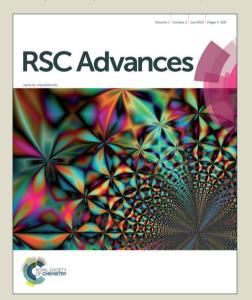


## RSC Advances

This article can be cited before page numbers have been issued, to do this please use: Y. Han, C. Yang, K. Wu, Y. Chen, B. Zhou and M. Xia, RSC Adv., 2015, DOI: 10.1039/C4RA16479B.



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. This Accepted Manuscript will be replaced by the edited, formatted and paginated article as soon as this is available.

You can find more information about *Accepted Manuscripts* in the **Information for Authors**.

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard <u>Terms & Conditions</u> and the <u>Ethical guidelines</u> still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.



Published on 30 January 2015. Downloaded by Gazi Universitesi on 03/02/2015 19:05:04

Cite this: DOI: 10.1039/c0xx00000x

www.rsc.org/xxxxxx

## **ARTICLE TYPE**

## A Facile Naphthalene-Based Fluorescent Chemodosimeter for Mercury **Ions in Aqueous Solution**

Yifeng Han,\* Chengyu Yang, Kai Wu, Yu Chen, Baocheng Zhou and Min Xia\*

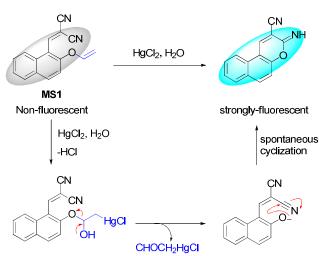
Received (in XXX, XXX) Xth XXXXXXXXX 20XX, Accepted Xth XXXXXXXX 20XX 5 DOI: 10.1039/b000000x

facile naphthalene-based fluorescence chemodosimeter, 2-((2-(vinyloxy)naphthalen-1-yl)methylene) malononitrile (MS1), for rapid, selective and sensitive detection of Hg<sup>2+</sup> by mercury-promoted hydrolysis of 10 vinylether group has been reported. The probe displayed a fast response time, and sensitive fluorescence response (100fold fluorescence enhancement) to the detection of Hg<sup>2+</sup> in aqueous solution.

Mercury, which is widely distributed in the environment such 15 as the air, soil, and water due to its use in batteries, dental amalgam, electrical apparatus, and industrial chemicals, is one of the most ubiquitous and poisonous heavy metals. Mercury ions are not biodegradable, and hence can concentrate through the food chain in the tissues of fish and marine mammals. 20 Excess mercury accumulation may induce strong damage to the central nervous system, various cognitive and motor disorders, and Minamata disease.<sup>2</sup> Due to the toxicity of Hg<sup>2+</sup>, the determination of mercury in biological and environmental samples is crucial both to the monitoring of environmental 25 pollution and to the diagnosis of clinical disorders.

In the past several years, considerable efforts have been made to develop fluorescent chemosensors for Hg<sup>2+</sup> based on the coordination of Hg<sup>2+</sup> to heteroatom-based ligands, Hg<sup>2+</sup> catalyzed desulfurization, and Hg<sup>2+</sup> promoted hydrolysis of 30 the vinyl ether group and  $\beta$ -alkynyl ether group. 3 However, most of them still have limitations such as interference from other coexisting metal ions, poor water-solubility, and laborious synthesis processes expensive chemicals.<sup>4</sup> Therefore, for practical applications, it is still desirable to 35 develop simple Hg<sup>2+</sup> sensors with good water solubility and high selectivity and sensitivity.

Compared with the typically-developed chemosensors,<sup>5</sup> fluorescent chemodosimeters, based-on highly specific chemical reactions between the dosimeters and the analytes, 40 have received much research attention due to their relatively higher selectivity.6 Recently, Peng, Talukdar, Wu, and Ahn's groups have reported fluorescent chemodosimeters based on "deprotection-cyclization strategy" for the detection of fluoride ions,6 while the development of chemodosimeters for 45 the specific determination of Hg<sup>2+</sup> is drawing increasing research efforts. However, among the few available Hg<sup>2+</sup> chemodosimeters reported,<sup>3</sup> most employ the pH-sensitive fluorescein or 7-amino coumarin as the fluorophore and their



Scheme 1 Hydrolysis of MS1 by mercury ions

pH-dependence may pose detection errors to the results. It is therefore strongly desirable to develop simple yet specific fluorescent chemodosimeters for Hg2+ that is immune to pH turbulence.

