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Graphical abstract

Design and Synthesis of Pyrazole - Oxindole Conjugates Targeting Tubulin Polymerization as New Anticancer Agents

Ahmed Kamal^{*1}, Anver Basha Shaik¹, Nishant Jain², Chandan Kishor², Ananthamurthy Nagabhushana ^{3,4}, Bhukya Supriya², G. Bharath Kumar¹, Sumit S. Chourasiya¹, Yerramsetty Suresh², Rakesh K Mishra³, Anthony Addlagatta^{*2}

A series of twenty one pyrazole linked oxindole conjugates comprising of a four ring scaffold (A, B, C and D) were synthesized and investigated for their antiproliferative activity.



Research Highlights

Design and Synthesis of Pyrazole - Oxindole Conjugates Targeting Tubulin Polymerization as New Anticancer Agents

- The new Pyrazole-oxindole conjugates were synthesized by Knoevenagel condensation
- Investigated for antiproliferative activity on different human cancer cell lines
- The lead congeners were taken for cellular tubulin polymerization assay
- Studies on Zebrafish embryos have been carried out
- Further, lead compounds have been docked against the tubulin structure

Design and Synthesis of Pyrazole - Oxindole Conjugates Targeting Tubulin Polymerization as New Anticancer Agents

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¹Medicinal Chemistry and Pharmacology, ² Centre for Chemical Biology (CSIR -Indian Institute of Chemical Technology, Hyderabad 500007, India. ³ CSIR - Centre for Cellular and Molecular Biology, Hyderabad 500007, India. ⁴CoE in Epigenetics, IISER-Pune, Pune 411021, India.

Abstract: A series of twenty one compounds with pyrazole and oxindole conjugates were synthesized by Knoevenagel condensation and investigated for their antiproliferative activity on different human cancer cell lines. The conjugates are comprised of a four ring scaffold; the structural isomers **12b** and **12c** possess chloro-substitution in the D ring. Among the congeners **12b**, **12c**, and **12d** manifested significant cytotoxicity and inhibited tubulin assembly. Treatments with **12b**, **12c** and **12d** resulted in accumulation of cells in G2/M phase, disruption of microtubule network, and increase in cyclin B1 protein. Zebrafish screening revealed that **12b**, and **12d** caused developmental defects. Docking analysis demonstrated that the congeners occupy the colchicine binding pocket of tubulin.

Keywords: Pyrazole-oxindole conjugates, tubulin depolymerization, zebrafish screening and molecular modeling.

Corresponding authors Tel.: +91-40-27193157; fax: +91-40-27193189, e-mail: <u>ahmedkamal@iict.res.in</u> (A. Kamal), <u>anthony@iict.res.in</u> (A. Addlagatta).

1. Introduction

Microtubules, composed of $\alpha\beta$ -tubulin heterodimers, are major constituents of the cytoskeleton in eukaryotic cells. The α - and β -tubulins are among the most highly conserved eukaryotic proteins [1]. Microtubules are pleiotropic in their function, particularly in organizing the spatial distribution of organelles in cells and chromosomes during cell division. Due to their essential functions in the cell, microtubules serve as an attractive drug target [2]. The dynamic equilibrium between microtubule polymerization and depolymerization is central to most of microtubule mediated functions including cell division [3]. This attribute has been exploited by numerous natural products and synthetic molecules which function as tubulin binding agents (TBAs) [4]. The microtubules posses three sites for ligand binding the vinca domain, colchicine domain and taxol domain. However, occurrence of peripheral neuropathy is a major limitation in development of antimicrotubule agents as drugs [5]. Therefore, discovery of novel molecules is required to overcome multi-drug resistance and neuropathies.

Oxindoles are important pharmacophores that are known to enhance anticancer activity of some established molecules. Sunitinib (5) and indirubin (6) are established anticancer agents containing oxindole as the basic scaffold (Figure 1A) [6-8]. Recently, it has been observed that chromone-pyrimidine, chromone-indolinone, chromone-pyrazole, indole-pyrimidine, indole-indolinone and indole-pyrazole conjugates demonstrated profound growth inhibitory activity against different cancer cells [9]. Moreover substituted indolin-2-ones are potential inhibitors of p90 ribosomal S6 protein kinase [10]. In addition, tri and tetra substituted pyrazole derivates proved to have potent anticancer action due to the inhibition of p38 α MAP kinase [11]. Whereas 4-arylazo-3,5-diamino-1H-pyrazoles function as novel group of ATP antagonists with moderate potency against CDK2-cyclin E complex [12]. The anticancer effects of some pyrazole amide derivatives are mediated by inhibition of the Elk-3 pathway and effects microtubules [13].

The unique feature of microtubule-binding agents, in contrast to other categories of anticancer drugs, is their incredible structural complexity and diversity, which provides many possibilities for optimization and new scaffold design. Combretastatin

(1) and colchicine (2) possess a two ring and three ring scaffolds with a trimethoxy phenyl group for anchorage, which efficiently inhibit tubulin polymerization. Based on the ring geometry, the substituted methoxylbenzoyl-aryl-thiazole (SMART) (3) series contain a basic three ring scaffold and functions as tubulin inhibitors [14]. Due to limited water solubility, the core thizaole ring was modified to imidazole to generate 2-aryl-4-benzoyl-imidazole series (ABI) (4) (Figure 1A) [15]. Nevertheless, the three ring (A, B and C) based scaffolds can be exploited further to generate better antitubulin molecules that could affect tubulin polymerization. Earlier we reported that oxindole derived imidazopyrazines exhibit significant anticancer activity [16]. In the present study, we performed the synthesis of (Z)-3-((3-phenyl-1H-pyrazol-5-yl) methylene) indolin-2-ones employing Knoevenagel condensation to generate a four ring scaffold (A: methoxy/methylene dioxy, B: pyrazole, C, D: oxindole). The congeners (**12a-12u**) were functionalized at A ring and D ring with various substituents such as methoxy, methylene dioxy, chloro, fluoro and nitro groups (Figure 1A).

<insert figure 1 here>

2. Results and discussion

2.1. Chemistry

The Synthesis of (Z)-3-((3-phenyl-1H-pyrazol5yl)methylene)indolin-2-one analogues 12 (a-u) described in the study are outlined in Scheme 1. The final step has been carried out by means of Knoevenagel condensation between an equimolar mixture of oxindole/indolinone and 3-substituted phenyl-1H-pyrazole-5-carbaldehydes 11 (a-d) in the presence of piperdine in ethanol [6], [9-10]. The key intermediates 3-substituted phenyl-1H-pyrazole-5-carbaldehydes 11 (a-d) was prepared in four sequential steps. Initially substituted acetophenones 7 (a-d) reacted with diethyl oxalate in the presence of sodium ethanolate in ethanol yielded ethyl 2,4-dioxo-4-(substituted phenyl)butanoates 8 (a-d). This was further cyclised with NH₂-NH₂.2HCl in ethanol to produce ethyl 3-substituted phenyl-1H-pyrazole-5-carboxylates 9 (a-d) in good yields [17-18]. The obtained carboxylates were reduced to (3-substitutedphenyl-1H-pyrazol-5yl) methanols 10 (a-d) by LiAlH₄. These were selectively oxidized to 3-substituted phenyl-1H-pyrazole-5-carbaldehydes 11(a-d) by IBX in DMSO.

<insert scheme 1 here>

3. 1. Evaluation of biological activity

3.1.1. Antiproliferative activity

The cell lines used were, HeLa (cervical carcinoma), A549 (non-small cell lung cancer), MCF-7 (breast cancer) and DU145 (prostate carcinoma) cell lines; the results are shown in Table 1. To determine the structure-activity relationships on the phenyl A ring and the oxindole D ring, we introduced various groups on both the rings. Among the compounds investigated 12b, 12c, and 12d showed promising anticancer activity against all the cell lines tested. Nocodazole was employed as standard drug. Interestingly the active compounds 12b and 12c are structural isomers with chloro substitutions at 5 and 6 of D ring with a mono methoxy at A ring. These structural isomers 12b and 12c exhibited IC₅₀ values of 6 μ M and 3 μ M respectively; with a maximal activity in HeLa cells (The IC_{50} value for each compound is the average of all four cancer cell lines). In contrast, **12d** (IC₅₀: 7.5 μ M) with a mono methoxy substitution A and D ring also exhibited strong activity in HeLa cells. The presence of bulkier groups in the A and D ring of 12k, (IC₅₀: 23.5 μ M) 12n (IC₅₀: 16.7 μ M) and 12s $(IC_{50}: 20.3 \mu M)$ decreased the activity of the molecules. The replacement of chloro with flouro in the compounds 12t and 12u in the D ring showed deleterious effect on the activity with IC₅₀ of 28.7 μ M and 33 μ M respectively. The compounds devoid of any substitutions (12a, 12g and 12h) in the ring D demonstrated better antiproliferative activity, when compared to 12e, 12l, 12o and 12p carrying an electron withdrawing group. Based on structural variation, the optimal order of substitutions on the A ring is mono methoxy > dimethoxy > 3, 4-(methylenedioxy) > trimethoxy. The compounds **12b**, (IC₅₀: 8.1 μ M), **12c** (IC₅₀: 5.9 μ M) and **12d** (IC₅₀: 9.2 μ M) displayed remarkable inhibition of tubulin polymerization, which positively correlated with antiproliferative activity (Table 2). As expected, Nocodazole significantly inhbited tubulin polymerization in our assays. Intriguingly, the compound 12b and 12c displayed differences in their binding interactions with tubulin due to the shift in chloro substitution. In comparison, the other analogues showed partial effect on tubulin polymerization. Therefore, results suggest that the presence of a monomethoxy group in A ring and chloro or methoxy group in D ring resulted in promising molecules, which exhibit potent anticancer activity and down regulate tubulin polymerization (Figure 1B) and their order of activities are 12c > 12b > 12d.

<insert table 1 here>

3.1.2. Effect on cell cycle arrest

To elucidate whether the cytoxicity induced by the derivatives was due to cell cycle arrest, we performed flow cytometry analysis for derivatives that exhibited potent cytotoxicity and significantly inhibited tubulin assembly. HeLa cells were treated with **12b**, **12c** and **12d** at 5 μ M and 10 μ M for 24 h. The cells accumulated in G2/M phase in a concentration dependent manner. Cells treated with **12c** at 10 μ M showed 79% arrest of cells in G2/M phase. In comparison, HeLa cells treated with **12b** or **12d** at 10 μ M resulted in an increase of mitotic cells by 60% and 72% respectively (Figure 2).

