



Contents lists available at ScienceDirect

Bioorganic & Medicinal Chemistry Letters

journal homepage: www.elsevier.com/locate/bmclSpiro[pyrrolidine-3,3'-oxindoles] as 5-HT₇ receptor ligandsÁdám Andor Kelemen^a, Grzegorz Satała^b, Andrzej J. Bojarski^b, György M. Keserű^{a,*}^a Medicinal Chemistry Research Group, Research Centre for Natural Sciences, Hungarian Academy of Sciences, Magyar tudósok körútja 2, H3660, 23-3650, 96, 1117 Budapest, Hungary^b Department of Medicinal Chemistry, Institute of Pharmacology, Polish Academy of Sciences, 12 Smętna Street, 31-343 Krakow, Poland

ARTICLE INFO

Keywords:

Oxindole
5-HT₇R
G-protein coupled receptor
Oxidative spiro-rearrangement
Pharmacophore model

ABSTRACT

Here we report the design and synthesis of spiro[pyrrolidine-3,3'-oxindole] derivatives representing a novel scaffold of 5-HT₇ receptor ligands. The synthesized analogues were validated as low nanomolar ligands showing selectivity in a panel of related serotonin receptor subtypes including 5-HT_{1A}R, 5-HT_{2A}R and 5-HT₆R.

The serotonin receptor subtype-7 (5-HT₇R) is expressed in both the central nervous system (CNS) and peripheral tissues, and coupled positively to G_{αs} (activation raises cAMP levels)¹ or G_{α12} protein. The 5-HT₇R plays role in the regulation of body temperature, sleep-wake rhythm, circadian rhythm, and mood. Thus the receptor has become rapidly an important target for several important CNS-related indications with proved in vivo efficacy in the animal models of depression,^{2,3} sleep disorders,⁴ anxiety,⁵ learning and memory deficits,⁶ and autism spectrum disorders.⁷

To date the candidate drug with the pyrazolo[3,4-d]azepine core (JNJ-18308683 **1**) has reached the clinic (a phase 2 study is currently recruiting⁸), and a number of selective and non-selective antagonists¹ (e.g. SB-656104 **2**, DR-4004 **3**) and agonists have been proved as investigational compounds (see Fig. 1 for examples).

An early 3D pharmacophore model was built on the basis of thirty known 5-HT₇R antagonists including DR-4004 and analogues, SB-258719 (structure not shown) and analogues, SB-269970 (structure not shown) analogue etc.⁹ The 5-HT₇R binding pharmacophore has been updated¹⁰ using broad structure-activity relationship (SAR) data and structure-based docking validated by site-directed mutagenesis experiments. Pharmacophore features of 5-HT₇R ligands include an aromatic ring (in stacking interactions with Phe^{3.28} and Tyr^{7.43}), two further hydrophobic regions HYD₁ and HYD₂ (facing towards Phe^{6.52}), a hydrogen bond acceptor (binding to Ser^{5.42} and/or Thr^{5.43}) next to HYD₁,

and a positive ionizable moiety located at 5–6 Å distance from HYD₁ that contacts Asp^{3.32} (Fig. 2).

Tetrahydrobenzindoles (e.g. DR-4004 (**3**))^{11,12} and corresponding oxindoles^{13,14} were validated as potent 5-HT₇R scaffolds. As an attempt to synthesize new, selective and less lipophilic compounds we designed spiro[pyrrolidine-3,3'-oxindoles] (**4**) as potential 5-HT₇R ligands. Aryl-piperazine analogues attached to the common spiro[pyrrolidine-3,3'-oxindole] core could provide ligands with more favourable ADME characteristics by exchanging the carbocycle of the tetrahydrobenzindole to the more polar and less rigid pyrrolidine ring. In order to design selective compounds against other serotonin receptors (5-HT_{1A}R, 5-HT_{2A}R, 5-HT₆R) we used the pharmacophore models by López *et al.*^{9,10} and further modified by Medina *et al.*¹⁵ These models suggest that decreasing the distance between PI (positive ionizable) and HBA (H-bond acceptor) features, facilitating interactions to Ser^{6.55} (Ala^{6.55} at 5-HT_{1A}R), introducing polar substituents at HYD₁-HYD₂ to contact Arg^{7.36} (that is absent in 5-HT_{1A}R) would be beneficial. Following these suggestions we set our objective for exploring selectivity drivers around the spiro[pyrrolidine-3,3'-oxindoles] core.