It is known that  $Hg^{2+}$  catalyzes hydrolysis of vinylether to form the corresponding hydroxyl group. We proposed that the Hg<sup>2+</sup> ion promoted hydrolysis of the vinyl enol ether group in MS1 would generate the hydroxy intermediate, which will readily spontaneous cyclize to form a highly 60 fluorescent chemodosimeter (Scheme 1).

Our research group is actively engaged in the development of novel selective and sensitive fluorescent probes for heavy

Scheme 2 Synthesis of MS1: (a) 1, 2-dibromoethane/K<sub>2</sub>CO<sub>3</sub>, acetone, 65 reflux, 3 h, 62%; (b) t-BuOK/DMSO, rt, 12 h, 46%; (c) CH<sub>2</sub>(CN)<sub>2</sub>/ piperidine, enthanol, rt, 1h, 48%

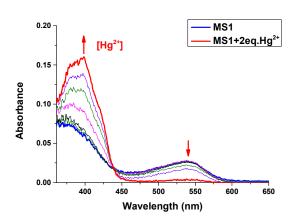
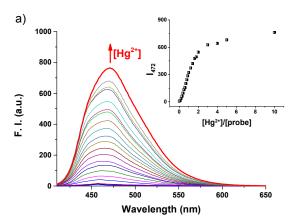
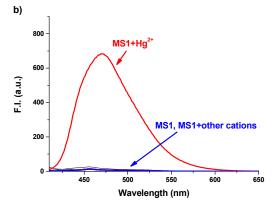


Fig. 1 Absorption spectra of MS1 (20  $\mu$ M) in PBS buffer solution (10 mM, pH 7.4, containing 1% CH<sub>3</sub>CN) in the presence of different concentrations of Hg<sup>2+</sup> (0-2.0 equiv.).

5 metal ions. 8 Herein, we report the synthesis and properties of deprotection-cyclization reaction based fluorescent chemodosimeter (MS1) that shows high selectivity and sensitivity for Hg<sup>2+</sup>.





10 Fig. 2 (a) Fluorescence spectra of MS1 (10  $\mu$ M) in PBS buffer solution (pH 7.4, containing 1% CH<sub>3</sub>CN) in the presence of different concentrations of Hg<sup>2+</sup> (0-50  $\mu$ M) ( $\lambda_{ex}$  = 395 nm). Inset: fluorescence intensity changes as a function of Hg<sup>2+</sup> concentration. (b) Emission spectra of MS1 (10 µM) in PBS buffer solution (pH 7.4, containing 1% 15 CH<sub>3</sub>CN) in the presence of various metal ions ( $\lambda_{ex} = 395$  nm, 5.0 eq. of Hg<sup>2+</sup>, and 10.0 eq. of Co<sup>2+</sup>, Cr<sup>3+</sup>, Cu<sup>2+</sup>, Fe<sup>2+</sup>, Fe<sup>3+</sup>, K<sup>+</sup>, Mg<sup>2+</sup>, Mn<sup>2+</sup>, Na<sup>+</sup>, Na<sup>+</sup>,  $^{1}$ , Pb<sup>2+</sup>, Sn<sup>4+</sup>, Ag<sup>+</sup>, Ca<sup>2+</sup>, and Zn<sup>2+</sup>, respectively).

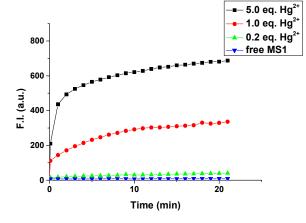


Fig. 3 Time-dependent fluorescence intensity changes of MS1 (10  $\mu$ M) 20 upon addition of various concentration of Hg<sup>2+</sup> (0, 0.2, 1.0, 5.0 equiv. each) in PBS buffer solution (pH 7.4, containing 1% CH<sub>3</sub>CN) ( $\lambda_{ex}$  = 395

As shown in Scheme 2, MS1 can be readily prepared in three convenient steps under facile conditions with high yield with commercially available 2-hydroxy-1naphthaldehyde. The product (MS1) was well characterized by <sup>1</sup>H, <sup>13</sup>C NMR, and HR-MS (ESI†).

We firstly assessed the UV-vis spectroscopic properties of MS1 in PBS buffer solution (10 mM, pH = 7.4, containing 1% 30 CH<sub>3</sub>CN). MS1 (20  $\mu$ M) displayed a moderate UV-vis absorption around 538 nm. Upon addition of Hg<sup>2+</sup> (0-2 equiv.), the absorption band at 538 nm decreased and a new band at 399 nm appeared instantly with an isosbestic point at 439 nm, which is owing to the loss of vinyl enol ether group 35 and the formation of cyclic compound (Fig. 1).

