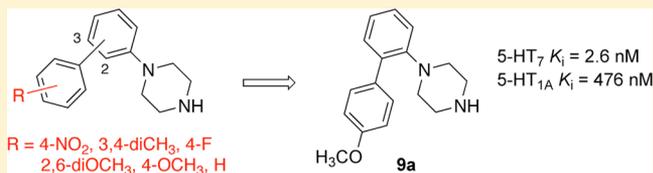


Investigations on the 1-(2-Biphenyl)piperazine Motif: Identification of New Potent and Selective Ligands for the Serotonin₇ (5-HT₇) Receptor with Agonist or Antagonist Action in Vitro or ex VivoEnza Lacivita,[†] Daniela Patarnello,[†] Nikolas Stroth,^{||} Antonia Caroli,[‡] Mauro Niso,[†] Marialessandra Contino,[†] Paola De Giorgio,[†] Pantaleo Di Pilato,[†] Nicola A. Colabufo,[†] Francesco Berardi,[†] Roberto Perrone,[†] Per Svenningsson,^{||} Peter B. Hedlund,[§] and Marcello Leopoldo^{*†}[†]Dipartimento Farmaco-Chimico, Università degli Studi di Bari "A. Moro", via Orabona, 4, 70125, Bari, Italy[‡]Department of Physics, Sapienza University, piazzale A. Moro, 5, 00185, Rome, Italy[§]Department of Molecular Biology, The Scripps Research Institute, La Jolla, California 92037, United States^{||}Center for Molecular Medicine, Department of Neurology and Clinical Neuroscience, Karolinska Institute and Karolinska University Hospital, 17176 Stockholm, Sweden

Supporting Information

ABSTRACT: Here we report the design, synthesis, and 5-HT₇ receptor affinity of a set of 1-(3-biphenyl)- and 1-(2-biphenyl)piperazines. The effect on 5-HT₇ affinity of various substituents on the second (distal) phenyl ring was analyzed. Several compounds showed 5-HT₇ affinities in the nanomolar range and >100-fold selectivity over 5-HT_{1A} and adrenergic α_1 receptors. 1-[2-(4-Methoxyphenyl)phenyl]piperazine (**9a**) showed 5-HT₇ agonist properties in a guinea pig ileum assay but blocked 5-HT-mediated cAMP accumulation in 5-HT₇-expressing HeLa cells.



INTRODUCTION

The serotonin 5-HT₇ receptor was identified starting in 1993 by the application of targeted molecular biology techniques. It has been described in various species and remains the last 5-HT receptor to be discovered. The 5-HT₇ receptor is localized in discrete areas of the brain and in the periphery. Within the central nervous system (CNS), high levels of this receptor have been detected in the hippocampus, thalamus, and hypothalamus (especially within the suprachiasmatic nucleus).¹ Nearly two decades after its discovery, much information is available on the pathophysiological role of 5-HT₇ in the CNS.²

Pharmacological blockade of the 5-HT₇ receptor by **1** (SB-269970) (Figure 1) or inactivation of the 5-HT₇ receptor gene leads to an antidepressant-like behavioral profile in rodent forced swim test and tail suspension test. It has also been suggested that the atypical antipsychotic drug amisulpride exerts its antidepressant action through blockade of 5-HT₇ receptors.³ Recently, a study showed that the pharmacological blockade of 5-HT₇ receptors produced a faster antidepressant-like response than the commonly prescribed antidepressant fluoxetine.⁴ This is particularly interesting, since antidepressants with a faster onset of action are an unmet need in depression therapy.

