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# Effect of betulinic acid and its ionic derivatives on M-MuLV replication

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## ABSTRACT

Murine leukemia virus (MuLV) is a retrovirus known causing leukemia and neurological disorders in mice, and its viral life cycle and pathogenesis have been investigated extensively over the past decades. As a natural antiviral agent, betulinic acid is a pentacyclic triterpenoid that can be found in the bark of several species of plants (particularly the white birch). One of the hurdles for betulinic acid to release its antiviral potency is its poor water solubility. In this study, we synthesized more water-soluble ionic derivatives of betulinic acid, and examined their activities against Moloney MuLV (M-MuLV). The mouse fibroblast cells stably infected with M-MuLV, 43D cells, were treated with various doses of betulinic acids and its derivatives, and the viral structural protein Gag in cells and media were detected by western blots. Two ionic derivatives containing the benzalkonium cation were found to inhibit the virus production into media and decreased Gag in cells. However, a cell proliferation assay showed that the benzalkonium cation through the inhibition of metabolism in 43D cells. Interestingly, all of these betulinic acid compounds exhibited a minimum impact on the processing and release of Gag from 43D cells, which outlines the differences of viral maturation between MuLV and human immunodeficiency virus.

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## 1. Introduction

Retroviruses comprise a large and diverse family of enveloped RNA viruses defined by common taxonomic denominators including the structural, compositional, and replicative properties [1]. Murine leukemia viruses (MuLVs) are simple retroviruses and encode only three polyproteins that are used in the assembly of progeny virus particles; Gag (structural protein), Pol (three retroviral enzymes, i.e. protease, reverse transcriptase, and integrase), and Env [2]. MuLVs are composed of diverse strains in both endogamous and exogenous viruses, and some of them are causative agents of leukemia and neurological disorders in mice [3]. Moloney murine leukemia virus (M-MuLV) is a well-studied replication-competent oncogenic retrovirus of gammaretrovirus genus which has enabled the understandings of general phenomenon of leukemogenesis; therefore, M-MuLV has been considered as a model retrovirus [4]. The viruses bind to the host cell via interactions between the envelope protein and its receptor on target cells. Subsequently, the penetrated viral cores are endocytosed and the uncoating occurs. The single stranded viral RNA genome is reverse transcribed and cDNA is integrated into the host genome by integrase. In the late phase, viral transcripts make the viral proteins. At the same time, the unspliced viral RNA works as the viral genome. These viral materials are trafficked to plasma membranes and assembled, and new viruses bud from the membranes at the end.

Betulinic acid (3 $\beta$ -hydroxyl-lup-20(29)-en-28-oic acid) is a natural pentacyclic lupine type triterpene that can be extracted from plants (e.g. birch tree bark) [5]. This natural compound has a number of medically relevant biological properties such as antiinflammatory, anti-cancer, and anti-viral activities [5–8]. A major obstacle with the releasing of the antiviral potency of betulinic acid is due to its poor solubility in aqueous solutions and common organic solutions (such as esters, alcohols, and ethers). As a matter of fact, the solubility of betulinic acid in water is only approximately

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 $0.02 \ \mu$ g/ml at room temperature [9]. In common organic solvents, the solubility of betulinic acid is also fairly low, e.g. 1% (w/v) in ethanol and 5% (w/v) in DMSO at 25 °C [10]. Some derivatives of betulinic acid had shown improved water solubility, as well as enhanced biological activities when compared with betulinic acid itself [6,11,12]. Recently, we developed new ionic derivatives of betulinic acid with a higher water solubility and thus a stronger antiviral activity [10,13,14]. In the present study, we further examined the effect of betulinic acid and its derivatives (see Fig. 1) on the retroviral replication of M-MuLV.

## 2. Materials and methods

**Synthesis of ionic derivatives of betulinic acid**. Betulinic acid, glycine methyl ester hydrochloride, *N*, *N'*-dicyclohexylcarbodiimide (DCC), 4-(dimethylamino)pyridine (DMAP), choline chloride, benzalkonium chloride, and Amberlyst A26 hydroxide form (Sigma-Aldrich, St. Louis, MO) were used to prepare ionic salts of betulinic acid conjugated with glycine (Fig. 1). The procedures for preparing ionic derivatives of betulinic acid are described previously [15,16]. In brief, betulinic acid (**BA1**, 204.4 mg), glycine methyl ester (104.0 mg), and triethylamine (8 mg) were dissolved in 50 ml of anhydrous tetrahydrofuran (THF) at room temperature. Into the reaction mixture, DCC (107.9 mg) and DMAP (30.3 mg) were added and the mixture was stirred at room temperature under argon for a



