

PII: S0960-894X(97)00164-9

## 3,7-FUNCTIONALIZED-10-METHYL PHENOTHIAZINE: A POTENTIAL TURN SCAFFOLD IN PEPTIDOMIMETICS

T. Kline,\*,§ E. Sieber-McMaster, W. F. Lau,<sup>¶</sup> and S. Natarajan

Bristol-Myers Squibb Pharmaceutical Research Institute. P.O. Box 4000, Princeton, New Jersey 08543

Abstract. Molecular modeling studies suggest that the phenothiazine nucleus, embedded in a peptide via attachments at the 3- and 7-positions, may be a possible surrogate for the  $\alpha$ -carbon backbone of five residue turns in a variety of proteins. The synthesis of the orthogonally-protected Fmoc 3-aminoethyl-7-carboxyethyl-10-methylphenothiazine (1) is described. © 1997 Elsevier Science Ltd.

Scaffolding of the backbone is an established way to probe for the active conformation of a protein or peptide.<sup>1</sup> A suitable scaffold should mimic the active backbone motif (e.g.,  $\alpha$  helix, turn, or  $\beta$  sheet) and should permit suitable functionalization to append the required sidechain groups. The phenothiazine nucleus was selected as a potential scaffold for the middle three residues for a five residue turn. Figure 1 shows how a 3,7-functionalized phenothiazine orients X and Y, the peptide chains at, respectively, the amino and carboxy termini of the scaffold, with respect to the *i* and *i*+4 residues flanking the turn. A ring-substituted phenothiazine might also be designed to add substituents congruent to the R<sub>(*i*+1)</sub>, R<sub>(*i*+2)</sub>, and R<sub>(*i*+3)</sub> sidechains, although  $\alpha$  carbon chirality would be lost.



Figure 1: Sequence of a peptide replaced by phenothiazine scaffold. The portions X and Y of the peptide (above) are outside the turn; residues with sidechains  $R_{(i)}$ - $R_{(i+4)}$  describe the turn. These functional groups can be substituted onto the phenothiazine (below).

A search of secondary structures gave over 75 proteins containing this turn/loop motif in which the backbone atoms of the *i*+1, *i*+2, and *i*+3 residues overlap well with the phenothiazine template. In these proteins the distances between  $C\alpha(i)$  and  $C\alpha(i+4)$  are within 9.2-9.6 Å, and the distances between  $C\alpha(i+1)$  and  $C\alpha(i+3)$  are between 6.9-7.1Å.<sup>2</sup> The corresponding distances in the phenothiazine compound are 9.6-12.3 Å and 7.3 Å. In Figure 2, the phenothiazine is shown superimposed on one example of this turn motif that spans Asp133-Asn137 in the calmodulin structure determined by Babu et al.<sup>3</sup> In this example, the congruent region of the protein is one of the calcium-binding sites.

<sup>&</sup>lt;sup>§</sup>Present address: PathoGenesis Corporation, 201 Elliot Avenue West, Suite 150, Seattle, Washington 98119. Direct all correspondence to this author.

<sup>&</sup>lt;sup>1</sup>Present address: Rhone-Poulenc Rorer, 500 Arcola Road, Collegeville, PA 19426-0800.



Figure 2: Stereo view of superimposed structures of calmodulin Asp133-Asn137 and 2-ethyl, 3-(2-N-methylamido)ethyl, 7-(2aceto)ethyl, 10-pentylphenothiazine. The view shows the region of the protein backbone corresponding to the template.

In our target, phenothiazine 1, the residues that compose the turn are replaced by the tricyclic ring system. Thus successful use of this surrogate requires that the sacrifice of the sidechains of i+1, i+2, and i+3 be compensated by the proper orientation of the *i* and i+4 sidechains. Positions 3 and 7, with orthogonal protection on the N- and C-termini, are the points of attachment to the peptide chains. It was expected that 1 would be amenable to both solution- and solid-phase chemistry.



Our initial attempts for the synthesis of 1 followed literature reports of phenothiazine formylation<sup>1,4</sup> and cyanomethylation.<sup>5</sup> In our hands, however, these reactions (Scheme I) gave disappointing yields and tedious separations. The generation of 2- cyanomethyl compound **5A** along with the desired regioisomer **5B** is likely to arise from an aryne intermediate.<sup>6</sup>



## Scheme I

A more efficient strategy uses essentially the same chemistry to append both the aminoethyl and carboxyethyl sidechains.<sup>7</sup> As shown in Scheme II, arylbromide 4 was readily alkylated by the Heck reaction<sup>8</sup> to the  $\alpha$ , $\beta$ -unsaturated ester 7.<sup>9</sup> Reduction,<sup>10</sup> saponification, and Curtius rearrangement gave 10, protected as a benzyl carbamate. Partial repetition of the sequence (regioselective bromination, Heck reaction, reduction) gave the

methyl ester 12, an intermediate (unlike 7) unstable to silica chromatography. The Cu/NaBH<sub>4</sub> system was highly sensitive to the impurities in crude 12, whereas Ni/NaBH<sub>4</sub> was not; the latter gave good yields of 13 that could then be purified. The Cbz protecting group at the N-terminus was exchanged for a Fmoc group as shown, and the ester saponified.



g.. NaBH<sub>4</sub> - NiCl<sub>2</sub>, MeOH-THF (7:3) 0 °C, 31% from 11 h. HBr/ AcOH i. 2.1 equiv. NaOH j. Fmoc-OSu, THF-dioxane-10% Na<sub>2</sub>CO<sub>3</sub>, 73% from 13

## Scheme II

The phenothiazine nucleus occurs in biologically-active compounds that span a wide spectrum of therapeutic effects; atoms 3, 7, and 10 are frequently (although not exclusively) the substituted positions.<sup>11</sup> By modeling studies, we have superimposed peptide-substituted 3-aminoethyl-7-carboxyethyl phenothiazines on a commonly-observed turn in diverse proteins. We then synthesized a scaffold having a methylated N-10 and orthogonally protected amino and carboxy groups at, respectively, the 3- and 7-sidechains in order to test the hypothesis that active peptidomimetics could be generated from this ring system. Results of incorporating scaffold 1 in a series of novel compounds will be presented in a subsequent report.

