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Satish Koppireddi, Jayaram Reddy Komsani, Sreenivas Avula, Sujitha Pombala, Satishbabu Vasamsetti, Srigiridhar Kotamraju, Rambabu Yadla

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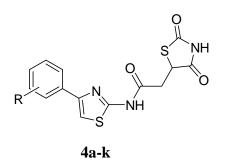
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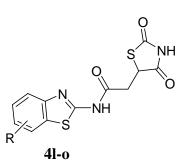
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A new series of *N*-thiazolyl-thiazolidinedione-2-acetamides are synthesized and evaluated for DPPH radical scavenging, SASA, LPI, EHI, IL-1ß and MCP-1 secretion inhibition activities. Some of them have exhibited good antioxidant and anti-inflammatory potential.

	••• •						
Anti-inflammatory activity							
	$IC_{50}(\mu M)$ values						
	<u>R</u>	<u>IL-1β</u>	<u>MCP-1</u>				
4h	o-Me	7.6	18.7				
4m	6-NO ₂	6.8	20.2				
4n	6-OMe	7.0	39.6				

Novel 2-(2,4-dioxo-1,3-thiazolidin-5-yl)acetamides as antioxidant and/or antiinflammatory compounds

Satish Koppireddi^a, Jayaram Reddy Komsani^a, Sreenivas Avula^a, Sujitha Pombala^b, Satishbabu Vasamsetti^b, Srigiridhar Kotamraju^b, Rambabu Yadla^{a,*}

^aFluoroorganics Division, CSIR-Indian Institute of Chemical Technology, Hyderabad-500 607, India; ^bChemical Biology Division, CSIR- Indian Institute of Chemical Technology, Hyderabad.

*Corresponding author. Tel.: +91-40-27193171; Fax: +91-40-27193185.

E-mail address: ryadla@yahoo.com (Rambabu Yadla).

ABSTRACT

A series of novel *N*-(4-aryl-1,3-thiazol-2-yl)-2-(2,4-dioxo-1,3-thiazolidin-5-yl)acetamides (4a-k) and *N*-(1,3-benzothiazol-2-yl)-2-(2,4-dioxo-1,3-thiazolidin-5-yl)acetamide derivatives (4l-o) are synthesized and evaluated for their anti-inflammatory and antioxidant activity (DPPH radical scavenging, superoxide anion scavenging, lipid peroxide inhibition, erythrocyte hemolytic inhibition). The structure of the compound 4j is confirmed by X-ray crystallography. Compounds 4k and 4l have exhibited good antioxidant activity in four assays, while compounds 4c, 4d, 4m, 4n and 4o have shown good DPPH radical scavenging efficacy. Compounds 4a, 4h, 4i, 4k, 4m and 4n have possessed excellent anti-inflammatory activity. *N*-[4-(o-Methoxyphenyl)-1,3-thiazol-2-yl]-2-(2,4-dioxo-1,3-thiazolidin-5-yl)acetamide (4k) and *N*-(6-nitro-/methoxy-1,3-benzothiazol-2-yl)-2-(2,4-dioxo-1,3-thiazolidin-5-yl)acetamide (4m and 4n) have exhibited both antioxidant anti-inflammatory activities.

Keywords:

Anti-inflammatory activity; Lipid peroxidation; Erythrocyte hemolysis; Superoxide anion and DPPH radical scavenging; *N-(1,3-Benzothiazol-2-yl)-2-(2,4-dioxo-1,3-thiazolidin-5-yl)acetamide*; *N-(4-Aryl-1,3-thiazol-2-yl)-2-(2,4-dioxo-1,3-thiazolidin-5-yl)acetamide*.

1. Introduction

Antioxidant therapies are gaining importance due to their ability to retard disease progression by reducing the damage caused by free radical oxidative stress in a patient [1]. Physiological levels of reactive oxygen species (ROS) play vital role as signaling molecules to mediate numerous biological functions causing alterations in cell growth, gene expression and host defense [2]. Under inflammatory conditions, presence of excess reactive oxygen species $(O_2^{-}, OH, H_2O_2, NO, ONOO^-)$ can initiate damage to nucleic acids, proteins, carbohydrates and lipids in many types of cells including macrophages [3]. Increased oxidative stress induced production of ROS, overwhelming the antioxidant defense system, has been implicated in the pathogenesis of various disorders including atherosclerosis [4,5], cancer [6,7], asthma [8], rheumatoid arthritis [9], ischemia-reperfusion injury [10], neurodegenerative diseases [11,12], inflammation [13], myocardial infarction and also aging [14,15].

Inflammation, on the other hand, is a multifactorial process. In response to infection stimulus, monocytes/ macrophage lineage cells activate and generate an inflammatory environment by secreting many pro-inflammatory cytokines. Among the cytokines, IL-1 β is a multipotent, proinflammatory and procoagulant cytokine affecting most cell types causing alzheimer's [16], rheumatoid arthritis [17], atherosclerosis [18], diabetes, etc. The onset of diabetes is believed to open a gateway to several other complications associated with increased oxidative stress and elevated inflammatory processes. Monocyte chemoattractant protein-1 (MCP-1) is a key regulator of monocyte recruitment to sites of inflammation and plays an important role in initiating local inflammation at the damaged site. MCP-1 levels are known to be elevated in various disorders including cardiovascular and neurodegeneration [19, 20]. In addition, MCP-1 induces several proinflammatory changes including secretion of cytokines and expression of adhesion molecules like intracellular adhesion molecule (ICAM), Vascular cell adhesion molecule (VCAM) etc., which in turn facilitates the binding of immune cells to endothelial cells and may promote an inflammatory environment within the vasculature [19]. It is desirable to develop small molecules containing two or three biologically active scaffolds possessing good antioxidant, anti-inflammatory and/or anti-diabetic activities [21,22].

Anti-diabetic drugs and drug candidates such as ciglitazones [23,24], englitazone [25], pioglitazone [26], rosiglitazone [27] and KRP-297 [28] contain thiazolidinedione scaffold. These compounds have been reported to be acting as ligands for peroxisome proliferator-activated

receptors (PPAR) [29]. Many compounds belonging to this class are believed to be potential anti-inflammatory agents [30]. Pyrazolyl-2,4-thiazolidinediones are reported to exhibit anti-inflammatory activity [31]. However, the exact mechanism of their action is not completely understood. Similarly, thiazole ring has been identified as a central feature of myriad natural products and their synthetic analogues have been pursued by medicinal chemistry researchers [32]. 2-Amino-1,3-thiazole based amide compounds have been reported as potential radical scavengers [33] and P53 inactivators [34]. 2-Amino-5-methyl-1,3-thiazole scaffold is present as an amide in the nonsteroidal anti-inflammatory drug, meloxicam [35]. By incorporating both these pharmacophores into a single molecule, we intend to synthesize a new series of 2-(2,4-dioxo-1,3-thiazolidin-5-yl)acetamides of 2-aminothiazoles and 2-aminobenzimidazoles having antioxidant and/or anti-inflammatory properties.

2 Results and Discussion

2.1. Chemistry

2-(2,4-Dioxo-1,3-thiazolidin-5-yl)acetic acid (1) is obtained in good yield by refluxing maleic anhydride and thiourea in concentrated hydrochloric acid for 5 h as reported earlier [36]. Iodine catalyzed condensation-cyclisation of thiourea with the respective acetophenone (2a-k) at 100°C for 8 h [37] has resulted in the formation of the corresponding 4-aryl-1,3-thiazol-2-amine (3a-k) in good yield. The target molecules viz., N-(4-aryl-1,3-thiazol-2-yl)-2-(2,4-dioxo-1,3thiazolidin-5-yl)acetamides 4a-k are synthesized in 65-71% yield by treating a dry dichloromethane (DCM) solution of 2-(2,4-dioxo-1,3-thiazolidin-5-yl)acetic acid (1), Nhydroxybenzotriazole (HOBT) and O-(1H-benzotriazol-1-yl)-N,N,N',N'-tetramethyluronium hexafluorophosphate (HBTU) with the corresponding 4-aryl-1,3-thiazol-2-amine (3a-k) and N,N-diisopropylethylamine (DIEA) in dry dimethylformamide (DMF) under stirring for 12 h. In a similar reaction, 2-(2,4-dioxo-1,3-thiazolidin-5-yl)acetic acid (1) is initially activated with HOBT and HBTU in dry DCM and then reacted in situ with commercially available 2aminobenzothiazoles 31-o in dry DMF in presence of DIEA for 12 h to obtain N-(1,3benzothiazol-2-yl)-2-(2,4-dioxo-1,3-thiazolidin-5-yl)acetamide derivatives 41-o in 54-61% yield. The detailed reaction sequence is depicted in Scheme 1. The thiazolidinedioneacetic acid and thiazolamine/ benzothiazolamine scaffolds present in the target molecules 4a-o are attached through an amide linkage.

> Scheme 1 <

The newly synthesized 2-(2,4-dioxo-1,3-thiazolidin-5-yl)acetamide derivatives 4a-o are well characterized by IR, ¹H NMR, ¹³C NMR and Mass spectroscopy including HRMS. The IR spectra of these compounds consisted of absorption bands ranging from 1660 - 1760 cm⁻¹ due to secondary amide C=O stretching, 3140 - 3450 cm⁻¹ of N-H stretching of amide group and at 3040 - 3130 cm⁻¹ due to the aromatic C-H stretching frequency. The presence of C-S bond stretching absorption is seen in the range 624 - 757 cm⁻¹. The ¹H NMR spectra of all the compounds showed signals for aromatic protons between δ 7.0 - 8.7 ppm. The two N-H protons appeared as two singlets around δ 11.5 and 12.5 ppm and are D₂O exchangeable. The thiazole ring proton signal appeared as a singlet at δ 6.9 - 7.9 ppm. Interestingly, the active methylene protons of acetamide compounds 4a-o are diastereotropic and their signals appeared as ABX pattern. The CHAHBHX pattern for all the thiazolidinedioneacetic acid derivatives is seen as doublet of doublet around δ 3.0 - 3.5 ppm and δ 4.6 ppm with coupling constants, J_{AB} = 16.8 - 17.9 H_Z, J_{AX} = 7.9 - 10.1 H_Z, J_{BX} = 3.1 - 4.1 H_Z. High coupling constant of J_{AB} compromises with the data of Takahashi [38]. The molecular formula and elemental composition are confirmed by high resolution mass spectral analysis. The structural assignment of these compounds is further supported by ¹³C NMR data. The physical characteristics of individual compounds, **4a-o** is presented in Table 1.

> Table 1 <

2.2. Crystal structure of compound 4j

In addition to the spectral characterization, a single crystal of compound **4j** is obtained from ethyl acetate/hexane (80:20). The crystal structure of compound **4j** has been determined by X-ray crystallography. The ORTEP view of compound **4j** is displayed in Fig. 1. The crystal data and

> Fig. 1 <

structural refinement parameters are presented in Table 2, while selected bond lengths and bond angles are listed in Table 3.

> Table 2 <

> Table 3 <

2.3. Biology

2.3.1. Antioxidant activities

The antioxidant efficacy of the newly synthesized thiazolidinedione derivatives is assayed by applying DPPH method. Superoxide anion scavenging activity (SASA), inhibition of lipid peroxidation (LPI) and hemolysis in erythrocyte membrane stabilization or erythrocyte hemolysis inhibition (EHI) assays are performed only for selected compounds showing good radical scavenging activity in DPPH method. The sample solutions are prepared at concentrations of 60, 80, 100 µg/mL and their EC₅₀ values are obtained. Ascorbic acid and luteolin are employed as standards in these assays. The results of the antioxidant activities of these compounds are shown in Table 3. The thiazolidinedione-5-acetamides of 4-(p-4-(p-bromophenyl)-thiazol-2-amine fluorophenyl)-thiazol-2-amine (**4**c), (**4d**), 4-(omethoxyphenyl)-thiazol-2-amine (4k) and all the benzimidazol-2-amine derivatives (4l-o) have exhibited good DPPH radical scavenging potential with EC_{50} values ranging from 20 - 60 μ M as compared to ascorbic acid (40.28 µM) and luteolin (44.18 µM) as positive controls. Compounds **4k** and **4l** have also shown good LPI and EHI along with moderate SASA. Among the structural features, the presence of benzothiazole moiety (41-o) and/or NO₂ or OMe substituent at suitable position as seen in 4h, 4k, 4m and 4n seem to be desirable.

$$>$$
 Table 4 $<$

2.3.2. Anti-inflammatory activity

The thiazolidinedione derivatives **4a-o** along with pioglitazone are assayed for their antiinflammatory potential in the presence of PMA (phorbol 13-myristate 12-acetate)-induced

inflammation using THP1 monocyte cell system (Table 5). This method is conventional, sensitive and well accepted for screening of newer anti-inflammatory agents. Compounds 4b, 4d, 4f, 4g with IC₅₀ values of 16.31 μ M, 33.12 μ M, 18.23 μ M and 23.4 μ M respectively, showed reasonable anti-inflammatory activities (Table 6). Compounds 4a and 4i showed good activity with IC₅₀ values of 9.6 µM and 11.72 µM. The thiazolidine-2,4-dione-5-acetamides of 4-(otolyl)-thiazol-2-amine (4h), 4-(o-methoxyphenyl)-thiazol-2-amine (4k), 6-nitro-benzimidazol-2amine (4m) and 6-methoxy-benzimidazol-2-amine (4n) have shown excellent IL-1 β inhibitory activity with IC₅₀ values between 6.83 μ M - 7.66 μ M with respect to piroxicam (21.33 μ M) as standard. However, compounds 4c, 4e, 4j, 4l, 4o and pioglitazone (Table 6) did not show appreciable activity. In terms of MCP-1 inhibition, compounds 4a, 4d, 4e, 4g, 4n and 4o with IC_{50} values of 30.4 μ M, 49.13 μ M, 31.7 μ M, 34.24 μ M, 39.6 μ M and 63.06 μ M, respectively showed reasonable activities against MCP-1 (Table 4). Compounds 4c, 4h, 4i, 4j, 4l and 4m have shown good MCP-1 inhibition activity with IC₅₀ values between 16.68 μ M – 20.29 μ M which are well comparable to pioglitazone with IC_{50} of 18.84 μ M. MCP-1 levels are significantly attenuated in pioglitazone treatment animals compared to placebo group in stent implantation animals [39]. However, neither pioglitazone nor the thiazolidinedione compounds have shown activity on PMA induced TNF-α production. This result corresponds with the report of Thieringer, *et al.*, which says that PPAR agonists do not inhibit TNF-α production [40].