betulinic acid / BA1



cholinium salt of betulinic acid-glycine [cholinium][BA-Gly] / BA2



[benzalkonium][betulinate] / BA4

duration of 48 h. After the reaction completed, the precipitate (dicyclohexylurea as byproduct) was filtered off and the filtrate was further evaporated under vacuum to remove THF. The crude product was then dissolved in 100 ml of ether/ethyl acetate (2:1, v/ v) and extracted with 100 ml of water to remove the excess watersoluble carbodiimide reagent and any remaining dicyclohexylurea. The organic layer was further extracted with 1.0 N HCl  $(2 \times 100 \text{ ml})$ to remove DMAP [17], saturated NaHCO<sub>3</sub> solution ( $1 \times 100$  ml), and lastly with water  $(1 \times 100 \text{ ml})$  [18,19]. The purified organic layer was then dried over Na<sub>2</sub>SO<sub>4</sub>. After filtration, ethyl acetate was evaporated under vacuum to yield the glycine methyl ester conjugate of betulinic acid. The methyl ester (298 mg) was dissolved in 100 ml of THF/H<sub>2</sub>O (4:1, v/v) solution, followed by the addition of LiOH (67.5 mg). The reaction mixture was then stirred at room temperature for 4 h under argon. Once the reaction completed, THF was evaporated under vacuum and the product obtained was dissolved in 200 ml of ethyl acetate, followed by sequential washing with water, 0.1 M HCl, and water once again. The organic layer was then dried with Na<sub>2</sub>SO<sub>4</sub>, and after filtration the solvent was removed under vacuum to yield glycine conjugate of betulinic acid (231.4 mg). The choline hydroxide was prepared from choline chloride following an anion-exchange column approach used previously [15]. The glycinylated betulinic acid, BA4, was dissolved in 100 ml of THF/H<sub>2</sub>O (4:1, v/v), followed by the addition of a five-fold molar excess of choline hydroxide (0.55 g). The reaction mixture



benzalkonium salt of betulinic acid-glycine [benzalkonium][BA-Gly] / BA3



[cholinium][betulinate] / BA5

Fig. 1. Structures of betulinic acid (BA1) and ionic derivatives (BA2–BA5).

was stirred at room temperature for a duration of 24 h. THF was removed by rotary evaporation. The crude product was extracted into 100 ml of ethyl acetate, and washed with 100 ml of distilled water to remove the excess choline hydroxide. The organic layer was dried and ethyl acetate was removed under vacuum to give the cholinium salt of betulinic acid-glycine([Choline][BA-Gly], (**BA2**, 171 mg) (Fig. 1). Using the above reaction strategies, we prepared the benzalkonium salt of glycinylated betulinic acid ([Bzk][BA-Gly], **BA3**, and cholinium betulinate ([Choline][BA], **BA5**) starting with betulinic acid. These compounds were used to treat mouse fibroblast cells.

**Treatment of 43D cells with betulinic acid and its derivatives.** Establishment of 43D mouse fibroblast cell line stably infected with the wild-type M-MuLV was described previously [20]. The 43D cells were cultured with DMEM media containing 10% Hyclone calf serum (GE Healthcare Life Sciences, PA), 100 IU/ml of Penicillin and 100 ug/ml of Streptomycin (Corning, VA) at 37 °C with 5% CO<sub>2</sub>. Cells were cultivated in 60 mm and 100 mm BioLite cell culture treated dishes (Thermo Scientific, MA, USA). The 43D cells were treated with betulinic acid, its derivatives, or benzalkonium chloride (BKC) for 24 h, and then the media were replaced. The cells and viruses were gathered 24 h further incubation after the second treatment of respective compounds.

