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Synthesis, structural characterization, antibiotics sensing and coordination chemistry of a fluorescent 4-amino-1,8-naphthalimide Tröger's base supramolecular scaffold

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ABSTRACT

The solvothermal synthesis and structural characterization of two new supramolecular coordination polymers featuring a unique V-shaped ligand bis-[*N*-(3-pyridyl)methyl)]-9,18-methano-1,8naphthalimide-[*b*,*f*][1,5]diazocine (**TBNap**), are reported. The ligand **TBNap** was synthesized as a racemic mixture and the molecular structure was analyzed crystallographically. **TBNap** showed a positive solvatochromism effect which was further supported by DFT calculations illustrating the D- π -A type fluorophore effects within the ligand structures. **TBNap** was further employed as a potential fluorescence probe for the discriminative sensing of structurally assorted antibiotics. A strong fluorescence quenching and preferential binding affinity were realized for nitro-containing antibiotics, i.e., Furazolidone, Nitrofurantoin and Nitrofurazone. The solvothermal reaction of **TBNap** with Cd(OAc)2·2H2O afforded a one-dimensional coordination polymer (TB-CP-1, *poly*-[Cd(**TBNap**)(OAc)2]). Similarly, reaction between **TBNap** and CdCl2.4H2O in DMF-C2H5OH rendered a 4,4'-2D coordination sheet (TB-CP-2, *poly*-[CdCl2(**TBNap**)]·DMF). The extended polymeric structures are further propagated into 3D supramolecular network through multiple face-to-face π - π interaction.

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Introduction

The rational design and construction of functional supramolecular coordination polymers (SCPs) have received considerable attention and significant advancement has been made in the past decades, due to their fascinating structural features and diverse applications [1,2]. Supramolecular self-assembly is a powerful synthetic method that provides a facile route to design intricate structures and materials from simple building blocks [3–5]. In general, SCPs are readily formed through self-

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assembly of a metal ion or metal clusters with multidentate organic/hybrid bridging linkers [6,7]. The functional properties of SCPs including high surface area, strong adsorption affinity, optical and magnetic properties can be systematically tailored for specific applications by the sensible selection of precursors and tuning of the reaction conditions [8-11]. In particular, the luminescent properties of SCPs can be modulated by incorporating stimuliresponsive fluorescent organic linkers such as tetraphenyl ethylene, imidazole, pyrazole, and 4,4'-bipyridine [12–14]. Luminescent SCPs through their selective response to analytes (emission enhancement or quenching) have been utilised in the sensing of toxic organic molecules and ions [15–18]. Amongst the myriad of SCPs, those with accessible nitrogen sites have attracted special attention because of their various application in gas storage, selective adsorption, and sequestration of environmental contaminants [14,19-25].

Antibiotics are well-known medicines widely used for the successful treatment and prevention of various bacterial infections in both humans and animals [26]. Beside their use as antibacterial agents, now antibiotics are considered harmful organic pollutants because of their pernicious effect on the natural environment and living organisms. The extensive use and inappropriate discarding of antibiotics have led to the excessive accumulation of antibiotics in the ecosystem posing serious health problems as well as damage to the environment[27]. For instance, it has been found that the long-term ingestion of antibiotics into human health causes several health-related issues such as hepatotoxicity, allergic reactions, and genetic disorders including cancers [28]. Therefore, the development of suitable chemical sensors for sensitive and selective detection of antibiotics is important for the improvement of human health and removal of environmental pollutions. Recently, fluorescence techniques have become an efficient sensing method for the trace detection of analytes because of their superior sensitivity, portability, and quick response for detection. A wide variety of fluorescence sensors such as luminescent metal-organic frameworks (LMOFs), covalent-organic polymers (COPs), small-molecules, quantum dots, and supramolecular gels are developed and successfully used for the trace detection of electronically assorted antibiotics in both organic and aqueous medium[29]. Among these sensors, small-molecule based fluorescence sensors are attractive because of their facile synthesis, easy purification, and enhanced solution processibility[30].

It has been reported that the introduction of Lewis basic nitrogen open sites in the backbone of