> Table 5 <

> Table 6 <

3. Conclusion

We have reported here a simple synthesis of a collection of novel 2,4-thiazolidinedione-5-acetamides of 4-aryl-1,3-thiazol-2-amines and 6-substituted-benzothiazol-2-amines. These compounds are screened *in vitro* for their DPPH radical scavenging activity along with IL-1 β and MCP-1 secretion inhibition efficacy. The compounds having DPPH radical scavenging potential are further assayed for their superoxide anion scavenging, lipid peroxidation inhibition and

erythrocyte hemolysis inhibition activities with reasonable success. Some of the compounds having methoxy- or nitro- substituent, especially on benzothiazole moiety have exhibited very good anti-inflammatory activity while showing good antioxidant potential. Further research on such molecules embedded with two biologically active scaffolds and exhibiting multiple activities may be useful in development of therapeutic applications where free radical induced inflammation plays a role in diseases.

4. Experimental Protocols

4.1. Synthetic Chemistry

All the reagents and solvents are purchased from commercial suppliers. Solvents are distilled and dried before use. Melting points are determined on the Veego (VMP-MP) melting point apparatus and are uncorrected. Merck 60 F_{254} silica gel coated glass sheets are used for thin layer chromatography. The compounds are purified by using column chromatography on silica gel (60-120 mesh) with ethyl acetate/ petroleum ether (1:2) as eluent. Infrared spectra are recorded on Perkin-Elmer FT-IR 1600 spectrometer. ¹H NMR and ¹³C NMR spectra are recorded on Bruker Avance 300 MHz and Bruker Inova 400 MHz spectrometer with TMS as internal standard. Chemical shifts (δ) are given in parts per million and coupling constants are given as absolute values expressed in Hertz. Mass spectral analysis using electrospray ionization (ESI) and high resolution mass spectrometer (QSTAR XL, Applied Biosystems/MDS Sciex, Foster City, CA, USA), equipped with an ESI source.

4.1.1. Synthesis of 2, 4-dioxo-1,3-thiazolidine-5-acetic acid (1). A mixture of thiourea (10 g, 131 mmol) and of maleic anhydride (12.8 g, 131 mmol) in 33 mL of concentrated hydrochloric acid is refluxed for 5 h and cooled to room temperature. The precipitate is separated by filtration, washed with cold water (5 mL) and crystallized from hot water to yield compound **1**. Yield 70%; mp 167-169 0 C; ¹H NMR (DMSO-d₆, 300 MHz) δ (ppm): 2.82-2.91 (dd, *J* = 17.7, 9.2 Hz, 1H, CH₂), 3.05-3.12 (dd, *J* = 17.5, 3.7 Hz, 1H, CH₂), 4.43-4.48 (m, 1H, CH), 11.95 (s, 1H, NH), 12.60 (s, 1H, COOH).

4.1.2. General procedure for synthesis of 2-amino-4-(substituted-phenyl)-1,3-thiazoles (3a-k).

A mixture of thiourea (50 mmol), the corresponding acetophenone (25 mmol) and iodine (25 mmol) is stirred at 100 0 C for 8 h. Then the reaction mixture is cooled, extracted with diethyl ether to remove excess of acetophenone, and then washed with aqueous sodium thiosulfate to remove excess iodine and later with cold water. The crude product is dissolved in hot water, filtered to remove sulphone, and the filtrate is basified with aqueous Na₂CO₃ to yield the corresponding 2-amino-4-(substituted-phenyl)-1,3-thiazole. The crude product is purified by recrystallization from alcohol.

4.1.2.1. 4-Phenyl-1,3-thiazol-2-amine (**3a**). Yield 70%; mp 146-148 °C (lit [41] mp 146-148 °C); ¹H NMR (DMSO-d₆, 300 MHz) δ (ppm): 5.13 (br s, 2H, NH₂, D₂O exchangeable), 6.66 (s, 1H, thiazole-H), 7.23-7.35 (m, 3H, Ar-H), 7.70-7.73 (m, 2H, Ar-H); ESI-MS(positive): 177.0 [M+H]⁺.

4.1.2.2. 4-(*p*-Chlorophenyl)-1,3-thiazol-2-amine (**3b**). Yield 68%; mp 164-166 °C (lit [37] mp 163-164 °C); ¹H NMR (DMSO-d₆, 300 MHz) δ (ppm): 6.23 (br s, 2H, NH₂, D₂O exchangeable), 6.70 (s, 1H, thiazole-H), 7.30 (d, *J* = 8.4 Hz, 2H, Ar-H), 7. 71 (d, *J* = 8.4 Hz, 2H, Ar-H); ESI-MS(positive): 211.0 [M+H]⁺.

4.1.2.3. 4-(*p*-*Fluorophenyl*)-1,3-thiazol-2-amine (**3***c*). Yield 69%; mp 108-110 °C (lit [42] mp 106 °C); ¹H NMR (DMSO-d₆, 300 MHz) δ (ppm): 5.94 (br s, 2H, NH₂, D₂O exchangeable), 6.60 (s, 1H, thiazole-H), 7.01-7.07 (m, 2H, Ar-H), 7.72-7.77 (m, 2H, Ar-H); ESI-MS(positive): 195.0 [M+H]⁺.

4.1.2.4. 4-(*p*-Bromophenyl)-1,3-thiazol-2-amine (**3d**). Yield 65%; mp 178-180 °C (lit [37] mp 180-181 °C); ¹H NMR (DMSO-d₆, 300 MHz) δ (ppm): 6.32 (br s, 2H, NH₂, D₂O exchangeable), 6.73 (s, 1H, thiazole-H), 7.45 (d, *J* = 8.4 Hz, 2H, Ar-H), 7. 65 (d, *J* = 8.4 Hz, 2H, Ar-H); ESI-MS(positive): 256.0 [M+H]⁺.

4.1.2.5. 4-(*p*-Iodophenyl)-1,3-thiazol-2-amine (**3e**). Yield 65%; mp 190-192 °C (lit [37] mp 176-177 °C); ¹H NMR (DMSO-d₆, 300 MHz) δ (ppm): 6.85 (s, 1H, thiazole-H), 7.44 (d, J = 8.4 Hz, 2H, Ar-H), 7.74 (d, J = 8.4 Hz, 2H, Ar-H); ESI-MS(positive): 303.0 [M+H]⁺.

4.1.2.6. 4-(o-Chlorophenyl)-1,3-thiazol-2-amine (**3***f*). Yield 66%; mp 142-144 °C (lit [43] mp 126 °C); ¹H NMR (DMSO-d₆, 300 MHz) δ (ppm): 5.15 (br s, 2H, NH₂, D₂O exchangeable), 7.03

(s, 1H, thiazole-H), 7.20-7.30 (m, 2H, Ar-H), 7.41 (d, J = 7.9 Hz, 1H, Ar-H), 7. 81 (d, J = 6.9 Hz, 1H, Ar-H); ESI-MS (positive): 211.0 [M+H]⁺.

4.1.2.7. 4-(*m*-Chlorophenyl)-1,3-thiazol-2-amine (**3g**). Yield 65%; mp 126-128 °C (lit [43] mp 112 °C); ¹H NMR (DMSO-d₆, 300 MHz) δ (ppm): 5.70 (s, 2H, NH₂, D₂O exchangeable), 6.73 (s, 1H, thiazole-H), 7.24-7.37 (m, 2H, Ar-H), 7.63 (d, *J* = 7.3 Hz, 1H, Ar-H), 7.78 (s, 1H, Ar-H); ESI-MS(positive): 211.0 [M+H]⁺.

4.1.2.8. 4-(o-Tolyl)-1,3-thiazol-2-amine (**3h**). Yield 68%; mp 81-83 °C (lit [37] mp 81-82 °C); ¹H NMR (DMSO-d₆, 300 MHz) δ (ppm): 2.43 (s, 3H, CH₃) 5.14 (br s, 2H, NH₂, D₂O exchangeable), 6.45 (s, 1H, thiazole-H), 7.18-7.25 (m, 3H, Ar-H), 7.50-7.53 (m, 1H, Ar-H);

ESI-MS(positive): 191.0 [M+H]⁺.

4.1.2.9. 4-(*m*-Tolyl)-1,3-thiazol-2-amine (**3i**). Yield 67%; mp 86-88 °C (lit [37] mp 79-92 °C); ¹H NMR (DMSO-d₆, 300 MHz) δ (ppm): 2.41 (s, 3H, CH₃) 6.86 (s, 1H, thiazole-H), 7.24 (d, J = 7.3 Hz, 1H, Ar-H), 7.32-7.45 (m, 3H, Ar-H); ESI-MS(positive): 191.0 [M+H]⁺.

4.1.2.10. 4-(*o*-Fluorophenyl)-1,3-thiazol-2-amine (**3j**). Yield 66%; mp 98-100 °C (lit [44] mp 95-96 °C); ¹H NMR (DMSO-d₆, 300 MHz) δ (ppm): 6.25 (s, 2H, NH₂, D₂O exchangeable) 6.92 (s, 1H, thiazole-H), 7.06-7.22 (m, 3H, Ar-H), 8.0-8.20 (m, 1H, Ar-H); ESI-MS(positive): 195.0 [M+H]⁺.

4.1.2.11. 4-(*o*-Methoxyphenyl)-1,3-thiazol-2-amine (**3**k). Yield 58%; mp 180-182 °C (lit [45] liquid); ¹H NMR (DMSO-d₆, 300 MHz) δ (ppm): 3.98 (s, 3H, CH₃), 6.96 (s, 1H, thiazole-H), 7.03-7.07 (m, 2H, Ar-H), 7.39-7.44 (m, 1H, Ar-H), 7.60-7.64 (m, 1H, Ar-H); ESI-MS(positive): 207.0 [M+H]⁺.

4.1.3. General procedure for the synthesis of N-(4-aryl-1,3-thiazol-2-yl)-2-(2,4-dioxo-1,3-thiazolidin-5-yl)acetamides (**4a-k**) and N-(1,3-benzothiazol-2-yl)-2-(2,4-dioxo-1,3-thiazolidin-5-yl)acetamide derivatives (**4l-o**).

The general protocol followed by us in obtaining 2-(2,4-dioxo-1,3-thiazolidin-5-yl)acetamide compounds **4a-o** is illustrated for the synthesis of N-(4-phenyl-1,3-thiazol-2-yl)-2-(2,4-dioxo-1,3-thiazolidin-5-yl)acetamide (**4a**) as an example. A solution of compound **1** (0.4 g, 2.55 mmol) in DCM (10 mL) is cooled to 0 °C and then charged with HOBT (0.36 g, 2.4 mmol), followed by HBTU (0.90 g, 2.4 mmol). The reaction mixture is stirred at 0 °C for 45 minutes.

After that, a solution of 4-phenylthiazol-2-amine, **3a** (0.29 g, 1.70 mmol) and DIEA (0.8 mL, 5.1 mmol) in a mixture of DCM (5 mL) and DMF (2.5 mL) is added drop wise over 5 minutes. The reaction temperature is initially maintained at 0 $^{\circ}$ C for 1 h and later at RT for 10 h. Completion of reaction is evidenced by TLC analysis. After evaporating the DCM solvent on rotavapor, the reaction mixture is diluted with 40 mL of distilled water and extracted with (3 x 15 mL) of ethyl acetate. The combined organic layer is washed with saturated aqueous sodium bicarbonate solution (2 x 10 mL), followed by saturated aqueous sodium chloride solution (2 x 20 mL). After drying over anhydrous sodium sulfate, the organic layer is filtered and the filtrate is stripped off the solvent. The crude product thus obtained is purified by column chromatography over silica gel. The rest of the 2-(thiazolidinedion-5-yl)acetamide derivatives (**4b-o**) are prepared similarly by reacting 2,4-dioxo-1,3-thiazolidine-5-acetic acid (**1**) with the appropriate amine (**3b-o**).

4.1.3.1. N-(4-Phenyl-1,3-thiazol-2-yl)-2-(2,4-dioxo-1,3-thiazolidin-5-yl)acetamide (4a).

IR (KBr) cm⁻¹: 3340, 3170 (NH), 3040 (Ar-H), 1750, 1692 (C=O), 1556 (C=C), 1407 (C-N), 1060; ¹H NMR (CDCl₃ + DMSO-d₆, 300 MHz) δ (ppm): 3.06-3.14 (m, 1H, CH₂), 3.38 (dd, *J* = 17.1, 3.5 Hz, 1H, CH₂), 4.56 (dd, *J* = 9.2, 3.5 Hz, 1H, CH), 7.20 (s, 1H, thiazole-H), 7.23-7.38 (m, 3H, Ar-H), 7.78 (d, *J* = 7.1 Hz, 2H, Ar-H); ¹³C NMR (CDCl₃ + DMSO-d₆, 75 MHz) δ (ppm): 37.5 (CH₂), 45.8 (CH), 106.7 (thiazole-C-5), 125.1, 127.0, 127.8, 133.7 (aromatic carbons), 149.0 (thiazole-C-4), 157.0 (thiazole-C-2), 167.0 (C=O), 171.4 (C=O), 174.0(C=O). HRMS-ESI m/z [M+H]⁺ calcd. for C₁₄H₁₂O₃N₃S₂: 334.0320; found: 334.0312.

4.1.3.2. N-[4-(p-Chlorophenyl)-1,3-thiazol-2yl]-2-(2,4-dioxo-1,3-thiazolidin-5-yl)acetamide

(*4b*). IR (KBr) cm⁻¹: 3379, 3136 (NH), 3042 (Ar-H), 1747, 1673 (C=O), 1154, 1075, 747 (C-S-C); ¹H NMR (CDCl₃ + DMSO-d₆, 300 MHz) δ (ppm): 3.15 (dd, *J* = 17.1, 8.6 Hz, 1H, CH₂), 3.38-3.40 (m, 1H, CH₂), 4.77 (dd, *J* = 8.6, 3.7 Hz, 1H, CH), 7.47 (d, *J* = 8.4 Hz, 2H, Ar-H), 7.72 (s, 1H, thiazole-H), 7.89 (d, *J* = 8.4 Hz, 2H, Ar-H); ¹³C NMR (CDCl₃ + DMSO-d₆, 75 MHz) δ (ppm): 37.1 (CH₂), 45.5 (CH), 107.0 (thiazole-C-5), 126.2, 127.6, 132.1 (aromatic carbons), 147.0 (thiazole-C-4), 156.8 (thiazole-C-2), 166.8 (C=O), 171.1(C=O), 174.4(C=O). HRMS-ESI m/z [M+H]⁺ calcd. for C₁₄H₁₁O₃N₃ClS₂: 367.9930; found: 367.9929.