As expected, MS1 alone is almost non-fluorescent ( $\lambda_{ex}$  = 395 nm,  $\Phi = 0.002$ , Table S1, ESI†) in neutral aqueous solution (10 mM PBS buffer, pH 7.4, containing 1% CH<sub>3</sub>CN), while the addition of increasing concentrations of Hg<sup>2+</sup> 40 gradually enhanced the fluorescent signal and ca. 100-fold increasing was observed when 5.0 equiv. of Hg<sup>2+</sup> was added (Fig. 2a, Table S1, ESI†), which was attributed to the cleavage of vinyl enol group by mercury ion promoted hydrolysis reaction and the formation of a highly fluorescent 45 cyclic compound (Scheme 1). Moreover, a blue-green fluorescent compound 5 have been isolated from MS1-Hg<sup>2+</sup> system (ESI†), which was agreed well with the proposed mercury induced deprotection-cyclization mechanism.

Subsequently, the time-dependence of MS1 fluorescence 50 was also evaluated in the presence of different concentration of Hg<sup>2+</sup>. The result shows that the fluorescence of all tested solutions remarkably increased to their maximum value within the 10 minutes. No changes in fluorescense were detected in the absence of  $Hg^{2+}$  (Fig. 3).

Further, the fluorescence titration of MS1 with various metal ions was conducted to examine the selectivity (Fig.2b). Much to our delight, the turn-on response of MS1 is highly specific for Hg<sup>2+</sup> and no obvious change of fluorescent emission was observed when it is treated with Co<sup>2+</sup>, Cr<sup>3+</sup> 60 Cu<sup>2+</sup>, Fe<sup>2+</sup>, Fe<sup>3+</sup>, K<sup>+</sup>, Mg<sup>2+</sup>, Mn<sup>2+</sup>, Na<sup>+</sup>, Ni<sup>2+</sup>, Pb<sup>2+</sup>, Sn<sup>4+</sup>, Ag<sup>+</sup>, Ca<sup>2+</sup>, and Zn<sup>2+</sup>. It should be mentioned that **MS1** still responds to Hg<sup>2+</sup> sensitively even in the presence of other relevant

Published on 30 January 2015. Downloaded by Gazi Universitesi on 03/02/2015 19:05:04

Published on 30 January 2015. Downloaded by Gazi Universitesi on 03/02/2015 19:05:04

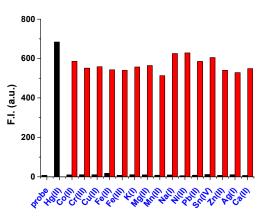
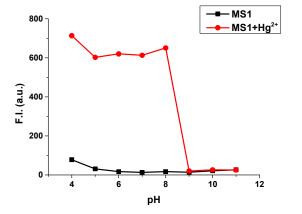


Fig. 4 Fluorescence responses of MS1 to various metal ions. Black bars represent the addition of 5.0 equiv. of Hg<sup>2+</sup> and 10.0 equiv. of the other appropriate metal ion to a 10  $\mu$ M solution of MS1. Red bars represent the 5 addition of 5.0 equiv. of Hg<sup>2+</sup> to the solutions containing MS1 (10  $\mu$ M) and the appropriated metals (10.0 equiv.).

competing ions (Fig. 4). Therefore, these results suggest that MS1 displays high selectivity toward Hg<sup>2+</sup> in neutral aqueous

- Moreover, the Hg<sup>2+</sup>-sensing ability of **MS1** at a wide range of pH values was investigated. As depicted in Fig. 5, MS1 alone is inert to pH in the range of 4.0-11.0. But in the presence of Hg<sup>2+</sup>, MS1 have no fluorescence response in the highly basic environment (pH  $\geq$  9) due to the reaction rate of 15 mercury ion-promoted hydrolysis of vinyl enol ether becomes slow at high pH value. However, satisfactory Hg<sup>2+</sup>-sensing abilities were exhibited in the range of pH from 4.0 to 8.0, indicating that MS1 could be used in neutral natural systems, or a mildly acidic or basic environment.
- For practical purposes, the detection limit of MS1 for the analysis of Hg<sup>2+</sup> was also an important parameter. The fluorescence titration curve revealed that the fluorescence intensity of MS1 at 470 nm increased linearly with the amount of Hg<sup>2+</sup> in the range of 0-5.0  $\mu$ M ( $R^2 = 0.994$ ) (Fig. S1, ESI†). 25 Thus, the detection limit of MS1 for Hg<sup>2+</sup> was calculated to be  $4.31 \times 10^{-8}$  M (Hg content = 8.8 ppb), which reveals the high sensitivity for the analysis of the mercury ions.