The 5-HT₇R pharmacophore suggested that we need a four-atom linker between the spiro carbon C-3 to the pyrrolidine-nitrogen and therefore connected this with two methylenes (Fig. 2) to the basic moiety of compounds with General Structure 4. Thus in line with other studies^{12,11,16–19} we focused on compounds with two atom spacer (Fig. 3, compound **5**, although compound **6** was also synthesized in

* Corresponding author.

E-mail address: kelemen.adam@tk.mta.hu (G.M. Keserű).<https://doi.org/10.1016/j.bmcl.2018.06.019>Received 15 April 2018; Received in revised form 1 June 2018; Accepted 11 June 2018
0960-894X/ © 2018 Published by Elsevier Ltd.

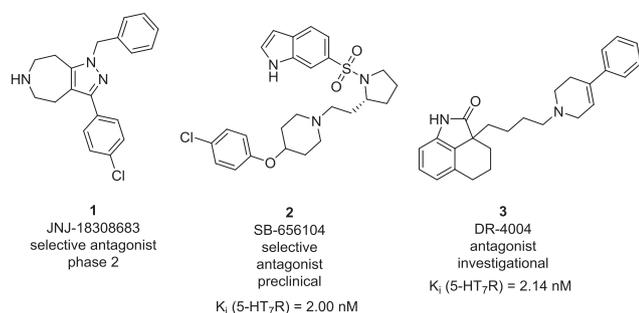


Fig. 1. Structures of the most prominent 5-HT₇R antagonists.

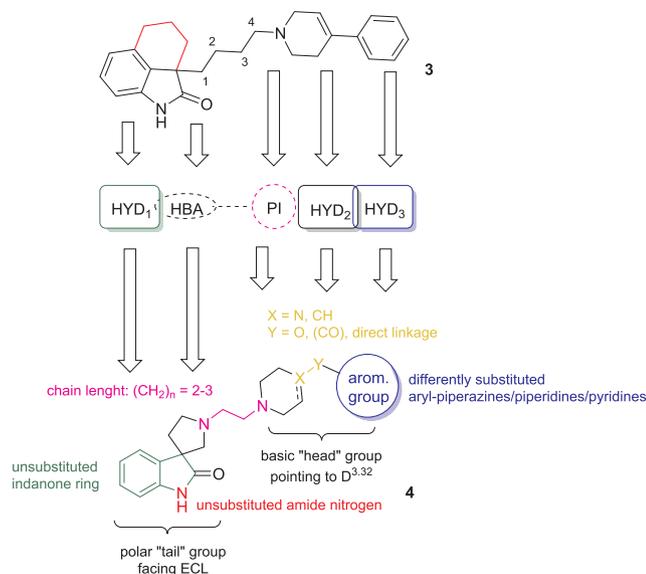
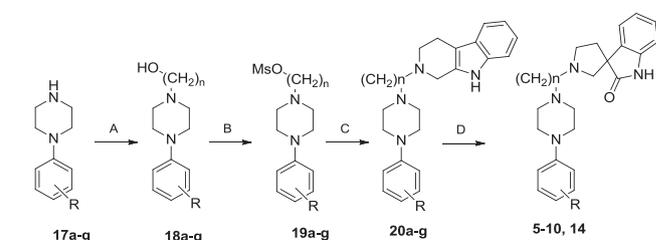


Fig. 2. Design concept for the SAR analysis of spiro[pyrrolidine-3,3'-oxindoles] (4).



Scheme 1. Synthesis of the aryl-piperazine derivatives (5–10, 14). Reagents and conditions: (A) 2-bromoethanol (or 3-bromopropanol in case of 6), K₂CO₃, MeCN, RT, overnight; (B) MsCl, TEA, DCM, reflux, 2 h; (C) tryptoline/tetrahydro-β-carboline, K₂CO₃, MeCN, reflux, overnight; (D) NBS, cc. AcOH, THF, water, 1.5 h, 0 °C.

order to investigate its effect on flexibility and HBA-PI distances.