4.1.3.3. *N-[4-(p-Fluorophenyl)-1,3-thiazol-2-yl]-2-(2,4-dioxo-1,3-thiazolidin-5-yl)acetamide* (*4c*). IR (KBr) cm⁻¹: 3240, 3162 (NH), 3041 (Ar-H), 1760, 1680 (C=O), 1560 (C=C), 1410 (C-N), 1090 (C-N); ¹H NMR (CDCl₃ + DMSO-d₆, 300 MHz) δ (ppm): 3.15-3.23 (m, 1H, CH₂), 3.37 (dd, J = 17.7, 3.7 Hz, 1H, CH₂), 4.59-4.62 (m, 1H, CH), 7.06-7.11 (m, 2H, Ar-H), 7.23 (s, 1H, thiazole-H), 7.81-7.85 (m, 2H, Ar-H), 11.96 (br s, 1H, NH, D₂O exchangeable), 12.37 (br s, 1H, NH, D₂O exchangeable); ¹³C NMR (CDCl₃ + DMSO-d₆, 75 MHz) δ (ppm): 35.5 (CH₂), 44.5 (CH), 105.8 (thiazole-C-5), 113.4, 113.7, 125.7, 129.0 (aromatic carbons), 146.2 (thiazole-C-4), 155.7 (thiazole-C-2), 166.2 (C=O), 170.4 (C=O), 173.7 (C=O). HRMS-ESI m/z [M+H]⁺ calcd. for C₁₄H₁₁O₃N₃FS₂: 352.0226; found: 352.0220.

4.1.3.4. N-[4-(p-Bromophenyl)-1,3-thiazol-2yl]-2-(2,4-dioxo-1,3-thiazolidin-5-yl)acetamide(4d). IR (KBr) cm⁻¹: 3371, 3140 (NH), 3043 (Ar-H), 1748, 1677 (C=O), 1155, 1065, 600 (C-S-C); ¹H NMR (CDCl₃ + DMSO-d₆, 300 MHz) δ (ppm): 3.14-3.18 (m, 1H, CH₂), 3.36 (dd, J = 17.1, 3.5 Hz, 1H, CH₂), 4.58 (dd, J = 8.6, 3.0 Hz, 1H, CH), 7.34 (s, 1H, thiazole-H), 7.49 (d, J = 8.3 Hz, 2H, Ar-H), 7.73 (d, J = 8.4 Hz, 2H, Ar-H), 11.95 (br s, 1H, NH, D₂O exchangeable), 12.39 (br s, 1H, NH, D₂O exchangeable); ¹³C NMR (CDCl₃ + DMSO-d₆, 75 MHz) δ (ppm): 36.3 (CH₂), 45.0 (CH), 106.9 (thiazole-C-5), 119.8, 126.1, 130.1, 132.1 (aromatic carbons), 146.8 (thiazole-C-4), 156.3(thiazole-C-2), 166.4 (C=O), 170.7 (C=O), 174.0 (C=O). HRMS-ESI m/z [M+H]⁺ calcd. for C₁₄H₁₁O₃N₃BrS₂: 411.9425; found: 411.9425.

4.1.3.5. *N-[4-(p-Iodophenyl)-1,3-thiazol-2-yl]-2-(2,4-dioxo-1,3-thiazolidin-5-yl)acetamide* (**4e**). IR (KBr) cm⁻¹: 3200, 3112 (NH), 3040 (Ar-H), 1747, 1680 (C=O), 1159, 1065, 625 (C-S-C); ¹H NMR (CDCl₃ + DMSO-d₆, 300 MHz) δ (ppm): 3.05 (dd, *J* = 17.1, 9.6 Hz, 1H, CH₂), 3.37 (dd, *J* = 17.3, 3.3 Hz, 1H, CH₂), 4.55-4.59 (m, 1H, CH), 7.25 (s, 1H, thiazole-H), 7.57 (d, *J* = 8.3 Hz, 2H, Ar-H), 7.68 (d, *J* = 8.4 Hz, 2H, Ar-H); ¹³C NMR (CDCl₃ + DMSO-d₆, 75 MHz) δ (ppm): 36.9 (CH₂), 45.4 (CH), 99.2(Ar-C-I), 107.2 (thiazole-C-5), 126.7, 132.9, 136.5 (aromatic carbons), 156.8 (thiazole-C-2), 169.9 (C=O), 174.4 (C=O). HRMS-ESI m/z [M+H]⁺ calcd. for C₁₄H₁₁O₃N₃IS₂: 459.9287; found: 459.9281.

4.1.3.6. N-[4-(o-Chlorophenyl)-1,3-thiazol-2yl]-2-(2,4-dioxo-1,3-thiazolidin-5-yl)acetamide(4f). IR (KBr) cm⁻¹: 3211, 3134 (NH), 3075 (Ar-H), 1756, 1701(C=O), 1138, 1039, 748 (C-S-C); ¹H NMR (CDCl₃ + DMSO-d₆, 300 MHz) δ (ppm): 3.09 (dd, J = 17.1, 9.2 Hz, 1H, CH₂), 3.40 (dd, J = 17.1, 3.7 Hz, 1H, CH₂), 4.61 (dd, J = 9.2, 3.7 Hz, 1H, CH), 7.25-7.37 (m, 2H, Ar-H) 7.44-7.47 (m, 2H, Ar-H, thiazole-H), 7.81-7.85 (m, 1H, Ar-H); ¹³C NMR (CDCl₃ + DMSO-d₆, 75 MHz) δ (ppm): 37.5 (CH₂), 45.8 (CH), 111.9 (thiazole-C-5), 126.0, 128.0, 129.6, 130.3, 131.0, 132.6 (aromatic carbons), 145.5 (thiazole-C-4), 156.0 (thiazole-C-2), 167.1 (C=O), 171.5(C=O), 174.7 (C=O). HRMS-ESI m/z $[M+H]^+$ calcd. for $C_{14}H_{11}O_3N_3ClS_2$: 367.9930; found: 367.9924.

4.1.3.7. *N-[4-(m-Chlorophenyl)-1,3-thiazol-2yl]-2-(2,4-dioxo-1,3-thiazolidin-5-yl)acetamide* (*4g*). IR (KBr) cm⁻¹: 3288, 3174 (NH), 3052 (Ar-H), 1753, 1676 (C=O), 1156, 1074, 741 (C-S-C); ¹H NMR (CDCl₃ + DMSO-d₆, 300 MHz) δ (ppm): 3.16 (dd, *J* = 17.1, 8.4 Hz, 1H, CH₂), 3.99 (dd, *J* = 14.1, 6.9 Hz, 1H, CH₂), 4.78 (dd, *J* = 8.6, 3.9 Hz, 1H, CH), 7.37 (d, *J* = 8.4 Hz, 1H, Ar-H), 7.44 (t, *J* = 7.7, 7.9 Hz, 1H, Ar-H), 7.82 (s,1H, thiazole-H), 7.85 (d, *J* = 7.7 Hz, 1H, Ar-H), 7.95 (s, 1H, Ar-H); ¹³C NMR (CDCl₃ + DMSO-d₆, 75 MHz) δ (ppm): 37.7 (CH₂), 45.9 (CH₂), 108.0 (thiazole-C-5), 123.3, 125.5, 127.0, 129.4, 133.7, 135.7 (aromatic carbons), 143.0 (thiazole-C-4), 158.9 (thiazole-C-2), 167.3 (C=O), 171.6 (C=O), 174.8 (C=O). HRMS-ESI m/z [M+H]⁺ calcd. for C₁₄H₁₁O₃N₃ClS₂: 367.9930; found: 367.9928.

4.1.3.8. *N*-[*4*-(*o*-*Tolyl*)-1,3-*thiazol*-2-*yl*]-2-(2,4-*dioxo*-1,3*thiazolidin*-5-*yl*)*acetamide* (**4h**). IR (KBr) cm⁻¹: 3291, 3155 (NH), 3054 (Ar-H), 1739, 1687 (C=O), 1167, 1064, 740 (C-S-C); ¹H NMR (CDCl₃ + DMSO-d₆, 300 MHz) δ (ppm): 2.44 (s, 3H, CH₃), 3.07(dd, *J* = 17.5, 9.3 Hz, 1H, CH₂), 3.42 (dd, *J* = 16.5, 4.1 Hz, 1H, CH₂), 4.59-4.61 (m, 1H, CH), 6.92 (s, 1H, thiazole-H), 7.19-7.21 (m, 3H, Ar-H), 7.53 (d, *J* = 7.2 Hz, 1H, Ar-H); ¹³C NMR (CDCl₃ + DMSO-d₆, 75 MHz) δ (ppm): 20.1 (CH₃), 37.1 (CH₂), 45.5 (CH), 109.5 (thiazole-C-5), 124.7, 126.8, 128.5, 129.8, 133.7, 134.7 (aromatic carbons), 148.8 (thiazole-C-4), 155.9 (thiazole-C-2), 166.7 (C=O), 171.2 (C=O), 174.4 (C=O). HRMS-ESI m/z [M+H]⁺ calcd. for C₁₅H₁₄O₃N₃S₂: 348.0477; found: 348.0466.

4.1.3.9. N-[4-(m-Tolyl)-1,3-thiazol-2-yl]-2-(2,4-dioxo-1,3-thiazolidin-5-yl)acetamide (**4i**).IR (KBr) cm⁻¹: 3275, 3154 (NH), 3050 (Ar-H), 1748, 1699 (C=O), 1161, 1091; ¹H NMR (CDCl₃ + DMSO-d₆, 300 MHz) δ (ppm): 2.39 (s, 3H, CH₃), 3.08 (dd, *J* = 17.1, 9.6 Hz, 1H, CH₂), 3.44 (dd, *J* = 17.1, 3.7 Hz, 1H, CH₂), 4.58 (dd, *J* = 9.8, 3.7 Hz, 1H, CH), 7.11 (d, *J* = 8.1 Hz, 1H, Ar-H), 7.17 (s, 1H, thiazole-H), 7.26 (t, *J* = 7.5 Hz, 1H, Ar-H), 7.59 (s, 1H, Ar-H), 7.63 (d, *J* = 6.2 Hz, 1H, Ar-H); ¹³C NMR (CDCl₃ + DMSO-d₆, 75 MHz) δ (ppm): 21.0 (CH₃), 36.9 (CH₂), 46.3 (CH), 108.0 (thiazole-C-5), 122.7, 126.2, 128.4, 128.5, 134.0, 137.7 (aromatic carbons), 148.9 (thiazole-C-4), 157.2 (thiazole-C-2), 168.1 (C=O), 172.4 (C=O), 175.7 (C=O). HRMS-ESI m/z [M+H]⁺ calcd. for C₁₅H₁₄O₃N₃S₂: 348.0477; found: 348.0470. 4.1.3.10. N-[4-(o-Fluorophenyl)-1,3-thiazol-2-yl]-2-(2,4-dioxo-1,3-thiazolidin-5-yl)acetamide(*4j*). IR (KBr) cm⁻¹: 3226, 3153 (NH), 3053 (Ar-H), 1739, 1687 (C=O), 1169, 1062, 739 (C-S-C); ¹H NMR (CDCl₃ + DMSO-d₆, 300 MHz) δ (ppm): 3.09 (dd, J = 17.1, 9.6 Hz, 1H, CH₂), 3.45 (dd, J = 17.1, 3.5 Hz, 1H, CH₂), 4.59 (dd, J = 9.8, 3.5 Hz, 1H, CH), 7.11-7.23 (m, 2H, Ar-H), 7.39 (d, J = 1.8 Hz, 1H, Ar-H), 7.56 (s, 1H, thiazole-H), 8.01 (t, J = 7.5, 7.7 Hz, 1H, Ar-H); ¹³C NMR (CDCl₃ + DMSO-d₆, 75 MHz) δ (ppm): 37.2 (CH₂), 45.6 (CH₃, 108.3 (thiazole-C-5), 111.1 (d, J = 13.7 Hz), 114.9, 115.2, 123.3, 128.0, (d, J = 8.8 Hz), 128.4 (aromatic carbons), 142.5 (thiazole-C-4), 156.0 (thiazole-C-2), 166.9 (C=O), 171.3 (C=O), 174.6 (C=O). HRMS-ESI m/z [M+H]⁺ calcd. for C₁₄H₁₁O₃N₃FS₂: 352.0226; found: 352.0220.

4.1.3.11. *N*-[4-(*o*-*Methoxyphenyl*)-1,3-thiazol-2-yl]-2-(2,4-dioxo-1,3-thiazolidin-5-yl)acetamide (*4k*). IR (KBr) cm⁻¹: 3251, 3184 (NH), 3070 (Ar-H), 1759, 1670 (C=O), 1115, 1056, 750 (C-S-C); ¹H NMR (CDCl₃ + DMSO-d₆, 400 MHz) δ (ppm): 3.09-3.14 (m, 1H, CH₂), 3.45 (dd, *J* = 16.8, 3.3 Hz, 1H, CH₂), 3.95 (s, 3H, OCH₃), 4.59 (dd, *J* = 10.1, 3.3 Hz, 1H, CH), 6.99-7.02 (m, 2H, Ar-H) 7.26 (t, *J* = 7.8 Hz, 1H, Ar-H), 7.59 (s, 1H, thiazole-H), 8.08 (d, *J* = 6.7 Hz, 1H, Ar-H) 11.8 (br s, 1H, NH, D₂O exchangeable), 12.1 (br s, 1H, NH, D₂O exchangeable); ¹³C NMR (CDCl₃ + DMSO-d₆, 75 MHz) δ (ppm): 36.9 (CH₂), 46.3 (CH₃, 55.3(OCH₃), 111.6 (thiazole-C-5), 111.8, 120.4, 122.4, 128.8, 155.7 (aromatic carbons), 144.7 (thiazole-C-4), 156.5 (thiazole-C-2), 168.0 (C=O), 172.5 (C=O), 175.7 (C=O). HRMS-ESI m/z [M+H]⁺ calcd. for C₁₅H₁₄O₄N₃S₂: 364.0426; found 364.0421.

4.1.3.12. N-(1,3-Benzothiazol-2-yl)-2-(2,4-dioxo-1,3-thiazolidin-5-yl)acetamide (41).

IR (KBr) cm⁻¹: 3192, 3110 (NH), 2924 (Ar-H), 1742, 1677 (C=O), 1182, 1069, 757 (C-S-C); ¹H NMR (CDCl₃ + DMSO-d₆, 300 MHz) δ (ppm): 3.39-3.40 (m, 1H, CH₂), 3.44-3.46 (m, 1H, CH₂), 4.62-4.67 (m, 1H, CH), 7.23 (t, *J* = 7.5 Hz, 1H, Ar-H), 7.35-7.40 (m, 1H, Ar-H), 7.68 (d, *J* = 7.9 Hz, 1H, Ar-H) 7.81 (d, *J* = 7.9 Hz, 1H, Ar-H). HRMS-ESI m/z [M+H]⁺ calcd. for C₁₂H₁₀O₃N₃S₂: 308.0164, found 308.0159.