30 Fig. 5 Effect of the pH on the fluorescence emission of MS1 (10  $\mu$ M) alone and MS1 (10  $\mu$ M) reacted with Hg<sup>2+</sup> (3.0 equiv.).

In conclusion, we have successfully developed a simple naphthalene-based fluorescense probe for Hg<sup>2+</sup> based on mercury triggered cleavage reaction under mild conditions. 35 The probe has the unique advantage of easy-preparation, good water solubility, and excellent selectivity and sensitivity response towards Hg<sup>2+</sup> in aqueous solution. We anticipate that the experimental results of this study will inspire the future design of metal-ion sensors in water for a variety of chemical 40 and biological applications.

This work was supported by the Zhejiang Provincial Natural Science Foundation of China (LY14B020016 and LQ13B020006) and the Program for Innovative Research 45 Team of Zhejiang Sci-Tech University (13060052-Y).

## Notes and references

Department of Chemistry, The Key Laboratory of Advanced Textile Materials and Manufacturing Technology, Zhejiang Sci-Tech University, 50 Hangzhou, 310018, China.

E-mail: zstuchem@gmail.com; Tel: +86-571-86843550; E-mail: xiamin@zstu.edu.cn.

- † Electronic Supplementary Information (ESI) available: Experimental 55 details, synthetic details of MS1, additional spectroscopic data, and copies of NMR spectra. See DOI: 10.1039/b000000x/
- (a) L. Magos, Met. Ions Biol. Syst., 1997, 34, 321-370; (b) M. F. Wolfe, S. Schwarzbach and R. A. Sulaiman, Environ. Toxicol. Chem., 1998, 17, 146-160; (c) P. B. Tchounwou, W. K. Ayensu, N. Ninashvili and D. Sutton, Environ. Toxicol., 2003, 18, 149-175; (d) P. Grandjean, P. Weihe, R. F. White and F. Debes, Environ. Res., 1998, 77, 165-172.
- (a) C. R. Baum, Curr. Opin. Pediatr., 1999, 11, 265-268; (b) E. K. Silbergeld, I. A. Silva and J. F. Nyland, Toxicol. Appl. Pharmacol., 2005, 207, S282-S292; (c) R. K. Zalups and S. Ahmad, J. Am. Soc. Nephrol., 2004, 15, 2023-2031; (d) Z. Zhang, X. Guo, X. Qian, Z. Lu and F. Liu, Kidney Int., 2004, 66, 2279-2282; (e) J. Huang, X. Ma, B. Liu, L. Cai, Q. Li, Y. Zhang, K. Jiang and S. Yin, J. Lumin., 2013, 141, 130-140.
- (a) M. Y. Berezin and S. Achilefu, Chem. Rev., 2010, 110, 2641-2684; (b) K. P. Carter, A. M. Young and A. E. Palmer, Chem. Rev., 2014, 114, 4564-4601; (c) E. M. Nolan and S. J. Lippard, Chem. Rev., 2008, 108, 3443-3480; (d) X. Chen, T. Pradhan, F. Wang, J. S. Kim and J. Yoon, Chem. Rev., 2012, 112, 1910-1956; (e) M. Kaur and D. H. Choi, Chem. Soc. Rev., 2015, 44, 58-77; (f) Z. Guo, S. Park, J. Yoon and I. Shin, Chem. Soc. Rev., 2014, 43, 16-29; (g) L. Yuan, W. Lin, K. Zheng, L. He and W. Huang, Chem. Soc. Rev., 2013, 42, 622-661; (h) J. Fan, M. Hu, P. Zhan and X. Peng, Chem. Soc. Rev., 2013, 42, 29-43; (i) J. Du, M. Hu, J. Fan and X. Peng, Chem. Soc. Rev., 2012, 41, 4511-4535; (j) W. Xuan, C. Chen, Y. Cao, W. He, W. Jiang, K. Liu and W. Wang, Chem. Commun., 2012, 48, 7292-7294; (k) M. Vedamalai and S. P. Wu, Org. Biomol. Chem., 2012, 10, 5410-5416; (1) J. Liu, Y. Q. Sun, P. Wang, J. Zhang and W. Guo, Analyst, 2013, 138, 2654-2660; (m) F. Song, S. Watanabe, P. E. Floreancig, and K. Koide, J. Am. Chem. Soc., 2008, 130, 16460-16461; (n) H. Jiang, J. Jiang, J. Cheng, W. Dou, X. Tang, L. Yang, W. Liu and D. Bai, New J. Chem., 2014, 38, 109-114; (o) L. Chen, L. Yang, H. Li, Y. Gao, D. Deng, Y. Wu and L. Ma, Inorg. Chem., 2011, 50, 10028-10032; (p) M. Saleem, R. Abdullah, A. Ali, B. J. Park, E. H. Choi, I. S. Hong and K. H. Lee, Anal. Methods, 2014, 6, 3588-3597.
- (a) M. Tian and H. Ihmels, Chem. Commun., 2009, 3175-3177; (b) N. Kumari, N. Dey, S. Jha and S. Bhattachary, ACS Appl. Mater. Interfaces, 2013, 5, 2438-2445; (c) S. Madhu, R. Kalaiyarasi, S. K. Basu, S. Jadhav and M. Ravikanth, J. Mater. Chem. C, 2014, 2,

