemistry, Technical University^a, Gellért tér 4, H-1521 Budapest XI Central Research Institute for Chemistry, Hungarian Academy of Sciences^b, Pusztaszeri ut 59-67, H-1525 Budapest II

Received January 3, 1983

Starting from the previously described keto ester 2, three raunescine analogues (7c, 9c, 10c) and one isoraunescine stereoisomer (10d) – all with a normal yohimbane skeleton – have been synthesized.

Synthese von Alkaloiden des Reserpin-Typs, II¹⁾. – Synthese von 18-Hydroxyyohimbin-Stereoisomeren und Raunescin-Derivaten mit normalem Molekülgerüst

Ausgehend von dem in der vorausgehenden Mitteilung beschriebenen Ketoester 2 wurden drei Raunescin-Derivate (7c, 9c, 10c) und ein Isoraunescin-Stereoisomer (10d) – alle mit normalem Yohimbangerüst – synthetisiert.

In our previous paper¹ we described the preparation of the keto ester 2 which can be used for the synthesis of 18-substituted yohimbine derivatives with D/E-*trans* as well as D/E-*cis* annulation. Proceeding with our studies in the field of reserpine-type alkaloids, we intended to convert 2 primarily into raunescine² analogues with a normal skeleton (1a-type compounds), so far unknown as natural substances.

Although Potier³⁾ and later Court⁴⁾ et al. reported the isolation of an "18-hydroxyyohimbine fraction" (one of the **1b** stereoisomers) from various Rauwolfia species, however, the exact steric structure of this alkaloid has not been given.

Thus, compounds of type 1 might be interesting not only as potentially biologically active substances, but also because of their possible identity with natural compounds.

A) Preparation of *trans*-2,3-substituted indolo[2,3-a]quinolizidines

In order to build up ring E the first task is to convert the keto function of 2 into a (methoxycarbonyl)methyl group in a way that the C-2 and C-3 substituents of the quinolizidine ring come into relative *trans* position. For this purpose, ketone 2 was reacted with the anion of diethyl [(methoxycarbonyl)methyl]phosphonate in dimethylformamide. According to our previous experience⁵⁾ under such conditions

there is no significant epimerization at C-3. The reaction produced the unsaturated ester 3 in about 80% yield. Investigation on compounds with related structures in the emetine and despirroloyohimbane series⁶⁾ revealed that if the C-3 substituent is in equatorial position the Z isomer is sterically overcrowded to be formed in observable quantities. According to the chemical shift of 1-H_{eq} in the proton spectrum ($\delta \approx 3.9$)^{6b)}, the exocyclic double bond has E geometry (see Experimental).



The catalytic hydrogenation of 3 proceeds with high stereoselectivity, from the reaction mixture only the *trans*-2,3 diester 4 could be isolated. The steric structure is confirmed by both spectroscopic data and chemical transformations. Some of the characteristic peaks of 3 and 4 in their ¹H and ¹³C NMR spectra appeared as doubled signals due to the fact that these products are mixtures of diastereomeric racemates.

The yohimbane skeleton was formed from 4 by Dieckmann ring closure carried out in benzene in the precence of potassium *tert*-butoxide. The resulting two diastereomers (5 and 6) could be separated by crystallization. NMR and mass spectrometric data indicated unambiguosly that the methoxycarbonyl group in both derivatives is in position 16, therefore the ring closure, as expected, took place with high regioselectivity.



Although 5 and 6 have a β -keto carboxylate structure, none of them shows any enolization tendency which can be attributed to the "*peri* effect". This phenomenon was first observed by *Wenkert*⁷ in the case of yohimbinone. The lack of enolization also supports the D/E-*trans* annulation in the skeleton of 5 and 6 because in the D/E-

cis fused allo and epi-allo series, enolization of the analogue β -keto esters is not hindered⁸⁾.

According to further information obtained from the ¹H NMR spectra, 18-OCH₃ is pseudoequatorial (α) in **5** and pseudoaxial (β) in **6** (see the respective coupling constants in Table 1). The stereochemistry at C-16 could not be deduced from the ¹H NMR spectra, since the signals of the protons in position 16 are overlapped by other signals. On the basis of the chemical shift values^{9,10} of C-16 and C-20, the C-16 substituent in compounds **5** and **6** proved to be in the same steric position (pseudoequatorial, α) as in yohimbinone, a natural keto ester⁹. Since the additional substituent, the C-18 methoxy group in compound **5** is also pseudoequatorial, no considerable γ effects are observed on C-16 and C-20 (δ = 61.49 and 37.93) in comparison with those of yohimbinone (δ = 61.8 and 37.9)⁹. The upfield shift of these carbon signals in compound **6** (δ = 58.17 and 33.54) is the consequence of the γ gauche effects of the pseudoaxial C-18 methoxy group. This effect on C-16 indicates, that the 16-H is pseudoaxial, consequently the methoxycarbonyl group is pseudoequatorial in compound **6**, too.

The chiral center C-18 of 6 can be epimerized under mild conditions in $2 \times \text{sodium}$ methoxide/methanol yielding the thermodynamically more stable compound 5.

Reduction of the keto ester 6 with $NaBH_4$ yielded two alcohols, 9a and 10a, in approximately equal amounts, whereas the reduction of 5 furnished two stereoisomers, 7a and 8a, in a ratio of about 10:1.



The ¹³C NMR spectra show that the configurations at C-16 and C-18 of the β -keto esters 5 and 6 remained intact during reduction. An alteration of configuration at either C-16 or C-18 should modify the C-20 chemical shifts of the alcohols relative to the respective β -keto esters (see Table 2).

The coupling constants of 17-H in the proton spectra testify that the alcohols 9a and 10a formed from 6 are C-17 epimers. In 9a – obtained in about 30% yield – the

hydroxy group is axial (α), whereas in its epimeric pair **10a** – formed in about 40% yield – it is in equatorial position. Comparison of the ¹³C NMR spectrum of **9a** with that of **10a** shows the axial hydroxy group of the former to exert γ gauche effects on C-19 and C-15.

In the ¹H NMR spectrum of the alcohol **7a** obtained from the keto ester **5** the coupling constants of 17-H indicate its equatorial (β) position, while those of the minor isomer **8a** are characteristic for an axial proton. The ¹H NMR data of **7f**, an acetylated derivative¹¹⁻¹⁴⁾ of **7a** is also in agreement with the stereochemical assignment of C-17 (17-H: δ = 5.80, both $J_{e,a}$ = 2.5 Hz). In the ¹³C NMR spectrum of **7a** the γ gauche effect of the axial 17-OH, similary to that in **9a**, causes upfield shifts of the C-19 and C-15 resonances.

Furthermore, a pure chemical proof could also be found in verifying the steric arrangement of **7a**. Weisenborn¹⁵ and later Patterson¹⁶ reported on the microbiological hydroxylation of natural yohimbine. The relative *cis* configuration of the two vicinal hydroxy functions of the isolated 18-hydroxyyohimbine was proved by easy acetonide formation. Similarly, upon acid treatment of the acetone solution of the diol **7b** obtained by ether splitting of **7a** (see Section B), the acetonide **7g** was obtained. The success of the acetonide formation and the coupling constants of 17-H and 18-H in the proton spectrum of **7g** (see Experimental) unambigously proves the *cis* (α , α) positions of both 17-OH and 18-OH in **7b**. Similarly, acetonide formation could be performed with compound **10b**, which also contains the substituents in *cis* (β , β) position. No reaction could be observed, however, with the diol **9b**, indicating the *trans* diaxial position of its C-17 and C-18 substituents.