4.1.3.13. N-(6-Nitro-1,3-benzothiazol-2-yl)-2-(2,4-dioxo-1,3-thiazolidin-5-yl)acetamide (**4m**). IR (KBr) cm⁻¹: 3275, 3193 (NH), 2917 (Ar-H), 1747, 1704 (C=O), 1165, 1044, 749 (C-S-C); ¹H NMR (CDCl₃ + DMSO-d₆, 300 MHz) δ (ppm): 3.17 (dd, *J* = 17.9, 8.9 Hz, 1H, CH₂), 3.50 (dd, *J* = 17.9, 2.9 Hz, 1H, CH₂), 4.62 (dd, *J* = 8.9, 3.9 Hz, 1H, CH), 7.81 (d, *J* = 8.9 Hz, 1H, Ar-H), 8.27 (d, *J* = 8.9 Hz, 1H, Ar-H), 8.77 (s, 1H, Ar-H), 11.96 (brs, 1H, D₂O exchangeable), 12.84 (brs, 1H, D₂O exchangeable); ¹³C NMR (CDCl₃ + DMSO-d₆, 75 MHz) δ (ppm): 37.2 (CH₂), 46.0 (CH₃), 118.8, 120.6, 121.7, 132.1, 143.0, 153.2, 162.8 (benzothiazole), 169.7 (C=O), 172.3 (C=O), 175.5 (C=O). HRMS-ESI m/z [M+H]⁺ calcd. for C₁₂H₉O₅N₄S₂: 353.0014; found 353.0015.

4.1.3.14. *N*-(6-*Methoxy*-1,3-*benzothiazol*-2-*yl*)-2-(2,4-*dioxo*-1,3-*thiazolidin*-5-*yl*)*acetamide* (**4n**). IR (KBr) cm⁻¹: 3207, 3094(NH), 3008 (Ar-H), 1738, 1675 (C=O), 1172, 1060, 639 (C-S-C); ¹H NMR (CDCl₃ + DMSO-d₆, 400 MHz) δ (ppm): 3.05 (dd, *J* = 17.1, 9.6 Hz, 1H, CH₂), 3.40 (dd, *J* = 17.1, 3.5 Hz, 1H, CH₂), 3.85 (s, 3H, OCH₃), 4.54 (dd, *J* = 9.4, 3.3 Hz, 1H, CH), 6.92-6.96 (m, 1H, Ar-H), 7.26 (s, 1H, Ar-H), 7.55 (d, *J* = 8.6 Hz, 1H, Ar-H); ¹³C NMR (CDCl₃ + DMSO-d₆, 75 MHz) δ (ppm): 38.8 (CH₂), 49.6 (CH), 53.4(OCH₃), 102.0, 112.0, 118.0, 130.6, 140.5 (benzothiazole), 153.9 (C=O), 168.3 (C=O), 188 (C=O). HRMS-ESI m/z [M+H]⁺ calcd. for C₁₃H₁₂O₄N₃S₂: 338.0269; found 338.0261.

4.1.3.15. *N*-(6-*Ethoxy*-1,3-*benzothiazol*-2-*yl*)-2-(2,4-*dioxo*-1,3-*thiazolidin*-5-*yl*)*acetamide* (**4o**). IR (KBr) cm⁻¹: 3205, 3100 (NH), 2928 (Ar-H), 1743, 1681 (C=O), 1175, 1060, 717 (C-S-C); ¹H NMR (CDCl₃ + DMSO-d₆, 300 MHz) δ (ppm): 1.39 (t, *J* = 6.9 Hz, 3H, CH₃), 3.06-3.15 (m, 1H, CH₂), 3.37 (dd, *J* = 17.1, 3.7 Hz, 1H, CH₂), 4.02 (q, *J* = 6.7, 6.9 Hz, 2H, OCH₂), 4.59 (dd, *J* = 9.2, 2.8 Hz, 1H, CH), 6.91 (dd, *J* = 11.1, 2.2 Hz 1H, Ar-H), 7.28 (s, 1H, Ar-H), 7.55 (d, *J* = 8.8 Hz, 1H, Ar-H); ¹³C NMR (CDCl₃ + DMSO-d₆, 75 MHz) δ (ppm): 13.21 (CH₃), 36.2 (CH₂), 44.7 (CH), 62.2(OCH₂), 103.0, 113.0, 119.0, 131.6, 141.5, 153.8, 154.1(benzothiazole), 166.7 (C=O), 170.5 (C=O), 173.9 (C=O). HRMS-ESI m/z [M+H]⁺ calcd. for C₁₄H₁₄O₄N₃S₂: 352.0426; found 352.0423.

4.1.4. X-ray structure analysis

X-ray data for the compound **4j** is collected at room temperature using a Bruker Smart Apex CCD diffractometer with graphite monochromated MoK α radiation (λ =0.71073Å) with ω scan method [46]. Preliminary lattice parameters and orientation matrices are obtained from four sets of frames. Unit cell dimensions are determined using 6121 reflections for **4j** data. Integration and scaling of intensity data are accomplished using SAINT program [46]. The structures are solved by Direct Methods using SHELXS97 [47] and refinement is carried out by full-matrix least-squares technique using SHELXL97 [47]. Anisotropic displacement parameters are included for all non-hydrogen atoms. All H atoms are positioned geometrically and treated as riding on their parent C atoms, with C-H distances of 0.93--0.97 Å, and with $U_{iso}(H) = 1.2U_{eq}$ or 1.5U_{eq} for methyl atoms. CCDC 905873 contains the supplementary crystallographic (C) data for These data be obtained free of charge this paper. can at www.ccdc.cam.ac.uk/conts/retrieving.html [or from the Cambridge Crystallographic Data Centre (CCDC), 12 Union Road, Cambridge CB2 1EZ, UK; fax: +44(0) 1223 336 033; email: deposit@ccdc.cam.ac.uk].

4.2. Biological experiments

4.2.1. Experiments in vitro

In the *in vitro* assays, each experiment is performed at least in triplicate and the standard deviation of absorbance is less than 10% of the mean.

4.2.2. DPPH radical scavenging assay

Antioxidant activity of synthesized thiazolidinedione-5-acetamide derivatives is assessed on the basis of the free radical scavenging effect of the stable 1,1-diphenyl-picrylhydrazyl (DPPH) following a previously described method [48] with some modifications. The diluted working solutions of the synthesized compounds are prepared in methanol (5, 10, 20, 40, 60 and 180 μ g/mL). One mL of methanol solution of DPPH (0.002 %) is mixed with 1 mL solution of test compound. The mixture is shaken vigorously and left to stand in dark for 30 min. Absorbance of the resulting solution is measured at 517 nm in a Lambda 25 UV-visible spectrophotometer (Perkin-Elmer, Shelton, CT, USA). Ascorbic acid and luteolin are used as positive controls. The radical scavenging ability is measured as a decrease in the absorbance of DPPH. Lower absorbance of the reaction mixture has indicated higher free radical scavenging activity. DPPH radical scavenging activity is calculated using the following formula: DPPH radical scavenging activity (%) = [(absorbance of control – absorbance of test sample)/ (absorbance of control)] x 100. Radical scavenging potential is expressed as EC₅₀ value, which represents the test compound concentration at which 50% of the DPPH radicals are scavenged [49].

4.2.3. Superoxide radical scavenging assay

The superoxide radical scavenging assay of samples is done according to the protocol described by Liu, *et al.*, [50]. The superoxide radical is generated by phenazine methosulfate - nicotinamide adenine dinucleotide (PMS/NADH) system, which reduces nitro blue tetrazolium (NBT) forming a purple coloured formazan. The reaction mixture consisted of 3.0 mL of 16 mM

tris-HCl buffer (pH 8.0) containing 78 mM NADH, 50 μ M NBT, 10 μ M PMS and various concentrations (5, 10, 20, 40, 60 and 180 μ g/mL) of test compound. After incubation for 5 min at room temperature the absorbance is read at 560 nm. Ascorbic acid is run in parallel as a positive control. The scavenging activity of superoxide radical (%) is calculated using the equation:

 $[(A_{560} of control - A_{560} of sample)/A_{560} of control] * 100$

4.2.4. Inhibition of lipid peroxidation assay

The inhibition of lipid peroxidation is assayed by measuring malondialdehyde (MDA) thiobarbituric acid adduct formation using egg yolk as the oxidizable substrate [51]. The yolk is taken out from an egg and same volume of PBS (0.1 M, pH 7.45) is added to it and stirred vigorously. The yolk suspension is mixed with 0.5 mL of test sample at various concentrations (50, 100, 150 and 300 μ g/mL), 0.2 mL of 25 mM FeSO₄ and 1.3 mL of PBS and incubated at 37 °C for 15 min before the reaction is quenched by adding 0.5 mL of 20% trichloroacetic acid and 1 mL of 0.8% thiobarbituric acid, and then the mixture is heated at 100 °C for 15 min. The samples are read at 532 nm. Ascorbic acid is used as positive control. The inhibition of lipid peroxidation is calculated from the following equation:

Inhibition effect (%) = $A_{control} - A_{sample} / A_{control} * 100\%$

4.2.5. Inhibition of erythrocyte hemolysis assay

The inhibition of erythrocyte hemolysis is measured according to the modified protocol described by Ng, et al., [52]. The human blood sample is collected in a tube with 3% sodium citrate. The blood sample is centrifuged at 3,000 g for 10 min to separate erythrocytes from the plasma and washed with PBS buffer (pH 7.4) and 1.25% erythrocyte suspension is prepared. Different concentrations of compounds (50, 100, 150 and 300 μ g/mL) are added to the mixture containing 1.0 mL erythrocyte suspension and 0.5 mL H₂O₂ (2.5 mM) and incubated at 37 °C for 30 min. After incubation, the reaction is terminated by incubating in ice water bath at 4 °C and then centrifuged at 3,000 g for 10 min. The absorbance of the precipitated protein is measured at 540 nm. Ascorbic acid is used as positive control. The inhibition rate of the hemolysis is calculated using the following equation:

Inhibition rate (%) =
$$(A_{H2O2} - A_{sample}) / (A_{H2O2} - A_{normal}) * 100$$

4.2.6. THP1 cell culture

THP1 monocytes are obtained from ATCC and grown in RPMI 1640 (Sigma) containing 10% FBS (Lonza), non essential amino acids, penicillin (100 U/mL), and streptomycin (100 μ g/mL) in 75-cm² filter vent flasks (Costar), incubated at 37°C in a humidified atmosphere containing 5% CO₂ and 95% air .

4.2.7. Measurement of cell viability

The effect of synthesized compounds on cell viability is determined by Trypan blue dye exclusion assay. THP1 cells are seeded at a density of 2 x 10^5 cells/ mL in 24-well plates in triplicates and are treated with test compounds (10μ M) for 48 h. At the end of the treatments, cells are harvested and resuspended in 0.4 % Trypan blue (Sigma) and live/dead cells are counted using cell countess chamber (Invitrogen).

4.2.8. Enzyme-linked immunosorbent assay for IL-1β and MCP-1 inhibition

To check the inhibitory effects of thiazole derivatives (**4a-o**) on PMA induced inhibition of inflammation, THP1 monocytes are seeded at a density of 2 x 10^5 cells/ mL in 24 well plates. Cells are pre-treated with 5, 10 and 20 μ M concentrations of compounds for 1 h before they are stimulated with 10 ng/mL of PMA. After 48 h of PMA stimulation supernatants are harvested and assayed for IL-1 β , MCP-1 and TNF- α using an enzyme-linked immunosorbent assay (ELISA) kit following the manufacturer's instructions (eBiosciences, San Diego, CA, USA). The absorbance in each well is measured with a microplate reader at 450 nm and corrected at 570 nm. Concentrations of cytokines released are obtained and the percentage of inhibition of cytokines production is calculated as compared to control conditions. IC₅₀ is also obtained as a mean of anti-inflammatory potency, expressing the predicted concentration of the correspondent compound able to reach 50% inhibition. The standard curves are calculated by plotting the pg/mL concentrations *versus* absorbance values from the standards, and are used to quantify the amount of cytokines released by the cells.

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activity data. The authors are grateful to the Council of Scientific and Industrial Research (CSIR), New Delhi, for the financial assistance.

Appendix A. Supplementary data

Supplementary data related to this article can be found at http://dx.doi.org/10.1016/j.ejmech.2013.

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Table 1

Structure and physicochemical characteristics of target compounds 4a-o.

	R N N N N N N N N N N N N N N N N N N N
4a-k	41-o

				D h			
Entry	Product ^a	R	Mol. Formula/ Mol. Wt.	$\mathbf{R_f}^{\mathrm{b}}$	C log P*	mp °C	Yield (%)
1	4 a	Н	$\begin{array}{c} C_{14}H_{11}N_{3}O_{3}S_{2}\\ 333 \end{array}$	0.51	2.28	236-238	71
2	4 b	p-Cl	$\begin{array}{c} C_{14}H_{10}N_{3}O_{3}S_{2}Cl\\ 367\end{array}$	0.56	3.00	>260	67
3	4c	<i>p</i> -F	$\begin{array}{c} C_{14}H_{10}N_{3}O_{3}S_{2}F\\ 351 \end{array}$	0.54	2.43	212-216	69
4	4d	<i>p</i> -Br	$C_{14}H_{10}BrN_3O_3S_2$ 412	0.57	3.15	252-254	67
5	4 e	<i>p</i> -I	$C_{14}H_{10}IN_3O_3S_2$ 459	0.59	3.41	242-244	62
6	4f	o-Cl	$C_{14}H_{10}N_3O_3S_2Cl$ 367	0.55	2.75	216-218	68
7	4 g	<i>m</i> -Cl	$\begin{array}{c} C_{14}H_{10}N_{3}O_{3}S_{2}Cl\\ 367\end{array}$	0.55	3.00	212-214	68
8	4h	o-Me	$C_{15}H_{13}N_3O_3S_2$ 347	0.50	2.48	192-194	69
9	4i	<i>m</i> -Me	$C_{15}H_{13}N_3O_3S_2$ 347	0.51	2.78	218-220	69
10	4j	o-F	$C_{14}H_{10}N_3O_3S_2F_{351}$	0.53	2.43	220-222	68
11	4k	o-OMe	$C_{15}H_{13}N_3O_4S_2$ 363	0.56	1.74	228-230	65
12	41	Н	$C_{12}H_9N_3O_3S_2$ 307	0.57	1.78	266-268	61
13	4m	6-NO ₂	$C_{12}H_8N_4O_5S_2$ 352	0.60	1.62	272-274	56
14	4n	6-OMe	$C_{13}H_{11}N_3O_4S_2$ 337	0.59	2.08	262-264	54
15	40	6-OEt	$C_{14}H_{13}N_3O_4S_2$ 351	0.58	2.61	246-248	58

^a All products are characterized by IR, NMR, ESI-MS and HRMS analysis; ^b Ethyl acetate: Petroleum ether (2:1);

* C log P values are calculated using ChemDraw Ultra 10.0.

Crystal data and structure refinement parameters for compound 4j.