<insert figure 2 here>

3.1.3. Effect of 12b, 12c and 12d on microtubule network

Presence of aberrant spindle fibers due to altered microtubule dynamics is a hallmark of cells treated with antimitotic agents [19]. In order to ascertain if the cell cycle arrest is due to spindle abnormality, HeLa cells treated with 5 μ M concentrations of **12b**, **12c** and **12d** were stained with tubulin antibody. The control treated cells exhibit an organized network of microtubules. In contrast cells exposed with **12b**, **12c** showed multipolar spindle fibers, and **12d** treated cells exhibit a metaphase arrest (Figure 3).

<insert figure 3 here>

3.1.4. Effect of 12b, 12c and 12d on cellular tubulin polymerization

These observations suggest that the aberrant spindle dynamics in cells treated with **12b**, **12c** and **12d** results in cell cycle arrest. Tubulin assembly assays reveal that the congeners inhibit microtubule polymerization, consequently we analyzed for their effect on cellular tubulin. To elucidate this, HeLa cells were treated with **12b**, **12c** and

12d at 5 μ M concentrations for 24 h. Subsequently, cells were permeablized to collect the soluble fraction and remaining cells were collected as the polymerized fraction. Immunoblot analysis revealed that the cells exposed to pyrazole-oxindole conjugates contained more tubulin in the soluble fraction of cells. Therefore, increased tubulin in soluble fraction of cells treated with pyrazole-oxindole conjugates corroborated with the inhibition of tubulin assembly and arrest of cells in G2/M phase (Figure 4A). To further validate our observations that pyrazole-oxindole conjugates function as microtubule inhibitors, cells treated with the selected analogues demonstrated increased amount of cyclin B1 protein, a G2/M marker (Figure 4B).

<insert figures 4A and 4B here>

3.2. Effect of 12b, 12c and 12d on zebrafish development

Zebrafish (Danio rerio) has emerged as a powerful model system for chemical genetics [20-22]. In order to validate anti-proliferative effect of these compounds *in vivo*, their effects on development of zebrafish embryos was examined. Most of the compounds screened had little or no effect on development at 25 μ M. However, embryos treated with **12b** exhibited marked delay in development. Its isomer **12c** has a mild effect on the development of zebrafish embryos, highlighting possible differential ADME (absorption, distribution, metabolism, excretion) properties of the ring substitutes *in vivo*. Embryos treated with **12a** also showed delay in the development. This could be due to better absorption and/or bioavailability of the compound *in vivo* or due to additional and/or different target/s and needs further investigation [23]. Microtubule inhibitors like nocadozole also exhibit such developmental delays (Figure 5). These results suggest that **12b** acts as an anti-mitotic compound in zebrafish embryos and could thus be a potential therapeutic scaffold along with **12c** and **12d**.

<insert figure 5 here>

3.3. Molecular modeling

In order to examine the interaction of these conjugates in colchicine binding domain of tubulin, Autodock has been employed and three lead compounds (12b, 12c

and **12d**) have been docked against the tubulin structure (PDB code: 3E22) [24-27]. The colchicine site is generally buried in the intermediate domain of the β -subunit, surrounded by two β -strands, two helices and a loop, which we label as A-site. Colchicine also interacts with parts of the neighboring α -subunit, which is labeled as Bsite, which is wider than the A-site. The A-ring in the current study binds in the A-site of the tubulin while the C and D-rings are placed near the B-site in a slightly different environment compared to the colchicine. A strong hydrogen bond is noted between the carbonyl of the C-ring in the oxindole moiety. Lys254 from β -subunit and Asn101 of α -subunit are involved in placing the oxindole moiety between the two subunits. Although the trimethoxy benzene group is identified as signature for binding to tubulin, we noted that monomethoxy derivatives of our molecules inhibit the tubulin polymerization better than the di and trimethoxy derivatives. This could be due to the strong hydrogen bond with the oxindole moiety in B-site that may affect the binding of the methoxy-substituted benzene in the A-site. Also, within the monomethoxy substituted molecule, inhibition efficiency of tubulin polymerization varied. The C and D rings are buried at the interface of the two subunits of tubulin (Figure 4). Specifically, the D ring is sandwiched between the Leu248 and Lys352 of β -subunit and Ser178 of α -subunit. This region is narrow and hydrophobic which limits the binding of larger or charged group on the D ring. A chlorine atom at position six in D ring inhibits the tubulin polymerization better than when it is placed at position five. This data also corroborates that all the nitro substituted molecules are comparatively less active (Table 1).

<insert figure 6 here>

4. Conclusion

In summary, we synthesized new class of pyrazole-oxindole conjugates comprised of a four ring scaffold, which was extensively modified at A and D ring. Majority of the compounds exhibited significant antiproliferative activity where in the presence of a single chloro or methoxy group at D ring was necessary for potent antitubulin activity. The compounds **12b**, **12c** and **12d** stalled cells in G2/M phase and prevented chromosomal separation. Higher levels of Cyclin B1 protein and tubulin in the soluble fraction of cells excellently corroborated with antitubulin activity of the compounds. Zebrafish screening assays demonstrated the congeners affect the normal development of embryos. Molecular modeling analysis revealed that the compounds dock to the colchicine binding site of tubulin. Based on the results, the design and synthesis of the compounds containing pyrazole-oxindole moiety are effective tubulin polymerization inhibitors, which can be further amenable for generation of different conjugates as potential anticancer drugs.

5. Experimental protocols

5.1. General

All chemicals and reagents were obtained from Aldrich (Sigma–Aldrich, St. Louis, MO, USA), Lancaster (Alfa Aesar, Johnson Matthey Company, Ward Hill, MA, USA), or Spectrochem Pvt. Ltd. (Mumbai, India) and were used without further purification. Reactions were monitored by TLC performed on silica gel glass plates containing 60 GF-254, and visualization was achieved by UV light or iodine indicator. Column chromatography was performed with Merck 60–120 mesh silica gel. ¹H NMR spectra were recorded on Bruker UXNMR/XWIN-NMR (300 MHz) or Inova Varian-VXR-unity (400, 500 MHz) instruments. ¹³C NMR spectra were recorded on Bruker UXNMR/XWIN-NMR (d) are reported in ppm downfield froman internal TMS standard. ESI spectra were recorded on a Micro mass Quattro LC using ESI+ software with capillary voltage 3.98 kV and ESI mode positive ion trap detector. High-resolution mass spectra (HRMS) were recorded on a QSTAR XL Hybrid MS–MS mass spectrometer. Melting points were determined with an Electro thermal melting point apparatus, and are uncorrected.

5.1.1. Preparation of ethyl 2,4-dioxo-4-(substituted phenyl)butanoates 8(a-d)

Diethyl oxalate (1.0 mol) was added to freshly prepared sodium ethanolate at 0 °C after 10 minutes substituted acetophenones **7(a-d)** (1.0 mol) were added slowly in small portions maintaining the temperature 0 °C. After completion of addition the

stirring was continued at room temperature for 4 h. the reaction mixture was neutralized by diluted H_2SO_4 and extracted with ethyl acetate to offered solid products **8(a-d)** (yield 85-90%) which were taken as such for the next step without purification

5.1.2. Preparation of ethyl 3-substituted phenyl-1H-pyrazole-5-carboxylates 9(a-d)

To each ethyl 2, 4-dioxo-4-(substituted phenyl) butanoates 8(a-d) (1.0 mol) which were obtained in the earlier step was added Hydrazine dihydrochloride (NH₂-NH₂.2HCl) (1.5 mol) in ethanol and heated to reflux for 3 h. The solvent was removed under vacuum then added water to the residue and the compound was extracted with ethyl acetate (50 ml X 4). The organic layer was dried on anhydrous NaSO₄ and evaporated the solvent to obtain crude product that was further purified by column chromatography using ethyl acetate and hexane. The pure compounds 9(a-d) were eluted at 30-40% of ethyl acetate with good yields.

5.1.2.1. ethyl 3-(4-methoxyphenyl)-1H-pyrazole-5-carboxylate (9a):

yellow colored solid; (yield 80.0%): $R_f = 0.3$ (30% ethyl acetate/hexane); ¹H NMR (300MHz, CDCl₃); δ 1.23-1.37 (t, 3H, J_1 =6.7 Hz, J_2 =7.5Hz, -CH₃), 3.81 (s, 3H, - OCH₃), 4.17-4.36 (q, 2H, J_1 =6.7 Hz, J_2 =7.5Hz, CH₂), 6.8 (s, 1H, ArH), 6.90 (d, 2H, J = 2.2Hz, ArH), 7.62 (d, 2H, J = 9.0Hz, ArH) ppm; MS (ESI) *m*/*z* 247[M+H].

5.1.2.2 ethyl 3-(3,4-dimethoxyphenyl)-1H-pyrazole-5-carboxylate 9(b):

Pale yellow colored solid; (yield 80.0%): $R_f = 0.3$ (30% ethyl acetate/hexane); ¹H NMR (300MHz, CDCl₃); δ 1.23-1.31 (t, 3H, J_1 =7.1 Hz, -CH₃), 3.85 (s, 3H, -OCH₃), 3.90 (s, 3H, -OCH₃) 4.19-4.32 (q, 2H, J_1 =7.1 Hz, -CH₂), 6.88 (d, 1H, J_1 =7.1 Hz, -ArH), 6.94 (s, 1H, ArH), 7.21-7.28 (m, 1H, ArH) 7.29-7.34 (m, 1H, ArH) 9.67 (brs, 1H, -NH) ppm; MS (ESI) m/z 277[M+H].

5.1.2.3 ethyl 3-(3,4,5-trimethoxyphenyl)-1H-pyrazole-5-carboxylate9(c):

Yellow colored solid; (yield 75.0%): $R_f = 0.3$ (40% ethyl acetate/hexane); ¹H NMR (300MHz, CDCl₃); δ 1.33-1.40 (t, 3H, J_1 =6.7 Hz, J_2 =7.5Hz, -CH₃), 3.88 (s, 3H, -OCH₃), 3.95 (s, 6H, -OCH₃) 4.33-4.45 (q, 2H, J_1 =6.7 Hz, J_2 =7.5Hz, CH₂), 7.01 (s, 2H, ArH), 7.05 (s, 1H, ArH) ppm; MS (ESI) *m/z* 307[M+H].