Several studies have suggested that 5-HT₇ receptors are involved in nociceptive processing. Activation of 5-HT₇ receptors exerts antinociceptive effects at the level of the spinal cord and pronociceptive effects in the periphery.⁵ Moreover,

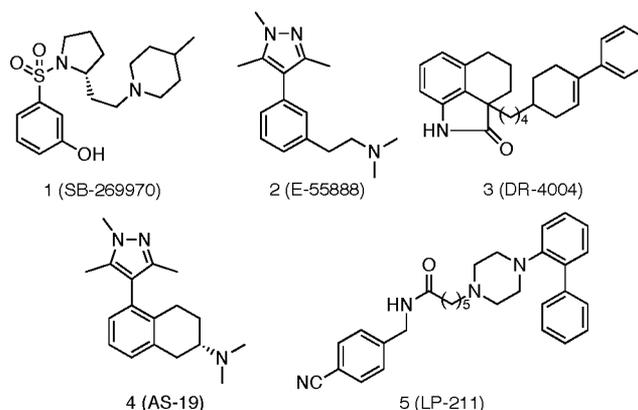


Figure 1. Structures of selective 5-HT₇ receptor agents.

since subcutaneous administration of the selective 5-HT₇ receptor agonist **2** (E-55888) increased the analgesic potency of oral morphine, it has been proposed that 5-HT₇ receptor agonists could be used as adjuvants of opioid analgesia.⁶

Animal studies of learning and memory have suggested a potentially important role for the 5-HT₇ receptor in cognitive processes.⁷ 5-HT₇ drugs such as **1**, **3** (DR-4004), and **4** (AS-19) (Figure 1) are able to reverse amnesia induced by post-

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training administration of scopolamine (a cholinergic antagonist). This supports the hypothesis that cholinergic, glutamatergic, and serotonergic systems interact in cognitively impaired animals. Moreover, it shows that the 5-HT₇ receptor can significantly influence cognitive dysfunction and therefore represents a potential therapeutic target for the treatment of memory dysfunction in cognitive disorders (schizophrenia, Alzheimer's disease, age-related decline).⁸ Collectively, the aforementioned findings underscore the importance of developing new 5-HT₇ receptor drugs.

Our research group has been involved for several years in the study of structure–activity relationships (SARs) of 4-substituted 1-arylpiperazine derivatives, the so-called “long-chain” arylpiperazines.^{9–13} Our studies have led to the identification of **5** (LP-211) (Figure 1) which showed high 5-HT₇ receptor affinity ($K_i = 0.58$ nM, rat cloned receptors, high selectivity over 5-HT_{1A} and D₂ receptors (324- and 245-fold, respectively), and agonist properties in an ex vivo assay of 5-HT₇ receptor activation. Disposition studies in mice evidenced that **5** undergoes N-dealkylation to **6a** (Table 1). This might be of relevance because N-unsubstituted 1-arylpiperazines can

reach the brain and may display pharmacological effects opposite to those of their parent drugs. Therefore, the final pharmacological effect of such drugs might result from the interplay between the neurochemical actions of the parent drug and of its active metabolite.¹⁴ For this reason, the affinities for a range of 5-HT receptors of **6a** and **5** were evaluated under the same experimental conditions. Compound **6a** displayed higher 5-HT₇ receptor affinity ($K_i = 1.4$ nM, human cloned receptor) and overall better selectivity profile than **5** ($K_i = 15$ nM, human cloned receptor).¹⁵ This was unexpected because the majority of the long-chain arylpiperazine derivatives described in the literature display higher 5-HT₇ receptor affinity than the N-(4)-unsubstituted counterparts.^{11,16,17} Thus, compound **6a** attracted our attention as a low molecular weight lead compound that could deliver a completely new set of selective 5-HT₇ receptor ligands.

CHEMISTRY

The synthesis of the target 1-arylpiperazine derivatives **6a,b**–**12a,b** (Table 1) required the key aniline intermediates **13a,b**–**17a,b**, **20a,b**, and **21a,b** (Scheme 1). Among these, **13a,b** were

