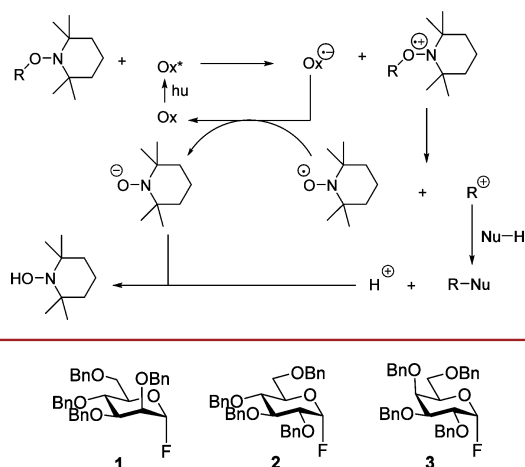
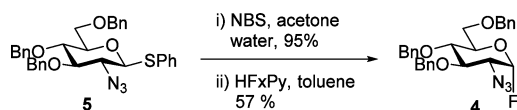


Scheme 2. Photocatalytic Cation Generation from Tempol Ethers and Tempol Glycosides



thioglucopyranoside **5** to the corresponding hemiacetal followed by treatment with HF·pyridine according to the method of Noyori et al. (Scheme 3).²⁷

Scheme 3. Synthesis of Glycosyl Fluoride **4**



Glycosyl fluorides **1–4** were then coupled to the persistent radical Tempol with activation by $\text{BF}_3\cdot\text{OEt}_2$ in acetonitrile at room temperature in the presence of tetramethylguanidine following the method of Sato.¹⁷ Manno-configured donor **1** gave a single α -glycoside **6**, whereas gluco- and galacto-configured glycosyl fluorides **2–4** afforded separable α,β mixtures of the corresponding glycosides **7–9** (Table 1) by a reaction whose mechanism remains to be clarified.

Table 1. Synthesis of Tempol Glycosides

entry	glycosyl fluorides	tempol glycosides (yield, %)
1	1	6 α (64) and 6 β (0)
2	2	7 α (38) and 7 β (37)
3	3	8 α (41) and 8 β (30)
4	4	9 α (22) and 9 β (63)

Adapting the method of Knowles,¹⁵ we photolyzed 0.1 M nitromethane solutions of the various Tempol glycosides with blue LEDs through Pyrex at room temperature in the presence of powdered 4 Å MS, 5 mol % of iridium photocatalyst **10**,¹⁵ and the model acceptor alcohols **11–13**, monitoring with TLC or ESI MS.

Catalyst **10** was selected for this study as the optimum system in the Knowles work¹⁵ on tertiary cation generation. Similarly, nitromethane was chosen as solvent by extrapolation from that study;¹⁵ indeed, a brief investigation of acetonitrile as solvent revealed it to be far inferior. This preliminary investigation was limited to the use of Tempol derivatives following Knowles,¹⁵ but

it is recognized that other hydroxylamine derivatives may eventually prove advantageous. Blank reactions conducted in the dark or by photolysis in the absence of **10** did not proceed and indicated the need for **10** and its photolytic activation. Experiments conducted in the absence of molecular sieves were marred by formation of significant amounts of the hydrolyzed donor (hemiacetals) but also proceeded significantly more slowly, suggesting involvement of the sieves in the reaction mechanism.

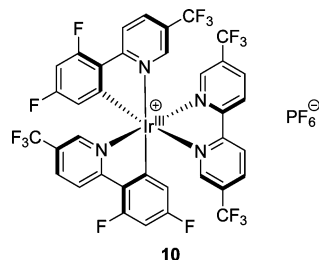
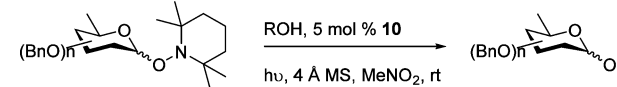
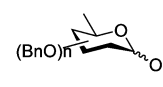
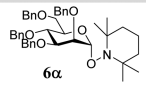

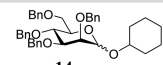
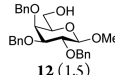
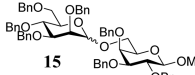
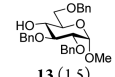
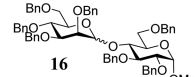
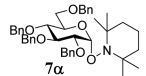
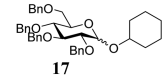
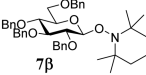
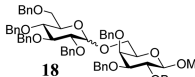
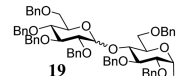
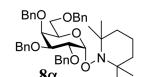
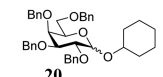
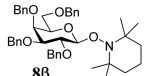
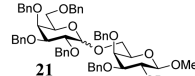
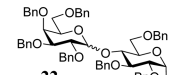
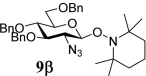
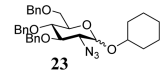
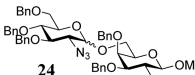
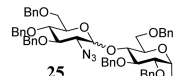