It should be noted that E2 elimination of water from 7a, which appears to be very probable due to the favourable stereo-electronic position¹⁷⁾, cannot be accomplished. When 7a was heated in 2 \times sodium methoxide/methanol, instead of elimination, only fast ester hydrolysis could be observed.

This result is presumably due to the neighbouring group participation of 17-OH in accelerating the hydrolysis, which phenomenon has been already observed in the family of yohimbine alkaloids¹⁸⁾. A similar failure was observed in the case of water elimination attempted with thionyl chloride, which was successful for yohimbine¹⁹⁾ and its analogues^{14,21)}.

The obtained stereoselective formation of **7a** from **5** was rather surprising and in contradiction with our previous experience⁵: similar sodium borohydride reduction of yohimbinone yielded primarily β -yohimbine containing 17-OH in equatorial (β) position.

One of the probable explanations for the unexpected stereoselectivity is, that the attack of the BH_4^{\ominus} anion on C-17 from the β side is preferred because of the steric hindrance caused by the α position of both C-16 and C-18 neighbouring substituents.

B) Synthesis of 18-hydroxyyohimbine stereoisomers

In the next step of the synthesis the C-18 methoxy groups of 7a - 10a were demethylated to yield 18-hydroxyyohimbine stereoisomers. The reaction was first performed with 48-% hydrogen bromide. Under the reaction conditions applied the ester group also hydrolyzed, and thus to obtain the target diols 7b - 10b subsequent esterification with diazomethane was required. According to ¹H and ¹³C NMR spectra of the resulting compounds no epimerization took place at the chirality centers during hydrogen bromide treatment (see Tables 1 and 2). Characteristic changes in the C-17, C-18, and C-19 carbon resonances of 7b - 10b in comparison with those of the respective 18-methoxy compounds 7a - 10a are due to the difference in the α and β substituent effects between the hydroxy and methoxy groups.

A dealkylation method, which has always been found to lead to retention served as an independent chemical proof for the stereochemistry of the 18-hydroxy compounds. Using boron(III) bromide²⁰⁾ reagent at 0°C in dichloromethane, from 7a uniformly 7b was obtained. However, the boron(III) bromide demethylation of compounds 9a and 10a containing an axial (β) methoxy group at C-18 was slower by one order than in the case of 7a with an equatorial (α) methoxy substituent. The reaction required larger excess of the reagent, proceeded with lower yield, and derivatives containing bromine in the aromatic nucleus were also formed. The difference observed between the rates of dealkylation of the epimers 7a – 10a is in good correlation with the steric position assumed for the substituents of ring E.

As noted above at least one of the 18-hydroxy derivatives of yohimbine is a natural compound. The isolation of "18-hydroxy-yohimbine" was reported^{3,4}) without exact steric assignment of either the ring system or the substituents. Based upon the denomination between the quotation marks this alkaloid must have been assumed to be an 18-hydroxy derivative of yohimbine with known steric structure, and therefore it should be identical either with 7b or 9b. If the alkaloid in question were a derivative of β -yohimbine, its identification could be achieved by comparison with 8b or 10b. Thus our synthetic stereoisomers 7b - 10b will be likely to help in solving this structure elucidation.

C) Formation of raunescine analogues with normal skeleton

In order to prepare the raunescine stereoisomers with normal skeleton, the diols 7b, 9b, and 10b were acylated with trimethoxybenzoyl chloride in pyridine.

According to our experiments these diols show different behaviour under the conditions of acylation. Whereas **7b** and **9b**, containing an axial (α) 17-OH group, could be transformed regioselectively into 18-monoacylated products **7c**, **9c**, in the case of **10b** with an equatorial (β) 17-OH function three trimethoxybenzoylated compounds **10c**, **d**, **e** were formed depending on the reaction conditions. The differences observed in regioselectivity can be interpreted on the basis of the steric differences between the starting diols. In general, the acylation of an axial hydroxy group involves higher transition state energy than that of an equatorial one. This fact explains the selective trimethoxybenzoylation of 7b into 7c. The different reactivity of the two axial hydroxy groups in 9b is due to the sterically more hindered position of the C-17 hydroxy substituent. The equatorial acylation could not predominate in 10b because of the steric hindrance of the neighbouring ester group, and as a result the reactivity of the equatorial hydroxy function becomes comparable with that of the axial one. Accordingly, under mild conditions a mixture of monoacyl isomers 10c and 10d could be obtained, whereas under forced conditions (excess of reagent, longer reaction time) the diacylated compound 10e was the main product.

The position of the acyl group was determined on the basis of the characteristic ¹H NMR shifts and the coupling constants of 17-H and $18-H^{12,14,21}$ (see Table 1). The structure of **10d** is also supported by the possibility of transacylation into **10c**. Thus, when **10d** was heated in methanol in the presence of silica gel adsorbent (Kieselgel PF₂₅₄₊₃₆₆; Merck) as mild acidic catalyst, the C-17 equatorial acyl group migrated into the axial position C-18. The acyl migration was accompanied by a deacylation side reaction. If the reaction is performed in methanolic hydrochloric acid¹⁴, the major product is the diol **10b**. Moreover, by trimethoxybenzoylation of **10d** the diacyl derivative **10e** can be obtained (see Scheme 2).



TMB-Cl = 3,4,5-trimethoxybenzoyl chloride

As a result of the above trimethoxybenzoylation reactions three raunescine derivatives with normal yohimbane skeleton (7c, 9c, 10c) and one isoraunescine stereoisomer **10d** were obtained.

To the best of our knowledge, no other linear method is known for the synthesis of raunescine stereoisomers with normal skeleton up to now.

The authors wish to thank J. Tamás for the mass spectra, P. Sándor for the NMR measurements, and G. Dörnyei for helpful discussions, the Hungarian Academy of Sciences and the Chinoin Pharaceutical and Chemical Works (Budapest) for financial support.

Experimental

IR spectra were recorded in KBr with a Spectromom 2000 spectrophotometer. The ¹H and ¹³C NMR spectra were obtained using a Varian XL-100-15 and a Jeol FX-100 Fourier transform instrument operating at 100 MHz for ¹H and 25 MHz for ¹³C NMR. Chemical shifts are reported as δ values downfield from TMS as internal standard in both cases. Mass spectra (MS) were recorded with an AEI MS 902 double-focusing instrument (70 eV, ion source temp. 150 °C, direct insertion). Melting points are uncorrected.

General procedures: All reactions were conducted under oxygen-free dry nitrogen and were monitored by the using Merck silica gel 60 F_{254} sheets and the following solvent systems: A = benzene-ethyl acetate (5:5), B = dichloromethane-methanol (10:0.5), C = dichloromethanemethanol (10:1). Spots were visualized with an UV lamp or iodine vapor. For the quantitative separations (plc) Merck silica gel 60 $PF_{254+366}$, for column chromatography Merck silica gel 60 (70 – 230 mesh) adsorbent was used.

Usual workup of the reaction mixtures was carried out by extraction of an aqueos solution at pH 8.5-9 with ether or dichloromethane. The combined organic layer was washed with water, dried with anhydrous MgSO₄, and finally the solvent was evaporated *in vacuo*.