permanently porous coordination polymers showed selective gas adsorption and enhanced uptake capacity for CO₂ owing to their basicity and polar C--N interactions [31-34]. However, the construction of SCPs with Lewis basic nitrogen open sites within the ligand skeleton is synthetically challenging due to the significant possibility of competitive binding of metal ions to these sites[12]. The design and use of synthetic constructs for the direct incorporation of noncoordinating open nitrogen-sites into SCPs are rather challenging to achieve [35,36]. Semi-rigid/flexible functionalised pyridyl ligands have been proven to be excellent bridging moieties for building up SCPs because the coordination requirements of the metal ions can be fulfilled through backbone bending of an appropriate twist angle, enabling various conformations to be adopted [37,38]. Here we used a 4-amino-1.8-naphthalimide derived Tröger's base supramolecular scaffold (**TBNap**) to assemble two nitrogen-rich SCPs with Lewis base open sites. TBNaps are fascinating chiral scaffolds comprising of a methano-1,5-diazocine ring placing two 1,8-naphthalimide moieties almost at right angles to each other giving rise to a unique V-shaped structure possessing a large internal hydrophobic cavity (Figure 1) [39-45]. Over the past several years, we have been interested in employing TBNaps as a bifunctional scaffold to generate various self-assembled structures, soft-materials, and extended polymeric networks for their application within the material and medicinal chemistry fields [46-52]. In this article we present the synthesis, structural characterisation and solvatochromism of TBNap, bis-[N-(3-pyridyl)methyl)]-9,18-methano -1,8-naphthalimide-[*b*,*f*][1,5]diazocine. Additionally, the antibiotic sensing propensity of TBNap is explained in detail. We recently have reported [52] two coordination polymers based on TBNap and in continuation, herein we report two new SCPs, poly-[Cd(**TBNap**)(OAc)₂] (**TB-CP-1**) and poly-[CdCl₂ (TBNap)]·DMF (TB-CP-2) generated via coordinationdriven self-assembly of Cd(II) metal ion and TBNap. The present study demonstrates the solvothermal synthesis, structural and thermal characterisation of TB-CP-1 and TB-CP-2.

Results and discussion

Synthesis and Structural Characterisation of TBNap. The ligand **TBNap** was synthesised from the corresponding amino derivative, 4-amino-1,8-naphthalimide (**Nap**) following the procedure reported in our recent literature [52]. The precursor **Nap** was obtained by catalytic hydrogenation of 4-nitro-1,8-naphthalic anhydride with 10%



Figure 1. Structure of dipyridyl Tröger's base linker (TBNap).

Pd/C at 3 atm of H₂ in dimethylformamide. The **Nap** was then converted into TBNap, by using 1.5 equivalent of paraformaldehyde in neat trifluoroacetic acid under an inert atmosphere over 12 hours at room temperature. The resulting reaction mixture was basified (pH > 10)using aq.NH₃, followed by extraction into dichloromethane, which gave the desired dipyridyl-TBNap as a racemic mixture in 83% yield after trituration with cold diethyl ether.[‡] The formation of **TBNap** was fully characterised by using various conventional spectroscopic techniques such as FT-IR, multinuclear [¹H and ¹³C] NMR, and HRMS [for details see the Experimental Section and Figures S1-S4 (ESI)]. The [1]H NMR spectrum confirmed the identity of TBNap by the well-separated doublet of doublets between 5.17 and 4.67 ppm corresponding to the methylene protons of the Tröger's base cleft (Figure S1, ESI). The high-resolution mass spectrometry (HRMS) analysis of **TBNap** showed one sharp peak at m/z = 643.2069 corresponding to the protonated molecular ion $[M + H]^+$ (Figure S3, ESI)[52].

The molecular structure of **TBNap** was further confirmed by using X-ray diffraction analysis. Suitable single crystals of **TBNap** were obtained by recrystallisation from a hot DMF solution after standing at room temperature for 48 hours. Crystallographic data and refinement parameters are summarised in Table S2 (ESI). The X-ray diffraction analysis of yellow plate crystals, which were isolated by filtration, provided a structural model in the triclinic P-1 space group. The diffraction data were modelled as a two-component inversion twin as presented in Figure 2A. The asymmetric unit contained two unique molecules of the ligand each with full chemical occupancy. The structural model clearly shows that the **TBNap** molecule adopts the unique bent to 'V-shaped' conformation, with a mean interplanar angle of 113.921(10)° between the two 1,8-naphthalimide ring systems. The 3-pyridyl substituents attached to the imide nitrogen atom sites face outwards from the cleft, twisted at a mean interplanar angle of 62.394(8)°, no solvent molecules or other guests were located within the lattice. The extended structure of **TBNap** features C-H···π interactions between adjacent parallel moieties as well as C-H--O interactions between 3-pyridyl C-H groups and neighbouring naphthalimide oxygen atoms (C-H-O interaction (2.5125(3) Å, C-O distance 3.0594(4) Å and C-H--O 151.557(3)°). Adjacent 1,8-naphthalimide groups are arranged into a dimer capped by 3-pyridyl moieties through offset face-toface π - π stacking interactions resulting in the head-totail arrangements of the TBNaps, as illustrated in Figure 2B.

Solvatochromism and Theoretical Study. The 4-amino-1,8-naphthalimide and its Tröger's base derivatives are found to be strongly coloured and fluorescent owing to the intramolecular charge-transfer (ICT) based excited state, which gives rise to the solvent polarity dependent large excited-state dipole [46–57]. To corroborate the effect of solvent polarity, the absorption, and



Figure 2. (A) Structure of Ligand **TBNap** as determined by X-ray crystallography. Two unique molecules exist within the unit cell. Hydrogen atoms are omitted for clarity. (B) Intermolecular interactions between adjacent moieties of **TBNap**. Two unique molecules are coloured red and blue, respectively. The orientation illustrates the packing to form a capped dimer with offset face to face π - π stacking interactions between groups.