5.1.2.4 ethyl 3-(benzo[d][1,3]dioxol-5-yl)-1H-pyrazole-5-carboxylate9(d):

Pale yellow colored solid; (yield 75.0%): $R_f = 0.3$ (40% ethyl acetate/hexane); ¹H NMR (500MHz, CDCl₃); δ 1.33-1.40 (t, 3H, J_1 =6.7 Hz, J_2 =7.5Hz, -CH₃), 4.33-4.45 (q, 2H, J_1 =6.7 Hz, J_2 =7.5Hz, CH₂), 6.0 (s, 2H, OCH₂O), 6.86 (d, 1H, J = 8.3Hz, ArH)7.05 (s, 1H, ArH), 7.21-7.28 (m, 2H, ArH) ppm; MS (ESI) *m*/*z* 261[M+H].

5.1.3. Preparation of (3-substitutedphenyl-1H-pyrazol-5-yl)methanols 10(a-d):

To the ethyl 3-substituted phenyl-1H-pyrazole-5-carboxylates 9(a-d), obtained in the above step was added LiAH₄ (0.5 mol) in dry THF at 0 °C and stirred for 1h at room temperature. Added saturated NH₄Cl solution drop wise to quench the unreacted LiAlH₄ and removed the THF under vacuum then extracted with ethyl acetate (100 ml X 4). The organic layer was dried on anhydrous NaSO₄ and evapourated ethyl acetate to obtain color less solid products of (3-substitutedphenyl-1H-pyrazol-5-yl)methanols 10(a-d)(yield 70-80%). The alcohols produced in this step were pure, and no further purification was required. These compounds were taken as such for the next step.

5.1.4. Preparation of 3-subtitutedphenyl-1H-pyrazole-5-carbaldehydes 11(a-d):

To the (3-substitutedphenyl-1H-pyrazol-5-yl)methanols 10(a-d) produced in the above step was added IBX (1.2 mol) in DMSO and stirred for 1 h at room temperature. Added ice cold water to the reaction mixture and extracted with ethyl acetate(50 ml X 4). The organic layer was dried on anhydrous NaSO₄ and evaporated the ethyl acetate to obtain pure corresponding 3-subtitutedphenyl-1H-pyrazole-5-carbaldehydes 11(a-d) in good yields (80-85%). The obtained carbaldehydes were as such taken in the next step for the synthesis of pyrazole-oxindole conjugates 12a-12u.

5.1.4.1. 3-(4-methoxyphenyl)-1H-pyrazole-5-carbaldehyde (11a):

3-(4-methoxyphenyl)-1H-pyrazole-5-carbaldehyde **11a** was prepared using above method by the addition of (3-(4-methoxyphenyl)-1H-pyrazol-5-yl)methanol **10a** (2.04g. 10 mmol) IBX (3.36g 1.2 mmol). Yellow colored solid; (1.71g. yield 85%): $R_f = 0.3$ (40% ethyl acetate/hexane); ¹H NMR (400MHz, CDCl₃); δ 3.85 (s, 3H, -OCH₃), 7.78-7.82 (m, 1H, ArH), 7.86-7.93 (m, 2H, ArH), 7.95-8.03 (m, 2H, ArH) 9.95 (s, 1H, CHO) ppm; MS (ESI) *m/z* 203[M+H].

5.1.4.2.3-(3, 4-dimethoxyphenyl)-1H-pyrazole-5-carbaldehyde (11b):

3-(3, 4-dimethoxyphenyl)-1H-pyrazole-5-carbaldehyde **11b** was prepared using above method by the addition of (3-(3,4-dimethoxyphenyl)-1H-pyrazol-5-yl)methanol **10b** (2.34g. 10 mmol) IBX (3.36g 1.2 mmol). Yellow colored solid; (1.85g. yield 82%): $R_f = 0.4$ (40% ethyl acetate/hexane); ¹H NMR (300MHz,CDCl₃); δ ppm; δ 3.87 (s, 3H, -OCH₃), 3.91 (s, 3H, -OCH₃), 6.93-7.04 (m, 1H, ArH), 7.27-7.47 (m, 2H, ArH), 7.86-8.10 (m, 1H, ArH) 9.93 (s, 1H, CHO) ppm; MS (ESI) *m/z* 233[M+H].

5.1.4.3.3-(3, 4, 5-trimethoxyphenyl)-1H-pyrazole-5-carbaldehyde (11c):

3-(3, 4, 5-trimethoxyphenyl)-1H-pyrazole-5-carbaldehyde **11c** was prepared using above method by the addition of (3-(3,4,5-trimethoxyphenyl)-1H-pyrazol-5-yl)methanol **10c** (2.64g. 10 mmol) IBX (3.36g 1.2 mmol). Yellow colored solid; (2.09g. yield 80%): $R_f = 0.3$ (40% ethyl acetate/hexane); ¹H NMR (400MHz, CDCl₃); δ 3.82 (s, 3H, -OCH₃), 3.92 (s, 6H, -OCH₃), 6.85-7.25 (m, 3H, ArH), 9.96 (s, 1H, CHO) ppm; MS (ESI) *m*/*z* 263[M+H].

5.1.4.4.3-(benzo[d][1,3]dioxol-5-yl)-1H-pyrazole-5-carbaldehyde (11d):

3-(benzo[d][1,3]dioxol-5-yl)-1H-pyrazole-5-carbaldehyde **11d** was prepared using above method by the addition of (3-(benzo[d][1,3]dioxol-5-yl)-1H-pyrazol-5-yl)methanol **10d** (2.18g. 10 mmol) IBX (3.36g 1.2 mmol). Yellow colored solid; (1.83g. yield 85%): $R_f = 0.3$ (40% ethyl acetate/hexane); ¹H NMR (300MHz,CDCl₃); δ 6.03 (s, 2H, -OCH₂O), 7.35-7.45 (m, 1H, ArH), 7.87-7.94 (m, 2H, ArH), 8.10-8.17 (m, 1H, ArH) 10.2 (s, 1H, CHO ppm; MS (ESI) *m/z* 217[M+H].

5.1.5. Synthesis of (Z)-3-((3-phenyl-1H-pyrazol-5-yl)methylene)indolin-2-one analogues **12(a-u**):

To the 3-subtituted phenyl-1H-pyrazole-5-carbaldehydes 11(a-d) prepared in the above step was added corresponding substituted oxindoles and catalytic amount of piperdine (1.0 ml) in ethanol. Heated the reaction mixture to reflux for 4 h at 85 °C. The solid compounds obtained in the reaction vessel were filtered and washed with ethanol for 4-5 times. After complete air drying the final compounds (Z)-3-((3-phenyl-1H-pyrazol-5-yl)methylene)indolin-2-one analogues 12(a-u) were obtained as pure solids (yield 75-80%).

5.1.5. 1. (Z)-3-((3-(4-methoxyphenyl)-1H-pyrazol-5-yl)methylene)indolin-2-one(12a):

This compound was prepared using the procedure described above by the addition of 3-(4-methoxyphenyl)-1H-pyrazole-5-carbaldehyde **11a** (150 mg 0.0495 mmol) and indolin-2-one (0.065 g 0.0495 mmol). The compound obtained as saffron colored solid Yield: 117 mg (75%); mp: 190-192 °C; ¹H NMR (300MHz, DMSO-d₆); δ 3.88 (s, 3H, - OCH₃), 7.05-7.09 (m, 4H, ArH), 7.19-7.26 (m, 1H, ArH), 7.42-7.49 (t, 1H, J_1 = 6.9Hz, J_2 = 7.7Hz, ArH), 7.50 (s, 1H, -CH), 7.62 (d, 1H, J = 7.5Hz, ArH) 7.68 (d, 1H, J = 8.8Hz, ArH) 9.42 (brs, 2H, -NH); ¹³C NMR (75 MHz, DMSO-d₆): δ 55.0, 109.3, 114.1, 114.3, 119.9, 120.9,125.3, 126.5, 126.6, 129.1, 129.6,140.3, 142.5, 159.0, 159.3, 169.3 ppm; IR (KBr) (v_{max} /cm⁻¹): v = 3136, 3058, 3003, 2963, 2832, 1897, 1673, 1598, 1516, 1344, 1252, 1204, 809 cm⁻¹; MS (ESI) *m*/*z* 318[M+H]; HR-MS (ESI) *m*/*z* for C₁₉H₁₆N₃O₂ calculated *m*/*z*: 318.1237, found *m*/*z*: 318.1227.

5.1.5.2. (Z)-5-chloro-3-((3-(4-methoxyphenyl)-1H-pyrazol-5-yl)methylene)indolin-2one(**12b**):

This compound was prepared using the procedure described above by the addition of 3-(4-methoxyphenyl)-1H-pyrazole-5-carbaldehyde **11a** (150 mg 0.0495 mmol) and 5chloroindolin-2-one (82.9 mg 0.0495 mmol). The compound obtained as yellow colored solid Yield: 135 mg (78%); mp: 196-198 °C; ¹H NMR (300MHz, DMSO-d₆); δ 3.85 (s, 3H, -OCH₃), 6.85-6.95 (m, 1H, ArH), 7.00 (t, 2H, *J* = 9.0Hz, ArH), 7.24 (d, 1H, *J*₁ = 6.9Hz, *J*₂ = 7.7Hz, ArH), 7.50 (s, 1H, ArH), 7.62 (d, 1H, *J* = 7.5Hz, ArH) 7.68 (d, 1H, *J* = 8.1Hz, ArH) 7.55 (s, 1H, ArH), 7.72-7.88 (m, 3H, ArH), 8.1-8.63 (m, 1H, ArH), 9.16 (brs, 1H, -NH) 10.6 (brs, 1H, -NH); ¹³C NMR (75 MHz, DMSO-d₆): δ 55.1, 107.5, 110.5, 114.1, 114.4, 121.0, 123.4, 124.2, 125.0, 125.7, 126.6,126.9, 129.0, 141.2, 159.0, 169.3 ppm; IR (KBr) (v_{max}/cm⁻¹): v = 3167, 3055, 2933, 1674, 1600, 1516, 1328, 1201, 1168, 1109, 949 cm⁻¹; MS (ESI) *m*/*z* 352[M+H]; HR-MS (ESI) *m*/*z* for C₁₉H₁₅O₂N₃ calculated *m*/*z*: 352.0847, found *m*/*z*: 352.0841.