3-{1,2,3,4,6,7,12,12ba-octahydro-2-[(methoxycarbonyl)methylene]indolo[2,3-a]quino-Methyl lizin-3-yl -2-methoxypropionate (3): To the stirred solution of a mixture of diethyl [(methoxycarbonyl)methyl]phosphonate (10 ml, 47 mmol) and sublimed potassium tert-butoxide (4.0 g, 35 mmol) in absol. DMF (50 ml) the ketone 2 (8.5 g, 24 mmol) in absol. DMF (40 ml) was added at 0°C. The reaction mixture was allowed to stand for about 12 h in the refrigerator (at 10° C), diluted with cold water (100 ml) and extracted with ether (3 \times 200 ml) at pH 8.5-9. The combined organic layer was washed, dried, and evaporated in vacuo. The crude material (9.1 g, 92%) contained about 80% of 2 (based on tlc, system A, $R_F 2 > 3$), and it was used without further purification for the next reaction step. An analytical sample was crystallized from methanol; m. p. 83-85°C, hydrochloride m. p. 156-158°C (methanol-ether). - IR: 1650 $(C = C, \text{ conj.}), 1712 (CO_2CH_3, \text{ conj.}), 1750 (CO_2CH_3), 2750 - 2850 (Bohlmann bands), 3300 \text{ cm}^{-1}$ (NH). $-{}^{1}$ H NMR (CDCl₃): $\delta = 3.37, 3.39$ (2s, 3H, OCH₃), 3.5 - 4.1 (m, 2H, 1-H_{e0}, 12b-H), 3.73, 3.75, 3.77 (3s, 6H, CO₂CH₃), 5.67, 5.71 (2s, 1H, =CH-), 6.9-7.5 (m, 4H, aromatic protons), 8.76, 8.86 (2s, 1 H, NH). $-{}^{1}$ H NMR (C₆D₆): $\delta = 3.14, 3.15$ (2s, 3 H, OCH₃), 3.2-4.1(m, 3H, 1-H_{ea}, 12b-H + ?), 3.35, 3.36, 3.40 (3s, 6H, CO₂CH₃), 3.92 (dd, $J_{gem} = 13$, $J_{a,e} = 13$ 3 Hz, 1-H_{eq} of one diastereomer), 5.66, 5.75 (2s, 1H, =CH-), 7.2-7.5 (m, 4H, aromatic protons), 8.44, 8.63 (2s, 1 H, NH). $-{}^{13}$ C NMR: see Table 3. - MS: m/e (rel. int.) = 412 (M^{\oplus}, 97), 411 (70), 397 (47), 381 (9), 353 (40), 309 (100), 296 (36), 295 (30), 184 (29), 183 (23), 182 (42), 171 (13), 170 (76), 169 (50), 156 (11), 144 (11), 130 (9).

Note: The geometry of the C=C bond in 3 was verified by comparison of its ¹H and ¹³C NMR data with that of model compound 11⁵⁾. ¹H NMR of 11 in C₆D₆: $\delta = 3.36$, 3.41 (2s, 6H, CO₂CH₃), 3.54 (m, 1H, 12b-H), 3.81 (dd, 1H, $J_{gem} = 13$, $J_{e,a} = 4$ Hz, 1-H_{eq}), 5.61 (s, 1H, = CH –), 7.0 – 7.55 (m, 4H, aromatic protons), 8.27 (s, 1H, NH). – ¹³C NMR of 11 see Table 3.

 $C_{23}H_{28}N_2O_5$ (412.5) Calc. C 66.96 H 6.84 N 6.79 Found C 66.93 H 6.82 N 6.84

Methyl $3-\{1,2,3,4,6,7,12,12b\alpha$ -octahydro- 2β -[(methoxycarbonyl)methyl]indolo-[2,3-a]quinolizin- 3α -yl]-2-methoxypropionate (4): Crude 3 (9.1 g) was hydrogenated in methanol (250 ml) in the presence of 10-% palladium on carbon to yield 4 which was isolated as hydrochloride (6.1 g, 67%); tlc: system A, $R_F 3 > 4$. Hydrochloride: m. p. 186–187°C (methanol-ether). – IR 1740 (CO₂CH₃), 3200 cm⁻¹ (NH). – Base 4: ¹H NMR (CDCl₃): $\delta = 3.39$, 3.40 (2s, 3 H, OCH₃), 3.72, 3.78 (2s, 6H, 2 CO₂CH₃), 7.05–7.55 (m, 4H, aromatic protons), 8.02 (s, 1 H, NH). – MS: *m/e* (rel. int.) = 414 (M[®], 100), 413 (90), 339 (40), 355 (20), 341 (14), 340 (6), 313 (20), 298 (4), 297 (7), 269 (5), 211 (30), 198 (7), 184 (15), 170 (30), 169 (30), 156 (20), 144 (20), 143 (15).

C23H30N2O5 HCl (451.0) Calc. C 61.25 H 6.93 N 6.21 Found C 61.15 H 6.79 N 6.29

18α-Methoxyyohimbinone (5) and 18β-Methoxyyohimbinone (6): Compound 4 (7.4 g, 18 mmol) was dissolved in absol. benzene (200 ml) and dried by azeotropic distillation. Sublimed potassium tert-butoxide (3.0 g, 27 mmol) was added to the stirred reaction mixture and refluxed for 2 h, protected from air and light. Then the reaction mixture was neutralized with an equivalent amount of glacial acetic acid (1.6 ml, 27 mmol), evaporated to dryness, and the residue was extracted with dichloromethane (5 × 50 ml) at pH 8.5. The combined organic extract was washed, dried, and evaporated *in vacuo*. The amorphous residue (5.7 g, 84%) was treated with methanol (40 ml) and the crystalline 5 (1.6 g, 23%), separated. The evaporated mother liquor was purified by column chromatography on silica gel with ethyl acetate to supply 6 (1.6 g, 23%) and an additional amount of 1.1 g (total 2.7 g, 39%) 5; tlc: system A, $R_F 6 > 4 > 5$.

5: m. p. $163 - 165 \,^{\circ}$ C (methanol). – IR: 1730, 1750 (CO₂CH₃, CO), 2730 – 2870 (Bohlmann bands), 3150, 3450 cm⁻¹ (NH). – ¹H NMR (CDCl₃): $\delta = 3.48$ (s, 3H, OCH₃), 3.87 (dd, 1H, $J_{a,e} = 5.5$, $J_{a,a} = 12$ Hz, 18-H), 3.85 (s, 3H, CO₂CO₃), 7.05 – 7.55 (m, 4H, aromatic protons), 7.85 (s, 1H, NH). – MS: m/e (rel. int.) = 382 (M^{\oplus}, 100), 381 (7), 367 (5), 351 (7), 324 (2), 323 (3.3), 237 (4.3), 235 (4), 224 (5.7), 223 (4), 211 (12), 184 (20), 170 (33), 169 (27), 150 (14), 144 (10), 143 (9.3).