Halo-scan, however, was planned to explore the substituent effects around the phenyl ring of the aryl-piperazine moiety comparing the profile of the unsubstituted 7 and the meta-Cl 8, ortho-Cl 9, para-Cl 5, and 3,4-dichloro-derivatives 10. Halo-scan methodology explores the functionalization of the ligands thus filling subpockets, and probing the impact of introducing lipophilic, and possibly H-bonding halogen atoms.²⁰

In order to explore the impact of the HYD₂/HYD₃ features on the 5-HT₇R affinity and selectivity we replaced the canonical aryl-piperazine by phenyl-, and phenoxy-piperidines 11, 12, 15, 5,6-dihydropyridine 13, and benzoyl-piperazine 16 moieties.

Aryl-piperazine derivatives 5–10, 14 were synthesized from the corresponding secondary amines 17a–g. First, cyclic secondary amines were alkylated by 2-bromoethanol (in case of 1-(4-chlorophenyl)piperazine 17a either by 3-bromopropanol) followed by the mesylation of the appropriate alcohols using mesyl chloride. A protecting group on the basic nitrogen of the tryptoline is necessary to avoid decomposition during the spiro-rearrangement reaction. As depicted in Scheme 1, *N*-alkylation by the corresponding mesylates leading to intermediates 20a–g provided the desired protection of the tetrahydro-β-carboline nitrogen for the final spirocyclization step to afford derivatives 5–10 and 14.²¹

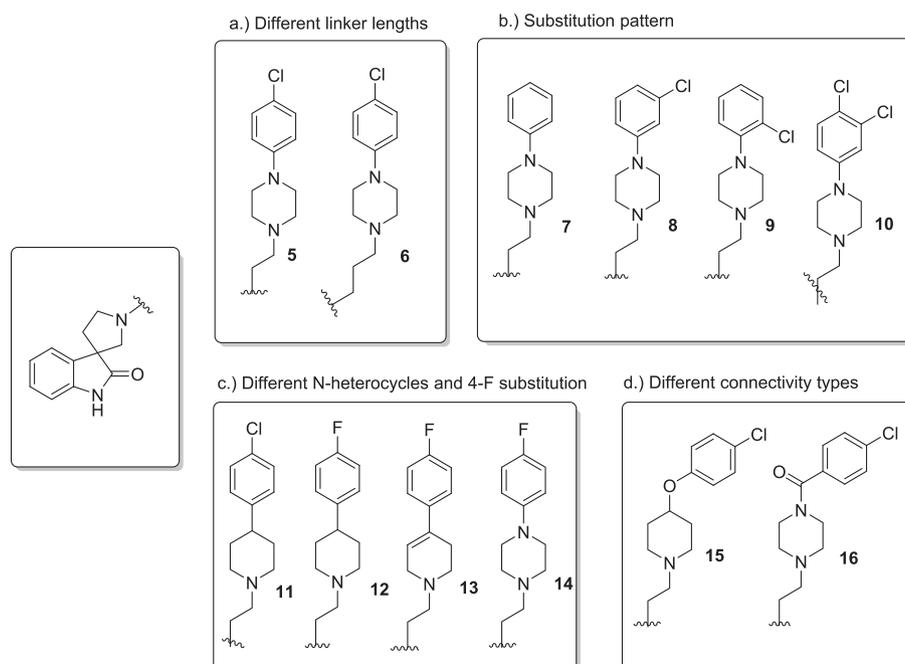
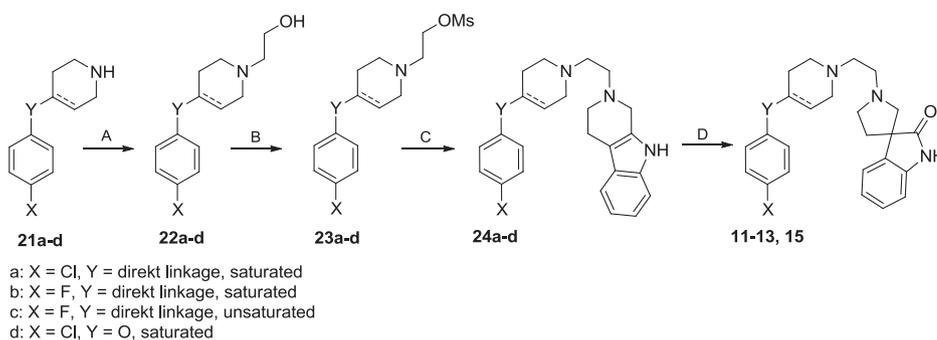
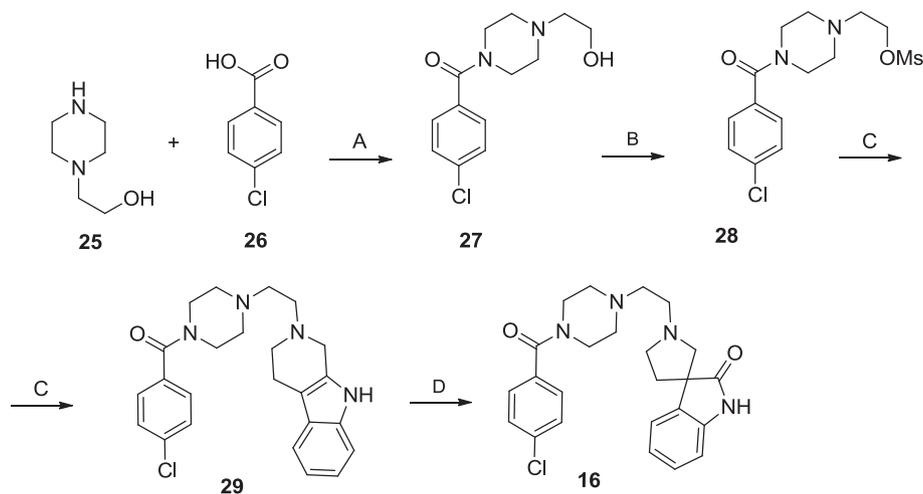