Detection of MuLV and beta-Tubulin. The sodium dodecyl sulfate (SDS) samples were prepared from the cells and the viruses released into media. The proteins were extracted from the cells with radioimmunoprecipitation assay (RIPA) buffer which is a common buffer used to lyse cultured mammalian cells. This buffer was composed of 20 mM Tris-HCl (pH 8.0), 150 mM NaCl, 1% NP-40, 1% sodium deoxycholate, 2.5 mM sodium pyrophosphate, and 1 x AMRESCO Protease Inhibitors (AMRESCO LLC, Solon OH, USA). In order to obtain SDS samples, 43D cells were incubated with 120 µl of RIPA on ice. The cell debris was span down by centrifugation at 13,000 rpm for 15 min at 4 °C. The supernatant was transferred into new micro-centrifuge tubes, and 4 x SDS buffer was added and boiled for 3 min. Separately, the viruses in media were concentrated by ultracentrifugation at 25,000 rpm with a Beckman SW28 ultracentrifuge rotor for 1 h at 4 °C [21]. All media were removed, and the viruses were suspended in 160  $\mu$ l of 1 x SDS sample buffer (62.5 mM Tris-HCl (pH 6.8), 2.5% SDS, 0.002% bromophenol blue,  $0.7135 \text{ M} \beta$ -mercaptoethanol, 10% glycerol). The viral proteins were detected by SDS-PAGE and western blots with the rabbit serum for M-MuLV p30<sup>CA</sup> [22], and anti-rabbit IgG conjugated with horseradish peroxidase. The images were acquired by the SuperSignal West Femto Maximum Sensitivity Substrate (Thermo Scientific) and Gel Doc<sup>TM</sup> XR + Gel Documentation System (Bio-Rad, CA, USA). To quantify the viral release efficiency, each Pr65<sup>Gag</sup> and p30<sup>CA</sup> (a precursor and a cleaved MuLV Gag proteins) band in the cells and media was quantified with the densitometry software AlphaEaseFC (Alpha Innotech, CA, USA), and the percentage of released p30<sup>CA</sup> divided by the total Gag proteins (Pr65<sup>Gag</sup> and p30<sup>CA</sup>) in cells and media was calculated [20]. Different exposures of the blots were analyzed to ensure that densitometry was in the linear range. Beta-Tubulin was detected by SDS-PAGE and western blots with antibeta-Tubulin (Cell Signaling Technology, MA) and goat antimouse IgG conjugated with horseradish peroxidase (ImmunoReagents, NC).

AlamarBlue Cell Viability Assay. Cell viability was assessed using the AlamarBlue Cell Viability Assay [23] from Bio-Rad following the manufacturer's protocols. Briefly,  $5 \times 10^3$  43D cells were placed on 96-well plates (100 µl of media/well). After overnight incubation, the cells were treated with DMSO, 10 µM, 30 µM and 100 µM of betulinic acid, its derivatives, or BKC for 48 h respectively, then 10 µl of AlamarBlue solution was added to the wells. Following 2-h incubation, the reaction was stopped by 50 µl

of 3% SDS. Absorbance of 570 nm/600 nm was measured by NanoDrop 2000 UV—Vis Spectrophotometer (Thermo Scientific) to determine reduction of resazurin dye. The DMSO treated cells were set to 100% and then the viability readings from the treated wells were calculated as a percentage of the control.

**Statistical analysis**. Students T-test was used to evaluate viral release efficiency from 43D cells treated with betulinic acid or its derivatives.

# 3. Results

To determine the inhibition of betulinic acid and its ionic derivatives against M-MuLV release, 43D cells were treated with 30 µM of these compounds for 48 h. Viral Gag protein is translated as poly-Gag, Pr65<sup>Gag</sup> which is further cleaved into some smaller Gag proteins such as p30<sup>CA</sup> through the maturation of viruses [2]. Since the cleavage of Gag proteins to yield p30<sup>CA</sup> occurs during budding/ viral release, the majority of Gag proteins in secreted viruses is p30<sup>CA</sup> (Fig. 2A). Poly-Gag proteins Pr65<sup>Gag</sup> in cells and p30<sup>CA</sup> in cells and viruses (media) were detected by SDS-PAGE and western blots with rabbit polyclonal anti-p30<sup>CA</sup> antibodies [22]. Compounds BA1, BA2, and BA5 did not show any significant impact on the intracellular Gag production and its processing, but compounds BA3 and BA4 decreased the intracellular Gag proteins. The signals for beta-Tubulin in the cells treated with compounds BA3 and BA4 exhibited a small decrease. Compounds BA3 and BA4 decreased the amount of p30<sup>CA</sup> in media, but the virus release efficiency was not affected by any of the compounds BA1-BA5 (Fig. 2B). Treatment of 10 µM of betulinic acids and its derivatives did not change Gag in



**Fig. 2.** Effect of betulinic acid and its ionic derivatives on MuLV replication. (A) 43D cells were treated with 30  $\mu$ M of the compounds for 48 h, and Gag proteins in cells and media were detected by western blots using the rabbit serum for anti-p30<sup>CA</sup>. (B) The intensity of Gag signals detected by western blots was quantified by immunodensitometry and the viral release efficiency was calculated. The figure shows the relative virus release efficiency (% Gag release) with standard deviation.

cells and in viruses as detected by western blots (data not shown).

