Accepted Manuscript

Investigation of biaryl heterocycles as inhibitors of Wee1 kinase

Anthony Mastracchio, Chunqiu Lai, Maricel Torrent, Kenneth Bromberg, Fritz G. Buchanan, Debra Ferguson, Velitchka Bontcheva, Eric F. Johnson, Loren Lasko, David Maag, Nirupama Soni, Alexander R. Shoemaker, Thomas D. Penning

PII:	S0960-894X(19)30226-4
DOI:	https://doi.org/10.1016/j.bmc1.2019.04.017
Reference:	BMCL 26384
To appear in:	Bioorganic & Medicinal Chemistry Letters
Received Date:	25 February 2019
Revised Date:	5 April 2019
Accepted Date:	8 April 2019



Please cite this article as: Mastracchio, A., Lai, C., Torrent, M., Bromberg, K., Buchanan, F.G., Ferguson, D., Bontcheva, V., Johnson, E.F., Lasko, L., Maag, D., Soni, N., Shoemaker, A.R., Penning, T.D., Investigation of biaryl heterocycles as inhibitors of Wee1 kinase, *Bioorganic & Medicinal Chemistry Letters* (2019), doi: https://doi.org/10.1016/j.bmcl.2019.04.017

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

-
-

Investigation of biaryl heterocycles as inhibitors of Wee1 kinase

- 2
- 3 Anthony Mastracchio^{a*}, Chunqiu Lai^a, Maricel Torrent^b, Kenneth Bromberg^a, Fritz G. Buchanan^a,
- 4 Debra Ferguson^a, Velitchka Bontcheva^a, Eric F. Johnson^a, Loren Lasko^a, David Maag^a, Nirupama
- 5 Soni^a, Alexander R. Shoemaker^a and Thomas D. Penning^a
- ⁶ ^aCancer Research, Discovery, Global Pharmaceutical Research and Development, AbbVie, 1
- 7 North Waukegan Road, North Chicago, Illinois 60064
- ⁸ ^aMolecular Modeling, Structural Biology, Discovery, Global Pharmaceutical Research and
- 9 Development, AbbVie, 1 North Waukegan Road, North Chicago, Illinois 60064
- ¹⁰ *Corresponding Author: Anthony Mastracchio (anthony.mastracchio@abbvie.com);
- 11 Number of figures: 3
- 12 Number of schemes: 3

CC

- 13 Number of tables: 1
- 14 Key words: Wee1, Cdk1, Cdc2, G1 checkpoint, G2/M checkpoint.
- 15 Running title: Investigation of biaryl heterocycles as inhibitors of Wee1 kinase

Wee1 is a key tyrosine kinase that helps control the G2/M checkpoint by preventing entry into 1 2 mitosis.^{1,2} This is of particular interest for cancer therapy since many cancer cells depend more heavily than do normal cells on a functional G2/M checkpoint for DNA repair.³⁻⁵ This suggests 3 that inhibition of Wee1 could induce premature entry into mitosis of DNA damaged cancer cells 4 leading to mitotic catastrophe and cell death.^{6,7} Following on this insight many cancer programs 5 have focused on the development of selective drug-like Wee1 inhibitors.^{8–10} AZD1775 (aka MK-6 7 1775, adavosertib), the first selective Wee1 inhibitor reported, is currently in clinical trials for the treatment of solid tumors. Clinical results for AZD1775 showed on-target efficacy as a single 8 agent and in combination thus providing additional support for targeting Wee1 in cancer.^{11–13} 9 10 In a previous account,¹⁴ we used a class of Wee1 inhibitors featuring an imino-11 dihydropyrimidone (1) that had emerged in the patent literature as a starting point. Then, aided 12 by molecular modeling, a series of compounds bearing a pyrimidine-based tricyclic scaffold 13 were design as exemplified by 2 and proved to be potent inhibitors of Wee1 (Figure 1). Our 14 studies also showed that the fused imidazole ring of 2 occupies a key region within the ATP 15 binding pocket in Wee1 adjacent to the bicyclic core that was not utilized by **1**. Encouraged by 16 these results we sought to explore a variety of biaryl cores that would 1) possess the key 17

elements for binding in the active site (i.e. proper hinge binder and pi interactions with phe433)

and 2) accommodate a substituent next to the bicyclic bridge (position *C*-8 or *N*-8). This