C22H26N2O4 (382.5) Calc. C 69.08 H 6.85 N 7.32 Found C 68.92 H 6.95 N 7.15

6: m. p. $125 - 127 \,^{\circ}$ C (methanol). – IR: 1700, 1740 (CO₂CH₃, CO), 2750 – 2870 (Bohlmann bands), 3150, 3500 cm⁻¹ (NH). – ¹H NMR (CDCl₃): $\delta = 3.32$ (s, 3H, OCH₃), 3.66 (t, 1H, $J_{e,a} = J_{e,e} = 3$ Hz, 18-H), 3.82 (s, 3H, CO₂CH₃), 7.05 – 7.55 (m, 4H, aromatic protons), 7.98 (s, 1H, NH).

C22H26N2O4 (382.5) Calc. C 69.08 H 6.85 N 7.32 Found C 68.85 H 7.01 N 7.34

Epimerization of 6 to 5: Compound 6 (40 mg, 0.1 mmol) was dissolved in 2 N methanolic sodium methoxide (5 ml) at -10° C, and the solution was kept at the same temperature for 72 h. Epimerization was monitored by tlc (system A, $R_F 6 > 5$). The resulting crystalline material was filtered off and recrystallized from methanol; m. p. 163–165 °C. According to spectral and chromatographic data the product was identical with compound 5 obtained from the ring closure of 4.

Note: Epimerization could not be accomplished at room temperature due to ring opening of 5 $(\rightarrow 4)$.

18α-Methoxyyohimbine (7a) and 18α-Methoxy-β-yohimbine (8a): To a stirred solution of 5 (1.2 g, 3.1 mmol) in methanol (80 ml) and dichloromethane (20 ml) sodium borohydride was added in small portions at 0°C. The reduction was followed by tlc (system B, $R_F 5 > 7a > 8a$). Subsequent to evaporation of the solvent *in vacuo* the residue was dissolved in a minimal amount of water and extracted with dichloromethane (4 × 20 ml) at pH 9.5. The combined organic layer was washed, dried, and evaporated *in vacuo*. The remaining material (1.15 g) was treated with methanol (50 ml) and the crystalline 7a (0.66 g, 55%) was separated. The evaporated mother liquor was purified by plc (system C, pretreated with ammonia, $R_F 7a > 8a$) to yield an additional amount of 0.18 g 7a (total 70%) and 8a (90 mg, 8%); tlc on alumina typ E: benzene-methanol (10:0.2), $R_F 7a > 8a$.

Note: The same ratio of 7a and 8a was obtained via catalytic (Pt/C) hydrogenation in glacial acetic acid (0.20 g of 5 and 0.20 g of Pt-C (10%) in 20 ml of solvent, 17 h).

7a: m. p. $210 - 212 \,^{\circ}$ C (methanol). – IR: 1730 (CO₂CH₃), 2750 – 2850 (Bohlmann bands), 3200, 3350 cm⁻¹ (OH, NH). – ¹H NMR (C₆D₆ + [D₆]DMSO): δ = 3.28 (s, 3 H, OCH₃), 3.45 (m, 1 H, $J_{a,e} = J_{a,e} = 2.5$ Hz, $J_{a,a} = 10$ Hz, 18-H_{ax}), 3.72 (s, 3 H, CO₂CH₃), 4.25 (m, 1 H, 17-OH), 4.43 (m, 1 H, $J_{e,a} = J_{e,a} = 2.5$ Hz, 17-H_{eq}), 7.05 – 7.65 (m, 4 H, aromatic protons), 10.70 (s, 1 H, NH). – MS: m/e (rel. int.) = 384 (M^{\oplus}, 100), 383 (100), 369 (2), 353 (4), 352 (2), 321 (2), 184 (25), 170 (25), 169 (30), 156 (20), 144 (15).

8a: m. p. $236 - 237 \,^{\circ}$ C (methanol). – IR: 1730 (CO₂CH₃), 2750 – 2850 (Bohlmann bands), 3350, 3400 cm⁻¹ (OH, NH). – ¹H NMR (CDCl₃): $\delta = 3.45$ (s, 3 H, OCH₃), 3.75 (m, 1 H, 17-H_{ax}), 3.84 (s, 3 H, CO₂CO₃), 7.05 – 7.45 (m, 4 H, aromatic protons), 7.76 (s, 1 H, NH). ¹H NMR (C₆D₆ + [D₆]DMSO): $\delta = 3.39$ (s, 3 H, OCH₃), 3.77 (s, 3 H, CO₂CH₃), 3.95 (m, 1 H, J_{a,a} = J_{a,a} = 10 Hz, 17-H_{ax}), 5.10 (m, 1 H, 17-OH), 7.12 – 7.57 (m, 4 H, aromatic protons), 10.92 (s, 1 H, NH). – MS: *m/e* (rel. int.) = 384 (M[®], 34), 383 (100), 369 (5.8), 367 (1.7), 353 (3.3), 211 (5), 184 (17), 169 (18.3).

O-Acetyl-18α-methoxyyohimbine (**7f**): To a solution of **7a** (126 mg, 0.33 mmol) in dry pyridine (3 ml) freshly distilled acetic anhydride (0.3 ml, 2.9 mmol) was added. After standing for about 12 h the crystalline product **7f** (90 mg, 64%) was filtered off; m. p. 266 – 268 °C (pyridine). – IR: 1230 (OCOCH₃), 1700 – 1735 (CO₂CH₃), 2780 – 2850 (Bohlmann bands), 3350 cm⁻¹ (NH). – ¹H NMR (C₆D₆ + [D₆]DMSO): δ = 1.97 (s, 3H, OCOCH₃), 3.28 (s, 3H, OCH₃), 3.72 (s, 3H, CO₂CH₃), 5.80 (t, 1 H, J_{c,a} + J_{c,a} = 6 Hz, 17-H_{cq}), 7.05 – 7.60 (m, 4H, aromatic protons), 10.75 (s, 1 H, NH). – MS: *m/e* (rel. int.) = 426 (M[⊕], 100), 425 (78), 411 (0.6), 397 (1.3), 395 (2.1), 384 (9), 383 (8.4), 367 (3.6), 307 (1.4), 184 (10), 170 (9.6), 169 (10), 156 (7.8), 144 (6).

C24H30N2O5 (426.5) Calc. C 67.58 H 7.09 N 6.57 Found C 67.46 H 6.95 N 6.27

18β-Methoxyyohimbine (9a) and 18β-Methoxy-β-yohimbine (10a): The keto ester 6 (2.3 g, 6 mmol) was dissolved in methanol (10 ml) and dichloromethane (40 ml). Sodium borohydride was added in small portions to the stirred reaction mixture at 0°C, and it was monitored by tlc (system B, R_F 10a > 9a). Workup including preparative tlc (cyclohexane-ether-methanol, 30: 50: 6, R_F 9a > 10a) supplied 9a (0.7 g, 30%) and 10a (1.1 g, 43%).

9a: m. p. $204 - 205 \,^{\circ}$ C (ether-petroleum ether) [**9a** · HCl: m. p. $253 - 255 \,^{\circ}$ C (methanol-ether)]. – IR: 1080 (C – OH), 1710 (CO₂CH₃), 2750 – 2850 (Bohlmann bands), 3200 – 3400 cm⁻¹ (OH, NH). – ¹H NMR (C₆D₆ + [D₆]DMSO): δ = 3.21 (s, 3H, OCH₃), 3.59 (q, 1H, J_{e,a} = J_{e,e} = J_{e,e} = 3 Hz, 18-H_{eq}), 3.70 (s, 3H, CO₂CH₃), 4.41 (t, 1H, J_{e,a} = J_{e,e} = 3 Hz, 17-H_{eq}), 7.13 – 7.65 (m, 4H, aromatic protons), 10.30 (s, 1H, NH). – MS: *m/e* (rel. int.) = 384 (M⁴), 100), 383 (100), 369 (1), 353 (2), 325 (4), 323 (1), 184 (10), 170 (10), 169 (15), 156 (10), 144 (8).