It was reported that betulinic acid suppressed some signaling pathways involved in the cellular metabolism and could induce cell death [24]. To assess the relationship between the inhibition of M-MuLV Gag production and the general cellular metabolism in 43D cells treated with these compounds, cell growth was measured by AlmarBlue cell proliferation assay which monitored the reducing environment of living cells [23]. The cells were treated with different concentrations of betulinic acids and its derivatives for 48 h. Compounds BA3 and BA4 reduced the cellular activities by 35% at 30  $\mu$ M and by 95% at 100  $\mu$ M, but other compounds did not show a strong inhibitory effect at 30 µM (Fig. 3), which was correlated with the beta-Tubulin in the 43D cells treated with  $30 \,\mu\text{M}$  and  $100 \,\mu\text{M}$  of compounds BA3 and BA4 for 48 h (Fig. 2A). These data suggest that compounds BA3 and BA4 could decrease the M-MuLV Gag production by inhibiting the cellular metabolisms. Since both compounds contain the benzalkonium cation, we tested the inhibitory effect of benzalkonium cation on cell growth. The 43D cells treated with BKC impaired metabolism in a dosedependent manner, and BA2 did not show synergistic effects with BKC (Fig. 4A). BKC decreased the signals for Gag and beta-Tubulin in cells and p30<sup>CA</sup> in media (Fig. 4B), which suggested that the benzalkonium cation inhibited viral production by decreasing the metabolism in 43D cells, and compounds BA3, BA4 and BKC seem not to affect intracellular Gag production and its processing.

## 4. Discussion

Our results demonstrated that two ionic derivatives of betulinic acid (compounds BA3 and BA4) reduced the cellular metabolisms, and decreased the MuLV production from 43D cells without alteration of the virus release efficiency or the maturation of viral particles. It was reported that both betulinic acid and platanic acid isolated from the leaves of Syzigium claviforum inhibited human immunodeficiency virus (HIV) replication in H9 lymphocyte cells [7]. A subsequent study [8] demonstrated that the major inhibitory steps in HIV-1 replication by betulinic acid and its derivatives are membrane fusion, viral assembly, and maturation. One betulinic acid derivative with alteration at C3, 3-O-(3,3-dimethylsuccinyl) betulinic acid/PA-457/DSB blocks a late step of virus replication through inhibiting the conversion of p25<sup>CA-SP1</sup> to p24 [25]. Cleavages of the precursor form of Gag including p24 and SP1 Gag spacer peptide is crucial to the structural alterations necessary for the formation of mature HIV-1 particles. The study using lentiviruses and their mutants responsible for DSB resistance indicated that Valine and Leucine immediately flanking the p25<sup>CA-SP1</sup> cleavage



**Fig. 3.** Cytotoxicity of betulinic acid and its ionic derivatives in 43D cells. 43D cells were treated with different doses of betulinic acid and its ionic derivatives for 48 h. The effect of these compounds on cell growth was measured by the AlmarBlue cell proliferation assay. The figure illustrates the relative cell proliferation to the control. The averages of two experiments are shown.

site determined the viral sensitivity to DSB [8,25,26]. We confirmed that betulinic acid inhibited the cleavage of  $p25^{CA-SP1}$ , and increased the amount of  $p25^{CA-SP1}$  by western blots (data not shown), but did not observe the inhibition of Gag cleavage in MuLV with betulinic acid and its ionic derivatives (Fig. 2A) although our ionic derivatives of betulinic acid displayed improved water solubilities, and exhibited a stronger antiviral activity against herpes simplex virus type-2 (HSV-2) than betulinic acid [14]. MuLV Gag is cleaved during virus maturation into MA, p12, p30<sup>CA</sup>, and NC [2], while amino acids flanking the MA-p12 are Leucine and Tyrosine, amino acids flanking the p12-p30<sup>CA</sup> are Alanine and Phenylalanine, and amino acids flanking the p30<sup>CA</sup>-NC are Leucine and Leucine. Since DSB failed to show any significant inhibition of HIV-1 PR in vitro using a synthetic peptide and recombinant Gag [25,27], this compound seems to target unique sequences in Gag to prevent viral maturation through unknown mechanisms. Our results are in agreement with the previous data indicating that DSB targeted the specific amino acid sequence flanking to the cleavage site, and suggest that amino acid sequence found in MuLV Gag and MuLV protease are not targeted by betulinic acid and its derivatives.