Table 2 reveals that good to excellent yields were obtained in all couplings conducted in this survey, with only modest differences in yield observed between the α - and β -anomers of a particular donor (compare Table 2, entries 4, 6, and 8 with 5, 7, and 9; and entries 10, 12, and 14 with 11, 13, and 15). Although the anomeric selectivity of the coupling reactions is modest, it is reproducibly a function of the anomeric configuration of the donor (compare Table 2, entries 4, 6, and 8 with 5, 7, and 9; and entries 10, 12, and 14 with 11, 13, and 15). Greater differences in selectivity between the two anomers of a given donor are observed with the more reactive cyclohexanol, which was additionally used in greater excess, and with the primary acceptor **12**, than with the less reactive secondary acceptor **13**. The implication is that glycosylation proceeds at least in part in a concerted manner in which displacement of the Tempol radical from the oxidatively activated donor is coupled with attack by the incoming acceptor. Such a concerted process can be interpreted as either a direct attack on the activated donor or an attack on the initially formed radical ion triplet before diffusive equilibration (Scheme 4). There is a steadily increasing body of kinetic evidence^{28–37} that a variety of stereoselective glycosylation reactions proceed by associative reaction pathways rather than by dissociatively free glycosyl oxocarbenium ions that have only been observed in super acidic media.^{22,23,38,39} In such classical glycosylation reactions, the associative pathways are more prevalent with more reactive acceptor alcohols,^{40–43} consistent with the observed pattern in Table 2. The hexafluorophosphate counterion is not considered to intervene directly in the reaction mechanism except for providing charge stabilization consistent with studies on the influence of a variety of counterions on the stereoselectivity of glycosylation reactions.^{38,44} The part of the reaction manifold that affords substitution with overall retention of configuration is best interpreted as taking place via solvent-separated ion triplets rather than via equilibration of the donor configuration followed by concerted displacement, as no evidence was found for donor anomerization in these reactions. The root cause of the observed acceleration of reaction rate by the 4 Å MS is presumably associated with their basic character⁴⁵ and the relative acidity⁴⁶ of the hydroxylamine Tempol.

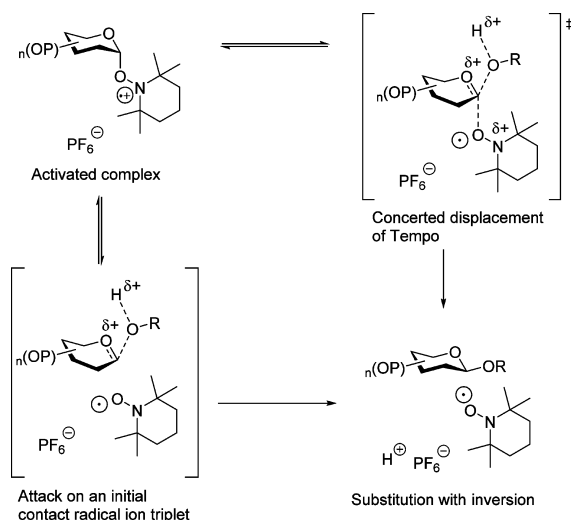
In summary, a new photocatalytic glycosylation method is presented that functions under irradiation through Pyrex by blue LEDs and in the absence of additives other than **10** and 4 Å MS. The method uses stable and readily accessible Tempol glycosides as donors and neither requires nor generates in situ any strongly

Table 2. Photoglycosylation of Alcohols with Donors 6–9 and Catalytic 10

<div style="display: flex; align-items: center; justify-content: center;"> <div style="text-align: center;">  </div> <div style="margin: 0 20px;"> $\xrightarrow[\text{h}\nu, 4 \text{ \AA MS, MeNO}_2, \text{rt}]{\text{ROH, 5 mol \% 10}}$ </div> <div style="text-align: center;">  </div> </div>					
6 - 9			14 - 25		
entry	donor	acceptor (equiv)	photolysis time (h)	product	yield, % ^a α:β-ratio ^b
1	 6α	 11 (3)	36	 14	80, 1.3:1
2	6α	 12 (1.5)	48	 15	62, 1.5:1
3	6α	 13 (1.5)	60	 16	64, 1:1
4	 7α	11 (3)	36	 17	96, 1:2.5
5	 7β	11 (5)	36	17	91, 1.2:1
6	7α	12 (1.5)	48	 18	88, 1:1
7	7β	12 (1.5)	48	18	79, 1.3:1
8	7α	13 (1.5)	48	 19	69, 1.3:1
9	7β	13 (1.5)	48	19	63, 1.6:1
10	 8α	11 (3)	36	 20	95, 1:1.6
11	 8β	11 (3)	36	20	87, 1.6:1
12	8α	12 (1.5)	48	 21	82, 1.2:1
13	8β	12 (1.5)	48	21	75, 1.4:1
14	8α	13 (1.5)	48	 22	77, 1.6:1
15	8β	13 (1.5)	48	22	59, 1.6:1
16	 9β	11 (3)	36	 23	86, 1.8:1
17	9β	12 (1.5)	72	 24	61, 1.4:1
18	9β	13 (1.5)	60	 25	57, 2:1