Figure 3. (A) UV-visible absorption and (B) normalised fluorescence emission spectra of **TBNap** in different polar solvents. (C) Corresponding photographs were taken in room light and under UV light.[52].

the fluorescence emission properties of TBNap was assessed in different solvents of varying polarity. The corresponding photophysical data are summarised in Table S1 (ESI). **TBNap** was fully soluble in CH₂Cl₂, THF, DMF, and DMSO, but sparingly soluble in toluene and CH₃CN and insoluble in H₂O. The electronic absorption spectra of **TBNap** in both polar and non-polar solvents showed a high-energy band at 345-348 nm which is ascribed to the π - π * transition and a low energy transition at 385–391 nm which is assigned to the ICT transition (Figure 3A). The absorption spectra of TBNap showed almost negligible changes in the absorption maxima with increasing polarity of medium and thus the effect of solvent polarity in the ground state was inconsequential. In contrast, the emission spectra of **TBNap** displayed a positive solvatochromism, where the emission maxima corresponding to the ICT transition was significantly red-shifted upon increasing solvent polarity (Figure 3B). This agrees with the nature of the excited state, for example, in the low-polar solvent toluene (P = 2.4) a strong blue-emission band observed, with λ_{max} at 469 nm, while in a polar solvent such as DMSO (P = 7.2) a yellow-green emission was observed with λ_{max} at 524 nm. Notably, a significant increase in Stokes shift from 83 to 133 nm was also observed along with noticeable spectral broadening upon increasing the solvent polarity. These results confirm that the electronically excited state of TBNap is more stabilised in highly polar solvents [52,58–62]. As is depicted in Figure 3C, the observed bathochromic shift was also clearly visible to the naked eye; a feature we have previously capitalised on for the discriminative detection of volatile organic pollutants[63].

To further validate the experimentally observed ICT transition, we performed theoretical calculations using the density functional theory (DFT) approach on **TBNap** structure to obtain the energy minimised structure and frontier molecular orbital diagrams. The DFT calculations



Figure 4. Frontier molecular orbitals (HOMO and LUMO) of TBNap.

were carried out at the M062X/6-31 G(d,p) [64] level using the Gaussian 09 program[65]. It revealed that the highest occupied molecular orbital (HOMO) (-7.59 eV) was primarily distributed on the Tröger's base (TB) unit itself (acting as a donor), while the lowest unoccupied molecular orbital (LUMO) (-1.79 eV) was more localised on the **Nap** unit (acting as an acceptor), as demonstrated in Figure 4. These results show that **TBNap** is a typical D- π -A type fluorophore and hence, strongly supports the 'push-pull' based ICT transition from the electron-



Figure 5. (A) The changes in the fluorescence emission spectra of **TBNap** upon the addition of different antibiotics in DMSO medium; (B) Competitive binding affinity of **TBNap** towards different antibiotics in the absence (red) and the presence (blue) of FZD antibiotic.

donating TB unit to the electron-accepting imide moiety [53–62]. Moreover, the structural features of the optimised geometry of **TBNap** concur well with the structure obtained from the X-ray diffraction analysis above.

Fluorescence Sensing of Antibiotics.

Inspired by its excellent photophysical characteristics, herein we have used TBNap as a small-molecular fluorescence probe for the detection of antibiotics. The following antibiotics were chosen for the fluorescence sensing studies: Nitrofurantoin (NFT), Nitrofurazone (NFZ), Sulphadiazine (SDZ), Sulphamethazine (SMZ), Chloramphenicol (CRP), Furazolidone (FZD), Dimetridazole (DMZ). To explore the selectivity of TBNap for antibiotics detection, we first measured the emission intensity of **TBNap** in the presence of different antibiotics listed above in the DMSO medium. As shown in Figure 5A, TBNap showed discriminative fluorescence sensing responses for different antibiotics. The nitrocontaining antibiotics such as FZD, NFT, and NFZ exhibited the highest fluorescence quenching due to their electron-deficient nature enhanced by the electronwithdrawing NO₂ groups. In sharp contrast, electronrich antibiotics such as SMZ, CRP, and SDZ displayed little enhancement in emission intensity. These studies confirmed the high selectivity of **TBNap** towards nitrocontaining antibiotics. To further corroborate the high selectivity of **TBNap**, competitive fluorescence studies using FZD as a representative example were performed in the presence of other interfering antibiotics. As illustrated in Figure 5B, where the initial emission intensity (red bar) of **TBNap** was moderately quenched (green bar) upon the addition of other interfering antibiotics. However, the subsequent mixing of FZD elicited a significant fluorescence quenching (blue bar) further confirming the preferential binding affinity of **TBNap** for nitro-containing antibiotics.