Empirical formula Formula weight Temperature (K) Color Wavelength (Å) Crystal system Space group	$C_{14}H_{10}FN_{3}O_{3}S_{2}$ 351.37 293 Colorless, needle 0.71073 Monoclinic $C24$
Temperature (K) Color Wavelength (Å) Crystal system Space group	Colorless, needle 0.71073 Monoclinic
Color Wavelength (Å) Crystal system Space group	0.71073 Monoclinic
Crystal system Space group	Monoclinic
Crystal system Space group	
Space group	
	C2/c
a(Å)	30.877(4)
b(Å)	5.7127(6)
c(Å)	18.357(3)
$\beta(deg)$	117.036(3)
Volume ($Å^3$)	2884.2(6)
Ζ	8
$D_{calc} (mg m^{-3})$	1.618
Absorption Coefficient (mm ⁻¹)	0.40
F(000)	1440.0
Crystal size (mm ³⁾	0.60 imes 0.20 imes 0.10
θ Range for data collection (deg)	2.3-28.0
Index ranges	$h=-36\rightarrow 36,$
	$k=-6 \rightarrow 6,$
	$l=-21 \rightarrow 21.$
Reflections collected	12857
Independent Reflections(Rint)	$2538 (R_{int} = 0.0346)$
Data/restraints/Parameters	2538/0/216
No. of parameters	216
Goodness-on-fit on F^2	1.18
Final R indices [$I > 2\sigma(I)$]	R1 = 0.0504, wR2 = 0.1241
R indices (all data)	R1 = 0.0527, wR2 = 0.1256
Largest diffraction peak and hole (Å e^3)	0.49 and -0.25
CCDC number	905873

D 11 (1			
Bond lengths		Bond angles	
S1-C1	1.766(3)	O1 - C2 - N1	124.7(3)
S1–C3	1.824(3)	O1 - C2 - C3	111.5(2)
O1–C2	1.216(3)	C2-N1-C1	118.5(3)
O2-C1	1.197(4)	N1-C1-S1	109.8(2)
N1-C1	1.383(4)	O2-C1-N1	125.2(3)
C2-N1	1.360(4)	O2-C1-S1	125.0(3)
O3–C5	1.217(3)	C4–C3– S1	114.3(2)
N2-C5	1.360(3)	O3 - C5 - N2	112.7(3)
N2-C6	1.379(4)	O3–C5– C4	124.3(2)
S2-C7	1.722(3)	N2-C6-S2	123.77(19)
S2-C6	1.729(3)	N3-C6-N2	120.4(2)
N3-C6	1.295(3)	N3–C6– S2	115.9(2)
N3-C8	1.381(4)	F1–C10–C9	118.5(3)
F1-C10	1.361(4)	F1-C10-C11	118.1(3)
		C14–C9–C8	119.8(3)

Selected bond distances (Å) and bond angles $(^{0})$ for the compound **4j**.

Compound				
	DPPH radical	Superoxide anion	Lipid peroxidation	Erythrocyte
	scavenging	scavenging	inhibition	hemolysis
	activity	activity		inhibition
4a	NA	NT	NT	NT
4b	NA	NT	NT	NT
4c	52.69	NA	NA	NA
4d	49.71	NA	NA	NA
4e	NA	NT	NT	NA
4f	NA	NT	NT	NT
4g	NA	NT	NT	NT
4 h	NA	NT	NT	NT
4i	90.16	NA	NT	NA
4j	82.50	NA	NA	NA
4k	58.68	79.94	131.79	96.62
41	52.18	101.18	149.70	107.28
4m	39.76	NA	NA	NA
4n	51.82	NA	NA	NA
4o	62.90	NA	NA	NA
Ascorbic acid ^b	40.28	21.01	139.97	96.625
Luteolin ^b	44.18	31.01	149.70	107.28

Antioxidant activities of the synthesized compounds (4a-o).

^a EC₅₀ values are mean of three experiments; ^b Positive control; NA: not active; NT: not tested.

Treatment conditions	Secreted IL-1β levels (pg/ mL)	Secreted MCP-1 levels (pg/ mL)
Without PMA	12 ± 0.38	28.12 ±3.18
PMA (10 ng/mL)	172 ± 0.71	3521.63 ±79.78
PMA+ Pioglitazone	165 ± 1.56	2588.33 ±11.64
(10 µM)		

Effect of pioglitazone on PMA-induced IL-1β, MCP-1 levels in THP1 cells.^a

^a 2 x 10^{5} /mL THP1 monocytes are seeded in 24 well cell culture plates and stimulated with 10 ng/ mL of PMA for IL-1 β , MCP-1 production. Cells are incubated for a period of 48 h. At the end of the treatment, conditioned media is collected and IL-1 β , MCP-1 levels are measured by ELISA as described in the materials and methods.

Compound	$\mathbf{IC_{50}}(\mu\mathrm{M})^{\mathrm{b}}$		•
Compound		MCP-1 secretion	
	inhibition efficacy	inhibition efficacy	-
4 a	9.6 ± 2.96	30.4 ± 3.4	
4b	16.31 ± 0.91	NA	
4 c	NA	17.58 ± 1.58	
4d	33.12 ± 5.57	49.13± 3.46	
4e	NA	31.7 ± 1.86	
4f	18.23 ± 1.53	26.49 ± 2.54	(
4g	23.4 ± 6.66	34.24 ± 1.86	
4 h	7.66 ± 0.46	18.79 ± 2.45	
4i	11.72 ± 4.05	18.94 ± 0.86	
4j	NA	16.68 ± 1.11	
4 k	7.23 ± 0.99	NA	
41	NA	20.8 ± 2.21	
4 m	6.83 ± 0.24	20.29 ± 3.21	
4 n	7.03 ± 0.06	39.6± 1.66	
4o	NA	63.06 ± 4.21	Ϋ́, Υ
Pioglitazone	NA	18.84 ± 2.16	
Piroxicam ^c	21.33 ± 3.2	NT	<i>*</i>

IL-1 β and MCP-1 secretion inhibition efficacy of the synthesized compounds (**4a-o**).^a

^a THP1 monocytes are pretreated with 5, 10 and 20 μ M concentrations of the above mentioned thiazole substituents (**4a-o**) for 1 h before simulation with 10 ng/ mL of Phorbol 13-myristate 12-acetate (PMA) to induce inflammation for a period of 48 h. At the end of the treatment, conditioned media is collected and levels of IL-1 β and MCP-1 are measured by ELISA as described in the materials and methods. ^b IC₅₀ values are Mean \pm SD of three independent experiments; ^c Piroxicam, a known anti-inflammatory agent is used as a positive control; NA: denotes activity >50 μ M; NT: indicates 'not tested'.

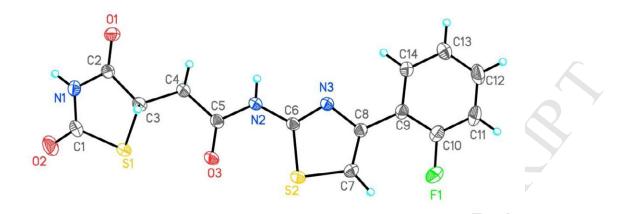
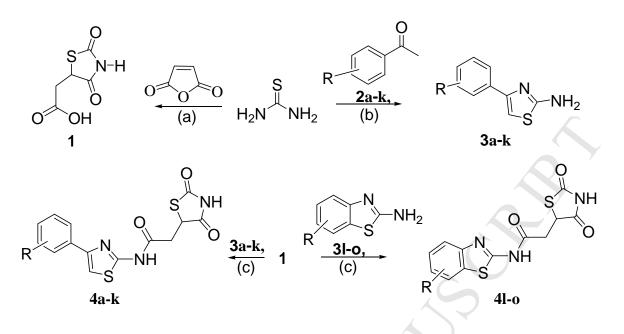
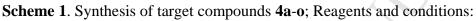


Fig. 1. An ORTEP view of the molecular structure and atom-numbering of compound **4j**. Displacement ellipsoids are drawn at the 30% probability level and H atoms are shown as small spheres of arbitrary radius.





- (a) Conc. HCl, $100 {}^{0}$ C, 5 h; (b) I₂, 90 0 C, 12 h;
- (c) *N*-Hydroxybenzotriazole (HOBT),

O-(1H-Benzotriazol-1-yl)-N,N,N',N'-tetramethyluronium hexafluorophosphate (HBTU),

and N,N-Diisopropylethylamine (DIEA), DCM, DMF, 0 0 C - RT, 12 h.

 $R = H, F, Cl, Br, I, Me, OMe, OEt, NO_2$.

Supporting information for

Novel 2-(2,4-dioxo-1,3-thiazolidin-5-yl)acetamides as antioxidant and/or anti-inflammatory compounds

Satish Koppireddi^a, Jayaram Reddy Komsani^a, Sreenivas Avula^a, Sujitha Pombala^b, Satishbabu Vasamsetti^b, Srigiridhar Kotamraju^b, Rambabu Yadla^{a,*}

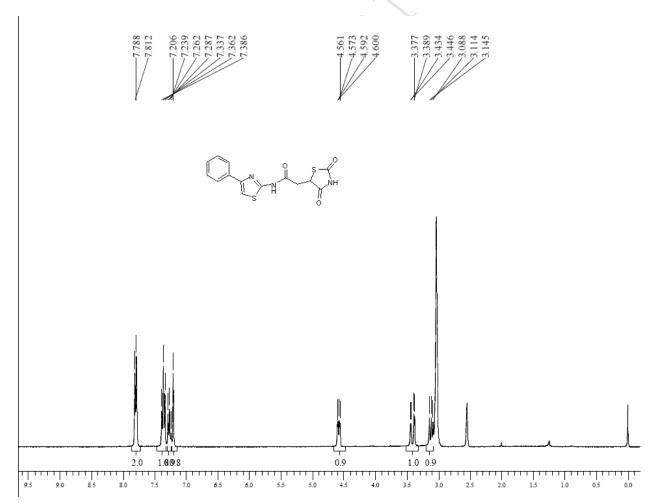
^aFluoroorganics Division, CSIR-Indian Institute of Chemical Technology, Hyderabad-500 607, India; ^bChemical Biology Division, CSIR- Indian Institute of Chemical Technology, Hyderabad.

**Corresponding author. Tel.:* +91-40-27193171; *Fax:* +91-40-27193185. *E-mail address:* <u>ryadla@yahoo.com</u> (Rambabu Yadla).

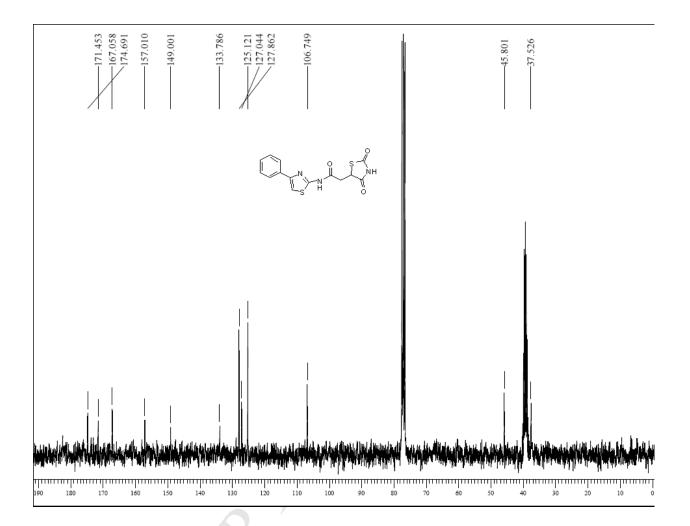
¹H NMR and ¹³C NMR spectra of *N*-thiazolyl-thiazolidinedione-2-acetamides 4a-o

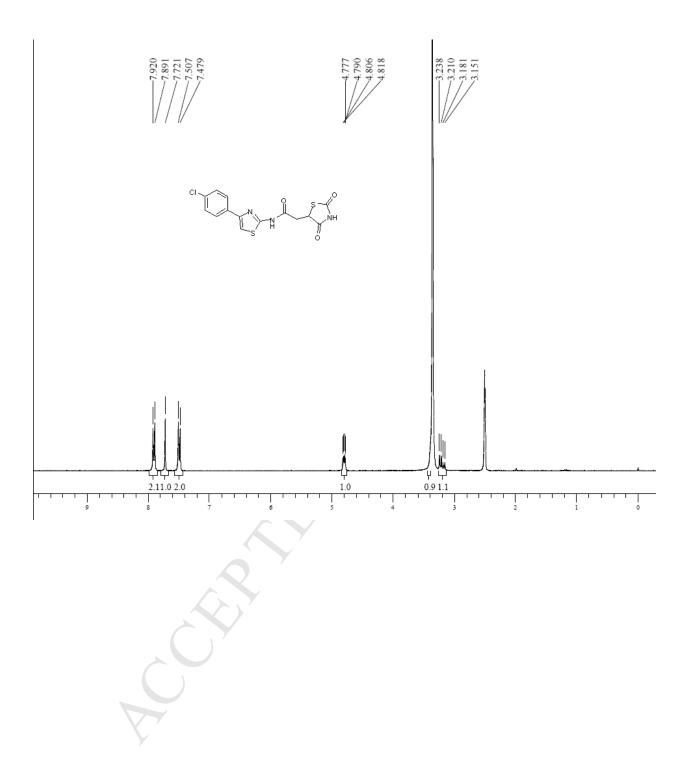
¹H NMR and ¹³C NMR charts of representative compounds (4a-4o)