10a: m. p. 225 – 228 °C (dichloromethane-petroleum ether) [**10a** · HCl: 241 – 242 °C (methanolether)]. – IR: 1725 (CO₂CH₃), 2780 – 2850 (Bohlmann bands), 3200 – 3400 cm⁻¹ (OH, NH). – ¹H NMR (C₆D₆ + [D₆]DMSO): δ = 3.23 (s, 3 H, OCH₃), 3.46 (q, 1 H, $J_{e,e} = J_{e,a} = J_{e,a} = 3$ Hz, 18-H_{eq}), 3.77 (s, 3 H, CO₂CH₃), 3.97 (dd, 1 H, $J_{a,e} = 3$ Hz, $J_{a,a} = 11$ Hz, 17-H_{ax}), 7.12 – 7.65 (m, 4H, aromatic protons), 10.60 (s, 1 H, NH). – MS: *m/e* (rel. int.) = 384 (M^{\oplus}, 100), 383 (100), 369 (2), 353 (5), 325 (5), 323 (1), 184 (10), 170 (15), 169 (25), 156 (15), 144 (8).

18α-Hydroxyyohimbine (7b): a) Compound 7a (0.5 g, 13 mmol) was dissolved in 48-% aqueous hydrogen bromide solution (20 ml) and kept at 100 °C for 3 h. The solvent was removed under reduced pressure and the dried (over P_2O_5 in vacuo) residue refluxed in absol. methanol (50 ml). After evaporation the remaining material was treated with a minimal amount of water and extracted with dichloromethane (2 × 20 ml) at pH 9.5. The combined organic layer was

washed, dried, and evaporated *in vacuo*, and the crude material obtained was purified by plc (system C, pretreated with ammonia) to give 7b (0.3 g, 62%).

b) To a solution of **7a** (0.25 g, 0.65 mmol) in absol. dichloromethane (100 ml) boron tribromide (0.90 g, 0.34 ml, 3.6 mmol) was added at 0 °C. The reaction mixture was stirred for 4 h at 0 °C and kept in a refrigerator for about 12 h. Then the solution was treated with 10-% ammonium hydroxide solution, the organic phase was separated, washed, dried, and evaporated *in vacuo*. The crude material (0.17 g) was purified by plc (system C, R_F **7a** > **7b**) to supply **7b** (0.13 g, 54%).

7b: m. p. $221 - 223 \circ C$ (ethyl acetate). ~ IR: 1020 (C – OH), 1720 (CO₂CH₃), 3250 cm⁻¹ (OH, NH). – ¹H NMR (CDCl₃): $\delta = 3.65$ (br. m, 1 H, $J_{a,a} + J_{a,e} + J_{a,e} = 16$ Hz, 18-H_{ax}), 3.78 (s, 3 H, CO₂CH₃), 4.18 (t, 1 H, $J_{e,a} = J_{e,a} = 2$ Hz, 17-H_{eq}), 6.95 – 7.45 (m, 4H, aromatic protons), 9.05 (s, 1 H, NH). – MS: m/e (rel. int.) = 370 (M[®], 100), 369 (97), 353 (3), 352 (4), 351 (4), 339 (6), 338 (8), 337 (7), 321 (4), 311 (5), 211 (8), 184 (21), 170 (7), 169 (22), 156 (15).

18α-Hydroxy-β-yohimbine (**8b**), 18β-Hydroxyyohimbine (**9b**), and 18β-Hydroxy-β-yohimbine (**10b**): **8b**, **9b**, and **10b** were obtained according to method a); **7b**: 55 – 60% yield; tlc: system C, R_F **10b** > **9b**; **7b** > **8b**.

8b: m. p. $185 - 187 \,^{\circ}$ C (dec., amorphous). - IR: 1050 (C - OH), 1710 (CO₂CH₃), 3350 cm⁻¹ (br., OH, NH). - ¹H NMR (CDCl₃): $\delta = 3.80$ (s, 1 H, CO₂CH₃). - MS: *m/e* (rel. int.) = 370 (M[®], 96), 369 (100), 355 (4), 353 (4), 337 (5), 311 (7), 309 (6), 293 (4), 240 (8), 184 (20), 169 (31), 156 (23).

9b: m. p. $174 - 176 \,^{\circ}$ C (dec., ethyl acetate). - IR: 1030 (C - OH), 1715 (CO₂CH₃), 3350 cm⁻¹ (br., OH, NH). - ¹H NMR (CDCl₃ + [D₆]DMSO): $\delta = 3.78$ (s, 3 H, CO₂CH₃), 3.98 (q, 1 H, $J_{e,a} = J_{e,e} = 3$ Hz, 18-H_{eq}), 4.02 (t, 1 H, $J_{e,a} = J_{e,e} = 3$ Hz, 17-H_{eq}), 7.0 - 7.45 (m, 4 H, aromatic protons), 8.73 (s, 1 H, NH). - MS: m/e (rel. int.) = 370 (M[⊕], 99), 369 (100), 355 (2), 353 (2), 315 (2), 311 (8), 240 (4), 184 (20), 170 (22), 169 (26), 156 (16).

10b: m. p. 182 – 184 °C (dec., ethyl acetate). – IR: 1710 (CO₂CH₃), 3350 cm⁻¹ (br., OH, NH). – ¹H NMR (CDCl₃): $\delta = 3.76$ (dd, 1 H, $J_{a,e} = 2.5$ Hz, $J_{a,a} = 10.5$ Hz, 17-H_{ax}), 4.04 (q, 1 H, $J_{e,e} = J_{e,a} = J_{e,a} = 3$ Hz, 18-H_{eq}), 7.0 – 7.5 (m, 4 H, aromatic protons), 7.90 (s, 1 H, NH). – MS: *m/e* (rel. int.) = 370 (M[®], 99), 369 (100), 355 (2), 353 (3), 351 (4), 311 (6), 240 (7), 184 (11), 170 (14), 169 (15), 156 (10).

Note: If the ether cleavage starting from 9a or 10a was carried out by method b) 9b or 10b could be isolated in a yield of 15% besides other products brominated at the indole part.

Spectral data of brominated diol side-products

Product from 9a (24%): IR: 710, 770, 1715 (CO₂CH₃), 3400 cm⁻¹ (br., OH, NH). – MS: m/e (rel. int.) = 450 (91), 449 (92), 448 (M^{\oplus}, 100), 447 (77), 410 (16), 409 (18), 391 (4.6), 389 (6), 384 (22), 383 (20), 341 (5), 264 (8), 262 (8), 250 (8), 249 (11), 248 (9), 247 (8).

Product from 10a (10%): IR: 720, 760, 1710 (CO_2CH_3), 3350 cm⁻¹ (br., OH, NH). - MS: m/e (rel. int.) = 450 (91), 449 (97), 448 (M^{\oplus}, 100), 447 (82), 410 (13), 409 (16), 391 (5.5), 389 (7), 384 (6), 383 (5.5), 341 (5.5), 264 (12), 262 (12), 250 (12), 249 (15), 248 (13), 247 (12).