To further determine the sensing propensity of **TBNaps** for antibiotic detections, a fluorescence titration experiment was performed using FZD. As shown in Figure 6, the initial emission intensity of **TBNap** at $\lambda_{max} = 524$ nm was quenched significantly upon the addition of FZD in increasing concentrations. No other



Figure 6. The extend of fluorescence quenching (left) of **TBNap** upon the addition of increasing concentration of FZD antibiotics and its corresponding Stern-Volmer plot (right).

remarkable spectral changes, such as a shift in λ_{max} , are observed. The change in emission intensity of **TBNap** was analysed by fitting the titration data to the well-known Stern-Volmer equation:

 $I_0/I = 1 + K_{SV}[Q]$

where, I_0 and I are the fluorescence intensity before and after the addition of analyte, respectively, [Q] is the molar concentration of analytes, and K_{SV} is the Stern-Volmer binding constant. From the fluorescence titration profile, a linear Stern-Volmer plot (Figure 6) was obtained and binding constant K_{SV} was determined as 3.42×10^4 M⁻¹, which is comparable to other reported fluorescence sensors for FZD detection [66] and the high K_{SV} value demonstrates preferential binding affinity of **TBNap** for antibiotics. The significant spectral overlap of the absorption spectrum of analyte FZD with emission spectra of sensor TBNap supports that an excited state energy transfer mechanism operates between **TBNap** and FZD (Figure S5, ESI). Therefore, the observed substantial fluorescence quenching of TBNap in presence of FZD is attributed to the efficient charge-transfer (CT) complex formation between electron-rich TBNap and electron-deficient FZD.

To investigate the fluorescence quenching response time to sense FZD by **TBNap**, we then performed a timedependent fluorescence titration study. The emission intensity of **TBNap** was monitored at different timeintervals and quenching efficiency was plotted against exposure time as shown in Figure S6 (ESI), which demonstrates a fast fluorescence sensing response for FZD detection. For instance, approximately 50% of quenching efficiency was observed within 60 s of exposure time and thus, **TBNap** can be considered a quick sensor to detect antibiotics. To determine the sensitivity of **TBNap**, a similar fluorescence titration was performed taking FZD at an extremely low concentration of FZD. The limit of detection (LOD) was estimated by using the following equation:

Limit of detection = $3\sigma/K$

Where σ is the standard deviation of the initial emission intensity of **TBNap** and *K* is the slope of the linear calibration curve. As depicted in Figure S7 (ESI), a plot of change in emission intensity as a function of FZD concentration gave a linear curve with slope $K = 13.92 \times 10^6$. The quantitative analysis of fluorescence titration data fitting into the above equation revealed that **TBNap** can indicate the presence of FZD as low as 6.82 ppm. These studies demonstrate the potential of **TBNap** for the fast and sensitive detection of antibiotics in solution.

Synthesis and structure of the *poly*-[Cd(TBNap)(OAc)₂] TB-CP-1. Having synthesised and structurally characterised, we subsequently explored the coordination chemistry of TBNap in the synthesis of two new supramolecular coordination polymers TB-CP-1 and TB-CP-2. These coordination polymers were each formed from the solvothermal reaction of **TBNap** and the appropriate Cd(II) metal salt in a 1:2 molar ratio in DMF or DMF-C₂H₅OH solvent mixtures as specified below. The solid-state structure of these polymers was analysed through single-crystal X-ray diffraction. The chemical composition and purity of the complexes were determined using elemental analysis and powder X-ray diffraction analysis, respectively. The reaction of **TBNap** and Cd(OAc)₂•2H₂O· in a 1:2 stoichiometry in DMF afforded X-ray diffraction quality yellow block crystals of **TB-CP-1** after heating at 100°C for 48 h. Analysis by single-crystal X-ray diffraction provided a structural model in the monoclinic P2/c space group, revealing a polymeric structure with the formula poly-[Cd(TBNap)(OAc)₂] as shown in Figure 7A. The asymmetric unit of *poly*-[Cd(**TBNap**)(OAc)₂] contains half of one molecule of **TBNap** and a Cd(II) ion exhibiting slightly distorted octahedral six-coordinate geometry; the coordination sphere being further occupied by two equivalent pyridyl nitrogen atoms and two bidentate acetato ligands. The nitrogen and oxygen atoms coordinate at Cd-N and Cd-O distances 2.32921(15) Å and 2.30594(18) Å for N1 and O2, respectively. All Cd(II) ions are in equivalent environments. The compound exhibits a densely packed structure and there are no solvent molecules or other quest molecules located within the lattice (Figure 7B). The **TBNap** molecule within poly-[Cd(**TBNap**)(OAc)₂] adopts the expected cleft conformation between the two naphthalimide rings (112.983(6)°). The bulk structure of the complex illustrates face-to-face π - π interactions between adjacent Nap groups at a mean interplanar angle of 2.648° and minimum interatomic distance of 3.8776(3) Å for C11...C11. There are also C-H...O interactions both between the acetate ligands and nearest TBNap C-H group which exhibit a relatively short C-O distance of 2.6684(14) Å and a C-H-O angle of 148° as well as between the 3-picolyl group and the adjacent TBNap unit; the C-H-O interaction has a distance of 2.4220 (13) Å and angle 172.50(12)°. There are also weak C-H $\dots\pi$ interactions between adjacent TBNap units with 2.8211(15) Å and angle 149.5°. Thermogravimetric analysis of an air-dried sample of TB-CP-1 corresponded with no solvent being present in the unit cell of the crystal as the sample remained stable with no mass loss occurring until 300°C. Above 300°C, decomposition of the remaining organic proportion of the framework is observed (Figure S8, ESI). The X-ray powder diffraction patterns of complex **TB-CP-1** convincingly match with the simulated powder pattern at 100 K of the crystal indicating that the crystallinity of the complex was