Fig. 3. Spiro[pyrrolidine-3,3'-oxindoles] designed for SAR studies.



Scheme 2. Synthesis of aryl-piperidine (**11–12**), and 5,6-dihydropyridine (**13**), and phenoxy (**15**) analogues. Reagents and conditions: (A) 2-bromoethanol, K_2CO_3 , MeCN, RT, overnight; (B) MsCl, TEA, DCM, reflux, 2 h; (C) tryptoline/tetrahydro- β -carboline, K_2CO_3 , MeCN, reflux, overnight; (D) NBS, cc. AcOH, THF, water, 1.5 h, 0 °C.



Scheme 3. Synthetic route leading to the amide-analogue (**16**). Reagents and conditions: (A) HATU, DIPEA, DMF, RT, overnight; (B) MsCl, TEA, DCM, reflux, 2 h; (C) tryptoline/tetrahydro- β -carboline, K_2CO_3 , MeCN, reflux, overnight; (D) NBS, cc. AcOH, THF, water, 1.5 h, 0 °C.

The synthesis of the aryl-piperidine **11–12**, the 5,6-dihydro-pyridine **13**, and the phenoxy **15** analogues (Scheme 2) was also started from the corresponding secondary amines **21a–d**, followed by the alkylation with 2-bromoethanol, to afford the alcohols **22a–d**. The crude mesylated derivatives **23a–d** were then used for alkylation of the tetrahydro- β -carboline to get **24a–d** that was spirocyclized to **11**, **12**, **13**, **15**, respectively.

1-(2-Hydroxyethyl)piperazine (**25**) was used as starting material for the synthesis of the benzoyl-piperazinyl analogue **16** (Scheme 3). The acylation²² of **25** with the 4-chlorobenzoic acid **26** gave the corresponding amide **27**, which was treated with mesyl-chloride to yield the mesylate **28**. After the alkylation of tetrahydro- β -carboline with **28**, the spirocyclization of intermediate **29** afforded **16**.

The synthesized compounds were investigated in competition binding assays against 5-HT₇R and other closely related serotonin receptor subtypes 5-HT_{1A}R, 5-HT_{2A}R and 5-HT₆R (Table 1).