^aCombined yield of the isolated anomers. ^bAnomeric ratios were determined by integration of the ¹H NMR spectra of the crude photolysates before purification.

Scheme 4. Concerted and Radical Ion Triplet Mechanisms for Glycosylation



electrophilic agents. Further work expanding the scope and selectivity of this novel glycosylation is underway in our laboratories.

■ ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: [10.1021/acs.orglett.7b00932](https://doi.org/10.1021/acs.orglett.7b00932).

Full experimental details and ^1H and ^{13}C NMR spectra for all new compounds (PDF)

■ AUTHOR INFORMATION

Corresponding Author

*E-mail: dcrich@chem.wayne.edu.

ORCID

David Crich: 0000-0003-2400-0083

Notes

The authors declare no competing financial interest.

■ ACKNOWLEDGMENTS

We thank the NIH (GM62160) for support of this work and acknowledge support from the NSF (MRI-084043) for the purchase of the 600 MHz NMR spectrometer in the Lumigen Instrument Center at Wayne State University.

■ REFERENCES

- (1) Hashimoto, S.; Kurimoto, I.; Fujii, Y.; Noyori, R. *J. Am. Chem. Soc.* **1985**, *107*, 1427.
- (2) Mao, R.-Z.; Guo, F.; Xiong, D.-C.; Li, Q.; Duan, J.; Ye, X.-S. *Org. Lett.* **2015**, *17*, 5606.
- (3) Mao, R.-Z.; Xiong, D.-C.; Guo, F.; Li, Q.; Duan, J.; Ye, X.-S. *Org. Chem. Front.* **2016**, *3*, 737.
- (4) Nakanishi, M.; Takahashi, D.; Toshima, K. *Org. Biomol. Chem.* **2013**, *11*, 5079.
- (5) Iwata, R.; Uda, K.; Takahashi, D.; Toshima, K. *Chem. Commun.* **2014**, *50*, 10695.
- (6) Kimura, T.; Eto, T.; Takahashi, D.; Toshima, K. *Org. Lett.* **2016**, *18*, 3190.
- (7) Furuta, T.; Takeuchi, K.; Iwamura, M. *Chem. Commun.* **1996**, 157.