Figure 7. (A) Structure of complex **TB-CP-1** with labelling scheme for unique heteroatoms. (B) Intermolecular interactions in the structure of **TB-CP-1** showing face to face π - π stacking interactions between adjacent complexes. Hydrogen atoms are omitted for clarity.

retained in the bulk structure (Figure S9, ESI). Additionally, Le Bail refinement of the powder diffraction data was undertaken, indicating a good agreement between the bulk room temperature data and singlecrystal calculated data, a good degree of phase purity (Figure S10, ESI).

Synthesis and Structure of poly-[CdCl₂ (TBNap)]·DMF (TB-CP-2): The reaction between **TBNap** and CdCl₂•4H₂O in a 1:2 stoichiometry ratio in DMF-EtOH (in 2:1 volume ratio) solutions afforded the diffraction quality yellow block crystals of TB-CP-2 after heating the solution at 100°C for 24 hours. The crystals were isolated by filtration after washing with fresh DMF. Analysis by single-crystal X-ray diffraction provided a structural model in the monoclinic $P2_1/c$ space group, revealing a polymeric structure with the formula poly-[CdCl₂(TBNap)]·DMF as shown in Figure 8A. The asymmetric unit contains one molecule of TBNap and a Cd(II) ion exhibiting octahedral coordination geometry. The other sites in the coordination sphere are occupied by two sets of two equivalent equatorial pyridyl nitrogen atoms and two axial chlorido ligands. The two nitrogen atoms coordinate at Cd-N distances 2.44700(8) Å and 2.40086(6) Å for N1 and N6, respectively. The TBNap molecule adopts the expected cleft conformation between the two naphthalimide rings, with a mean inter-planar angle between the naphthalene fragments of 112.4373(16)°. There are also three chemically equivalent partially occupied DMF solvent molecules within the unit cell. The DMF molecules were disordered and the most realistically modelled as chemical occupancy of 0.25 and 0.5 each. The solvent molecules in the extended structure align along the c axis perpendicular to the 2D sheet. The extended structure, as illustrated in Figure 8B, is also a 4,4'- two-dimensional coordination polymer containing offset face-to-face π - π interactions between adjacent Naps groups at a mean interplanar angle of 4.165° with the minimum interatomic distance of 3.77517(12) Å for C13--C13 as illustrated in Figure 8B. There are also anion $\dots \pi$ interactions between the naphthalimide groups and chlorido ligands (π-H···Cl distance 2.75668(9) Å) as well as weak intermolecular interactions between the complex and solvent molecules. Thermogravimetric analysis of an air-dried sample TB-



Figure 8. (A) Structure of complex **TB-CP-2** with labelling scheme for unique heteroatoms. (B) Extended structure in the structure of **TB-CP-2** illustrating the formation of a 4,4'-2D coordination polymer. Hydrogen atoms and partially occupied solvent molecules are omitted for clarity.

CP-2 showed an initial weight loss of 8% occurring in the range 30-180°C, this corresponds to the evaporation of 1 DMF molecule per unit cell. There is a more gradual loss of a further 4% below 380°C, presumably representing a mixture of mobile solvent molecules trapped within the freshly isolated framework which are lost on extended standing in the air. Above 300°C, decomposition of the remaining organic proportion of the framework was observed (Figure S11, ESI). The experimentally observed powder diffraction pattern of the complex TB-CP-2 matches well with the simulated pattern of the crystal at 100 K confirming that the phase purity is retained in the bulk compound (Figure S12, ESI). Additionally, Le Bail refinement was carried out on the powder diffraction indicating a good agreement between the structural and powder data and a good degree of phase purity (Figure S13, ESI).



Figure 9. Overlaid structures of **TBNap** and **TB-CP-1** illustrating the difference in the dimensionality of the discrete ligand structure and the coordination polymer. Hydrogens and partially occupied solvents are omitted for clarity.