The unsubstituted phenylpiperazine derivative **7** of the spiro[pyrrolidine-3,3'-oxindole] core showed reasonably high affinity towards the target, however its selectivity was moderate. Halo-scan around the phenyl ring revealed that the 5-HT₇R affinity and 5-HT_{1A}R and 5-HT₆R selectivities are increasing from ortho **9** – meta **8** – para **5** direction. In fact, the para-Cl derivate (**5**) showed low nanomolar affinity for the target and more than hundred-fold selectivity against two of the three other serotonin receptors. Selectivity against these receptors was further improved for the para-F analogue **14**. As compared to compound **5**, the longer ([CH₂]₃) alkyl chain in **6** increased the distance between the PI and HBA features, accounting for an overall loss of affinity for the 5-

HT₇R. Selectivity against 5-HT_{1A}R decreased, however the selectivity against the 5-HT_{2A}R subtype has improved. **6** showed lower selectivity (65.6-fold) towards 5-HT₆R than **7** (163.3-fold).

Starting from the most active 4-fluoro derivative (**14**) the 5,6-dihydropyridine derivative (**13**) showed decreased selectivity (91.8-fold) against 5-HT_{1A}R. Replacing the piperazine ring by piperidine (**12**) gave 10-times lower affinity that was further confirmed by the corresponding 4-chloro analogue (**11**). Selectivities against 5-HT_{1A}R and 5-HT_{2A}R did not change significantly, but the 5-HT₆R selectivity decreased ten-times. Similar to **11** and **12**, introduction of the phenoxy-piperidine moiety (**15**) improved the affinity towards 5-HT₆R. Finally, the benzoyl-piperazine derivative (**16**) showed reduced affinity to 5-HT₇R, however, it has the highest selectivity against 5-HT_{2A}R (32.8-fold).

In summary, preliminary SAR data demonstrates that spiro[pyrrolidine-3,3'-oxindoles] are potent and selective 5-HT₇R ligands. We confirmed that the 2-methylene linker ensures the ideal distance between HYD1 and HYD2 and therefore it is beneficial for the 5-HT₇R affinity. Although previous findings^{13,23} suggested that the para substitution at the aryl-piperazine moiety may abolish 5-HT₇R affinity we showed that it provides spiro[pyrrolidine-3,3'-oxindoles] with reasonable affinity and selectivity against 5-HT_{1A}R and 5-HT₆R. Actually, the 4-fluoro analogue (**14**) showed the best affinity and most remarkable selectivity against 5-HT_{1A}R and 5-HT₆R. Selectivity against 5-HT_{2A}R might be improved by replacing the phenyl substituent of the piperazine by a benzoyl group. The present results show the potential of this novel chemotype and validate its further optimization for more detailed *in vivo* characterization in diseases models.

Table 1

Serotonergic activity of the spirooxindole derivatives as measured in binding assays of four serotonin receptors (K_i values are in μM). 5-HT₇R affinities and selectivities are shown in bold and italics, respectively. Assay reference compounds: serotonin in 5-HT_{1A}R and 5-HT₇R assays, chlorpromazine in 5-HT_{2A}R and methiothepine in 5-HT₆R assay. Results were expressed as means of at least three separate experiments ($\text{SD} \leq 24\%$).

ID	Structure	5-HT _{1A}	5-HT _{2A}	5-HT ₆	5-HT ₇
5		3.755 <i>129.5</i>	0.068 <i>2.3</i>	4.737 <i>163.3</i>	0.029
6		2.891 <i>43.1</i>	0.715 <i>10.7</i>	5.559 <i>83.0</i>	0.067
7		6.005 <i>60.1</i>	0.496 <i>5.0</i>	6.556 <i>65.6</i>	0.100
8		2.687 <i>48.9</i>	0.351 <i>6.4</i>	2.005 <i>36.5</i>	0.055
9		1.096 <i>4.5</i>	0.452 <i>1.8</i>	2.843 <i>11.6</i>	0.245
10		4.054 <i>106.7</i>	0.270 <i>7.1</i>	1.131 <i>29.8</i>	0.038
11		14.900 <i>52.5</i>	0.404 <i>1.4</i>	4.577 <i>16.1</i>	0.284
12		52.010 <i>189.1</i>	0.259 <i>0.9</i>	5.285 <i>19.2</i>	0.275
13		3.949 <i>91.8</i>	0.075 <i>1.7</i>	6.267 <i>145.7</i>	0.043
14		14.160 <i>674.3</i>	0.059 <i>2.8</i>	4.744 <i>225.9</i>	0.021
15		13.790 <i>114.0</i>	0.343 <i>2.8</i>	1.693 <i>14.0</i>	0.121
16		48.240 <i>91.2</i>	17.330 <i>32.8</i>	7.746 <i>14.6</i>	0.529