Table 1. Binding Affinities of Target Compounds

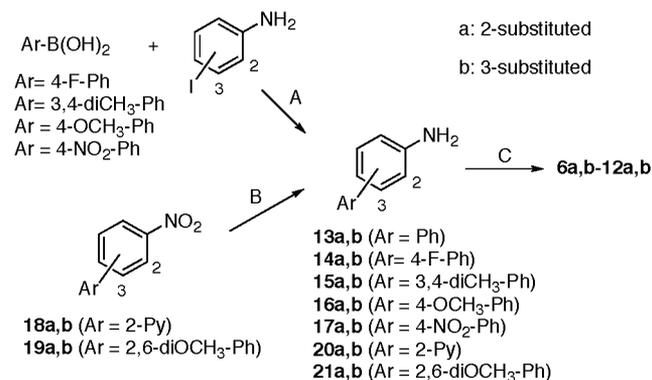
		K_i , nM \pm S.E.M. ^a			
compd	pos	Ar ²	5-HT ₇	5-HT _{1A}	α_1
6a	2		1.4 ^b	99 ^b	59.1 \pm 2.5
6b	3		25 \pm 1.7	72.1 \pm 3.3	(21%) ^c
7a	2		2.7 \pm 1.5	(15%) ^d	(14%) ^c
7b	3		109 \pm 64	63.2 \pm 3.1	(3%) ^c
8a	2		1.9 \pm 0.6	(22%) ^d	165 \pm 2
8b	3		93 \pm 2	(34%) ^d	(7%) ^c
9a	2		2.6 \pm 0.1	476 \pm 3	156 \pm 2
9b	3		> 5000	(42%) ^d	(29%) ^c
10a	2		7.0 \pm 1.2	(16%) ^d	105 \pm 2
10b	3		13 \pm 4	31.4 \pm 2.3	(8%) ^c
11a	2		23 \pm 6	(23%) ^d	(11%) ^c
11b	3		80 \pm 4	14.9 \pm 2.3	(27%) ^c
12a	2		96 \pm 11	(21%) ^d	(10%) ^c
12b	3		7.1 \pm 1.7	149 \pm 2	(3%) ^c
22a	2		18.4 \pm 6.5	--	--
22b	3		8.2 \pm 1.9	--	--
23a	2		6.9 \pm 0.3	--	--
23b	3		1.4 \pm 0.1	17.4 \pm 1.5	--
5-CT			0.3 \pm 0.1	--	--
5-HT			--	10 \pm 1.5	--
Phentolamine			--	--	16 \pm 2

^aValues are the mean \pm SEM from three independent experiments.

^bData taken from ref 15. ^cDisplacement of [³H]prazosin from rat cerebral cortex membranes by a single concentration of test compound (100 nM). Data are the mean of two independent experiments.

^dDisplacement of [³H]-8-OH-DPAT from human cloned 5-HT_{1A} receptors by a single concentration of test compound (100 nM). Data are the mean of two independent experiments.

Scheme 1. Synthesis of Target Compounds **6a,b**–**12a,b**^a



^aReagents: (A) Pd(dppf)Cl₂ and 5 N NaOH or tetrakis(triphenylphosphino)palladium(0) and 2 M Na₂CO₃; (B) ammonium formate, 10% Pd/C; (C) bis(2-chloroethyl)amine hydrochloride, K₂CO₃, KI.

commercially available, whereas the remaining anilines were prepared according to literature methods by cross-coupling reaction of 2- or 3-iodoaniline and the appropriate benzeneboronic acid under Suzuki conditions (**14a,b**–**17a,b**) or through catalytic reduction of nitroderivatives **18a,b** and **19a,b** (anilines **20a,b** and **21a,b**). The target compounds **6a,b**–**12a,b** were obtained by condensing the intermediate anilines **13a,b**–**17a,b**, **20a,b**, and **21a,b** with bis(2-chloroethyl)amine hydrochloride.

RESULTS AND DISCUSSION

Structure–Activity Relationships for 5-HT₇ Receptor.

The notion that N-(4)-unsubstituted 1-arylpiperazines can bind to the 5-HT₇ receptor was reported by Shen et al. in 1993.¹⁸ In particular, it was shown that 1-(1-naphthalenyl)piperazine, 1-(2-methoxyphenyl)piperazine, and 1-(3-chlorophenyl)piperazine can bind at 5-HT₇ receptors with moderate to good affinities ($K_i = 83$ nM, 243 nM, and 352 nM, respectively). Over the years, the above-mentioned 1-arylpiperazines were used as starting points for the development of several classes of long-chain arylpiperazine derivatives.