Structural correlation

The structural arrangements of the two newly synthesised polymeric complexes TB-CP-1 and TB-CP-2 presented herein differ from that of the discrete TBNap ligand structure. The complex poly-[Cd(TBNap)(OAc)₂] forms a one-dimensional coordination polymer whereas poly-[CdCl₂(TBNap)]·DMF forms a two dimensional 4,4' sheet. Furthermore, in both the complexes, upon the coordination of pyridyl nitrogen to the Cd(II) metal ion, there is a significant twisting component in the folding of the equivalent pyridyl groups compared to the **TBNap** ligand structure. In **TBNap**, the plane fold angle is 61.090(8)° and twist angle is 15.646(6)°, whereas in poly-[CdCl₂(**TBNap**)]·DMF the plane fold angle is 72.286(2)° and plane twist angle is 30.787(4)°; in poly-[Cd(**TBNap**)(OAc)₂] the plane fold angle is 8.146(5)° and twist angle is 125.380(8)°. These distinct structural differences are clearly illustrated in overlaid structures given in Figure 9 and Figure S14 (ESI). Moreover, there are a few meaningful conclusions that can be drawn between comparing these structures with those we have previously reported of **TBNap** with CoCl₂ and Cd(NO₃)₂•4H₂O[52]. The solvothermal reaction of **TBNap** and CoCl₂ in DMF yielded a one-dimensional extended polymeric structure in which the ligand TBNap adopts an expected cleft shape confirmation with a mean inter-planar angle of 81.1° between the naphthalene moieties. The two pyridyl groups of TBNap coordinating to Co(II) are twisted inwards towards the cleft and this endowed a zig-zag nature to the polymeric network. Similarly, the reaction of TBNap and Cd(NO₃)₂•4H₂O resulted in a one-dimensional polymeric structure consisting of two coaxial zig-zag chains with structurally unique **TBNap** in a more compact cleft conformation. These studies demonstrate that by judicious selection of reacting components and tuning the reaction conditions, several structurally unique supramolecular coordination polymers with interesting functional properties can be generated. We are in the process of investigating these possibilities further.

Conclusions

In summary, we have synthesised two new Cd(II) supramolecular coordination polymers

poly-[Cd(**TBNap**)(OAc)₂] (**TB-CP-1**) and poly-[CdCl₂ (**TBNap**)]·DMF (**TB-CP-2**) from a close to 'V-shaped' fluorescent naphthalimide derived Tröger's base ligand and structurally characterised by using X-ray diffraction analysis. The solid-state structural analysis of the ligand **TBNap** showed that the structure had a mean interplanar angle of 113.921(10)° between the two **Nap** ring systems, which is a somewhat larger angle than we have seen for some other TBNap structures, the solution studies demonstrated that TBNap displayed a positive solvatochromism, due to the 'push-pull' based ICT transition, in different solvents with varying polarity.[‡] The highly emissive **TBNap** was used as a promising fluorescence probe for antibiotic sensing in solution. The fluorescence titration studies demonstrated the preferential and strong binding affinity of TBNap towards nitrocontaining antibiotics and detect antibiotics at ppmlevel. The observed strong fluorescence quenching is ascribed to the excited energy transfer from TBNap to antibiotics. The coordination polymers both TB-CP-1 and TB-CP-2 were not realised for antibiotics sensing owing to their poor stability in solution and they were found to decompose in the DMSO solution. The coordination properties of TBNap were analysed in the solidstate. The crystallographic analysis revealed that **TB-CP** -1 is a 1D coordination polymer exhibiting a densely packed extended structure with no solvent or guest molecules located within the crystal lattice. All Cd(II) ions are in an equivalent environment exhibiting distorted octahedral geometry occupied by two equivalent pyridyl nitrogen atoms and two bidentate acetato ligands. The structural analysis unveiled that TB-CP-2 is a 4,4'-2D coordination polymer network containing three partially occupied DMF solvent molecules within the lattice. The octahedral Cd(II) coordination geometry is occupied by two sets of two equivalent equatorial pyridyl nitrogen atoms and two axial chlorido ligands. In each case, ligand TBNap adopts the expected cleft conformation between the two naphthalimide rings and substantial face-to-face π - π interactions between the adjacent chains leading to the structurally interesting supramolecular coordination networks. These results are of major importance in demonstrating the versatility of TBNap in fluorescence sensing and constructing coordination polymers with nitrogen-rich open sites for application in energy and environmental-related areas, and such investigations are currently underway in our laboratories.

Experimental section

Materials and General Methods. All reagents, solvents, and starting materials were purchased from various commercial sources, were of reagent grade, and were used as received. Solvents used were HPLC grade unless otherwise stated. 4-Nitro-1,8-naphthalic anhydride, 3-picolylamine, palladium on carbon (10 wt% loading), paraformaldehyde, trifluoroacetic acid (TFA), antibiotics, and all-metal salts used to make polymers were purchased from Sigma-Aldrich and used as received. The melting point was determined using an Electrochemical IA9000 digital melting point apparatus in an unsealed capillary tube.

the procedure reported in the literature [52,67].

The elemental analysis for C, H, and N was performed on an Exeter analytical CE-450 elemental analyser.

FT-IR spectra were recorded in the range 4000-550 cm⁻¹ on a Perkin-Elmer spectrometer equipped with a universal ATR sampling accessory.

All NMR spectra were recorded on a Bruker-DPX-Avance spectrometer operating at 600 MHz for [1]H NMR and 150 MHz for [13]C NMR in the commercially available deuterated solvent. Chemical shifts are reported in parts-per-million (ppm) relative to the internal solvent (CD_3)₂SO = 2.5 ppm signal. All NMR data were processed with Bruker Win-NMR 5.0, Topspin, and MestReNova software. Multiplicities were abbreviated as follows: singlet (s), doublet (d), doublet of doublet (dd), triplet (t), multiplet (m).