- 2534-2544; (d) M. Tian, L. Liu, Y. Li, R. Hu, T. Liu, H. Liu, S. Wang and Y. Li, Chem. Commun., 2014, 50, 2055-2057.
- 5 (a) P. Dinake, P. E. Prokhorova, V. S. Talanov, R. J. Butcher and G. G. Talanova, Tetrahedron Lett., 2010, 51, 5016-5019; (b) J. H. Kim, J. Y. Noh, I. H. Hwang, J. J. Lee and C. Kim, Tetrahedron Lett., 2013, 54, 4001-4005; (c) P. Srivastava, R. Ali, S. S. Razi, M. Shahid, S. Patnaik and A. Misra, Tetrahedron Lett., 2013, 54, 3688-3693; (d) L. N. Neupane, J. Y. Park, J. H. Park and K. H. Lee, Org. Lett., 2013, 15, 254-257; (e) M. Kumar, N. Kumar, V. Bhalla, H. Singh, P. R. Sharma and T. Kaur, Org. Lett., 2011, 13, 1422-1425; (f) M. Kumar, S. I. Reja and V. Bhalla, Org. Lett., 2012, 14, 6084-
- 6 (a) Y. Peng, Y. M. Dong, M. Dong and Y. W. Wang, J. Org. Chem., 2012, 77, 9072-9080; (b) A. Roy, D. Kand, T. Saha and P. Talukdar, Chem. Commun., 2014, 50, 5510-5513; (c) D. Kim, S. Singha, T. Wang, E. Seo, J. H. Lee, S. J. Lee, K. H. Kim and K. H. Ahn, Chem. Commun., 2012, 48, 10243-10245; (d) J. T. Yeh, P. Venkatesan and S. P. Wu, New J. Chem., 2014, 38, 6198-6204.
- (a) M. Santra, B. Roy and K. H. Ahn, Org. Lett., 2011, 13, 3422-3425; (b) J. Jiang, W. Liu, J. Cheng, L. Yang, H. Jiang, D. bai and W. Liu, Chem. Commun., 2012, 48, 8371-8373; (c) Y. S. Cho, K. H. Ahn, Tetrahedron Lett., 2010, 51, 3852-3854; (d) S. Zhang, J. Geng, W. Yang and X. Zhang, RSC Adv., 2014, 4, 12596-12600.
- (a) K. Wu, Y. Gao, Z. Yu, F. Yu, J. Jiang, J. Guo and Y. Han, Anal. Methods, 2014, 6, 3560-3563; (b) X. Li, Y. Gong, K. Wu, S. H. Liang, J. Cao, B. Yang, Y. Hu and Y. Han, RSC Adv., 2014, 4, 36106-36109; (c) X. Li, C. Yang, K. Wu, Y. Hu, Y. Han and S. H. Liang, Theranostics, 2014, 4, 1233-1238; (d) J. Liu, K. Wu, S. Li, T. Song, Y. Han and X. Li, Dalton Trans., 2013, 42, 3854-3859; (e) J. Liu, K. Wu, X. Li, Y. Han and M. Xia, RSC Adv., 2013, 3, 8924-8928.

Published on 30 January 2015. Downloaded by Gazi Universitesi on 03/02/2015 19:05:04