20 substituent could in turn be used as a vector for modulation of compound properties and to

21 further explore this region of the binding pocket. Modeling studies suggested that biaryl system

such as 3 or 4 could fulfill these requirements. Consequently, we sought to explore structure-

activity relationships for these biaryl cores bearing substituents at *C*-8 or *N*-8.¹⁵ Building on the insights we gained during our exploration of tricyclic compounds **2**, we selected a bis-Cl aryl moiety at the *C*-6 or *N*-6 position and an arylpiperazine or cyclopropyl tetrahydroisoquinoline in the amino aryl region as preferred scaffolds to evaluate the various bicyclic cores. Our findings indicate that both 4-pyridones (**3**) and 2-pyridones (**4**) may serve as interesting lead series for the development of potent and selective Wee1 inhibitors with desirable pharmacokinetic properties as well as antitumor efficacy in a murine xenograft model.



Previously disclosed work:

8

9 **Figure 1.** Modification strategies of novel and selective Wee1 inhibitors.

10 Schemes 1 & 2 outline the general synthesis of analogs related to biaryls **3** and **4**. As seen in

11 Scheme 1, the known pyrimidine ester **5** was hydrolyzed under standard conditions to give the

1 corresponding acid which in turn was treated with thionyl chloride to afford acyl chloride **6**.

2 Next, an *in situ* generated organocuprate of compound **7** was added to yield ketone **8**. At this

- 3 point, the desired substituent at R¹can be introduced by treatment of **8** with an appropriately
- 4 substituted primary amine or aniline followed by condensation/cyclization with DMF-DMA at
- 5 elevated temperature. The methyl thioether group of **10** was oxidized by mCPBA, setting the

6 stage for the final S_NAr reaction with the desired aminoaryl substrate to provide analogs of 4-





Scheme 1. Synthesis of 4-pyridone series. Reagents and conditions: (a) 5 molar aqueous NaOH,
THF, 60°C, 16 hrs, 89% yield; (b) SOCl₂, DMF (cat), 90°C, 3 hrs, 99% yield; (c) CuCN, LiCl, THF, –
25°C to rt, 79% yield; (d) amine, 60°C, IPA, 16 hrs, (for R = Me, 91% yield): (e) DMF-DMA, 150°C,

1	90 minutes, (for R = Me, 76% yield); (f) <i>m</i> -CPBA, CH ₂ Cl ₂ , 30 minutes, rt; (g) Ar-NH ₂ , TFA, MeCN,
2	100°C, 16 hrs (for R = Me and Ar = tetrahydroisoquinoline analog, two steps 65% yield).
3	
4	2-Pyridone analogs of 4 were prepared as outlined in Scheme 2. The synthesis commences with
5	either commercially available pyrimidine ester 15 or with pyrimidine ester 14 , the latter being
6	obtained through the condensation of ethyl 3-oxopentanoate 12 with methyl
7	carbamimidothioate 13 and DMF–DMA at elevated temperature. The esters were then
8	converted to arylamides 16 via a three-step sequence involving saponification, acyl chloride
9	formation then treatment with 2,6-dichloroaniline. The 2-pyridone ring can then be forged
10	after treatment of compound 16 with Vilsmeier-Haack reagent 18 in DMF at 60°C. For 17b a
11	methyl group at C-8 was obtained but for 17a an additional molecule of Vilsmeier-Haack
12	reagent was added at C-8 to yield aldehyde 19. This aldehyde can then act as an ideal functional
13	handle to introduce various substituents for structure-activity studies near the bridged
14	pyrimidine region as shown in Scheme 2. Finally the aminoaryl moiety was installed using the
15	same synthetic sequence we described for the 4-pyridone series.