Acknowledgments: The authors would like to thank Ms. Yolanda Pan for NMR data, the Analytical R & D Department for mass spectra and elemental analyses, and Ms. Jeannette Manello for literature searches. We appreciate the efforts of Dr. C. A. Meyers for his interest and encouragement in the work and critical reading of this manuscript.

## **References and Notes:**

- This subject is reviewed periodically. See, for example, excellant summary articles by (a) Farmer, P. S. In Drug Design; Ariens, E. J., Ed.; Academic: San Diego, 1980; Vol X, pp 119-143, (b) Kahn, M. Synlett 1993, 821, (c) Olson, G. L.; Bolin, D. R.; Bonner, M. P.; Bos, M.; Coo, C. M.; Fry, D. C.; Graves, B. J.; Hatada, M.; Hill, D. E.; Kahn, M.; Madison, V. S.; Rusiecki, V. K.; Sarabu, R.; Sepinwall. J.; Vincent, G. P.; Voss, M. J. Med. Chem. 1993, 36, 3039, (d) McDowell, R. S.; Artis, D. R. In Annual Reports in Medicinal Chemistry; Bristol, J. A., Ed.; Academic: San Diego, 1995; Vol 30, pp 265-274.
- 2. A search was carried on SYBYL's binary protein database using the protein database search option: SYBYL Molecular Modeling System, Version 6.1, TRIPOS Assoc., St. Louis, MO, U.S.A. Segments from various proteins in which the inter-C $\alpha$  distance between residue *i* and *i*+4 was between 9.2 to 9.6 Å and where the inter-C $_{\alpha}$  distance between residue *i*+1 and *i*+3 was between 6.9 Å and 7.1 Å were identified. When these segments (75 segments obtained from various proteins) were overlaid with C $\alpha$  atoms of the calmodulin segment (3cln) Asp 133-Asn 137, they were found to have RMS deviation of  $\leq 0.5$ Å.
- 3. Babu, Y. S.; Bugg, C. E.; Cook, W. J. J. Mol. Biol. 1988, 204, 191.
- 4. Farcasan, V.; Oprean, I.; Bodea, C. Rev. Roumaine de Chemie 1970, 15, 1433.
- 5. Biehl, E.; DePaul S.; Khanapure, S. P.; Self, J. L.; Siriwardane, U.; Taylor, S.; Tran, L. K.; Tschantz, M. A. Heterocycles 1990, 31, 2209.
- 6. The regio isomers were separated by flash chromatography on silica gel (5% ethyl acetate/hexane). The assignments were based on distinct <sup>1</sup>H NMR signals for the CH<sub>2</sub> protons:  $\delta$  3.83 for **5A**, the earlier-eluting fraction, and  $\delta$  3.64 for **5B**. This is consistent with literature reports (ref 4) for this reaction and <sup>1</sup>H NMR assignments for a compound analogous to **5B**.
- 7. This general strategy was used to prepare a dibenzofuran β turn mimetic: Diaz, H.; Kelly, J. W. Tetrahedron Lett. **1991**, 32, 5725.
- 8. Zebovitz, T. C.; Heck, R. F. J. Org. Chem. 1977, 42, 3907.
- 9. On a large (25 g, 86 mmol) scale, a substantial amount of dealkylation to the  $\alpha$ ,  $\beta$ -unsaturated acid occurred, giving a ratio of 1.3:1 acid to ester. Although the mechanism was not rigorosuly investigated, we attribute this to incompletely neutralized HBr. Following realkylation to 7 with KHCO<sub>3</sub> and CH<sub>3</sub>I, Scheme II was resumed and proceeded as shown.
- 10. Narisada, M.; Horibe, I.; Watanabe, F.; Takeda, K. J. Org. Chem. 1989, 54, 5308.
- Antiinflammatory: Fortin R.; Guindon, Y.; Lau, C. K.; Rokach, J.; Yoakim, C. Eur. Patent 138 481, 1984; Chem. Abstr. 1984, 104, 15081. Walford, G.; Shen, T-Y.; Witzel, B. E.; Greenwald, R. Fr. Demand FR 2053020 710521, 1970; Chem. Abstr 1970, 76, 140759. J. McDermed. US Patent 4 681 878, 1987; Chem. Abstr 1987, 108, 26974. Antimicrobial: Shah, V. H.; Baxi, A. J.; Parikh, A. R. J. Int. Chemist 1986, 58, 165. Tozer, T.; Tuck, L. D.; Cymerman C. J. J. Med. Chem. 1969, 12, 294. Neurological: Olney, J. U.S. Patent 4 833 138A, 1987; Chem. Abstr. 1987, 112, 30668. Antitumor: Korosi, J.; Csaba, G. Ger Offen DE1906527 691204, 1969; Chem. Abstr. 1969, 72, 55477.

(Received in USA 3 February 1997; accepted 13 March 1997)