¹H NMR spectrum of **4a**

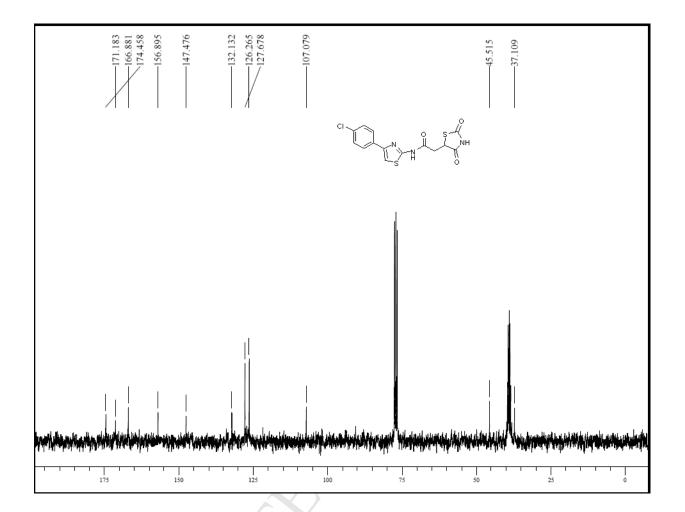


¹³C NMR spectrum of **4a**

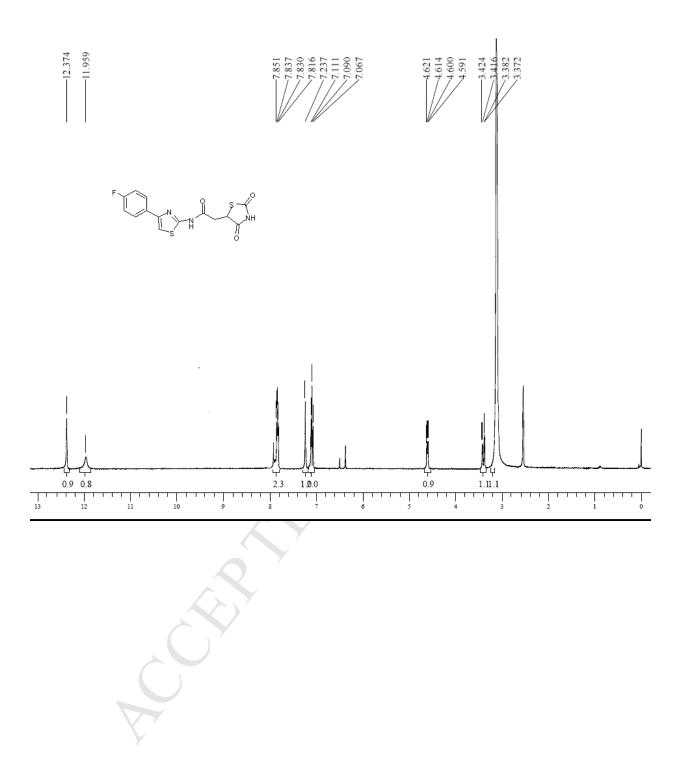




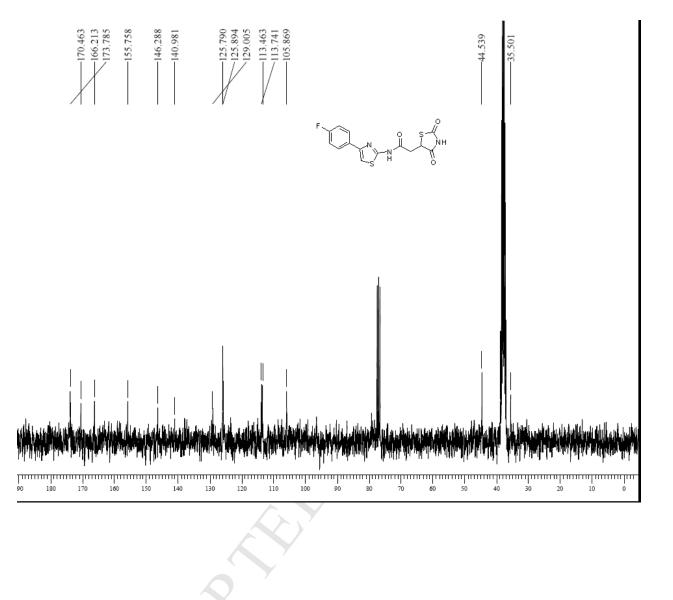
¹³C NMR spectrum of **4b**



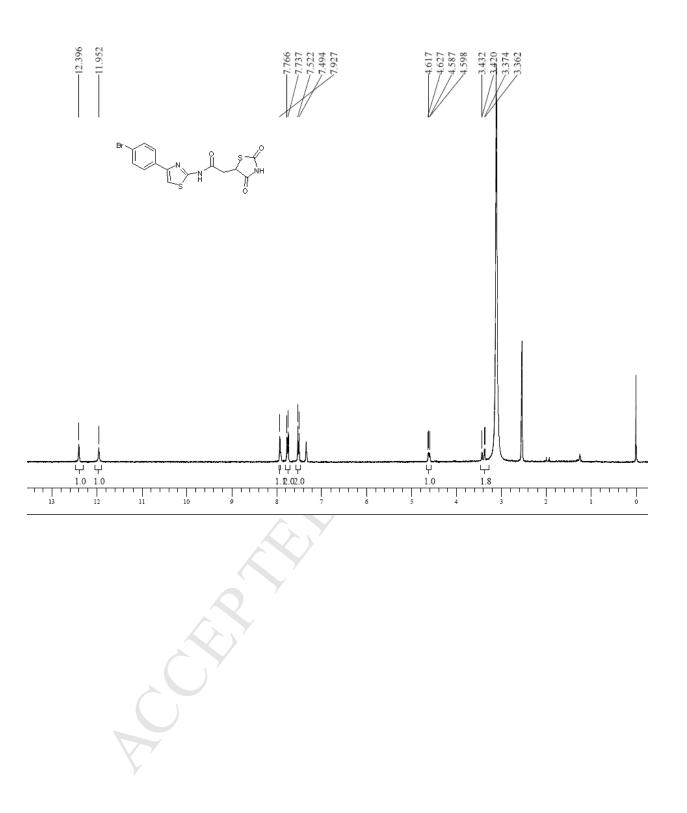
¹H NMR spectrum of 4c



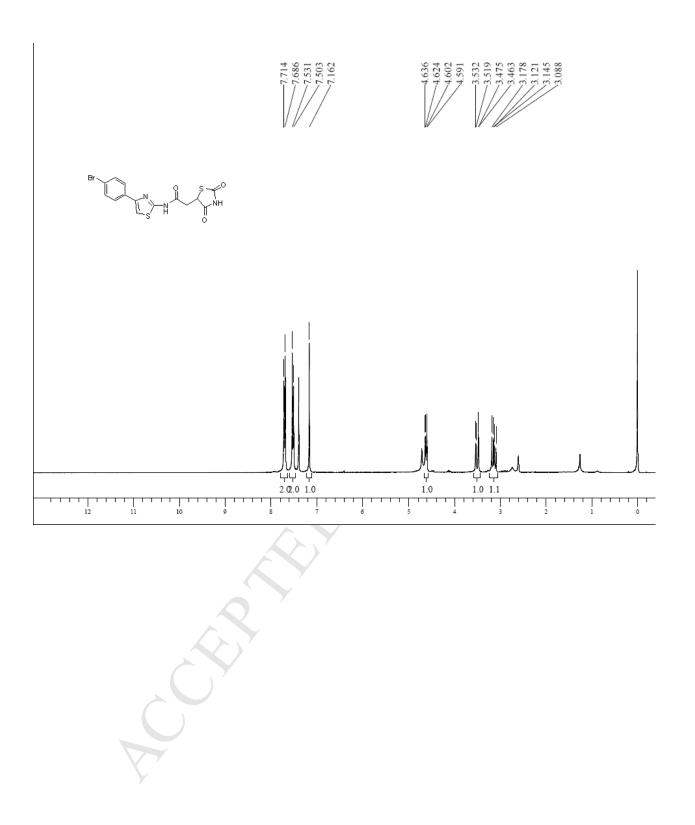
¹³C NMR spectrum of **4c**



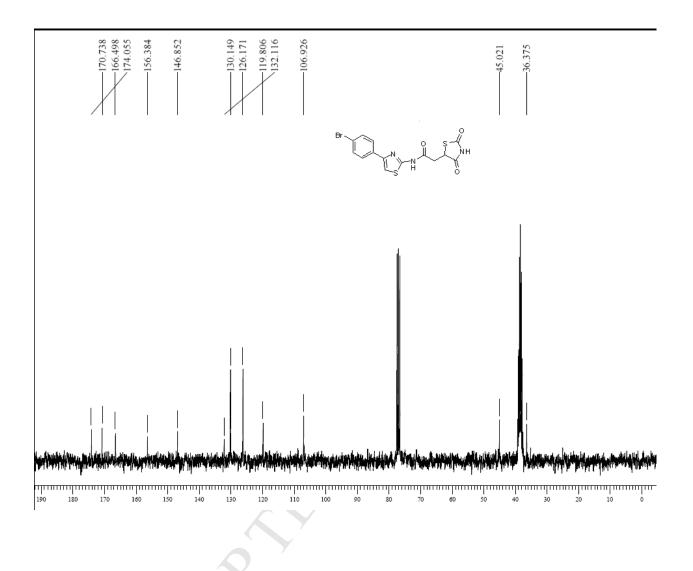
¹H NMR spectrum of **4d**

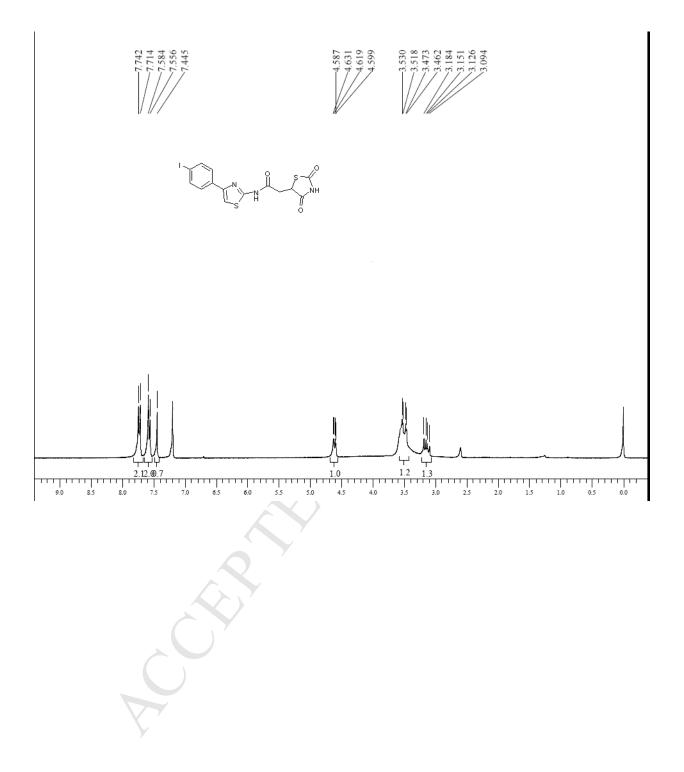


 1 H NMR spectrum of **4d** (D₂O exchanged)



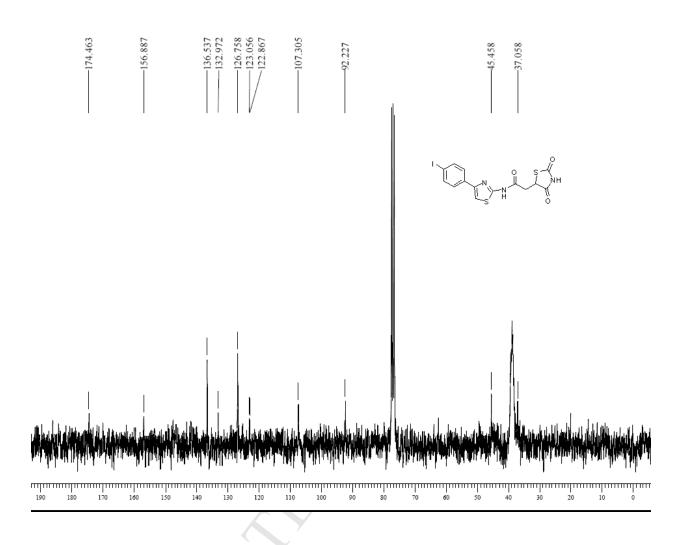
¹³C NMR spectrum of **4d**

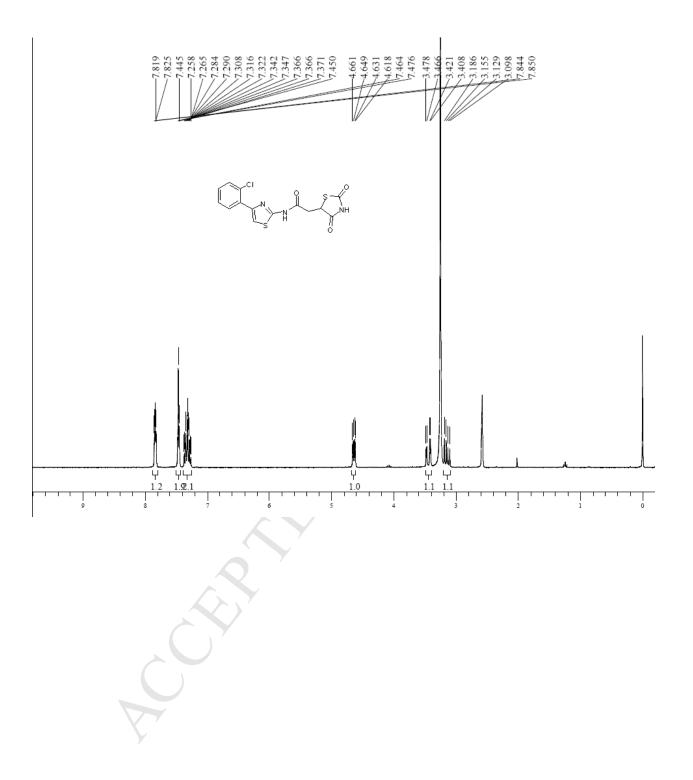




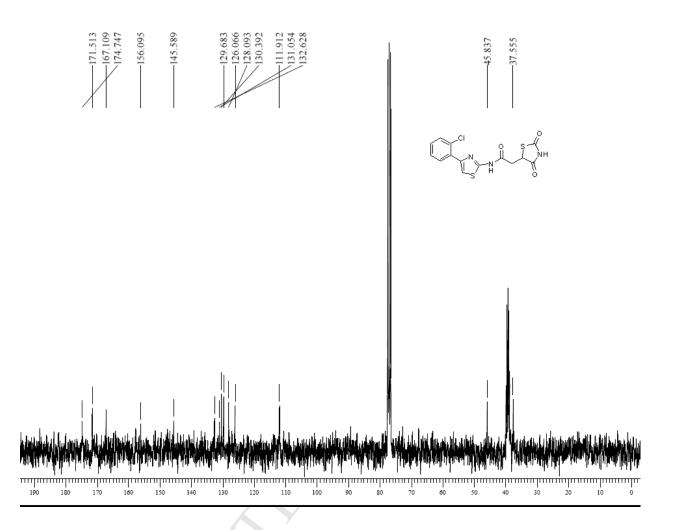
¹³C NMR spectrum of **4e**

¹H NMR spectrum of **4f**

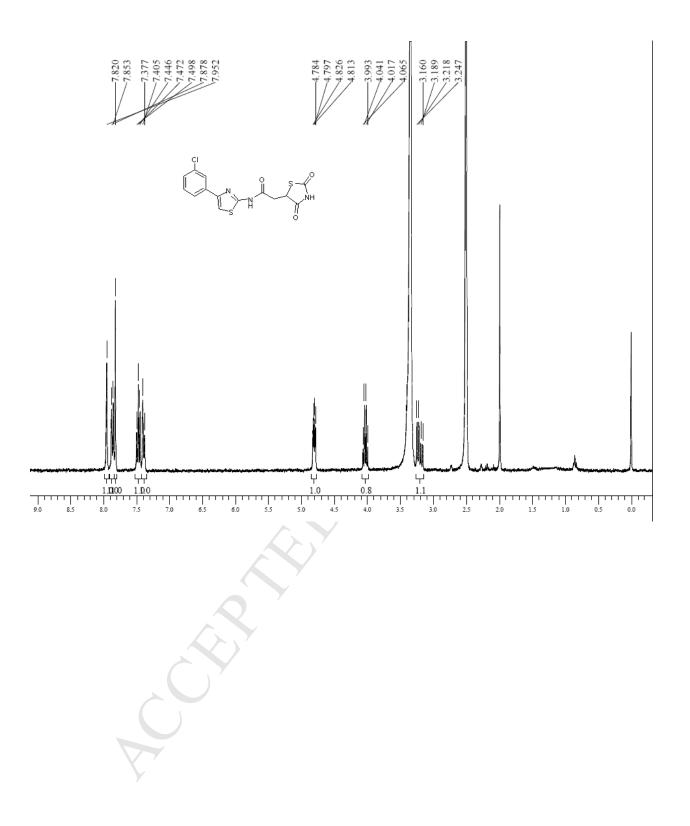