Scheme 2. Synthesis of 2-pyridone series. Reagents and conditions: (a) 13, DMF-DMA, 80°C, 10
minutes then methyl carbamimidothioate, reflux, 16 hrs, 52% yield; (b) 1 molar aqueous LiOH,
MeOH, THF, rt, 2 hrs, (for 16, 86% yield; for 15, 74% yield); (c) SOCl₂, DMF (cat), dioxane, rt then
2,6-dichloroaniline, 100°C, 16 hrs, (for 17a, 40% yield; for 17b, 29% yield); (d) DMF, 60°C, 3 hrs,
(for 19, 29% yield, for 20, 43%): (e) Ph₃PCH₃Br, KOt-Bu, THF, rt, 48 hrs, 62% yield; (f) NaBH₄,
CeCl₃*7H₂O, CH₂Cl₂/MeOH (3:1), 0°C, 30 minutes, 70 % yield; (g) DAST, CH₂Cl₂, 3 hrs, rt, 61%

yield; (h) *m*-CPBA, CH_2Cl_2 , 30 minutes, rt; (i) Ar-NH₂, TFA, MeCN, 100°C, 16 hrs. (for R = Me, two steps 81% yield)

3

4 Table 1 shows the structures of the compounds evaluated and their respective affinities

5 presented as K_i values which was assessed in a routine 6-point TR-FRET binding assay. Activities

6 in functional antiproliferative cell viability assays (H1299 cell line) and in a pCDK1 ELISA assay

7 measuring the ability of Wee1 to phosphorylate CDK1 at Tyr 15, both utilizing the NCI-H1299

8 cell line (human non-small cell lung carcinoma), are also shown. In addition, mouse microsomal

9 stability data (Cl(mic), L/hr/Kg) are also included in Table 1 for initial assessment of metabolic

MA

10 clearance.¹⁶

11

Compound	Structure	Ki (nM)	H1299 EC ₅₀ (μM)	pCDK1 EC ₅₀ (μM)	CL(mic) (L/hr/Kg) Mouse
2	N N N N K F H N	1.1	0.31	0.060	38.1
26	N N N N Cl Cl Cl Cl H N H N H N H	0.3	0.52	0.079	16.2
27	$\begin{array}{c} N \\ N \\ N \\ N \\ N \\ N \\ H \\ M \\ H \\ M \\ M \\ M \\ M \\ M \\ M \\ M$	1.0	0.46	0.022	34.9

12 **Table 1**. Biaryl analogs.¹⁷





Most analogs yielded equal or better binding affinity than the lead molecule 2 suggesting that
1) the proposed cores can retain binding requirements to the protein similar to compound 1
and 2 and that 2) these biaryl systems are likely to participate in productive pi-pi interaction
with Phe 433 as suggested by our docking studies.

6

7 Compounds **26-35** explored the effect of substituent variation at *N*-8 on binding affinity and cell

8 antiproliferative activity for the 4-pyridone series. Computational modeling of compound **26**

predicted that a range of substitutions could be accommodated at N-8 since this vector is 1 2 oriented towards an open area between Phe433 located at the base of the ATP binding site, and Val313 located in the Gly-rich loop (Figure 2).¹⁸ The experimental results validated our 3 computational model and showed that binding affinity remained largely unaffected when 4 neutral, polar or hydrogen bond donor groups are present (compounds 26-28 & 30-35). 5 However, when a basic nitrogen was introduced (compound **29**) a significant reduction in 6 7 binding affinity was observed. One hypothesis for the drop in affinity with amine **29** is that the 8 amine substituent sits on a large hydrophobic patch with no polar residues to stabilize the net charge. Exploration of the N-8 substituent did confirm that this vector could be utilized for 9 modulation of physicochemical properties as a range of cell permeability and microsomal 10 clearance was obtained with variations of the stereoelectronic nature of the substituent. In 11 general it was observed that small non-polar alkyl substituents offer the best balance of cell 12 13 activity and compound stability. This can be seen with methyl analog 33 and ethyl analog 34 where both compounds compare well with first generation compound 2 in terms of binding 14 affinity ($K_i = 1.1$ nM vs 0.3 nM for 33), cell viability (EC₅₀ = 0.31 μ M vs 0.10 μ M for 33) and in the 15 PD assay pCDK1 ELISA (EC₅₀ = 0.060 μ M vs 0.011 μ M for **33**). This establishes the 4-pyridone 16 core as a suitable ring system for productive binding interaction in the active site of Wee1. 17