Acknowledgments

The authors are grateful to Ágnes Gömörö and László Drahos (MS Proteomics Research Group, Core Technologies Centre) for exact mass (MS) measurements.

This work was supported by the National Brain Research Program grant (2017-1.2.1-NKP-2017-00002) and by statutory funds of the Institute of Pharmacology, Polish Academy of Sciences in Krakow.

A. Supplementary data

Supplementary data (synthetic procedures, analytical data, description of the cell-based assays) associated with this article can be found, in the online version, at <http://dx.doi.org/10.1016/j.bmcl.2018.06.019>.

References

- Nikiforuk A. *CNS Drugs*. 2015;29:265–275.
- Mullins U, Gianutsos G, Eison AS. *Neuropsychopharmacology*. 1999;21:352–367.
- Sleight AJ, Carolo C, Petit N, Zwingelstein C, Bourson A. *Mol Pharmacol*. 1995;47:99–103.
- Lopez-Rodriguez M, Benhamu B, Morcillo M, Porras E, Lavandera J, Pardo L. *Curr Med Chem Nerv Syst Agents*. 2004;4:203–214.
- Galici R, Boggs JD, Miller KL, Bonaventure P, Atack JR. *Behav Pharmacol*. 2008;19:153–159.
- Meneses A, Terrón JA. *Behav Brain Res*. 2001;121:21–28.
- Ciranna L, Catania MV. *Front Cell Neurosci*. 2014;8:250.
- U.S. National Library of Medicine. U.S. National Library of Medicine. <https://clinicaltrials.gov/> (accessed December 19, 2016).
- López-Rodríguez ML, Porras E, Benhamú B, Ramos JA, Morcillo MJ, Lavandera JL. *Bioorg Med Chem Lett*. 2000;10:1097–1100.
- López-Rodríguez ML, Porras E, Morcillo MJ, et al. *J Med Chem*. 2003;46:5638–5650.
- Kikuchi C, Nagaso H, Hiranuma T, Koyama M. *J Med Chem*. 1999;42:533–535.
- Kikuchi C, Ando T, Watanabe T, et al. *J Med Chem*. 2002;45:2197–2206.
- Volk B, Barkóczy J, Hegedus E, et al. *J Med Chem*. 2008;51:2522–2532.
- Volk B, Gacsályi I, Pallagi K, et al. *J Med Chem*. 2011;54:6657–6669.
- Medina RA, Sallander J, Benhamú B, et al. *J Med Chem*. 2009;52:2384–2392.
- Deau E, Robin E, Voinea R, et al. *J Med Chem*. 2015;58:8066–8096.
- Perrone R, Berardi F, Colabufo NA, Lacivita E, Leopoldo M, Tortorella V. *J Med Chem*. 2003;46:646–649.
- Pittala V, Pittala D. *Mini-Reviews. Med Chem*. 2011;11:1108–1121.
- Leopoldo M, Lacivita E, Berardi F, Perrone R, Hedlund PB. *Pharmacol Ther*. 2011;129:120–148.
- Shah P, Westwell AD. *J Enzyme Inhib Med Chem*. 2007;22:527–540.
- Kelemen Á.A, Satala G, Bojarski AJ, Keseru GM. *Molecules*. 2017;22:2221.
- Zhuang R, Gao L, Lv X, et al. *Eur J Med Chem*. 2017;126:1056–1070.
- Leopoldo M, Berardi F, Colabufo NA, et al. *J Med Chem*. 2004;47:6616–6624.