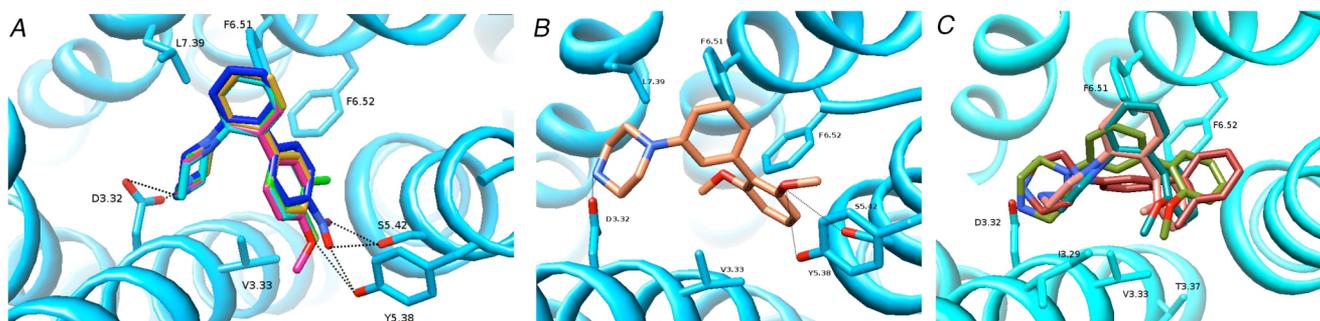


Figure 2. (A) Main interactions of **6a** (yellow), **7a** (cyan), **8a** (green), **9a** (magenta), and **10a** (blue) within the 5-HT₇ receptor binding site. The protonated piperazine N(4) nitrogen forms an ionic interaction with D3.32 at the distance of 2.57 Å. The oxygen of methoxy group of **9a** forms hydrogen-bond contacts with S5.42 and Y5.38 at distances of 3.4 and 3.1 Å, respectively. The same pattern of interactions is shown by **7a** and **10a**. (B) Main interactions of **12b** within the 5-HT₇ receptor binding site. (C) Main interactions of **22a,b** and **23a,b** within the 5-HT₇ receptor binding site.

In many cases the introduction of a large substituent on the basic nitrogen of the piperazine ring had a beneficial effect on the affinity and specificity for the 5-HT₇ receptor.^{11,16,17} Therefore, it was unexpected that removal of the pendant substituent from the basic nitrogen of **5** afforded the potent and selective 5-HT₇ ligand **6a**. Since no studies dealing with N-(4)-unsubstituted 1-aryl piperazines as 5-HT₇ receptor ligands can be found in the literature, we decided to explore the SARs of a set of compounds structurally related to **6a**. For our study, we have evaluated the 1-(2-biphenyl)piperazine derivatives **6a–12a**, focusing on the aromatic ring denoted as Ar² in the general structure reported in Table 1.

In particular, we wanted to explore if introduction of substituent(s) that could modify the electronic properties or the steric hindrance of the Ar² group had an effect on 5-HT₇ receptor affinity. We also evaluated the affinity for the 5-HT₇ receptor of the corresponding 1-(3-biphenyl)piperazine counterparts **6b–12b**. The 1-(4-biphenyl)piperazine framework was not taken into consideration at this stage because a previous study showed that 1-(4-substituted-phenyl)piperazines were nearly devoid of 5-HT₇ receptor affinity.¹¹ The binding affinity values of the target compounds at the 5-HT₇ receptor are shown in Table 1. In comparison of the 5-HT₇ affinity values of **6a** (Ar² = phenyl), **7a** (Ar² = 4-fluorophenyl), **8a** (Ar² = 3,4-dimethylphenyl), **9a** (Ar² = 4-methoxyphenyl), and **10a** (Ar² = 4-nitrophenyl), no great differences can be noted. This indicates that there is no preference between electron-rich and electron-poor aromatic rings in that position. Introduction of an aza group in **6a** afforded compound **11a** (Ar² = 2-pyridyl), causing a >10-fold loss in affinity. The presence of substituents in different positions of the Ar² ring had different effects on 5-HT₇ receptor affinity. In particular, **8a** (Ar² = 3,4-dimethylphenyl) and **9a** (Ar² = 4-methoxyphenyl) displayed affinities in the same range as **6a**, suggesting that the binding pocket of the receptor is large enough to accommodate such substituents. By contrast, a marked decrease in affinity was observed in the case of **12a** (Ar² = 2,6-dimethoxyphenyl). Next, we evaluated the effect on 5-HT₇ receptor affinity of shifting the Ar² ring from 2- to 3-position (derivatives **6b–12b**). This modification reduced the 5-HT₇ receptor affinity (except for **12b**) to various extents: compound **10b** showed only half the affinity of its isomer **10a**, whereas **9b** was devoid of 5-HT₇ affinity ($K_i > 5000$ nM), contrary to the isomer **9a** ($K_i = 2.6$ nM). A notable exception to this trend was derivative **12b** ($K_i =$