Figure 2. Structure and ligand interaction plot of compound 26 docked into the Wee1 kinase binding site. The model predicts two key hydrogen bond interactions with the backbone polar atoms of the hinge (Cys379) and an additional hydrogen bond interaction with the side chain of gate keeper residue Asn376. The biaryl core is also predicted to engage in pi-pi interactions with Phe433.

7

Given the similar shape and electronic properties between 4-pyridones and 2-pyridones we decided to evaluate the latter to see if this new ring system could also be used as template for generation of potent Wee1 inhibitors. We determined that indeed the 2-pyridone analogs retain good enzymatic potency and cell potency. Both series offered a similar activity profile and the different chemical structures of the two could be utilized to introduce different yet complimentary substituents at position 8 of the bicyclic ring system.

14

15 Following these results, compound **33** was selected for advanced profiling. The compound

- showed desirable PK properties in mouse (iv $t_{1/2}$ = 2.7 h; iv CL = 1.5 L/h/kg; po AUC = 6.2 μ M*h;
- 17 po F = 44%) and was thus selected to study its ability to potentiate the efficacy of the DNA-

damaging agent irinotecan in BRCA1 deficient, p53 deficient MX-1 TNBC xenograft model. As
shown in Figure 3, the combination of compound **33** with irinotecan achieved 88% tumor
growth inhibition (TGI) when compared to vehicle and 57% TGI when compared to irinotecan
alone on day 43. Moreover, the durability of the combination response was highlighted by 48%
tumor growth delay when compared to irinotecan alone (using 1000 mm³ as an endpoint) and
therefore supports the enhanced efficacy of irinotecan by compound **33** *in vivo*. ¹⁹



Figure 3. Tumor growth inhibition with compound 33 in combination with irinotecan.²⁰ MX-1
brei (homogenized tumor cell suspension) was sub-cutaneously injected into the flank of SCID
mice. After the tumor volume reached ~350 mm³, mice were treated with vehicle (black line),
10 mkd (mg/kg/day) irinotecan Q4Dx4 (closed circles), or 10 mkd irinotecan Q4Dx4 + 25 mkd
compound 33 QD 2 days on 2 day off for 4 cycles (open squares). Dosing began on day 24.
Tumor volumes were measured twice a week.