APCI-ESI mass spectra were acquired on a Bruker microTOF-Q III spectrometer interfaced with a Dionex UltiMate 3000 LC or direct insertion probe. The instrument was operated in the positive or negative mode as required. Agilent tuning mix APCI-TOF was used to calibrate the system. The m/z values were recorded over a range of 100–1600. MicroTof control and HyStar software were used to carry out the analysis. HPLC-grade CH₃CN or CH₃OH or DMSO were used as carrier solvents.

Thermogravimetric analysis (TGA) of the complexes was performed on an analyser equipped with an ultramicro balance with a sensitivity of 0.1 μ g. The temperature range was from 25°C to 800°C with a scan rate of 10° C/min under N₂ purge.

X-ray powder diffraction patterns were collected using a Bruker D2 Phaser instrument with Cu Ka radiation ($\lambda = 1.5418$ Å). Samples were ground and mounted on silicon sample holders, and data were collected in the 20 range 5 – 55° at room temperature. The patterns were compared with those simulated from the single-crystal data collected at 100 K.

UV-visible absorption spectra were recorded in 1 cm quartz cuvettes (Hellma) either on a Varian Cary 50 spectrometer or recorded using a Thermo Scientific evolution 201 spectrometers. Baseline correction was applied for all spectra.

Emission spectra were recorded either on a Varian Cary Eclipse Fluorimeter or recorded using the Perkin Elmer-6500 Fluorimeter. The temperature was kept constant throughout the measurements at 298 K by using a thermostated unit block.

Synthetic procedure

Bis-[N-(3-pyridyl)methyl)]-9,18-methano

-1,8-naphthalimide-[*b*,*f*][1,5]diazocine (TBNap)[52]. Compound Nap (200 mg, 0.66 mmol, 1 equiv.) and paraformaldehyde (30 mg, 0.99 mmol, 1.5 equiv.) were flushed with argon. Trifluoroacetic acid (4 mL) was added at 0°C and the solution was stirred at room temperature for 12 h under an inert atmosphere. The reaction mixture was then added dropwise to aqueous ammonium hydroxide (100 mL) at 0°C until a pH > 10was achieved. Dichloromethane $(2 \times 200 \text{ mL})$ was added, and the organic layer was extracted and washed with saturated NaHCO₃ (2 \times 100 mL), brine (2 \times 100 mL) and H_2O (2 \times 100 mL). The solution was dried over MgSO₄ and the solvents were removed under reduced pressure to isolate compound **TBNap** (186 mg, 0.27 mmol, 83%) as a bright yellow solid after trituration with cold-diethyl ether. Anal. Calcd (%) for $C_{39}H_{26}N_6O_4 \cdot 0.4CH_2Cl_2$: C, 69.94; H, 3.99; N, 12.42: Found C, 70.21; H, 3.91; N, 12.53. Melting point 283-285°C (decomp.). HRMS (APCI) m/z: calcd for $C_{39}H_{27}N_6O_4$ [M + H⁺] 643.2094, found 643.2069; [1]H NMR (600 MHz, (CD₃)₂SO) δ 8.75--8.73 (2 H, d, J = 8.9 Hz, Ar-H), 8.53 (2 H, s, Pyridyl-H), 8.49-8.48 (2 H, d, J = 7.3 Hz, Ar-H), 8.40-8.39 (2 H, d, J = 6.2 Hz, Pyridyl-H), 8.13 (2 H, s, Ar-H), 8.00–7.93 (2 H, m, Ar-H), 7.64–7.62 (2 H, d, J = 8.0 Hz, Pyridyl-H), 7.26–7.23 (2 H, dd, J = 7.9, 4.8 Hz, Pyridyl-H), 5.17 (4 H, s, Pyridyl-CH₂), 5.18–5.15 (2 H, d, J = 17.4 Hz, NCH₂), 4.72 (2 H, s, NCH₂), 4.68–4.65 (2 H, d, J = 17.4 Hz, NCH₂); [13]C NMR (150 MHz, (CD₃)₂SO) δ 163.97, 163.39, 149.82, 149.53, 148.73, 135.65, 133.45, 131.48, 130.97, 129.87, 128.05, 127.68, 127.22, 126.71, 123.93, 122.76, 117.94, 66.52, 57.26, 4100; FT-IR v_{max} (ATR, cm⁻[1]) 3038, 2156, 1971, 1702, 1654, 1624, 1594, 1581, 1517, 1478, 1460, 1428, 1405, 1378, 1335, 1319, 1231, 1180, 1168, 1120, 1098, 1027, 975, 934, 869, 828, 786, 761, 732, 711, 674, 630, 613,586.