5.1.5.3. (Z)-6-chloro-3-((3-(4-methoxyphenyl)-1H-pyrazol-5-yl)methylene)indolin-2one(**12c**):

This compound was prepared using the procedure described above by the addition of 3- (4-methoxyphenyl)-1H-pyrazole-5-carbaldehyde **11a** (150 mg 0.0495 mmol) and 6- chloroindolin-2-one (82.9 mg 0.0495 mmol). The compound obtained as pale yellow colored solid Yield: 132 mg (76%); mp: 190-192 °C; ¹H NMR (300 MHz, DMSO-d₆);

δ 3.83 (s, 3H, -OCH₃), 6.83-7.08 (m, 4H, ArH), 7.59 (d, 1H, J = 8.1Hz, ArH), 7.65 (s, 1H, ArH), 7.72 (d, 1H, J = 8.1Hz, ArH), 7.78 (d, 1H, J = 8.4Hz, ArH) 7.91-7.96 (m, 1H, ArH), 9.03 (brs, 1H, -NH)10.51(brs, 1H, -NH); ¹³C NMR (75 MHz, DMSO-d₆): δ 55.1, 109.3, 114.1, 114.4, 120.6, 126.28, 126.5, 126.6, 127.4, 133.7, 143.8, 159.3, 169.3 ppm; IR (KBr) (v_{max} /cm⁻¹): v = 3155, 3027, 2966, 2933, 2836, 1676, 1593, 1515, 1455, 1344, 1251, 1205, 1175, 1063, 1025 cm⁻¹; MS (ESI) *m/z* 352[M+H]; HR-MS (ESI) *m/z* for C₁₉H₁₅O₂N₃ calculated *m/z*: 352.0847, found *m/z*: 352.0844.

5.1.5.4. (Z)-5-methoxy-3-((3-(4-methoxyphenyl)-1H-pyrazol-5-yl)methylene)indolin-2one(**12d**):

This compound was prepared using the procedure described above by the addition of 3-(4-methoxyphenyl)-1H-pyrazole-5-carbaldehyde **11a** (150 mg 0.0495 mmol) and 5methoxyindolin-2-one (80 mg 0.0495 mmol). The compound obtained as saffron colored solid Yield: 137 mg (80%); mp: 208-210 °C; ¹H NMR (300MHz, DMSO-d₆); δ 3.87 (s, 3H, -OCH₃), 3.79 (s, 3H, -OCH₃), 6.73-6.79 (m, 2H, ArH), 7.01 (d, 2H, *J* = 8.4Hz, ArH), 7.06-7.30 (m, 1H, ArH), 7.42 (s, 1H, =CH), 7.69-7.83 (m, 2H, ArH) 7.85 (s, 1H, ArH), 8.82 (brs, 1H, -NH) 10.9 (brs, 1H, -NH); ¹³C NMR (75 MHz, DMSO-d₆): δ 55.2, 55.5, 105.9, 109.4, 114.2, 114.4, 122.7, 125.9, 126.7, 126.6, 134.0, 136.1, 154.4, 155.0, 159.1, 159.4, 169.5 ppm; IR (KBr) (v_{max}/cm⁻¹): v = 3448, 3120, 3069, 2995, 2956, 2828, 1677, 1610, 1516, 1337, 1247, 1207, 1167, cm⁻¹; MS (ESI) *m*/*z* 352[M+H]; HR-MS (ESI) *m*/*z* for C₁₉H₁₅O₂N₃ calculated *m*/*z*: 352.0847, found *m*/*z*: 352.0844.

5.1.5.5. (Z)-3-((3-(4-methoxyphenyl)-1H-pyrazol-5-yl)methylene)-5-nitroindolin-2one(**12e**):

This compound was prepared using the procedure described above by the addition of 3-(4-methoxyphenyl)-1H-pyrazole-5-carbaldehyde **11a** (150 mg 0.0495 mmol) and 5-nitroindolin-2-one (88 mg 0.0495 mmol). The compound obtained as brown red colored solid Yield: 138 mg (77%); mp: 218-220 °C;

¹H NMR (300MHz, DMSO-d₆); δ 3.80 (s, 3H, -OCH₃), 7.01-7.11 (m, 3H, ArH), 7.20 (s, 1H, ArH), 7.63 (s, 1H, =CH), 7.78 (d, 1H, *J* = 8.6Hz, ArH), 8.17-8.28 (m, 2H, ArH) 10.04 (brs, 1H, -NH); ¹³C NMR (75 MHz, DMSO-d₆): δ 55.2, 107.9, 109.4, 114.5, 121.7, 121, 122.1, 123.1, 126.3, 126.8, 128.6, 142.0, 148.1, 159.5, 169.8 ppm; IR (KBr)

 $(v_{max}/cm^{-1}): v = 3293, 3142, 2932, 2834, 1697, 1613, 1518, 1332, 1250, 1178, 1125 cm^{-1};$ MS (ESI) *m/z* 363[M+H]; HR-MS (ESI) *m/z* for C₁₉H₁₅O₄ N₄ calculated *m/z*: 363.1087, found *m/z*: 363.1082.

5.1.5.6.(*Z*)-5-chloro-3-((3-(3,4,5-trimethoxyphenyl)-1H-pyrazol-5-l)methylene)indolin-2-one(**12***f*):

This compound was prepared using the procedure described above by the addition of 3-(3, 4, 5-trimethoxyphenyl)-1H-pyrazole-5-carbaldehyde **11c** (150 mg 0.0572 mmol) and 5-chloroindolin-2-one (95 mg 0.0572 mmol). The compound obtained as yellow colored crystal Yield: 192 mg (82%); mp: 198-200 °C; ¹H NMR (400MHz, DMSO-d₆); δ 3.69 (s, 3H, -OCH₃), 3.86 (s, 6H, -OCH₃), 6.90 (d, 1H, *J*= 8.3Hz, ArH), 7.17 (s, 2H, =CH, ArH), 7.27 (s, 1H, ArH), 7.30-7.39 (m, 1H, ArH), 7.50 (s, 1H, ArH) 7.86-7.96 (m, 1H ArH) 9.18 (brs, 1H, -NH) 10.71 (brs, 1H, -NH); ¹³C NMR (75 MHz, DMSO-d₆): δ 55.9, 60.1, 102.9, 108.3, 110.7, 120.1, 123.4, 124.4, 125.2, 126.9, 126. 129.2, 137.7, 141.2, 153.3, 169.2 ppm; IR (KBr) (v_{max}/cm⁻¹): v = 3115, 2947, 2832, 1683, 1612, 1587, 1545, 1513, 1463, 1427, 1377, 1346, 1237, 1178, 1132, 1070 cm⁻¹; MS (ESI) *m/z* 412[M+H]; HR-MS (ESI) *m/z* for C₂₁H₁₉O₄ N₃Cl calculated *m/z*: 412.1058, found *m/z*: 412.1044.

5.1.5.7.(*Z*)-3-((3-(3,4,5-trimethoxyphenyl)-1H-pyrazol-5-yl)methylene)indolin-2one(**12g**):

This compound was prepared using the procedure described above by the addition of 3-(3, 4, 5-trimethoxyphenyl)-1H-pyrazole-5-carbaldehyde **11c** (150 mg 0.0572 mmol) and indolin-2-one (76 mg 0.0572 mmol). The compound obtained as brown colored solid Yield: 168 mg (78%); mp: 208-210 °C; ¹H NMR (300 MHz, DMSO-d₆); δ 3.72 (s, 3H, -OCH₃), 3.88 (s, 6H, -OCH₃), 6.86-7.01 (m, 1H, ArH), 7.15 (s, 2H, =CH, ArH), 7.22-7.32 (m, 1H, ArH), 7.47 (s, 1H, ArH), 7.67-7.82 (t, 1H, *J* = 7.5Hz, ArH) 8.23 (m, 1H ArH) 9.18 (s, 1H, -ArH) 10.54 (brs, 1H, -NH) 11.12 (brs, 1H, -NH); ¹³C NMR (75 MHz, DMSO-d₆): δ 54.2, 58.2, 100.9, 101.5, 105.9, 107.6, 108.2, 118.1, 119.1, 119.8, 123.8, 124.2, 127.3, 127.8, 138.5, 140.7, 151.3, 151.5, 166.7, 167.7 ppm; IR (KBr) (v_{max}/cm⁻¹): v = 3114, 3054, 2935, 2827, 1677, 1600, 1544, 1513, 1471, 1349, 1127, 1075, 1009 cm⁻¹; MS (ESI) *m*/*z* 378[M+H]; HR-MS (ESI) *m*/*z* for C₂₁H₂₀O₄ N₃ calculated *m*/*z*: 378.14483, found *m*/*z*: 378.1441. 5.1.5.8. (Z)-3-((3-(3,4-dimethoxyphenyl)-1H-pyrazol-5-yl)methylene)indolin-2one(**12h**):

This compound was prepared using the procedure described above by the addition of 3-(3, 4-dimethoxyphenyl)-1H-pyrazole-5-carbaldehyde **11b** (150 mg 0.0659 mmol) and indolin-2-one (86 mg 0.0659 mmol). The compound obtained as yellow colored solid Yield: 182 mg (80%); mp: 196-198 °C; ¹H NMR (300 MHz, DMSO-d₆); δ 3.82 (s, 3H, -OCH₃), 3.87 (s, 3H, -OCH₃), 6.87-7.13 (m, 3H, ArH), 7.25 (s, 1H, =CH), 7.34-7.46 (m, 2H, ArH), 7.73 (t, 1H, *J* = 7.5Hz, ArH), 8.24 (s, 1H ArH) 9.03 (s, 1H, -NH) 11.09 (brs, 1H, -NH); ¹³C NMR (75 MHz, DMSO-d₆): δ 55.5, 55.6, 108.7, 108.9, 109.4, 111.8, 111.9, 129.21, 129.6, 148.7, 148.9, 149.0, 169.6 ppm; IR (KBr) (v_{max}/cm⁻¹): v = 3163, 3024, 2836, 1678, 1604, 1516, 1471, 1423, 1380, 1348, 1308, 1266, 1228, 1201, 1153, 1020 cm⁻¹; MS (ESI) *m*/z 348 [M+H]; HR-MS (ESI) *m*/z for C₂₀H₁₈O₃ N₃ calculated *m*/z: 348.13427, found *m*/z: 348.13318.