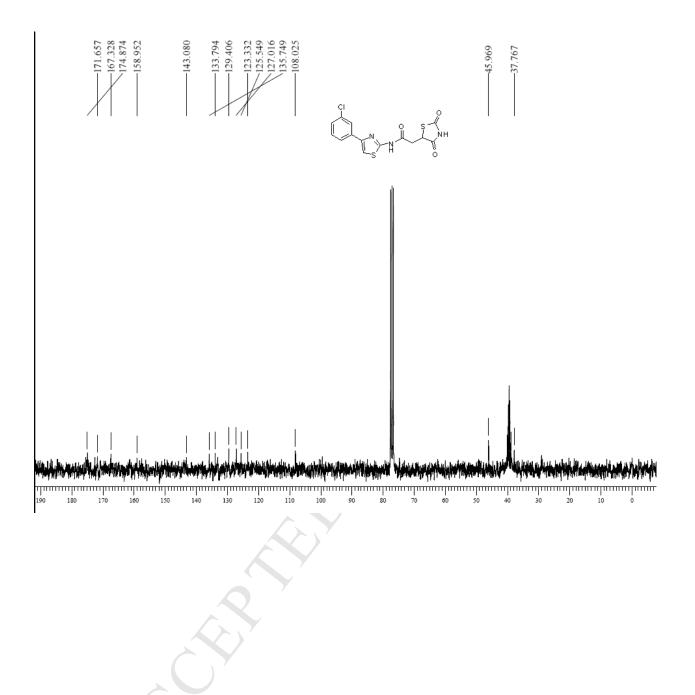
¹³C NMR spectrum of **4f**



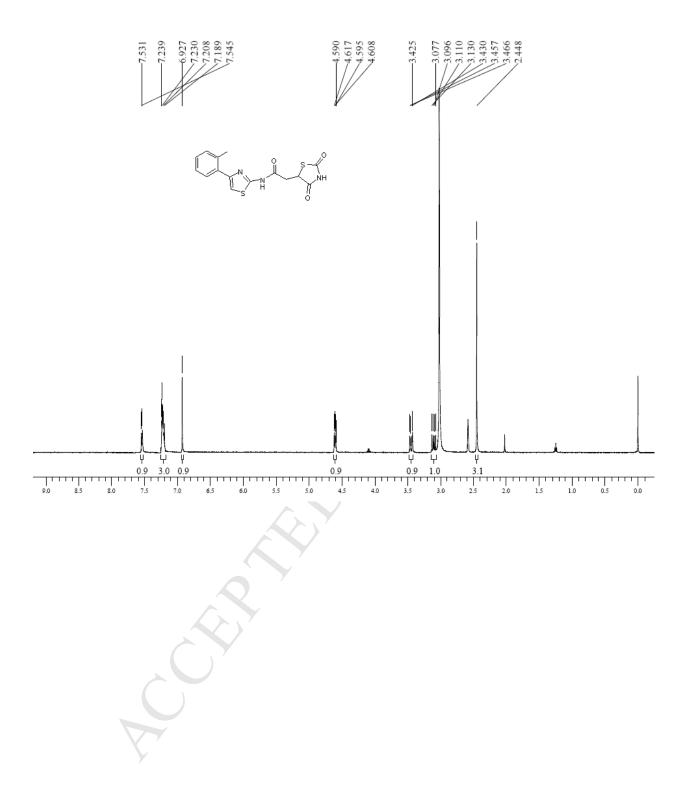
¹H NMR spectrum of **4g**



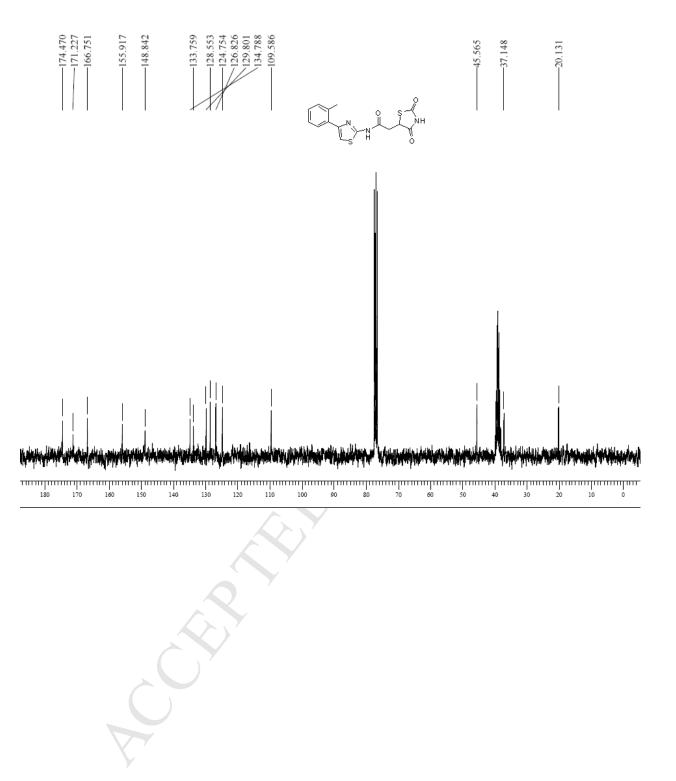
¹³CNMR spectrum of **4g**



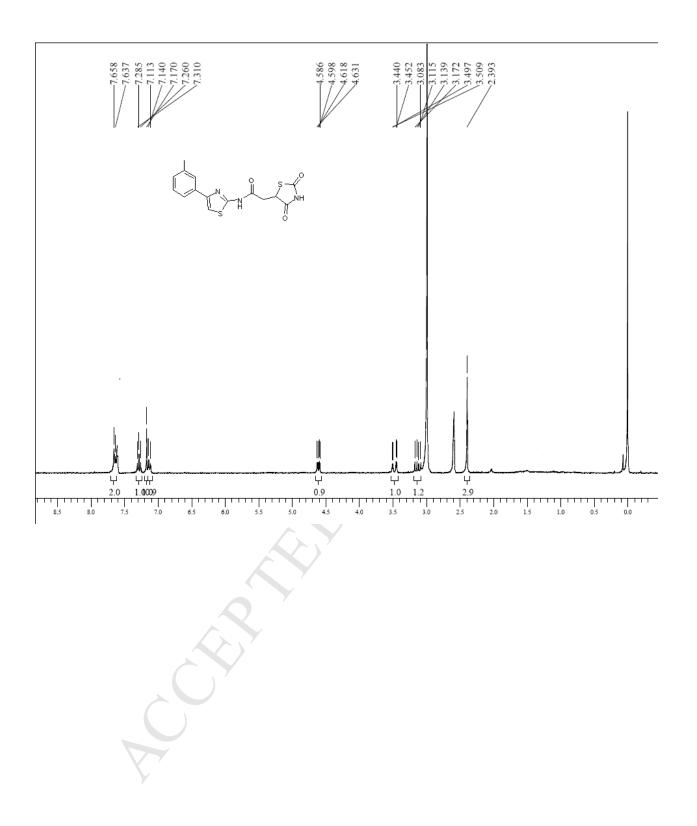
¹H NMR spectrum of **4h**



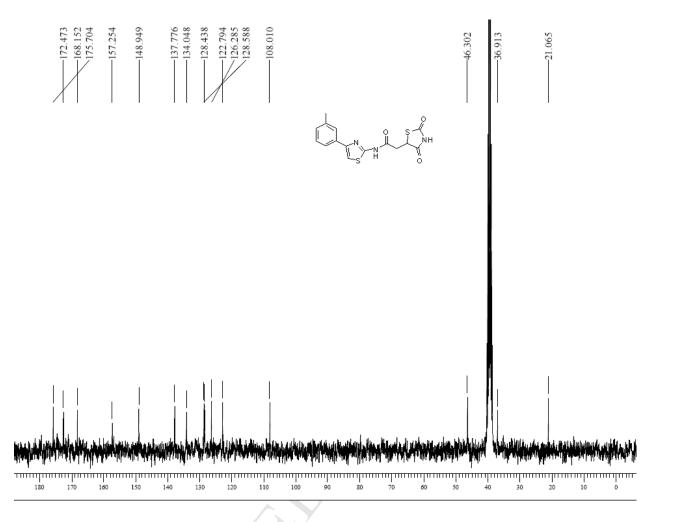
¹³C NMR spectrum of **4h**



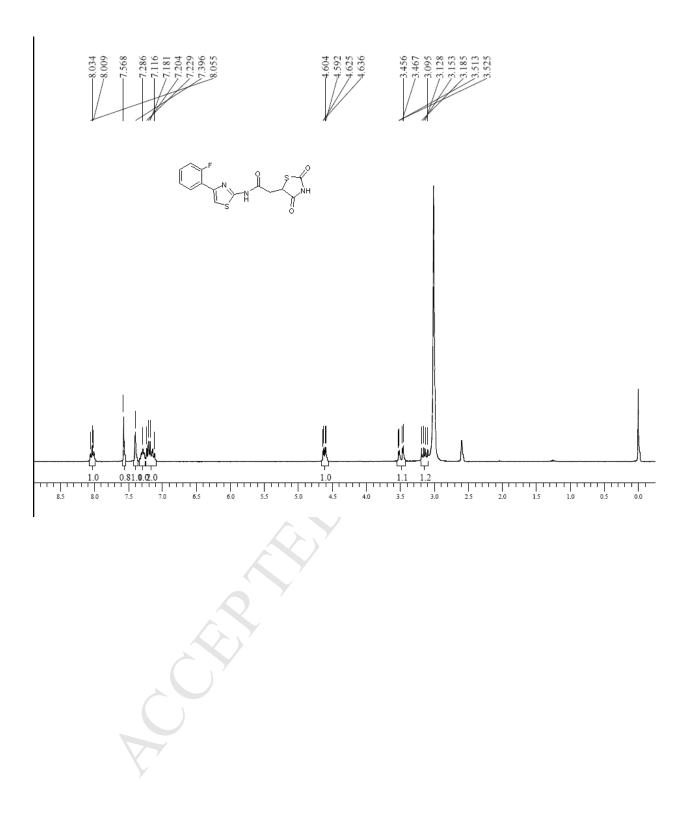
¹H NMR spectrum of **4i**



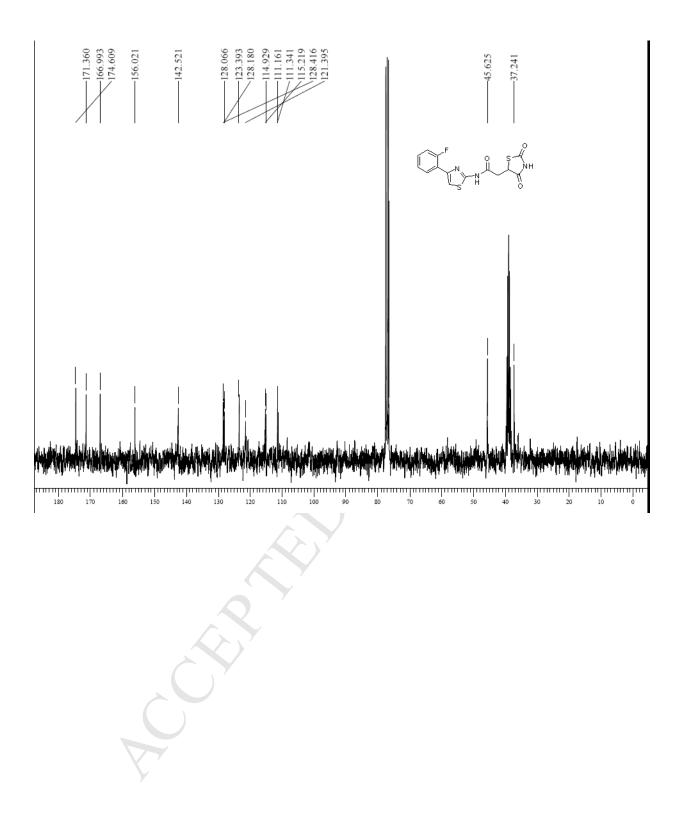
¹³C NMR spectrum of **4i**



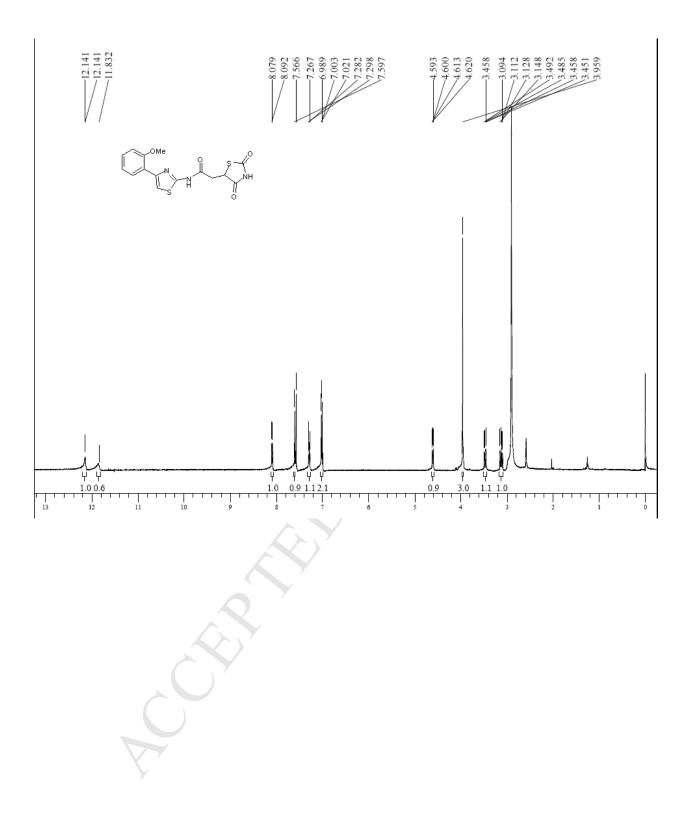
¹H NMR spectrum of **4**j



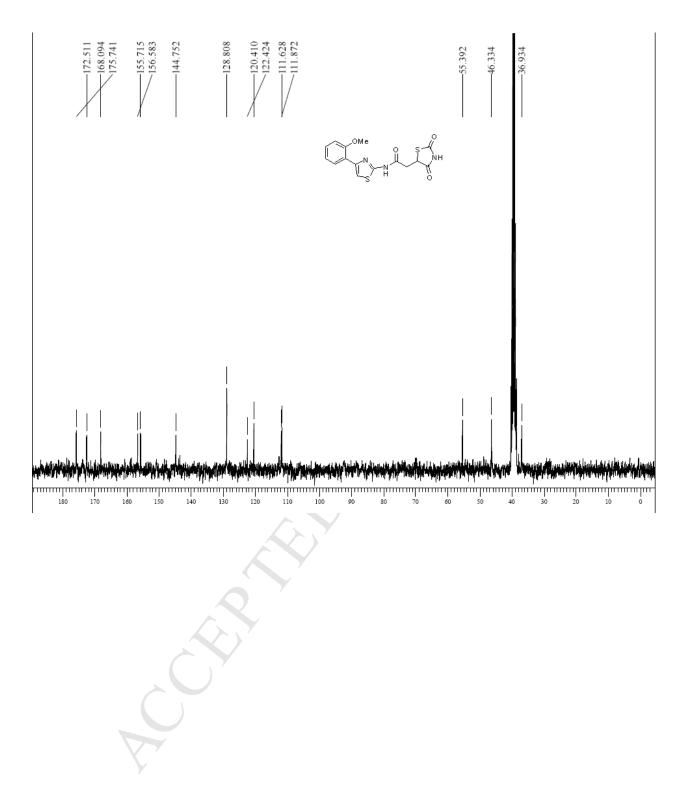
¹³C NMR spectrum of **4**j



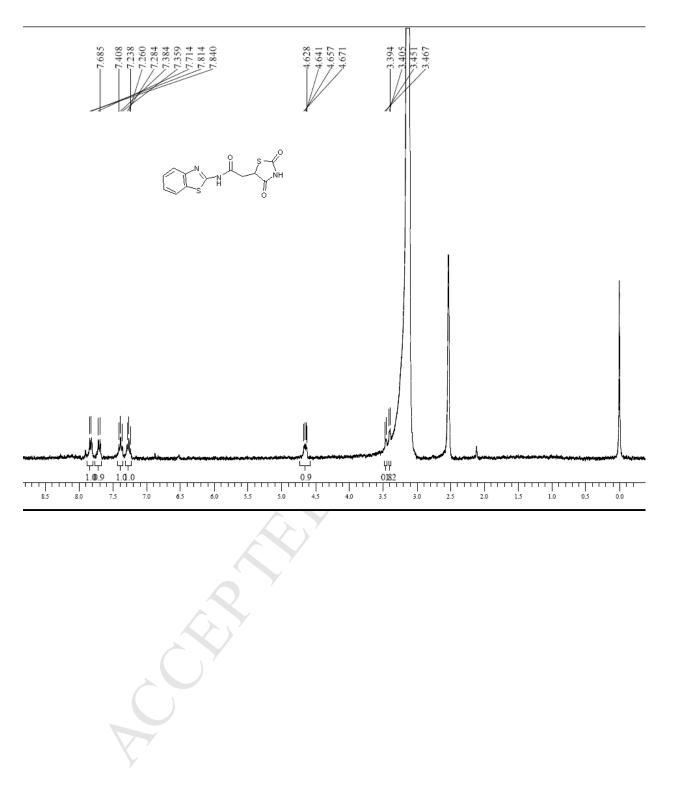
¹H NMR spectrum of **4**k



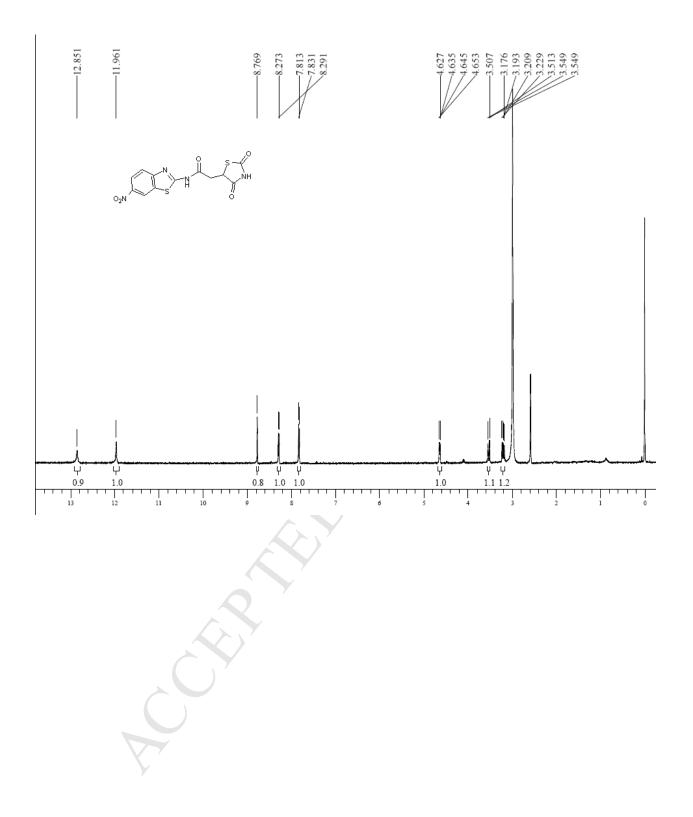