1	In summary, we designed and synthesized 3 new series of pyrimidine-based biaryl derivatives
2	and evaluated their binding affinity and activity in cells. The data provided shows that both 4-
3	pyridone and 2-pyridone cores are suitable replacements for the tricyclic core and allow for a
4	new substitution pattern that can be used for evaluating substituents at position C-8 or N-8 of
5	the bicyclic ring system. Our studies have also identified 4-pyridone 33 as a viable Wee1
6	inhibitor with improved <i>in vitro</i> activity in cell over benchmark compound 2 . This compound
7	also exhibits good drug-like properties and potentiated the anti-proliferative activity of
8	irinotecan in vivo when dosed orally in an MX-1 xenograft model. These findings suggest that
9	pyridinone-based biaryl core such as 3 and 4 may serve as a suitable template for the
10	development of potent and efficacious Wee1 inhibitors.
11	Acknowledgments
11 12	Acknowledgments We would like to acknowledge the work Nirupama Soni for the generation of the Wee1 TR-FRET
11 12 13	Acknowledgments We would like to acknowledge the work Nirupama Soni for the generation of the Wee1 TR-FRET binding assay data.
11 12 13 14	Acknowledgments We would like to acknowledge the work Nirupama Soni for the generation of the Wee1 TR-FRET binding assay data. Disclosures:
11 12 13 14 15	AcknowledgmentsWe would like to acknowledge the work Nirupama Soni for the generation of the Wee1 TR-FRETbinding assay data.Disclosures:All authors are employees of AbbVie. The design, study conduct, and financial support for this
 11 12 13 14 15 16 	AcknowledgmentsWe would like to acknowledge the work Nirupama Soni for the generation of the Wee1 TR-FRETbinding assay data.Disclosures:All authors are employees of AbbVie. The design, study conduct, and financial support for thisresearch were provided by AbbVie. AbbVie participated in the interpretation of data, review,
 11 12 13 14 15 16 17 	AcknowledgmentsWe would like to acknowledge the work Nirupama Soni for the generation of the Wee1 TR-FRETbinding assay data.Disclosures:All authors are employees of AbbVie. The design, study conduct, and financial support for thisresearch were provided by AbbVie. AbbVie participated in the interpretation of data, review,and approval of the publication.
 11 12 13 14 15 16 17 18 	Acknowledgments We would like to acknowledge the work Nirupama Soni for the generation of the Wee1 TR-FRET binding assay data. Disclosures: All authors are employees of AbbVie. The design, study conduct, and financial support for this research were provided by AbbVie. AbbVie participated in the interpretation of data, review, and approval of the publication.
 11 12 13 14 15 16 17 18 19 	Acknowledgments We would like to acknowledge the work Nirupama Soni for the generation of the Wee1 TR-FRET binding assay data. Disclosures: All authors are employees of AbbVie. The design, study conduct, and financial support for this research were provided by AbbVie. AbbVie participated in the interpretation of data, review, and approval of the publication.
 11 12 13 14 15 16 17 18 19 20 	Acknowledgments We would like to acknowledge the work Nirupama Soni for the generation of the Wee1 TR-FRET binding assay data. Disclosures: All authors are employees of AbbVie. The design, study conduct, and financial support for this research were provided by AbbVie. AbbVie participated in the interpretation of data, review, and approval of the publication. References

Lundgren, K.; Walworth, N.; Booher, R.; Dembski, M.; Kirschner, M.; Beach, D. mik1 and
 wee1 cooperate in the inhibitory tyrosime phosphorylation of cdc2. *Cell* 1991, *64*, 1111 1122.