5.1.5.9. (Z)-5-chloro-3-((3-(3,4-dimethoxyphenyl)-1H-pyrazol-5-yl)methylene)indolin-2-one(**12i**):

This compound was prepared using the procedure described above by the addition of 3-(3, 4-dimethoxyphenyl)-1H-pyrazole-5-carbaldehyde **11b** (150 mg 0.0659 mmol) and 5-chloroindolin-2-one (110 mg 0.0659 mmol). The compound obtained as yellow colored solid Yield: 193 mg (77%); mp: 198-199 °C; ¹H NMR (500 MHz, DMSO-d₆); δ 3.80 (s, 3H, -OCH₃), 3.84 (s, 3H, -OCH₃), 6.88 (d, 1H, *J* = 8.3Hz, ArH), 7.01 (d, 1H, *J* = 8.4Hz, ArH), 7.19 (s, 1H, =CH), 7.26-7.42 (m, 2H, ArH), 7.46 (s, 1H, ArH), 7.52 (s, 1H ArH), 7.85-7.99 (m, 1H, ArH), 9.21 (brs, 1H, -NH) 10.7 (brs, 1H, -NH); ¹³C NMR (75 MHz, DMSO-d₆): δ 55.5, 55.6, 107.7, 108.7, 108.9,110.6, 117.9, 112, 124.2, 125.1, 125.8, 127.0, 129.1, 141.2, 143.5, 147.3, 148.9, 149, 169.2 ppm; IR (KBr) (v_{max}/cm⁻¹): v = 3118, 2955, 2829, 1680, 1609, 1522, 1437, 1382, 1343, 1263, 1234, 1157, 1064, 1026 cm⁻¹; MS (ESI) *m*/*z* 348 [M+H]; HR-MS (ESI) *m*/*z* for C₂₀H₁₇O₃ N₃Cl calculated *m*/*z*: 382.0953, found *m*/*z*: 382.0950.

5.1.5.10. (Z)-6-chloro-3-((3-(3,4-dimethoxyphenyl)-1H-pyrazol-5-yl)methylene)indolin-2-one(**12***j*):

This compound was prepared using the procedure described above by the addition of 3-(3, 4-dimethoxyphenyl)-1H-pyrazole-5-carbaldehyde **11b** (150 mg 0.0659 mmol) and

6-chloroindolin-2-one (110 mg 0.0659 mmol). The compound obtained as yellow colored solid Yield: 191 mg (76%); mp: 196-198 °C; ¹H NMR (500 MHz, DMSO-d₆); δ 3.80 (s, 3H, -OCH₃), 3.85 (s, 3H, -OCH₃), 6.88 (m, 1H, ArH), 7.0-7.14 (m, 2H, ArH), 7.17 (s, 1H, =CH), 7.36 (d, 1H, J = 8.1 Hz ArH), 7.42 (s, 1H, ArH), 7.48 (s, 1H ArH), 7.78-7.88 (m, 1H, ArH), 9.1 (brs, 1H, -NH) 10.7 (brs, 1H, -NH); ¹³C NMR (75 MHz, DMSO-d₆): δ 55.4, 55.5, 107.2, 109.0, 109.3, 112.0, 120.5, 124.0, 126.2, 127.4, 133.6, 143.4, 143.8, 147.2, 148.9, 169.3 ppm; IR (KBr) (v_{max} /cm⁻¹): v = 3120, 2951, 2829, 1680, 1605, 1522, 1437, 1382, 1343, 1250, 1234, 1157, 1064, 1026 cm⁻¹; MS (ESI) *m*/*z* 348 [M+H]; HR-MS (ESI) *m*/*z* for C₂₀H₁₇O₃ N₃Cl calculated *m*/*z*: 382.0950.

5.1.5.11. (Z)-3-((3-(3,4-dimethoxyphenyl)-1H-pyrazol-5-yl)methylene)-5methoxyindolin-2-one(**12k**):

This compound was prepared using the procedure described above by the addition of 3-(3, 4-dimethoxyphenyl)-1H-pyrazole-5-carbaldehyde **11b** (150 mg 0.0659 mmol) and 5-methoxyindolin-2-one (107 mg 0.0659 mmol). The compound obtained as saffron colored solid Yield: 196 mg (79%); mp: 192-195 °C; ¹H NMR (300 MHz, DMSO-d₆); δ 3.77 (s, 3H, -OCH₃), 3.79 (s, 3H, -OCH₃), 3.84 (s, 3H, -OCH₃), 6.70-6.93 (m, 1H, Ar**H**), 6.98-7.20 (m, 2H, =C**H**, Ar**H**), 7.31-7.52 (m, 3H, Ar**H**), 7.85 (s, 1H, Ar**H**), 8.84 (s, 1H, Ar**H**), 10.3 (brs, 1H, -N**H**) 10.9 (brs, 1H, -N**H**); ¹³C NMR (75 MHz, DMSOd₆): δ 55.4, 105.9, 107.2, 109.9, 108.9, 108.7, 111.9, 112.0, 114.5, 117.8, 122.7, 125.9, 134.11, 136.2, 149.0, 154.4, 155.0, 169.5 ppm; IR (KBr) (v_{max}/cm⁻¹): v = 3121, 3000, 2912, 2839, 1676, 1610, 1517, 1479, 1435, 1381, 1336, 1302, 1274, 1205, 1230, 1165, 1131, 1062 cm⁻¹; MS (ESI) *m*/*z* 378 [M+H]; HR-MS (ESI) *m*/*z* for C₂₁H₂₀O₄ N₃ calculated *m*/*z*: 378.14483, found *m*/*z*: 378.14395.

5.1.5.12. (Z)-3-((3-(3,4-dimethoxyphenyl)-1H-pyrazol-5-yl)methylene)-5-nitroindolin-2-one(**12l**):

This compound was prepared using the procedure described above by the addition of 3-(3, 4-dimethoxyphenyl)-1H-pyrazole-5-carbaldehyde **11b** (150 mg 0.0659 mmol) and 5-nitroindolin-2-one (117 mg 0.0659 mmol). The compound obtained as yellow colored solid Yield: 193 mg (75%); mp: 174-176 °C; ¹H NMR (300 MHz, DMSO-d₆); δ 3.80 (s, 3H, -OCH₃), 3.86 (s, 3H, -OCH₃), 7.06 (d, 3H, *J* = 8.6 Hz ArH), 7.24 (s, 1H, =CH), 7.40 (d, 1H, J = 8.3 Hz Ar**H**), 7.45 (s, 1H, Ar**H**), 7.63 (s, 1H Ar**H**), 8.12-8.34 (m, 1H, Ar**H**), 10.07 (brs, 1H, -N**H**) 11.2 (brs, 1H, -N**H**); ¹³C NMR (75 MHz, DMSO-d₆): δ 55.4, 55.5, 108.9, 112.0, 118.0, 120.9, 121.9, 122.1, 123.1, 126.3, 128.6, 141.9, 143.7, 147.1, 148.1, 149.0, 169.7 ppm; IR (KBr) (ν_{max} /cm⁻¹): $\nu = 3166$, 3011, 2908, 1689, 1605, 1523, 1467, 1434, 1341, 1385, 1263, 1240, 1197, 1161, 1141, 1076, 1029 cm⁻¹; MS (ESI) *m*/*z* 393 [M+H]; HR-MS (ESI) *m*/*z* for C₂₀H₁₇O₅ N₄ calculated *m*/*z*: 393.11935, found *m*/*z*: 393.11945.

5.1.5.13. (*Z*)-6-chloro-3-((3-(3,4,5-trimethoxyphenyl)-1H-pyrazol-5-yl)methylene)indolin-2-one(**12m**):

This compound was prepared using the procedure described above by the addition of 3-(3, 4, 5-trimethoxyphenyl)-1H-pyrazole-5-carbaldehyde **11c** (150 mg 0.0572 mmol) and 6-chloroindolin-2-one (95 mg 0.0572 mmol). The compound obtained as yellow colored solid Yield: 183 mg (75%); mp: 184-186 °C; ¹H NMR (500 MHz, DMSO-d₆); δ 3.69 (s, 3H, -OCH₃), 3.84 (s, 6H, -OCH₃), 6.82-6.99 (m, 1H, ArH) 7.03-7.12 (m, 1H, ArH), 7.15(s, 1H, =CH), 7.25 (s, 1H, ArH), 7.48 (s, 1H, ArH) 7.70-7.94 (m, 1H, ArH), 9.09 (d, 1H, *J* = 8.3 Hz ArH), 10.7 (brs, 1H, -NH) 11.2 (brs, 1H, -NH); ¹³C NMR (75 MHz, DMSO-d₆): δ 55.8, 56.0, 60.0, 102.6, 102.8, 108.0, 109.3, 110.0, 120.6, 121.5, 121.7, 124.1, 124.3, 126.2, 127.5, 128.3, 133.7, 139.2, 141.4, 143.4, 147.43, 151.5, 153.3, 153.1, 168.6, 169.4 ppm; IR (KBr) (v_{max}/cm⁻¹): v = 3115, 2947, 2832, 1683, 1612, 1587, 1545, 1513, 1463, 1427, 1377, 1346, 1236, 1179, 1132, 1070, 1039 cm⁻¹; MS (ESI) *m/z* 412 [M+H]; HR-MS (ESI) *m/z* for C₂₁H₁₉O₄ N₃Cl calculated *m/z*: 412.1058, found *m/z*: 412.1065.

5.1.5.14. (Z)-5-methoxy-3-((3-(3,4,5-trimethoxyphenyl)-1H-pyrazol-5yl)methylene)indolin-2-one(**12n**):

This compound was prepared using the procedure described above by the addition of 3-(3, 4, 5-trimethoxyphenyl)-1H-pyrazole-5-carbaldehyde **11c** (150 mg 0.0572 mmol) and 5-methoxyindolin-2-one (93 mg 0.0572 mmol). The compound obtained as saffron colored solid Yield: 195 mg (81%); mp: 208-210 °C; ¹H NMR (300 MHz, DMSO-d₆); δ 3.70, (s, 3H, -OCH₃), 3.77 (s, 3H, -OCH₃), 3.87 (s, 6H, -OCH₃), 6.72-6.94 (m, 1H, ArH) 7.14 (s, 1H, ArH), 7.16(s, 1H, =CH), 7.43 (s, 1H, ArH), 7.85 (s, 1H, ArH) 10.8 (brs, 1H, -NH); ¹³C NMR (75 MHz, DMSO-d₆): δ 55.5, 56.0, 60.0, 102.7, 102.9, 106.0, 109.4, 114.6, 115.1, 122.5, 126.0, 125.9, 134.0, 136.2, 137.6, 153.1, 154.3, 154.9, 169.4 ppm; IR (KBr) (v_{max} /cm⁻¹): v = 3121, 2942, 2837, 1678, 1592, 1544, 1511, 1481, 1430, 1383, 1339, 1302, 1269, 1238, 1201, 1171, 1124, 1085 cm⁻¹; MS (ESI) *m*/*z* 408 [M+H]; HR-MS (ESI) *m*/*z* for C₂₂H₂₂O₅ N₃ calculated *m*/*z*: 408.15540, found *m*/*z*: 408.15567.