7.1 nM), which has higher affinity than its isomer **12a** ($K_i = 96$ nM).

To rationalize the SARs, we developed a pharmacophore model (see Supporting Information). Matrix distances between the pharmacophoric features (i.e., a positively charged nitrogen atom and two aromatic rings that corresponded to Ar¹ and Ar²) were in general agreement with those reported in the pharmacophore models by Kolaczowski et al.¹⁹ (the general “affinity” hypothesis) and by Badarau et al.²⁰ (the “second hypothesis”). The analysis of aligned target compounds clearly indicated that the pharmacophoric groups of compounds **6a–12a** matched the features of the pharmacophore model more closely than those of compounds **6b–12b** (Figure SI3, Supporting Information). Next, docking simulations were conducted by use of the Autodock4 program²¹ to analyze the binding mode of **6a,b–12a,b**. Kolaczowski et al.¹⁹ defined the 5-HT₇ receptor binding site as constituted by the interaction point of the positively charged nitrogen atom and two pockets: one localized between transmembrane helices (TMHs) 4–6 in a deep cavity and the other one between TMHs 7–3 in the extracellular exposed area. According to our docking studies, the compounds reported in Table 1 preserve the crucial ionic interaction between protonated piperazine N(4) nitrogen with D3.32 (according to Ballesteros and Weinstein)²² and the CH– π interaction between the phenyl ring attached to the piperazine ring (Ar¹) with F6.51 (Figure 2A). Moreover, the compounds with the highest affinities also showed CH– π interaction between Ar² and F6.52. The binding mode analysis suggested that the TMH 4–6 pocket is not large enough to accommodate the Ar² group when it is in the 3-position of Ar¹, thus explaining the lower affinity of **6b–11b** as compared to **6a–11a**. In particular, the conformations proposed for **9b** do not allow the crucial ionic interaction with D3.32. On the other hand, a different binding scenario could be hypothesized for compound **12b**: the methoxy group formed H-bonding interactions with Y5.38 and S5.42 that drove Ar² into a hydrophobic pocket formed by I3.29, V3.33, and F6.52 (Figure 2B). Consequently, the structure-based conformation of **12b** showed that the orientation and distances of the essential triplet features were comparable with those observed for the compounds with highest affinity. To check this further, we have studied an additional small set of compounds having a 2-methoxy- or a 2-methyl group on Ar², with Ar² in either 2- or 3-position on Ar¹ (compounds **22a,b** and **23a,b**, Table 1). These compounds were prepared following the synthetic routes

depicted in Scheme 1 (see Supporting Information). As in the case of compounds **12a,b**, 1-(3-biphenyl)piperazine derivatives **22b** and **23b** showed higher 5-HT₇ receptor affinity than the 1-(2-biphenyl) counterparts **22a** and **23a**, but in this case, a small difference was shown. Docking analysis suggested that the methoxy or methyl substituent in 2-position of compounds **22a,b** and **23a,b** interacts with the pocket created by I3.29, V3.33, M3.34, T3.37, and I4.37 residues and allows optimal interaction of Ar¹ and Ar² with F6.51 and F6.52, respectively (Figure 2C). Therefore, affinity data and docking analysis of compounds **12a,b**, **22a,b**, and **23a,b** indicate that the introduction of small substituents in one or both ortho-positions of Ar² enables the molecules to adopt the optimal conformation for interaction through hydrophobic or hydrogen-bonding interactions. All in all, the binding mode analysis evidenced that the interaction of biphenyl piperazines with the 5-HT₇ receptor is driven essentially by steric and hydrophobic requirements. In the case of compound **12b**, the hydrogen-bond formation with Y5.38 and S5.42 is fundamental for the optimal orientation of Ar².