¹³C NMR spectrum of **4**k



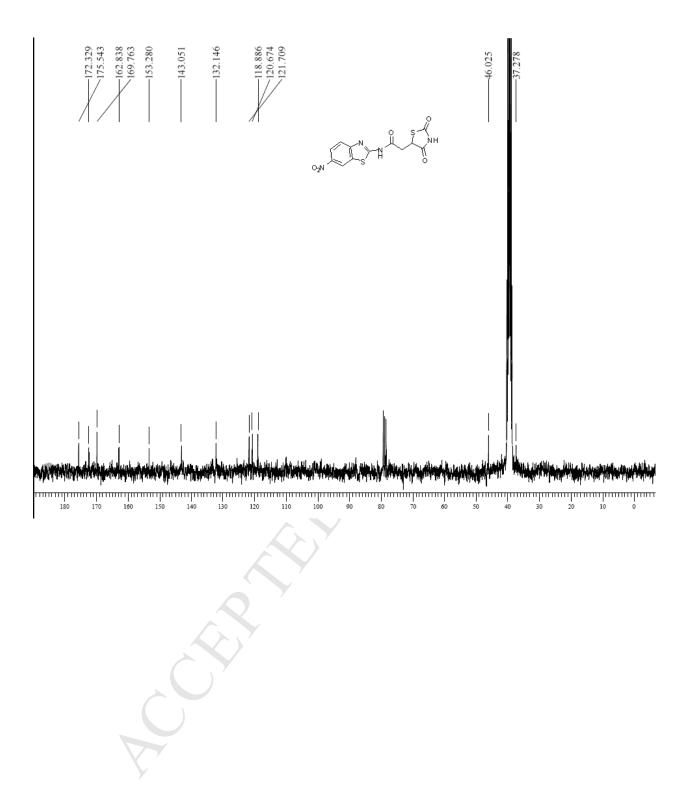
¹H NMR spectrum of **4**l



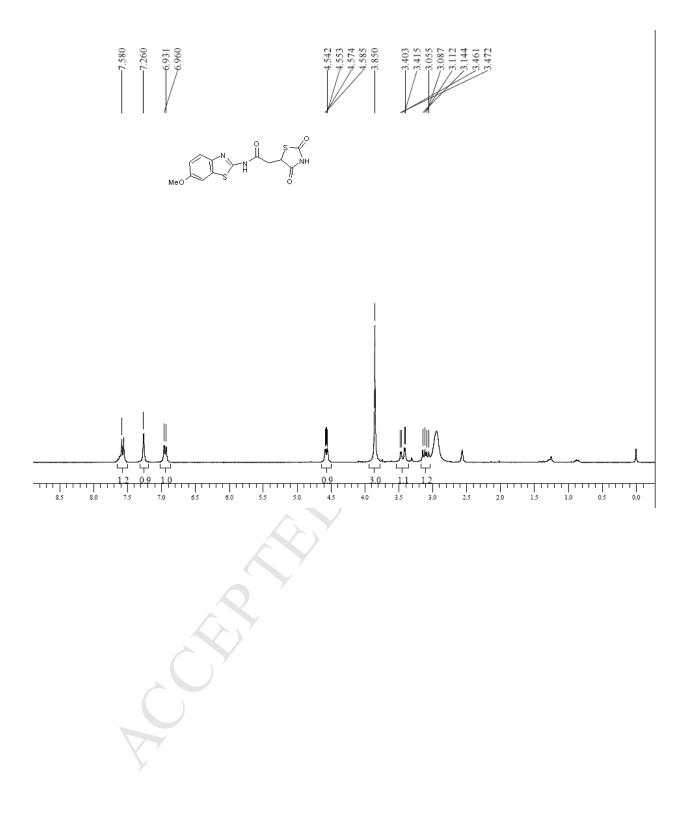
¹H NMR spectrum of 4m



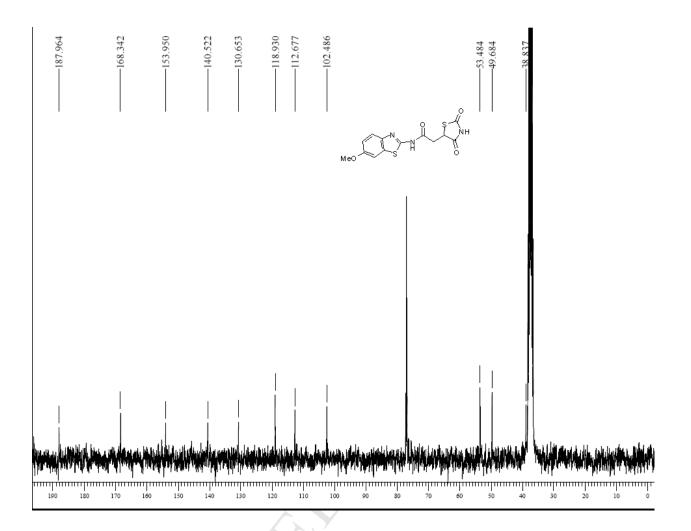
¹³C NMR spectrum of **4m**



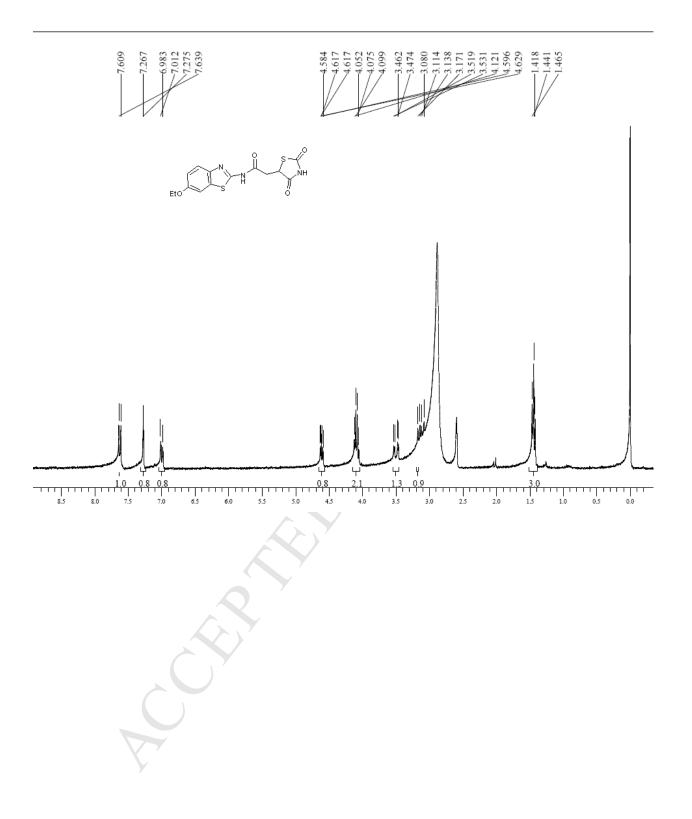
¹H NMR spectrum of **4n**



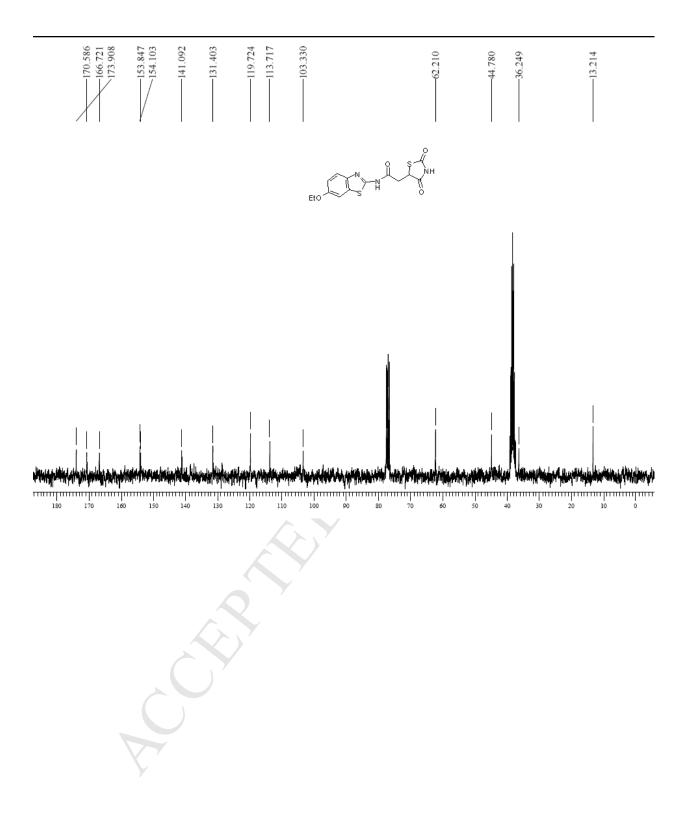
¹³CNMR spectrum of **4n**



¹H NMR spectrum of **40**



¹³C NMR spectrum of **40**



High lights

- > Novel N-thiazolyl/ benzothiazolyl-thiazolidinedione-2-acetamides are prepared.
- ➤ Essayed for DPPH RSA, SASA, LPI, EHI, IL-1ß & MCP-1 secretion inhibition.
- > Some of them show both anti-inflammatory and antioxidant potential.
- Structure of one of the target compounds is confirmed by X-ray.