5.1.5.15. (Z)-5-nitro-3-((3-(3,4,5-trimethoxyphenyl)-1H-pyrazol-5yl)methylene)indolin-2-one(**12o**):

This compound was prepared using the procedure described above by the addition of 3-(3, 4, 5-trimethoxyphenyl)-1H-pyrazole-5-carbaldehyde **11c** (150 mg 0.0572 mmol) and 5-nitroindolin-2-one (101 mg 0.0572 mmol). The compound obtained as light yellow colored solid Yield: 190 mg (79%); mp: 203-205 °C;

¹H NMR (300 MHz, DMSO-d₆); δ 3.86 (s, 3H, -OCH₃), 6.69-6.94 (m, 2H, ArH) 7.14 (d, 1H, ArH), 7.25 (s, 1H, =CH), 7.43 (s, 1H, ArH), 7.85 (s, 1H, ArH) 10.8 (brs, 1H, -NH) 14.45 (brs, 1H, -NH); ¹³C NMR (75 MHz, DMSO-d₆): δ 56.0, 60.0, 102.7, 102.9, 108.7, 109.2, 121.7, 122.0, 123.2, 123.7, 126.2, 128.3, 137.7, 141.9, 143.7, 147.0, 148.1, 153.2, 169.6 ppm; IR (KBr) (v_{max}/cm⁻¹): v = 3121, 2942, 2837, 1678, 1592, 1544, 1511, 1481, 1430, 1383, 1339, 1302, 1269, 1238, 1201, 1171, 1124, 1085 cm⁻¹; MS (ESI) *m*/*z* 423 [M+H]; HR-MS (ESI) *m*/*z* for C₂₁H₁₉O₅ N₄ calculated *m*/*z*: 423.12230, found *m*/*z*: 423.12240.

5.1.5.15. (Z)-3-((3-(benzo[d][1,3]dioxol-5-yl)-1H-pyrazol-5-yl)methylene)-5nitroindolin-2-one(**12**p):

This compound was prepared using the procedure described above by the addition of 3-(benzo[d][1,3]dioxol-5-yl)-1H-pyrazole-5-carbaldehyde **11d** (150 mg 0.0694 mmol) and 5-nitroindolin-2-one (123 mg 0.0694 mmol). The compound obtained as light yellow colored solid Yield: 195 mg (75%); mp: 196-198 °C;

¹H NMR (500 MHz, DMSO-d₆); δ 6.11 (s, 2H, -OCH₂O), 6.97-7.16 (m, 2H, ArH) 7.23 (s, 1H, =CH), 7.38 (d, 1H, J = 8.1 Hz ArH), 7.45 (s, 1H, ArH) 7.61 (s, 1H, ArH) 8.14-8.34 (m, 1H, ArH) 10.0 (s, 1H, ArH)11.29 (brs, 1H, -NH) 14.13 (brs, 1H, -NH); ¹³C NMR (75 MHz, DMSO-d₆): δ 101.3, 105.7, 108.4, 108.8, 109.3, 119.3, 121.7, 122.1, 122.4, 123.1, 126.3, 128.4, 141.9, 147.8, 148.1, 147.4, 147.0, 169.7 ppm; IR (KBr) (v_{max} /cm⁻¹): v = 3117, 2917, 1684, 1620, 1549, 1515, 1461, 1416, 1337, 1244, 1187, 1127, 1079, 1039 cm⁻¹; MS (ESI) m/z 377 [M+H]; HR-MS (ESI) m/z for C₁₉H₁₃N₄O₅ calculated m/z: 377.32440, found m/z: 377.32410.

5.1.5.17. (Z)-3-((3-(benzo[d][1,3]dioxol-5-yl)-1H-pyrazol-5-yl)methylene)-6chloroindolin-2-one(**12q**):

This compound was prepared using the procedure described above by the addition of 3-(benzo[d][1,3]dioxol-5-yl)-1H-pyrazole-5-carbaldehyde **11d** (150 mg 0.0694 mmol) and 6-chloroindolin-2-one (115 mg 0.0694 mmol). The compound obtained as light yellow colored solid Yield: 202 mg (80%); mp: 210-212 °C; ¹H NMR (300 MHz, DMSO-d₆); δ 6.11 (s, 2H, -OCH₂O), 6.80-7.11 (m, 3H, ArH) 7.25-7.39 (m, 2H, ArH) 7.45 (s, 1H, =CH), 7.65-7.85 (m, 3H, ArH) 8.26 (s, 1H, ArH) 8.90 (brs, 1H, -NH); 13 C NMR (75 MHz, DMSO-d₆): δ 100.9, 101.1, 105.4, 105.6, 108.3, 108.5, 109.2, 118.8, 119.0, 120.4, 124.1, 133.7, 141.3, 143.7, 146.9, 147.3, 147.5, 147.7, 169.3 ppm; IR (KBr) (v_{max}/cm^{-1}) : v = 3160, 3092, 1697, 1614, 1512, 1471, 1439, 1381, 1312, 1240, 1199, 1168, 1057 cm⁻¹; MS (ESI) *m/z* 366 [M+H]; HR-MS (ESI) *m/z* for $C_{19}H_{13}ClN_{3}O_{3}$ calculated m/z: 366.06513, found m/z: 366.06509. (Z)-3-((3-(benzo[d][1,3]dioxol-5-yl)-1H-pyrazol-5-yl)methylene)-5-5.1.5.18. chloroindolin-2-one(12r):

This compound was prepared using the procedure described above by the addition of 3-(benzo[d][1,3]dioxol-5-yl)-1H-pyrazole-5-carbaldehyde **11d** (150 mg 0.0694 mmol) and 6-chloroindolin-2-one (115 mg 0.0694 mmol). The compound obtained as yellow colored solid Yield: 202 mg (80%); mp: 208-210 °C; ¹H NMR (300 MHz, DMSO-d₆); δ 6.10 (s, 2H, -OCH₂O), 6.80-7.11 (m, 2H, ArH) 7.20-7.32 (m, 3H, ArH) 7.35 (s, 1H, =CH), 7.51-7.74 (m, 2H, ArH) 7.81 (s, 1H, ArH) 8.46 (s, 1H, ArH) 9.21 (brs, 1H, -NH); ¹³C NMR (75 MHz, DMSO-d₆): δ 100.9, 101.1, 105.4, 105.6, 108.3, 108.5, 109.2, 118.8, 119.0, 120.4, 124.1, 133.7, 141.3, 143.7, 146.9, 147.3, 147.5, 147.7, 169.3 ppm; IR (KBr) (ν_{max} /cm⁻¹): ν = 3160, 3092, 1697, 1614, 1512, 1471, 1439, 1381, 1312, 1240, 1199, 1168, 1057 cm⁻¹; MS (ESI) *m*/*z* 366 [M+H]; HR-MS (ESI) *m*/*z* for C₁₉H₁₃ClN₃O₃ calculated *m*/*z*: 366.06513, found *m*/*z*: 366.06509.

5.1.5.19. (Z)-3-((3-(benzo[d][1,3]dioxol-5-yl)-1H-pyrazol-5-yl)methylene)-5methoxyindolin-2-one(**12s**): This compound was prepared using the procedure described above by the addition of 3-(benzo[d][1,3]dioxol-5-yl)-1H-pyrazole-5-carbaldehyde **11d** (150 mg 0.0694 mmol) and 5-methoxyindolin-2-one (113 mg 0.0694 mmol). The compound obtained as yellow colored solid Yield: 207 mg (83%); mp: 205-207 °C; ¹H NMR (500 MHz, DMSO-d₆); δ 3.86 (s, 3H, -OCH₃), 6.08 (s, 2H, -OCH₂O), 6.70-6.94 (m, 2H, ArH) 6.93-7.14 (m, 1H, ArH) 7.16 (d, 1H, *J* = 11.8, ArH), 7.27-7.54 (m, 3H, =CH, ArH) 7.86 (s, 1H, ArH) 8.85 (brs, 1H, NH) 10.98 (brs, 1H, -NH); ¹³C NMR (75 MHz, DMSO-d₆): δ 53.5, 99.4, 103.7, 103.9, 106.8, 108.7, 111.9, 112.0, 114.5, 117.8, 122.7, , 134.11, 136.2, 147.5, 149.0, 154.4, 155.0, 169.5 ppm; IR (KBr) (v_{max}/cm⁻¹): v = 3445, 3120, 3002, 2825, 2722, 1678, 1614, 1545, 1511, 1466, 1437, 1397, 1333, 1273, 1241, 1205, 1164 cm⁻¹; MS (ESI) *m*/*z* 362 [M+H]; HR-MS (ESI) *m*/*z* for C₂₁H₁₅N₃O₄ calculated *m*/*z*: 362.11034, found *m*/*z*: 362.11060.

5.1.5.20. (Z)-3-((3-(benzo[d][1,3]dioxol-5-yl)-1H-pyrazol-5-yl)methylene)-5fluoroindolin-2-one(**12t**):

This compound was prepared using the procedure described above by the addition of 3-(benzo[d][1,3]dioxol-5-yl)-1H-pyrazole-5-carbaldehyde **11d** (150 mg 0.0694 mmol) and 5-fluoroindolin-2-one (104 mg 0.0694 mmol). The compound obtained as yellow colored solid Yield: 176 mg (73%); mp: 185-187 °C; ¹H NMR (500 MHz, DMSO-d₆); δ 6.08 (s, 2H, -OCH₂O), 6.74-7.22 (m, 4H, ArH) 7.25-7.91 (m, 3H, ArH, =CH) 8.24 (s, 1H, ArH) 10.52 (brs, 1H, NH) 11.11 (brs, 1H, -NH); ¹³C NMR (75 MHz, DMSO-d₆): δ 85.0, 99.49, 103.9, 106.8, 117.2, 117.4, 124.9, 132.0, 145.7, 146.1, 167.9 ppm; IR (KBr) (v_{max}/cm⁻¹): v = 3127, 1858, 1680, 1611, 1545, 1511, 1466, 1435, 1396, 1335, 1243, 1198, 1165, 1110, 1041 cm⁻¹; MS (ESI) *m/z* 362 [M+H]; HR-MS (ESI) *m/z* for C₂₁H₁₅N₃FO₄ calculated *m/z*: 362.11034, found *m/z*: 362.11060.