1	2.	Parker, L. L.; Piwnica-Worms, H. Inactivation of the p34cdc2-cyclin B complex by the
2		human WEE1 tyrosine kinase. <i>Science</i> 1992 , <i>257</i> , 1955-1957.
3	3.	Kastan, M. B.; Bartek, J. Cell-cycle checkpoints and cancer. Nature 2004, 432 316-323.
4	4.	Bucher, N.; Britten, C. D. G2 checkpoint abrogation and checkpoint kinase-1 targeting in
5		the treatment of cancer. Br. J. Cancer 2008, 98, 523–528.
6	5.	Chen, T.; Stephens, P. A.; Middleton, F. K.; Curtin, N. J. Targeting the S and G2
7		checkpoint to treat cancer. Drug Discov. Today 2012 , 17, 194–202.
8	6.	Do, K.; Doroshow, J. H.; Kummar, S. Wee1 kinase as a target for cancer therapy. <i>Cell</i>
9		<i>Cycle</i> 2013 , <i>12</i> , 3159–3164.
10	7.	Matheson, C. J.; Backos, D. S.; Reigan, P. Targeting WEE1 kinase in cancer. Trends
11		Pharmacol. Sci. 2016 , 37, 872–881.
12	8.	Mizuarai, S.; Yamanaka, K.; Itadani, H.; Arai, T.; Nishibata, T.; Hirai, H.; Kotani, H.
13		Discovery of gene expression-based pharmacodynamic biomarker for a p53 context-
14		specific anti-tumor drug Wee1 inhibitor. <i>Mol Cancer</i> 2009, 8, 34.
15	9.	Palmer, B. D.; Thompson, A. M.; Booth, R. J.; Dobrusin, E. M.; Kraker, A. J.; Lee, H. H.;
16		Lunney, E. A.; Mitchell, L. H.; Ortwine, D. F.; Smaill, J. B.; Swan, L. M.; Denny, W. A. 4-
17		Phenylpyrrolo[3,4-c]carbazole-1,3(2H,6H)-dione inhibitors of the checkpoint kinase
18		wee1. Structure-activity relationships for chromophore modification and phenyl ring
19		substitution. J. Med. Chem. 2006 , 49, 4896-4911.
20	10.	Palmer, B. D.; Smaill, J. B.; Rewcastle, G. W.; Dobrusin, E. M.; Kraker, A.; Moore, C. W.;
21		Steinkampf, R. W.; Denny, W. A. Structure-activity relationships for 2-anilino-6-
22		phenylpyrido[2,3-d]pyrimidin-7(8H)-ones as inhibitors of the cellular checkpoint kinase
23		Wee1. Bioorg. Med. Chem. Lett. 2005, 15, 1931-1935.
24	11.	Kreahling, J. M.; Foroutan, P.; Reed, D.; Martinez, G.; Razabdouski, T.; Bui, M. M.;
25		Raghavan, M.; Letson, D.; Gillies, R. J.; Altiok, S. Wee1 inhibition by MK-1775 leads to
26		tumor inhibition and enhances efficacy of gemcitabine in human sarcomas. PLOS 2013,
27		<i>8</i> , e57523.
28	12.	Do, K.; Wilsker, D.; Ji, J.; Zlott, J.; Freshwater, T.; Kinders, R. J.; Collins, J.; Chen, A. P.;
29		Doroshow, J. H.; Kummar, S. Phase I study of single-agent AZD1775 (MK-1775), a Wee1
30		kinase inhibitor, in patients with refractory solid tumors. J. Clin. Oncol. 2015, 33, 3409-
31		3415.
32	13.	Leijen, S.; van Geel, R. M.; Pavlick, A. C.; Tibes, R.; Rosen, L.; Razak, A. R.; Lam, R.;
33		Demuth, T.; Rose, S.; Lee, M. A.; Freshwater, T.; Shumway, S.; Liang, L. W.; Oza, A. M.;
34		Schellens, J. H.; Shapiro, G. I. Phase I study evaluating WEE1 inhibitor AZD1775 as
35		monotherapy and in combination with gemcitabine, cisplatin, or carboplatin in patients
36		with advanced solid tumors. J. Clin. Oncol. 2016, 34, 4371–4380.
37	14.	Tong, Y.; Torrent, M.; Florjancic, A. S.; Bromberg, K. D.; Buchanan, F. G.; Ferguson, D. C.;
38		Johnson, E. F.; Lasko, L. M.; Maag, D.; Merta, P. J.; Olson, A. M.; Osterling, D. J.; Soni, N.;
39		Shoemaker, A. R.; Penning, T. D.; Pyrimidine-based tricyclic molecules as potent and
40		orally efficacious inhibitors of wee1 kinase. ACS Med. Chem. Lett. 2015, 6, 58-62.
41	15.	Squire, C. J.; Dickson, J. M.; Ivanovic, I.; Baker, E. N. Structure and inhibition of the
42		human cell cycle checkpoint kinase, wee1a kinase: an atypical tyrosine kinase with a key
43		role in CDK1 regulation. Structure 2005, 13, 541-550.

- 1 16. Mouse microsomal stability was used as a routine assay for translation of clearance into 2 the efficacy models.
- 3 17. Note that the number of values for all data collected for each compounds were at least
 4 N=2.
- 518.The coordinates used for docking were obtained from the Protein Data Bank with the
entry code 1X8B
- For evidence of on-target activity see TR-FRET kinome profiling data of compound 33 in
 supporting information.
- 9 20. In house tumor efficacy studies showed that Wee1 inhibitor alone, in similar mouse

- 10 xenograft tumor studies, shows only modest tumor growth inhibition thus supporting
- 11 that Wee1 inhibition synergizes the action of Irinotecan. For an example of such studies 12 see reference (14).