poly-[Cd(TBNap)(OAc)₂] (TB-CP-1). Compound TBNap (20 mg, 31.1 µmol) and cadmium(II) acetate dihydrate (16.6 mg, 62.2 µmol) were combined in 3 mL of DMF. The reaction mixture was briefly sonicated, sealed in a Teflon-capped vial and heated at 100°C for 48 h. Yellow crystals were isolated by filtration, washed with fresh DMF (2 × 2 mL) and dried in air. Yield: 12 mg (60%). m.p. > 300°C. Anal. Calcd (%) for C₄₃H₃₂N₆O₈Cd: C, 59.15; H, 3.69; N, 9.62: Found C, 57.66; H, 3.88; N, 9.38. FT-IR v_{max} (ATR, cm⁻[1]) 2982, 1696, 1657, 1619, 1596, 1570, 1546, 1510, 1482, 1458, 1421, 1401, 1376, 1352, 1333, 1321, 1297, 1254, 1234, 1196, 1166, 1126, 1086, 1050, 1030, 1010, 959, 933, 911, 847, 836, 806,785, 756, 708, 676, 663, 645, 622, 581, 572, 554.

poly-[CdCl₂(TBNap)]-DMF (TB-CP-2). Compound TBNap (20 mg, 31.1 µmol) and cadmium(II) chloride (11.4 mg, 62.2 µmol) were combined in a mixture of DMF-EtOH (3 mL in 2:1 volume ratio). The mixture was briefly sonicated, sealed in a Teflon-capped vial, and heated at 100°C for 24 h. Yellow crystals were isolated by filtration, washed with fresh DMF (2 × 2 mL) and dried in air. Yield: 8.5 mg (43%). m.p. > 300°C. Anal. Calcd (%) for C₃₉H₂₆Cl₂N₆ O₄Cd·DMF: C, 52.16; H, 3.82; N, 9.36: Found C, 53.94; H, 4.14; N, 9.09. FT-IR v_{max} (ATR, cm⁻[1]) 2978, 1695, 1650, 1619, 1595, 1569, 1510, 1479, 1458, 1434, 1403, 1373, 1333, 1318, 1301, 1256, 1232, 1169, 1152, 1129, 1105, 1089, 1050, 1023, 1010, 961, 912, 870, 835, 784, 756, 705, 661, 641, 621, 581.

Crystallography

Structural and refinement parameters are presented in Table S2. All diffraction data were measured using a Bruker APEX-II Duo dual-source instrument using graphite-monochromated Mo Ka ($\lambda = 0.71073$ Å) as specified. Datasets were collected using ω and ϕ scans with the samples immersed in NVH immersion oil and maintained at a constant temperature of 100 K using a Cobra cryostream. The data were reduced and processed using the Bruker APEX-3 suite of programs [68] Multi-scan absorption corrections were applied using SADABS[69]. The diffraction data were solved using SHELXT and refined by full-matrix least-squares procedures using SHELXL-2015 within the OLEX-2 GUI [70,71]. All nonhydrogen atoms were refined with anisotropic displacement parameters. All carbon-bound hydrogen atoms were placed in calculated positions and refined with a riding model, with isotropic displacement parameters equal to either 1.2 or 1.5 times the isotropic equivalent of their carrier atoms[72]. Crystals of ligand TBNap were processed as a single component, the data was processed with PLATON TwinRotMot[72]. The final refinement was carried out on the merged HKLF 5 file, twin law HKLF 5 BASF [0.243(4)]. The R_{int} value for the individual domain was 6.87. Crystals of poly-[Cd(TBNap(OAc)₂] (TB-**CP-1**) were twinned following the twin law HKLF 5 BASF [0.1494(7)]. The data was processed with TWINABS and the final refinement was carried out on, the merged HKLF 5 files[73]. The R_{int} value for the individual domain was 0.0283. The data was rotated from the first domain by 179.2 degrees about reciprocal axis -0.157 1.000 - 0.016 and real axis -0.079 1.000 - 0.013. The Twin law used to convert hkl from first to this domain (SHELXL TWIN matrix): $-0.978\ -\ 0.312\ -\ 0.006\ -\ 0.155\ 0.975\ -\ 0.026$ 0.024 - 0.028 - 0.997. Crystals of poly-[CdCl₂ (**TBNap**)]-DMF (**TB-CP-2**) provided good quality diffraction data and the model was refined to convergence with only minimal restraints, mostly involving the partially occupied solvent molecules. Particular refinement strategies for each structure, including the specific use of restraints, are provided in the combined crystallographic information files. CCDC 1864799 – 1864801.

Associated content

Supporting Information.

Spectroscopic [¹H, 13C NMR, FTIR, and APCI-MS] characterisation, TGA, powder diffraction, antibiotics sensing, and crystallographic refinement data are available.

Accession codes

CCDC 1,864,799 (**TBNap**), 1864800 (**TB-CP-1**) and 1864801 (**TB-CP-2**) contains the supplementary crystal-lographic data for this paper.

Foot note

‡While in the current study we used **TBNap** in its racemic form, we have previously demonstrated that such structures can be resolved[49]. While not reported herein [53–57], the solvent effect for the **Nap** itself is generally larger than seen for the **TBNaps** structures as the lone pair of the nitrogen is more delocalised with the ring structures. This can also be seen in the ground and the excited states, as λ_{max} occurs at longer wavelengths.

Notes

Any additional relevant notes should be placed here.

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Disclosure statement

No potential conflict of interest was reported by the authors.

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The manuscript was written through the contributions of all authors. All authors have approved the final version of the manuscript.

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