5.1.5.21. (Z)-5-fluoro-3-((3-(4-methoxyphenyl)-1H-pyrazol-5-yl)methylene)indolin-2-one(**12u**):

This compound was prepared using the procedure described above by the addition of 3-(4-methoxyphenyl)-1H-pyrazole-5-carbaldehyde **11a** (150 mg 0.0495 mmol) and 5fluoroindolin-2-one (74 mg 0.0694 mmol). The compound obtained as yellow colored solid Yield: 124 mg (75%); mp: 190-192 °C; ¹H NMR (500 MHz, DMSO-d₆); δ 3.86 (s, 3H, -OCH₃), 6.75-7.25 (m, 5H, ArH) 7.55 (s, 1H, ArH), 7.60-7.95 (m, 2H, ArH, =CH) 8.04 (s, 1H, ArH) 10.98 (brs, 1H, NH) 13.88 (brs, 1H, -NH); ¹³C NMR (75 MHz, DMSO-d₆): δ 85.0, 99.49, 103.9, 106.8, 117.2, 117.4, 124.9, 132.0, 145.7, 146.1, 167.9 ppm; IR (KBr) (v_{max} /cm⁻¹): v = 3010, 2965, 2928, 2809, 2702, 1679, 1611, 1577, 1517, 1481, 1461, 1435, 1384, 1337, 1284, 1249, 1201, 1169, 1105, 1054, 1030 cm⁻¹; MS (ESI) *m*/*z* 336 [M+H]; HR-MS (ESI) *m*/*z* for C₁₉H₁₅N₃O₂F calculated *m*/*z*: 336.11414, found *m*/*z*: 336.11414.

5.2. Cell Cultures, Maintenance and Antiproliferative Evaluation:

All cell lines used in this study were purchased from the American Type Culture Collection (ATCC, United States). A549, MCF-7, and HeLa were grown in Dulbecco's modified Eagle's medium (containing 10% FBS in a humidified atmosphere of 5% CO2 at 37 °C). DU145 cells were cultured in Eagle's minimal essential medium (MEM) containing non-essential amino acids, 1 mM sodium pyruvate, 10 mg/mL bovine insulin, and 10% FBS. Cells were trypsinized when sub-confluent from T25 flasks/60 mm dishes and seeded in 96-wel plates. The synthesized test compounds were evaluated for their *in vitro* antiproliferative in four different human cancer cell lines. A protocol of 48 h continuous drug exposure was used, and a MTT cell proliferation assay was used to estimate cell viability or growth. The cell lines were grown in their respective media containing 10% fetal bovine serum and were seeded into 96-well microtiter plates in 200 µL aliquots at plating densities depending on the doubling time of individual cell lines. The microtiter plates were incubated at 37 °C, 5% CO2, 95% air, and 100% relative humidity for 24 h prior to addition of experimental drugs. Aliquots of 2 μ L of the test compounds were added to the wells already containing 198 μ L of cells, resulting in the required final drug concentrations. For each compound, four concentrations (1, 10, 100, and 1000 µM) were evaluated, and each was done in

triplicate wells. Plates were incubated further for 48 h, and the assay was terminated by the addition of 10 μ L of 5% MTT and incubated for 60 min at 37 °C. Later, the plates were air-dried. Bound stain was subsequently eluted with 100 μ L of DMSO, and the absorbance was read on an multimode plate reader (Tecan M200) at a wavelength of 560 nm. Percent growth was calculated on a plate by plate basis for test wells relative to control wells. The above determinations were repeated thrice. The growth inhibitory effects of the compounds were analyzed by generating dose response curves as a plot of the percentage surviving cells versus compound concentration. The sensitivity of the cancer cells to the test compound was expressed in terms of IC₅₀, a value defined as the concentration of compound that produced 50% reduction as compared to the control absorbance. IC₅₀ values are indicated as means \pm SD of three independent experiments [28].

5.2.1. Analysis of Cell Cycle:

HeLa cells in 60 mm dishes were incubated for 24 h in the presence or absence of test compounds **12b**, **12c** and **12d** at 5μ M or 10 μ M concentrations. Cells were harvested with Trypsin-EDTA, fixed with ice-cold 70% ethanol at 4 °C for 30 min, ethanol was removed by centrifugation and cells were stained with 1 mL of DNA staining solution [0.2 mg of Propidium Iodide (PI), and 2 mg RNase A] for 30 min as described earlier . The DNA contents of 20,000 events were measured by flow cytometer (BD FACSCanto II). Histograms were analyzed using FCS express 4 plus [28].

5.2.2. Tubulin polymerization assay:

An *in vitro* assay for monitoring the time-dependent polymerization of tubulin to microtubules was performed employing a fluorescence-based tubulin polymerization assay kit (BK011, Cytoskeleton, Inc.) according to the manufacturer's protocol. The reaction mixture in a final volume of 10 μ l in PEM buffer (80 mM PIPES, 0.5 mM EGTA, 2 mM MgCl₂, pH 6.9) in 384 well plates contained 2 mg/mL bovine brain tubulin, 10 μ M fluorescent reporter, 1 mM GTP in the presence or absence of test compounds at 37°C. Tubulin polymerization was followed by monitoring the fluorescence enhancement due to the incorporation of a fluorescence reporter into microtubules as polymerization proceeds. Fluorescence emission at 420 nm (excitation wavelength is 360 nm) was measured for 1 h at 1-min intervals in a multimode plate reader (Tecan M200). To determine the IC₅₀ values of the compounds against tubulin polymerization, the compounds were pre-incubated with tubulin at varying concentrations (1, 5, 10 and 20 μ M). Assays were performed under similar conditions as employed for polymerization assays as described above[28].

5.2.3. Western blot Analysis of Soluble versus Polymerized Tubulin and cyclin B1:

Cells were seeded in 12-well plates at 1×10^5 cells per well in complete growth medium. Following treatment of cells with respective compounds (**12b**, **12c and 12d**) for duration of 24 h, cells were washed with PBS and subsequently soluble and insoluble tubulin fractions were collected. To collect the soluble tubulin fractions, cells were permeablized with 200 µL of pre-warmed lysis buffer [80 mM Pipes-KOH (pH 6.8), 1 mM MgCl₂, 1 mM EGTA, 0.2% Triton X-100, 10% glycerol, 0.1% protease inhibitor cocktail (Sigma-Aldrich)] and incubated for 3 min at 30 °C. Lysis buffer was gently removed, and mixed with 100 µL of 3×Laemmli's sample buffer (180 mM Tris-

Cl pH 6.8, 6% SDS, 15% glycerol, 7.5% β-mercaptoethanol and 0.01% bromophenol blue). Samples were immediately heated to 95 °C for 3 min. To collect the insoluble tubulin fraction, 300 µL of 1×Laemmli's sample buffer was added to the remaining cells in each well, and the samples were heated to 95°C for 3 min. Equal volumes of samples were run on an SDS-10 % polyacrylamide gel and were transferred to a nitrocellulose membrane employing semidry transfer at 50 mA for 1h. Blots were probed with mouse anti-human α -tubulin diluted 1:2,000 ml (Sigma) and stained with rabbit anti-mouse secondary antibody coupled with horseradish peroxidase, diluted 1:5000 ml (Sigma). Bands were visualized using an enhanced Chemiluminescence protocol (Pierce) and radiographic film (Kodak.). For cyclin B1 immunoblots, Cells were seeded in 12-well plates at 1×10^5 cells per well in complete medium and treated with different concentrations of 12b, 12c and 12d for 24h. After treatment, cells were washed twice with phosphate-buffered saline and lysed in 1X SDS sample buffer. Proteins were separated, transferred, probed and analyzed similar to tubulin. The primary anti-cyclin B1 antibody was employed at 1:1500 (Sigma) and horseradish peroxidase coupled goat anti-rabbit secondary antibody diluted 1:5,000 (Sigma) [28].

5.2.4. Immunohistochemistry of tubulin and Analysis of nuclear morphology

HeLa cells were seeded on glass cover slip, incubated for 24 h in the presence or absence of test compounds **12b**, **12c** and **12d** at a concentration of 5 μ M. Cells grown on coverslips were fixed in 3.5% formaldehyde in phosphate-buffered saline (PBS) pH 7.4 for 10 minutes at room temperature. Cells were permeablized for 6 minutes in PBS containing 0.5% Triton X-100 (Sigma) and 0.05% Tween-20 (Sigma). The permeablized cells were blocked with 2% BSA (Sigma) in PBS for 1h. Later, the

cells were incubated with primary antibody for tubulin from (sigma) at (1:200) diluted in blocking solution for 4h at room temperature. Subsequently the antibodies were removed and the cells were washed thrice with PBS. Cells were then incubated with FITC labeled anti-mouse secondary antibody (1:500) for 1h at room temperature. Cells were washed thrice with PBS and mounted in medium containing DAPI. Images were captured using the Olympus confocal microscope and analyzed with Provision software.

5.3. Zebrafish maintenance and Screening:

Wild type zebrafish (Danio rerio) were raised and maintained at 28.5°C with 14hr:10 hr light : dark cycle. 2-3 pairs of zebrafish were synchronously mated and embryos were pooled. The embryos were dechorinated and 3-4 embryos were dispensed in 200ul of E3 embryo medium (5mM NaCl, 0.17mM KCl, 0.44mM CaCl₂, 0.68 mM MgCl₂) into each well of a 96 well plate. The embryos were treated with the compound (10 & 25uM), nocadozole (5uM) or DMSO (1%) at 5hpf and incubated at 28.5°C. About 20 embryos were screened for each compound in biological replicates. They were observed after 28hpf with a Leica steromicroscope. Only those compounds that resulted in similar phenotypic defects in most of the embryos were considered as active [22].

5.4. Molecular Modelling:

AutoDock was used to dock 3,4,5-trimethoxybipheny derivatives in colchicine binding site of tubulin [24-26]. Initial Cartesian coordinates for the protein-ligand complex structure were derived from crystal structure of tubulin (PDB ID: 3E22). The protein targets were prepared for molecular docking simulation by removing water molecules, bound ligands. Hydrogen atoms and Kollman charges were added to each

protein atom. Auto-Dock Tools (ADT) was used to prepare and analyze the docking simulations for the AutoDock program. Coordinates of each compound were generated using Chemdraw11 followed by MM2 energy minimization. Grid map in Autodock that defines the interaction of protein and ligands in binding pocket was defined. The grid map was used with 60 points equally in each x, y, and z direction. AutoGrid 4 was used to produce grid maps for AutoDock calculations where the search space size utilized grid points of 0.375 Å. The Lamarckian genetic algorithm was chosen to search for the best conformers. Each docking experiment was performed 100 times, yielding 100 docked conformations. Parameters used for the docking were as follows: population size of 150; random starting position and conformation; maximal mutation of 2 Å in translation and 50 degrees in rotations; elitism of 1; mutation rate of 0.02 and crossover rate of 0.8; and local search rate of 0.06. Simulations were performed with a maximum of 1.5 million energy evaluations and a maximum of 50000 generations. Final docked conformations were clustered using a tolerance of 1.0 Å root mean square deviation. The best model was picked based on the best stabilization energy. Final figures for molecular modeling were generated by using PyMol [27].