Selectivity over 5-HT_{1A} and Adrenergic α_1 Receptors.

Target compounds were counterscreened for their affinity against the serotonin 5-HT_{1A} receptor because it is colocalized with the 5-HT₇ receptor. Also, target compounds were evaluated for their adrenergic α_1 receptor affinity because it has been reported that the 1-(2-biphenyl)piperazine scaffold might have affinity for this receptor.²³ As far as the affinity for 5-HT_{1A} receptors is concerned, all the 2-biphenyl derivatives displayed poor affinities except **9a** (Ar = 4-methoxyphenyl, K_i = 476 nM). On the other hand, the 3-biphenyl derivatives showed a different trend. Compounds **6b**, **7b**, **10b**, and **11b** displayed good 5-HT_{1A} receptor affinity (72.1 nM < K_i < 14.9 nM), differently from **8b** and **9b**. With regard to the adrenergic α_1 receptor, all compounds showed weak affinities, except **6a** and **10a** (K_i = 59.1 nM and 105 nM, respectively). Taken together, these data indicated that the 1-(2-biphenyl)piperazine motif can deliver high-affinity 5-HT₇ ligands and that an appropriate substitution pattern around the Ar² ring can lead to selective 5-HT₇ receptor ligands.

Activation of 5-HT₇ Receptors in Guinea Pig Ileum. 5-HT₇ receptor activation is known to mediate relaxation of gastrointestinal smooth muscle. Previous studies have demonstrated that activation of 5-HT₇ receptors inhibits contractions induced by substance P in the guinea pig ileum.²⁴ We investigated the effect of compounds **6a–9a**, which showed the highest 5-HT₇ receptor affinity and selectivity values among the newly reported compounds, on substance P-induced contractions in the guinea pig ileum. The assays were conducted in the presence of a cocktail of non-5-HT₇ receptor antagonists (see Supporting Information) in order to elucidate whether or not the relaxation response in this particular tissue is due to direct activation of the 5-HT₇ receptor. For comparative purposes, we also tested **4**, a selective 5-HT₇ ligand that behaves as a potent (EC_{50} = 9 nM) partial 5-HT₇ receptor agonist (77% maximal effect compared to 5-HT) toward cAMP accumulation in HEK-293F/h5-HT₇ cells.²⁵ As already reported,¹¹ the 5-HT₇ agonist 5-CT was able to reduce 40% of the contraction induced by substance P, an effect that was competitively reverted by the antagonist **1**. Compounds **6a–9a** were able to induce relaxation of substance P-mediated contraction (Figure 3). However, the effects elicited by **6a** (0.5 μ M and 1 μ M) and **8a** (2 μ M) were not 5-HT₇ receptor-mediated because they were not reverted by **1** (3 μ M). On the

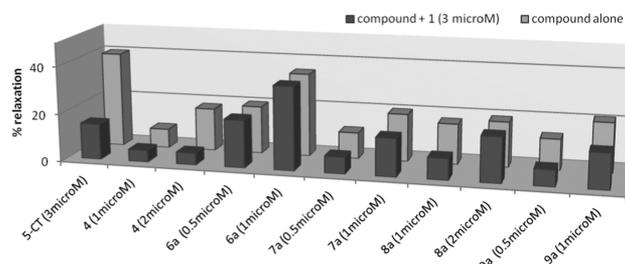


Figure 3. Percent relaxation of substance P-mediated contraction induced by 5-HT₇ agonists in isolated guinea pig ileum.

other hand, **7a** and **9a** were able to induce 5-HT₇ receptor-mediated relaxation at concentrations of 0.5 μ M and 1 μ M, because the effects were abolished by **1** (3 μ M). Finally, **4** induced relaxation at concentrations of 1 μ M and 2 μ M but not at 0.5 μ M (data not shown), the effects being reverted by **1** (3 μ M). All in all, these data indicate that compounds **7a** and **9a** behave as 5-HT₇ competitive agonists in this assay with specificity as good as **4**, contrary to **6a** and **8b**.