 18α -(3,4,5-Trimethoxybenzoyloxy)yohimbine (7c): To the solution of 7b (0.74 g, 1.9 mmol) in absol. pyridine (30 ml) 3,4,5-trimethoxybenzoyl chloride (1.33 g, 5.7 mmol) was added. The reaction mixture was kept at 100 °C for 1 h and the acylation monitored by tlc (system C, R_F 7c > 7b). After removing the solvent *in vacuo* the residue was dissolved in dichloromethane (70 ml) and stirred with saturated aqueous K₂CO₃ solution for 20 min. The organic layer was separated, washed, dried, and evaporated *in vacuo*. The crude material (0.74 g) was purified by column chromatography on silica gel with dichloromethane-methanol (100:2) to yield 7c

(350 mg, 31%); m. p. 168 – 169°C (ethyl acetate). – IR: 1710 – 1730 (ester CO), 2760, 2810, 2840 (Bohlmann bands), 3480 (NH), 3520 cm⁻¹ (OH). – ¹H NMR (CDCl₃): δ = 3.81, 3.90 (2s, 3 H, 9H, CO₂CH₃, OCH₃), 4.50 (t, 1 H, $J_{e,a} = J_{e,a} = 2.5$ Hz, 17-H_{eq}), 5.11 (m, 1 H, $J_{a,e} = J_{a,e} = 2.5$ Hz, $J_{a,a} = 12$ Hz, 18-H_{ax}), 7.05 – 7.55 (m, 6H, aromatic protons), 7.94 (s, 1 H, NH). – MS: m/e (rel. int.) = 564 (M[®], 33), 563 (18), 549 (1), 547 (1), 533 (1), 505 (0.5), 370 (2), 369 (2), 366 (10), 365 (9), 352 (40), 351 (40), 294 (1.6), 293 (4), 281.5 (3), 226 (70), 212 (90), 211 (40), 197 (80), 195 (55), 186 (8), 184 (45), 170 (16), 169 (42), 167 (7), 157 (9), 156 (30), 154 (100).

C31H36N2O8 (564.6) Calc. C 65.94 H 6.43 N 4.96 Found C 65.62 H 6.27 N 5.11

18β-(3,4,5-Trimethoxybenzoyloxy)yohimbine (9c): Starting with 9b (50 mg, 0.14 mmol) in absol. pyridine (5 ml) using trimethoxybenzoyl chloride (92 mg, 0.4 mmol) the above mentioned sequence afforded 90 mg of crude 9c which was purified by preparative tlc (system C, R_F 9c > 9b) to give 9c (25 mg, 34%); m. p. 165 – 166 °C (ethyl acetate-petroleum ether). – IR: 1715 (ester CO), 3200 – 3400 cm⁻¹ (OH, NH). – ¹H NMR (CDCl₃): δ = 3.80, 3.87, 3.89 (3s, 12 H, CO₂CH₃, OCH₃), 4.26 (dd, 1 H, $J_{e,a} = J_{e,e} = 3$ Hz, 17-H_{eq}), 5.29 (q, 1 H, $J_{e,a} = J_{e,e} = 3$ Hz, 18-H_{eq}), 7.0 – 7.50 (m, 6 H, aromatic protons), 7.88 (s, 1 H, NH). – MS: *m/e* (rel. int.) = 564 (M[⊕], 100), 563 (60), 549 (3), 533 (3), 434 (2), 352 (30), 351 (30), 293 (4), 281.5 (4), 226 (20), 212 (60), 195 (30), 184 (20), 170 (20), 169 (20), 156 (20), 144 (10).

C31H36N2O8 (564.6) Calc. C 65.94 H 6.43 N 4.96 Found C 65.48 H 6.17 N 5.12

 18β -(3,4,-5-Trimethoxybenzoyloxy)- β -yohimbine (10c) and Methyl 18β -hydroxy- 17β -(3,4,5trimethoxybenzoyloxy)yohimban- 16α -carboxylate (10d): a) The solution of 10b (0.30 g, 0.8 mmol) in absol. pyridine (20 ml) was treated with trimethoxybenzoyl chloride (0.54 g, 2.4 mmol) at 100 °C for 1 h. The reaction was followed by tlc (system C, R_F 10c > 10d > 10b). After evaporation of the solvent *in vacuo* the residue was dissolved in dichloromethane (30 ml) and stirred with saturated aqueous K_2CO_3 solution for 20 min. The organic phase was separated, washed, dried, and evaporated *in vacuo*. The residue was purified by plc (system C) to supply 10d (50 mg, 11%) and 10c (125 mg, 26%).

10c: m. p. $153 - 156 \,^{\circ}$ C (ethyl acetate-petroleum ether). – IR: 1710 - 1730 (ester CO), 2750 bis 2800 (Bohlmann bands), 3300 – 3400 (OH, NH). – ¹H NMR (CDCl₃): $\delta = 3.83$, 3.88 (2s, 3 H, 9H, CO₂CH₃, OCH₃), 4.08 (dd, 1 H, $J_{a,e} = 3$, $J_{a,a} = 11$ Hz, 17-H_{ax}), 5.52 q, 1 H, $J_{e,e} = J_{e,a} = J_{e,a} = 3$ Hz, 18-H_{eq}), 7.0 – 7.54 (m, 6H, aromatic protons), 7.85 (s, 1 H, NH). – MS: m/e (rel. int.) = 564 (M[®], 100), 563 (58), 549 (2), 547 (1), 533 (1.4), 505 (1.5), 370 (5), 369 (6), 366 (1.5), 365 (1), 352 (30), 351 (30), 294 (6), 293 (8), 281.5 (6), 226 (23), 212 (35), 211 (16), 197 (16), 195 (50), 186 (25), 184 (20), 170 (24), 169 (25), 167 (40), 157 (40), 156 (30), 154 (10).

C31H36N2O8 (564.6) Calc. C 65.94 H 6.43 N 4.96 Found C 65.54 H 6.21 N 5.18

10d: m. p. $217 - 218 \,^{\circ}$ C (ethyl acetate). - 1710 - 1730 (ester CO), 2750 - 2800 (Bohlmann bands), $3300 - 3450 \,^{-1}$ (OH, NH). - 1 H NMR (CDCl₃): $\delta = 3.67$, 3.88 (2s, 3H, 9H, CO₂CH₃, OCH₃), 4.38 (q, 1H, $J_{e,e} = J_{e,a} = J_{e,a} = 3$ Hz, $18 - H_{eq}$), 5.22 (dd, 1H, $J_{a,e} = 3$ Hz, $J_{a,a} = 10$ Hz, $17 - H_{ax}$), 7.0 - 7.5 (m, 6H, aromatic protons), 8.05 (s, 1H, NH). - MS: m/e (rel. int.) = 564 (M[⊕], 100), 563 (55), 549 (1.8), 547 (0.9), 533 (1.6), 505 (1.4), 370 (8), 369 (14), 366 (1.5), 365 (0.8), 352 (23), 351 (26), 294 (7), 293 (8), 281.5 (6), 226 (40), 212 (35), 211 (30), 197 (20), 195 (40), 186 (3), 184 (25), 170 (20), 169 (25), 167 (6), 157 (8), 156 (22), 154 (7).