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Figure Captions

Figure 1A. Structures of some anticancer molecules Combretastatin (1), colchicine (2), substituted methoxylbenzoyl-aryl-thiazole (SMART) (3), 2-aryl-4-benzoyl-imidazole series (ABI) (4), Sunitinib (5), indirubin (6) and pyrazole-oxindole conjugates (12a-12u).

Figure 1B. SAR of pyrazole-indole conjugates (see table1 for R₁, R₂)

Figure 2. Anti-mitotic effects of 12b, 12c, and 12d by FACS analysis:

Induction of cell cycle G2/M arrest by compound **12b**, **12c**, and **12d**. HeLa cells were harvested after treatment at 5 mM and 10 μ M for 24h. Untreated cells and DMSO treated cells served as controls. The percentage of cells in each phase of cell cycle was quantified by flow cytometry.

Figure 3. Effect of 12b, 12c, and 12d on microtubules and nuclear condensation: HeLa cells were independently treated with 12b, 12c and 12d at 5 μ M concentrations for 24h. Following the termination of experiment, cells were fixed and stained for tubulin. DAPI was used as counter stain. The merged images of cells stained for tubulin and DAPI are represented.

Figure 4. Distribution of tubulin in polymerized vs soluble fractions as analyzed by immunobloting in 12b, 12c 12d treated HeLa cells: Figure 4A. HeLa cells were treated with 5 mM of 12b, 12c and 12d for 24h. Tubulin was detected by Immunoblot analysis. Figure 4B. Hela and cells were treated with 5mM concentrations of compound 12b, 12c, and 12d for 24h. Subsequently, whole lysates were prepared and analyzed for Cyclin-B1. A non-specific band at 90kDa was used as control.

Figure 5. Effect of microtubule inhibitors on development of zebrafish embryos: Zebrafish embryos were treated with (a) DMSO (1%), 12a, 12b, 12c and 12d or Nocodazole at 5hpf. The embryos were imaged at 30hpf to observe morphological changes.

Figure 6. *Panel A*, Docking pose of 12b (yellow), 12c (magentas), 12d (cyan). The green surface represents β -chain while the grey line diagram indicates the α -chain of tubulin. The 4-methoxybenzene moiety of all the compounds is buried in hydrophobic pocket of colchicine binding domain (A-site). *Panel B*, Stick representation of selected amino acids of α and β subunits of tubulin interacting with compound 12b, 12c and

12d. Grey color for the b-chain and blue for the a-chain of tubulin was used for representation. Red dotted lines indicate the hydrogen bonds.

Table 1: Structures of the compounds 12a to 12u and *invitro* antiproliferative activity.**Table 2.** Antitubulin activity of compounds 12a to 12u

Scheme Caption

Scheme 1. Synthesis of Pyrazole - Oxindole Conjugates (12a-12u): *Reagents and conditions*: (i) NaOEt/EtOH, 4 h, 0°C-rt, 68%; (ii) NH₂-NH₂.2HCl/ EtOH, 3 h, reflux, 72–79%; (iii) LiAlH₄/THF, 1 h 0°C-rt, 51–70%. (iv) IBX/DMSO, 1h, rt, 51–70% (v) oxindoles/EtOH, 4 h, reflux, 51–70%

Tables

Table 1. Structures of the compounds 12a to 12u and *invitro* antiproliferative activity.



	R ₁	R ₂	IC ₅₀ <u>+</u> SEM (μM)				
			HeLa	A549	MCF7	DU145	Average
12a	4-OCH ₃	Н	18.0 <u>+</u> 0.6	14.0 <u>+</u> 0.4	15.0 <u>+</u> 0.3	21.0 <u>+</u> 0.5	17
12b	4-OCH ₃	5-Cl	5.0 <u>+</u> 0.7	6.0 <u>+</u> 1.3	9.3 <u>+</u> 0.3	4.0 <u>+</u> 2.1	6.0
12c	4-OCH ₃	6-Cl	2.4 ± 0.8	3.0 <u>+</u> 0.6	3.6 <u>+</u> 0.4	3.0 <u>+</u> 1.9	3.0
12d	4-OCH ₃	5-OCH ₃	7.1 <u>+</u> 0.3	8.0 <u>+</u> 0.2	6.0 <u>+</u> 0.3	9.0 <u>+</u> 3.2	7.5
12e	4-OCH ₃	5- NO ₂	33.0 <u>+</u> 0.4	22.0 <u>+</u> 0.3	24.0 ± 2.1	26.0 ± 4.6	26.2
12f	3,4,5(OCH ₃) ₃	5-Cl	12.0 <u>+</u> .1	27.0 <u>+</u> 0.1	44.0 <u>+</u> .7	21.0 ± 2.1	26.0
12g	3,4,5(OCH ₃) ₃	Н	21.0 <u>+</u> 0.2	21.0 <u>+</u> 0.2	27.0 ± 2.1	35.0 <u>+</u> 5.1	26.0
12h	3,4,(OCH ₃) ₂	Н	21.0 <u>+</u> 0.2	29.0 <u>+</u> 0.3	36.0 <u>+</u> 2.8	31.0 <u>+</u> 6.7	29.2
12i	3,4,(OCH ₃) ₂	5-Cl	10.0 <u>+</u> 0.8	24.0 <u>+</u> 0.1	16.0 <u>+</u> 0.1	21.0 <u>+</u> 1.2	17.7
12j	3,4,(OCH ₃) ₂	6-Cl	11.0 <u>+</u> 0.2	18.0 <u>+</u> 5.6	27.0 <u>+</u> 0.2	13.9 <u>+1</u> .6	17.4
12k	3,4,(OCH ₃) ₂	5-OCH ₃	13.0 <u>+</u> 0.3	15.0 <u>+</u> 1.6	32.0 <u>+</u> 0.7	34.0 <u>+</u> 2.3	23.5
12l	3,4,(OCH ₃) ₂	5-NO ₂	36.0 <u>+</u> 0.6	42.0 <u>+</u> 2.4	42.0 ± 2.2	28.0 <u>+</u> 5.9	37.0
12m	3,4,5(OCH ₃) ₃	6-Cl	11.0 <u>+</u> 0.4	33.0 <u>+</u> 0.6	22.0 <u>+</u> 0.4	27.0 <u>+</u> 2.7	23.2
12n	3,4,5(OCH ₃) ₃	5-OCH ₃	16.0 <u>+</u> 0.3	33.0 <u>+</u> 1.3	8.8 <u>+</u> 0.6	9.1 <u>+</u> 0.3	16.7
120	3,4,5(OCH ₃) ₃	5-NO ₂	30.0 <u>+</u> 0.5	22.0 <u>+</u> 0.3	16.0 <u>+</u> 0.4	20.0 <u>+</u> 0.6	22.0
12p	3,4-(OCH ₂ O)	5-NO ₂	24.0 <u>+</u> 0.3	37.0 <u>+</u> 0.4	17.0 <u>+</u> 0.6	36.0 <u>+</u> 0.6	28.5
12q	3,4-(OCH ₂ O)	6-Cl	7.3 <u>+</u> 1.8	27.0 <u>+</u> 0.5	18.0 <u>+</u> 0.2	24.0 <u>+</u> 4.5	19.0
12r	3,4-(OCH ₂ O)	5-Cl	12.0 <u>+</u> 0.1	29.0 <u>+</u> 1.8	44.0 <u>+</u> 0.7	24.0 ± 2.1	27.2
12s	3,4-(OCH ₂ O)	5-OCH ₃	13.5 <u>+</u> 1.2	24.0 <u>+</u> 3.2	9.8 <u>+</u> 0.3	34.0 <u>+</u> 1.1	20.3
12t	3,4-(OCH ₂ O)	5-F	32.0 <u>+</u> 0.4	26.0 <u>+</u> 3.1	36.0 <u>+</u> 1.5	21.0 <u>+</u> 5.2	28.7
12u	4-OCH ₃	5-F	22.0 ± 0.2	29.0 <u>+</u> 1.8	36.0 <u>+</u> 3.1	25.0 <u>+</u> 6.7	28.0
Nocodazole		1.1 <u>+</u> 0.5	0.89 ± 0.01	1.7 <u>+</u> 0.65	3.2 <u>+</u> 0.83	1.72	

Cell lines were treated with different concentrations of compounds for 48 h as described under Materials and Methods. Cell viability was measured employing MTT assay. IC50 values are indicated as the mean of three independent experiments.

Compound	Inhibition of Tubulin polymerization(µM)	Compound	Inhibition of Tubulin polymerization(µM)
12a	38.9 <u>+</u> 1.90	12l	18.0 <u>+</u> 0.71
12b	8.10 <u>+</u> 0.61	12m	22.4 <u>+</u> 1.23
12c	5.90 <u>+</u> 0.78	12n	16.0 <u>+</u> 1.10
12d	9.2 <u>+</u> 1.10	120	15.3 <u>+</u> 4.10
12e	21.0 <u>+</u> 1.10	12p	14.7 <u>+</u> 2.10
12f	25.3 <u>+</u> 2.30	12q	14.5 <u>+</u> 2.30
12g	18.0 <u>+</u> 0.63	12r	35.3 <u>+</u> 2.30
12h	23.0 <u>+</u> 1.90	12s	12.0 <u>+</u> 0.89
12i	33.8 <u>+</u> 1.60	12t	24.0 <u>+</u> 0.56
12j	18.9 <u>+</u> 0.89	12u	25.0 <u>+</u> 0.59
12k	22.1 <u>+</u> 2.50	Nocodazole	2.2 <u>+</u> 0.66

 Table 2. Antitubulin activity of compounds 12a to 12u

Effect of congeners on tubulin polymerization. IC_{50} values for **12a** to **12u** were determined from the tubulin polymerization assays.







B

 \mathbf{Y}



A ring, 4-methoxy is the best, when compared to 3,4-dimethoxy, 3,4,5trimethoxy and 3,4-(methylenedioxy) substituted ring.









Figure 4B





Nocodazole



Figure 5



Figure 6



Scheme 1. Synthesis of Pyrazole - Oxindole Conjugates (12a-12u): *Reagents and conditions*: (i) NaOEt/EtOH, 4 h, 0°C-rt, 85-90%; (ii) NH₂-NH₂.2HCl/ EtOH, 3 h, reflux, 65-70%; (iii) LiAlH₄/THF, 1 h 0°C-rt, 70-80%. (iv) IBX/DMSO, 1h, rt, 80-85% (v) oxindoles/EtOH, 4 h, reflux, 73-85%.

Scheme