Activity at 5-HT₇ Receptors Stably Expressed in HeLa Cells. Compounds **6a** and **9a** were active in the guinea pig ileum assay, but the effect of **6a** appeared not to be specific to 5-HT₇ (no blockade with **1**). To further characterize their function, we therefore tested both compounds against 5-HT_{7(a)} receptors stably expressed in HeLa cells. Neither **6a** nor **9a** stimulated cAMP accumulation (data not shown), and both compounds behaved as antagonists instead. Thus, cAMP accumulation induced by 1 μ M 5-HT was inhibited by **6a** and **9a** with IC_{50} values of 531 and 438 nM, respectively (Figure 4A). To confirm their inhibitory action at recombinant

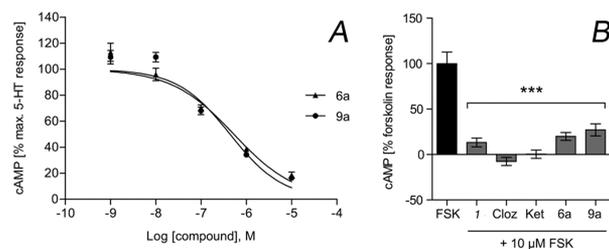


Figure 4. Effects of compounds **6a** and **9a** on (A) 5-HT- and (B) forskolin-induced cAMP accumulation in vitro.

5-HT_{7(a)}, the compounds were compared with established receptor antagonists regarding their capacity to block forskolin-induced cAMP. Similar to **1**, clozapine, and ketanserin, **6a** and **9a** significantly inhibited cAMP accumulation, thus providing further evidence for antagonist effects, at least in the context of a heterologous system (Figure 4B).

CONCLUSIONS

The main aim of the present study was exploration of the SARs of a set of 1-(2-biphenyl)- and 1-(3-biphenyl)piperazines, toward the goal of identifying new high-affinity 5-HT₇ ligands. This goal was achieved, as several compounds with K_i s in the low nanomolar range were identified. The SARs showed that various substituents can be introduced in the distal phenyl ring of the 1-(2-biphenyl)piperazine scaffold without affecting 5-HT₇ receptor affinity. On the other hand, the 1-(3-biphenyl)piperazine scaffold appears to be more sensitive to the presence and position of substituents. Considering the selectivity over 5-

HT_{1A} and α_1 receptors, it is noteworthy that, despite their low molecular weight, several ligands herein reported showed affinities 2 orders of magnitude lower than those at 5-HT₇ receptors. All in all, affinity and computational data indicate that both 1-(2-biphenyl)- and 1-(3-biphenyl)piperazine scaffolds can deliver high-affinity 5-HT₇ ligands or can be starting points for development of newer long-chain arylpiperazines capable of binding at 5-HT₇ receptors.

Investigation of signaling through the 5-HT₇ receptor yielded interesting results. While compounds **9a** and **10a** induced relaxation of guinea pig ileum in the same fashion as 5-CT and the selective 5-HT₇ receptor agonist **4**, compounds **6a** and **9a** failed to stimulate cAMP accumulation in 5-HT_{7a}-expressing HeLa cells. Instead, inhibition was observed when cells were cotreated with 5-HT and **6a** or **9a**. Moreover, similar to established antagonists of 5-HT₇,²⁶ both compounds inhibited forskolin-stimulated cAMP. Thus, at least in the context of our heterologous system, **6a** and **9a** displayed antagonist properties.