- (8) Griffin, G. W.; Bandara, N. C.; Clarke, M. A.; Tsang, W.-S.; Garegg, P. J.; Oscarson, S.; Silwanis, B. A. *Heterocycles* **1990**, *30*, 939.
- (9) Cumpstey, I.; Crich, D. *J. Carbohydr. Chem.* **2011**, *30*, 469.
- (10) Wever, W. J.; Cinelli, M. A.; Bowers, A. A. *Org. Lett.* **2013**, *15*, 30.
- (11) Spell, M.; Wang, X.; Wahba, A. E.; Conner, E.; Ragains, J. *Carbohydr. Res.* **2013**, *369*, 42.
- (12) Yuan, X.; Cheng, S.; Shi, Y.; Xue, W. *Synthesis* **2014**, *46*, 331.
- (13) Yu, Y.; Xiong, D.-C.; Mao, R.-Z.; Ye, X.-S. *J. Org. Chem.* **2016**, *81*, 7134.
- (14) Spell, M. L.; Deveau, K.; Bresnahan, C. G.; Bernard, B. L.; Sheffield, W.; Kumar, R.; Ragains, J. R. *Angew. Chem., Int. Ed.* **2016**, *55*, 6515.
- (15) Zhu, Q.; Gentry, E. C.; Knowles, R. R. *Angew. Chem., Int. Ed.* **2016**, *55*, 9969.
- (16) Kancharla, P. K.; Kato, T.; Crich, D. J. *Am. Chem. Soc.* **2014**, *136*, 5472.
- (17) Sato, S.; Kumazawa, T.; Matsuba, S.; Onodera, J.-i.; Aoyama, M.; Obara, H.; Kamada, H. *Carbohydr. Res.* **2001**, *334*, 215.
- (18) Lambert, J. B.; Mark, H. W. *J. Am. Chem. Soc.* **1978**, *100*, 2501.
- (19) Glaudemans, C. P. J.; Fletcher, H. G. *J. Am. Chem. Soc.* **1965**, *87*, 4636.
- (20) Mootoo, D. R.; Konradsson, P.; Udodong, U.; Fraser-Reid, B. *J. Am. Chem. Soc.* **1988**, *110*, 5583.
- (21) Douglas, N. L.; Ley, S. V.; Lucking, U.; Warriner, S. L. *J. Chem. Soc., Perkin Trans. 1* **1998**, 51.
- (22) Bohé, L.; Crich, D. C. *R. Chim.* **2011**, *14*, 3.
- (23) Martin, A.; Arda, A.; Désiré, J.; Martin-Mingot, A.; Probst, N.; Sinaÿ, P.; Jiménez-Barbero, J.; Thibaudau, S.; Blériot, Y. *Nat. Chem.* **2015**, *8*, 186.
- (24) Zeng, J.; Vedachalam, S.; Xiang, S.; Liu, X.-W. *Org. Lett.* **2011**, *13*, 42.
- (25) López, J. C.; Uriel, C.; Guillaumon-Martin, A.; Valverde, S.; Gómez, A. M. *Org. Lett.* **2007**, *9*, 2759.
- (26) López, J. C.; Bernal-Albert, P.; Uriel, C.; Valverde, S.; Gómez, A. M. *J. Org. Chem.* **2007**, *72*, 10268.
- (27) Hayashi, M.; Hashimoto, S.; Noyori, R. *Chem. Lett.* **1984**, *13*, 1747.
- (28) Crich, D. *Acc. Chem. Res.* **2010**, *43*, 1144.
- (29) Huang, M.; Garrett, G. E.; Birlirakis, N.; Bohé, L.; Pratt, D. A.; Crich, D. *Nat. Chem.* **2012**, *4*, 663.
- (30) Huang, M.; Retailleau, P.; Bohé, L.; Crich, D. *J. Am. Chem. Soc.* **2012**, *134*, 14746.
- (31) Adero, P. O.; Furukawa, T.; Huang, M.; Mukherjee, D.; Retailleau, P.; Bohé, L.; Crich, D. *J. Am. Chem. Soc.* **2015**, *137*, 10336.
- (32) D'Angelo, K. A.; Taylor, M. S. *J. Am. Chem. Soc.* **2016**, *138*, 11058.
- (33) Khomutnyk, Y. Y.; Argüelles, A. J.; Winschel, G. A.; Sun, Z.; Zimmerman, P. M.; Nagorny, P. J. *Am. Chem. Soc.* **2016**, *138*, 444.
- (34) Wurst, J. M.; Liu, G.; Tan, D. S. *J. Am. Chem. Soc.* **2011**, *133*, 7916.
- (35) Hosoya, T.; Takano, T.; Kosma, P.; Rosenau, T. *J. Org. Chem.* **2014**, *79*, 7889.
- (36) Hosoya, T.; Kosma, P.; Rosenau, T. *Carbohydr. Res.* **2015**, *411*, 64.
- (37) Kwan, E. E.; Park, Y.; Besser, H. A.; Anderson, T. L.; Jacobsen, E. N. *J. Am. Chem. Soc.* **2017**, *139*, 43.
- (38) Bohé, L.; Crich, D. *Carbohydr. Res.* **2015**, *403*, 48.
- (39) Bohé, L.; Crich, D. *Nat. Chem.* **2016**, *8*, 99.
- (40) Krumper, J. R.; Salamant, W. A.; Woerpel, K. A. *J. Org. Chem.* **2009**, *74*, 8039.
- (41) Beaver, M. G.; Woerpel, K. A. *J. Org. Chem.* **2010**, *75*, 1107.
- (42) van der Vorm, S.; Hansen, T.; Overkleef, H. S.; van der Marel, G. A.; Codée, J. D. C. *Chem. Sci.* **2017**, *8*, 1867.
- (43) Hagen, B.; Ali, S.; Overkleef, H. S.; van der Marel, G. A.; Codée, J. D. C. *J. Org. Chem.* **2017**, *82*, 848.
- (44) Hasty, S. J.; Ranade, S. C.; Demchenko, A. V. *Reports Org. Chem.* **2014**, *4*, 1.
- (45) Kartha, K. P. R.; Mukhopadhyay, B.; Field, R. A. *Carbohydr. Res.* **2004**, *339*, 729.
- (46) Bartmess, J. E. In *Chemistry of Hydroxylamines, Oximes and Hydroxamic Acids*; Rappoport, Z., Liebman, J. F., Eds.; Wiley: Chichester, U.K., 2011; Vol. 2, pp 115–143.