C31H36N2O8 (564.6) Calc. C 65.94 H 6.43 N 4.96 Found C 65.72 H 6.34 N 5.21

b) Compound 10d (15 mg) in dry methanol (5 ml) was refluxed in the presence of silica gel $PF_{254+366}$ (30 mg) for 4 h. After filtration the solvent was evaporated *in vacuo* and the residue separated by tlc (system C, R_F 10c > 10d > 10b) to give 10c (3.5 mg), 10b (5 mg), and unreacted 10d (2 mg).

	17-H	18-H
5	_	$3.87 \text{ (ax)}, J_{a,c} = 5.5, J_{a,a} = 12$
6	-	3.66 (eq), $J_{e,e} = J_{e,a} = 3$
7a	4.43^{e} (eq), $J_{e,a} = J_{e,a} = 2.5$	3.45 (ax), $J_{a,a} = 10$, $J_{a,c} = J_{a,c} = 2.5$
8a	3.95^{e} (ax), $J_{a,a} = J_{a,a} = 10$	b)
9a	4.41^{e} (eq), $J_{ea} = J_{ee} = 3$	3.59 (eq), $J_{e,a} = J_{e,e} = J_{e,e} = 3$
10 a	3.97^{c} (ax), $J_{a,c} = 3$, $J_{a,a} = 11$	3.46 (eq), $J_{e,e} = J_{e,a} = J_{e,a} = 3$
7b	4.18 ^{c)} (eq), $J_{e_{a}} = J_{e_{a}} = 2$	3.65 (ax), $J_{aa} + J_{ae} + J_{ae} = 16$
9b	4.02^{d} (eq), $J_{ea} = J_{ea} = 3$	3.98 (eq), $J_{ea} = J_{ee} = J_{ee} = 3$
10b	$3.76^{\rm c}$ (ax), $J_{\rm a,c} = 2.5$, $J_{\rm a,a} = 10.5$	4.04 (eq), $J_{e,e} = J_{e,a} = J_{e,a} = 3$
7 c	$4.50^{\rm c}$ (eq), $J_{e,a} = J_{e,a} = 2.5$	5.11 (ax), $J_{a,a} = 12$, $J_{a,e} = J_{a,e} = 2.5$
9c	$4.26^{\rm c}$ (eq), $J_{e,a} = J_{e,e} = 3$	5.29 (eq), $J_{e,a} = J_{e,e} = J_{e,e} = 3$
10 c	4.08^{c} (ax), $J_{a,e} = 3$, $J_{a,a} = 11$	5.52 (eq), $J_{e,e} = J_{e,a} = J_{e,a} = 3$
10 d	5.22° (ax), $J_{a,e} = 3$, $J_{a,a} = 10$	4.38 (eq), $J_{e,e} = J_{e,a} = J_{e,a} = 3$
10e	5.47 ^d (ax), $J_{a,e} = 3$, $J_{a,a} = 10$	5.82 (eq), $J_{e,e} = J_{e,a} = J_{e,a} = 3$
	aje uju	*** ***

Table 1. Characteristic ¹H NMR data^{a)}

^{a)} Shifts are given in δ values relative to Me₄Si and J values in hertz. - ^{b)} Overlapped. - ^{c)} In CDCl₃ solution. - ^{d)} In CDCl₃ + [D₆]DMSO solution. - ^{e)} In C₆D₆ + [D₆]DMSO solution.

Carbon	6		10a	9b	10b	5	7a	7b
C-2	134.08	135.49	135.29	134.86	134.96	134.03	135.36	135.37
C-3	58,94	60.30	59.47	60.24	59.77	58.94	60.07	60.19
C-5	52.75	52.89	52.57	52.86	52.86	52.86	52.73	52.86
C-6	21.82	21.75	21.64	21.64	21.66	21.93	21.70	21.77
C-7	108.32	106.48	106.32	107.21	106.77	108.61	106.47	106.47
C-8	127.32	126.94	126.74	127.13	127.00	127.50	126.89	126.99
C-9	118.11	117.40	117.35	117.81	117.59	118.21	117.38	117.48
C-10	119.39	118.35	118.24	118.84	118.52	119.60	118.32	118.44
C-11	121.44	120.29	120.24	120.84	120.54	121.66	120.28	120.43
C-12	110.91	111.03	110.85	111.01	111.03	110.94	111.03	111.06
C-13	136,19	136.28	136.04	136.11	136.00	136.37	136.25	136.35
C-14	35.04	33.97	33.72	33.76	33.56	35.22	33.73	33.86
C-15	44.23	35.78	42.13	36.31	42.20	44.12	35.43	35.35
C-16	58.17	48.14	51.52	47.48	51.19	61.49	51.42	51.43
C-17	205.36	68.25	72.84	70.62	73.24	202.40	67.02	70.69
C-18	82.86	78.14	78.45	68.37	68.46	83.43	80.11	70.69
C-19	36.08	26.88	30.67	30.91	34.79	37.04	28.52	31.78
C-20	33.54	33.79	33.41	33.61	33.03	37.93	38.24	38.47
C-21	60.50	61.36	60.67	61.30	61.03	60.39	61.08	61.19
- CO -	170.00	173.69	174.34	175.34	175.02	169.02	172.41	172.71
- CO ₂ Me	52.18	51.22	51.22	51.72	51.61	52.21	51.32	51.64
$-OCH_3$	57.18	55.94	56.83	-	-	58.26	55.49	-

Table 2. ¹³C NMR chemical shifts^{a,b)}

^{a)} Chemical shifts (ppm) are relative to internal TMS in $CDCl_3$ solutions. - ^{b)} The small amounts of **8a** and **8b** were not sufficient to take their ¹³C NMR spectra.

	11	3
C-1	31.21	27.92 + 31.24
C-2	160.56	159.83 + 161.00
C-3	43.73	40.37 + 41.22
C-4	57.98	56.37 + 57.51
C-6	51.63	51.28 + 51.66
C-7	20.15	19.13 + 20.15
C-7a	107.56	107.26 + 107.44
C-7b	127.24	127.27
C-8	117.97	118.00
C-9	119.20	119.20
C-10	121.28	121.28
C-11	111.01	111.09
C-11 a	135.09	136.05
C-12a	133.44	133.09 + 133.44
С-12Ъ	59.18	58.39 + 59.18
- CO -	173.46	172.99
C(α)	31.61	78.13 + 78.54
C(B)	24.81	33.08 + 33.90
-CO-	167.61	167.76 + 167.84
= C - H	113.35	113.03 + 114.72
- OCH ₃		52.07 + 58.39
- CO ₂ CH ₃	51.22	51.28
$-CO_2CH_3$	51.63	51.66

Table 3. ¹³C NMR chemical shifts (δ values, CDCl₃, internal TMS)