It has previously been shown that a given 5-HT receptor ligand can act as agonist in one and antagonist in another receptor-expressing system. For example, the 5-HT₄ ligand 4-amino-5-chloro-2-methoxybenzoic acid 2-(1-piperidinyl)ethyl ester (ML 10302) is a partial agonist in guinea pig ileum but an antagonist of cAMP accumulation in guinea pig hippocampus.²⁷ Similarly, partial agonist as well as antagonist properties at central 5-HT_{1A} receptors have been described for 8-{2-[4-(methoxyphenyl)-1-piperazinyl]ethyl}-8-azaspiro[4.5]decane-7,9-dione (BMY 7378).²⁸ Our present results demonstrate that dual agonist/antagonist ligands also exist for the 5-HT₇ receptor, exemplified by compound **9a**. In light of this finding, we believe that the reinvestigation of agonists and antagonists previously published by our laboratory and others will tell us if the dual agonist/antagonist feature of **9a** is shared by other 5-HT₇ ligands. This might ultimately help to explain the observed inconsistencies in the role of 5-HT₇ receptors in the CNS.⁸

EXPERIMENTAL SECTION

General Procedure for the Preparation of Piperazines 6a,b–12a,b, 22a,b, and 23a,b. A mixture of the appropriate aniline (4.0 mmol), bis(2-chloroethyl)amine hydrochloride (0.71 g, 4.0 mmol), K₂CO₃ (0.55 g, 4.0 mmol), and KI (0.66 g, 4.0 mmol) in xylene (30 mL) was refluxed for 48 h. After cooling, the solvent was distilled off in vacuo and the residue was taken up with AcOEt (30 mL) and 5% aqueous NaOH solution (30 mL). The organic phase was separated and then washed with brine. The organic phase was dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. The crude product was chromatographed with CHCl₃/MeOH, 9:1, to afford the desired compounds as pale yellow oil. The purity of the tested compounds **6a,b–12a,b, 22a,b, and 23a,b** was assessed by RP-HPLC and combustion analysis. All compounds showed $\geq 95\%$ purity.

1-[2-(4-Methoxyphenyl)phenyl]piperazine (9a). Yield 30%. ¹H NMR (CDCl₃) δ 1.71 (br s, 1H, D₂O exchanged), 3.12 (br s, 8H), 3.84 (s, 3H), 6.94 (d, 2H, *J* = 8.8 Hz), 7.00 (d, 1H, *J* = 8.0 Hz), 7.10–7.15 (m, 1H), 7.22–7.30 (m, 2H), 7.46 (d, 2H, *J* = 8.8 Hz). GC-MS *m/z* 269 (M⁺ + 1, 11), 268 (M⁺, 56), 226 (100), 210 (43), 167 (24). The hydrochloride salt melted at 198–200 °C (from MeOH/Et₂O). Anal. (C₁₇H₂₀N₂O·HCl·0.5 H₂O) C, H, N.

Radioligand Binding Assays. Binding of [³H]-5-CT at human cloned 5-HT₇ receptor was performed according to Jasper et al.²⁹ Binding of [³H]-8-OH-DPAT at human cloned 5-HT_{1A} receptors was performed as previously described.³⁰ Binding of [³H]prazosin at α_1 -adrenoceptors was performed according to Glossmann and Hornung.³¹

Isolated Guinea Pig Ileum Assay. Inhibition of substance P-induced contraction in guinea pig ileum by 5-HT₇ receptor agonists was performed as previously described.¹¹

Measurement of Intracellular cAMP Accumulation. HeLa cells stably expressing human 5-HT_{7(a)} (kindly provided by Dr. Mark Hamblin) were cultured as described³² and seeded into 96-well plates before treatment. Intracellular cAMP was measured by a commercial enzyme-linked immunosorbent assay (ELISA; Abnova, Taipei City, Taiwan).

ASSOCIATED CONTENT

Supporting Information

Additional text, three figures, and two tables with synthetic details and spectral data for **6a,b–12a,b, 15a,b, 17a,b, and 20a,b–23a,b**; elemental analysis of **6a,b–12a,b, 22a,b, and 23a,b**; matrix distances; molecular modeling and molecular docking methods; PHASE alignment analysis; training set for pharmacophore model identification; list of inactive compounds; and biological methods and statistical analysis. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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Notes

The authors declare no competing financial interest.

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