Methyl 17β, 18β-bis(3, 4, 5-trimethoxybenzoyloxy)yohimban-16α-carboxylate (10e): a) The solution of 10b (0.38 g, 1 mmol) in absol. pyridine (10 ml) was treated with trimethoxybenzoyl chloride (1.3 g, 5.6 mmol) at 100 °C for 2 h. Similar workup including preparative tlc (system B, R_F 10e > 10b) gave 10e (0.15 g, 33%); m. p. 223 – 224 °C (ethyl acetate). – IR: 1710–1730 (CO₂CH₃), 2750–2850 (Bohlmann bands), 3200–3400 cm⁻¹ (NH). – ¹H NMR (CDCl₃ + [D₆]DMSO): δ = 3.66, 3.74, 3.84, 3.86, 388, (5s, 6H, 3H, 12H, CO₂CH₃, OCH₃), 5.47 (dd, 1H, $J_{a,e}$ = 3 Hz, $J_{a,a}$ = 10 Hz, 17-H_{ax}), 5.82 (q, 1H, $J_{e,e}$ = $J_{e,a}$ = 3 Hz, 18-H_{eq}), 6.95–7.45 (m, 8H, aromatic H), 9.42 (s, 1H, NH). – MS: *m/e* (rel. int.) = 758 (M[⊕], 25), 757 (8), 658 (1), 656 (1.3), 578 (11), 563 (1), 560 (1.4), 547 (15), 546 (40), 545 (19), 531 (1.5), 515 (1.1), 487 (0.7), 378.5 (2), 366 (2.2), 352 (8), 351 (18), 336 (14), 335 (22), 334 (8), 333 (8), 273 (4), 226 (38), 212 (100), 197 (47), 195 (42), 185 (10), 184 (25), 170 (13), 169 (19), 156 (25).

b) Compound **10d** (10 mg, 0.02 mmol) in absol. pyridine (3 ml) was heated with trimethoxybenzoyl chloride (17 mg, 0.07 mmol) at 100 °C for 4 h. Workup including plc yielded **10e** (6 mg).

Acetonide of 18α -Hydroxyyohimbine (7g): To the solution of 7b (80 mg, 0.2 mmol) in dry acetone (8 ml) a few drops of perchloric acid and 1 g of molecular sieves (3Å) were added. The reaction mixture was stirred at room temperature and monitored by tlc (system C, R_F 7g > 7b). At about 80-% conversion (≈ 4 h) solid potassium carbonate was added to neutralize, the salt filtered off, and the solvent evaporated *in vacuo*. The residue, after separation by plc (system C), gave 7g (40 mg, 35%); m. p. 236-237 °C (ethyl acetate). - IR: 1040, 1140, 1210 (characteristic bands), 1725 (CO₂CH₃), 2750-2850 (Bohlmann bands), 3350 cm⁻¹ (NH). - ¹H NMR (CDCl₃): $\delta = 1.34, 1.51$ (2s, 3H, 3H, CH₃), 3.35 (d, 1H, 12b-H), 3.79 (s, 3H, CO₂CH₃), 4.16 [m, 1H,

 $J_{18,19} = 12$ Hz (a,a), $J_{18,19} = 7$ Hz (a,e), $J_{18,17} = 4$ Hz (a,e), 18-H_{ax}], 4.42 (t, 1 H, $J_{e,a} = J_{e,a} = 4$ Hz, 17-H_{eq}), 7.0 - 7.5 (m, 4H, aromatic protons), 8.03 (s, 1 H, NH). – MS: m/e (rel. int.) = 410 (M^{\oplus}, 100), 409 (91.2), 395 (10.6), 351 (5.8), 349 (4.6), 197 (5.9), 194 (10.6), 170 (8.7), 169 (9.5), 156 (8.2), 144 (5.9), 70 (5.9), 61 (8.7), 45 (5.9), 43 (50.7).

Note: The acetonide of 10b could be obtained by a similar process, however, no acetonide formation could be detected in the case of 9b.

- ¹⁾ Part I: C. Szántay, G. Blaskó, K. Honty, E. Baitz-Gács, and L. Tőke, Liebigs Ann. Chem. 1983, 1269; Part III: C. Szántay, G. Blaskó, K. Honty, E. Baitz-Gács, J. Tamás, and L. Tőke, ibid. 1983, 1292.
- ²⁾ ^{2a)} C. F. Huebner and E. Schlittler, J. Am. Chem. Soc. 79, 250 (1959). ^{2b)} N. Hosansky and E. Schmith, J. Am. Pharm. Assoc. 44, 639 (1955).
- ³⁾ S. P. Majumdar, J. Poisson, and P. Potier, Phytochemistry 12, 1167 (1973).
- ⁴⁾ ^{4a)} M. M. Iwu and W. E. Court, Planta Med. 34, 390 (1978). ^{4b)} M. M. Iwu and W. E. Court, Phytochemistry 17, 1651 (1978). ^{4c)} N. N. Sabri and W. E. Court, Phytochemistry 17, 2023 (1978). ^{4d)} M. M. Iwu and W. E. Court, Planta Med. 38, 260 (1980). Unfortunately the authors could not provide us with a sample for comparison.
- 5) L. Toke, K. Honty, and C. Szántay, Chem. Ber. 102, 3248 (1969).
- ⁶⁾ ^{6a)} C. Szántay, L. Tőke, and P. Kolonits, J. Org. Chem. 31, 1447 (1966). ^{6b)} L. Szabó, K. Honty, L. Tőke, and C. Szántay, Chem. Ber. 105, 3231 (1972).
- ⁷⁾ E. Wenkert and B. G. Jackson, J. Am. Chem. Soc. 81, 5601 (1959).
- ⁸⁾ See Part III of this series: Liebigs Ann. Chem. 1983,1292.
- ⁹⁾ E. Wenkert, C.-J. Chang, H. P. S. Chawla, D. W. Cochran, E. W. Hagaman, J. C. King, and K. Orito, J. Am. Chem. Soc. **98**, 3645 (1976).
- ¹⁰⁾ K. Honty, E. Baitz-Gács, G. Blaskó, and C. Szántay, J. Org. Chem. 47, 5111 (1982).
- ¹¹⁾ J. D. Albright, L. A. Mitscher, and L. Goldman, J. Org. Chem. 28, 38 (1963).
- 12) W. E. Rosen and J. N. Shoolery, J. Am. Chem. Soc. 83, 4816 (1961).
- 13) L. Tőke, K. Honty, L. Szabó, G. Blaskó, and C. Szántay, J. Org. Chem. 38, 2496 (1973).
- 14) L. Szabó, I. Tóth, L. Tőke, and C. Szántay, Liebigs Ann. Chem. 1977, 642.
- 15) S. C. Pan and F. L. Weisenborn, J. Am. Chem. Soc. 80, 4749 (1958).
- ¹⁶ E. L. Patterson, W. W. Andres, E. F. Krause, R. E. Hartman, and L. A. Mitscher, Arch. Biochem. Biophys. 103, 117 (1963).
- ¹⁷⁾ M. M. Janot, R. Goutarel, E. W. Warnhoff, and A. LeHir, Bull. Soc. Chim. Fr. 1961, 637.
- ¹⁸⁾ M. J. Allen, J. Chem. Soc. 1960, 4904.
- ¹⁹⁾ J. D. Albright, L. A. Mitscher, and L. Goldman, J. Heterocycl. Chem. 7, 623 (1970).
- ²⁰⁾ J. F. W. McOmie, M. L. Watts, and D. E. West, Tetrahedron 24, 2289 (1968).
- ²¹⁾ I. Tóth, L. Szabó, M. Kajtár-Peredy, E. Baitz-Gács, L. Radics, and C. Szántay, Tetrahedron 34, 2113 (1978).

[2/83]