As shown in Fig. 3, compounds BA3 and BA4 reduced the cellular metabolism, and inhibited the growth of 43D cells, which was largely due to the amount of beta-Tubulin in cells treated with the compounds (Fig. 2). Compounds BA3 and BA4 are quaternary ammonium salts that contain the benzalkonium cation, and our experiments using BKC suggested that the benzalkonium cation in BA3 and BA4 showed cytotoxic effects (Fig. 4). Therefore, these



**Fig. 4.** Effects of benzalkonium chloride on 43D growth and MuLV replication. (A) 43D cells were treated with different doses of benzalkonium chloride (BKC) and the cholinium salt of betulinic acid-glycine, BA2 for 48 h. The effect of the compounds on cell growth was measured by the AlmarBlue cell proliferation assay. The figure illustrates the relative cell proliferation to the control. The averages of two experiments are shown. (B) 43D cells were treated with  $30 \,\mu$ M of the compounds for 48 h, and Gag proteins in cells and media were detected by western blots using the rabbit serum for anti-p $30^{CA}$ .

observations are consistent with our other data suggesting that cytotoxic effects of BA3 and BKC are comparable for prostate cancer cells (unpublished data). BKC has been reported to show antimicrobial activities against bacteria [28], HIV-1 [29] and fungi [30] through a variety of cellular changes such as cell membrane damage, adenosine triphosphate depletion, generation of oxidative stress, and cell apoptosis. Our data imply that such effects induced by BKC seem not affect maturation and release of MuLV particles (Fig. 4).

In summary, our current experiments using betulinic acid and its derivatives outline the differences of viral maturation between MuLV and HIV-1 through evaluating the MuLV production in cells dosed with betulinic acid compounds.

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## **Transparency document**

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## References

- T.E. Spencer, M. Palmarini, Application of next generation sequencing in mammalian embryogenomics: lessons learned from endogenous betaretroviruses of sheep, Anim. Reprod. Sci. 134 (2012) 95–103.
- [2] A. Rein, Murine leukemia viruses: objects and organisms, Adv. virol. 2011 (2011) 403419.
- [3] J.M. Coffin, S.H. Hughes, H.E. Varmus, Retroviruses, Cold Spring Harbor (New York), Cold Spring Harbor Laboratory Press, 1997.
- [4] H. Fan, Leukemogenesis by Moloney murine leukemia virus: a multistep process, Trends Microbiol. 5 (1997) 74–82.
- [5] R.H. Cichewicz, S.A. Kouzi, Chemistry, biological activity, and chemotherapeutic potential of betulinic acid for the prevention and treatment of cancer and HIV infection, Med. Res. Rev. 24 (2004) 90–114.
- [6] P. Yogeeswari, D. Sriram, Betulinic acid and its derivatives: a review on their biological properties, Curr. Med. Chem. 12 (2005) 657–666.
- [7] T. Fujioka, Y. Kashiwada, R.E. Kilkuskie, L.M. Cosentino, L.M. Ballas, J.B. Jiang, W.P. Janzen, I.S. Chen, K.H. Lee, Anti-AIDS agents, 11. Betulinic acid and platanic acid as anti-HIV principles from Syzigium claviflorum, and the anti-HIV activity of structurally related triterpenoids, J Nat Prod 57 (1994) 243–247.
- [8] C. Aiken, C.H. Chen, Betulinic acid derivatives as HIV-1 antivirals, Trends Mol. Med. 11 (2005) 31–36.
- [9] S. Jäger, K. Winkler, U. Pfüller, A. Scheffler, Solubility studies of oleanolic acid and betulinic acid in aqueous solutions and plant extracts of Viscum album L, Planta Med. 73 (2007) 157–162.
- [10] H. Zhao, C.L. Jones, J.V. Cowins, Lipase dissolution and stabilization in etherfunctionalized ionic liquids, Green Chem. 11 (2009) 1128–1138.

- [11] I. Baglin, A.-C. Mitaine-Offer, M. Nour, K. Tan, C. Cave, M.-A. Lacaille-Dubois, A review of natural and modified betulinic, ursolic and echinocystic acid derivatives as potential antitumor and anti-HIV agents, Mini Rev. Med. Chem. 3 (2003) 525–539.
- [12] R. Mukherjee, V. Kumar, S.K. Srivastava, S.K. Agarwal, A.C. Burman, Betulinic acid derivatives as anticancer agents: structure activity relationship, Anti-Canc. Agents Med. Chem. (Formerly Current Med. Chem.-Anti-Cancer Agents) 6 (2006) 271–279.
- [13] H. Zhao, G.A. Baker, Z. Song, O. Olubajo, T. Crittle, D. Peters, Designing enzymecompatible ionic liquids that can dissolve carbohydrates, Green Chem. 10 (2008) 696–705.
- [14] R.J. Visalli, H. Ziobrowski, K.R. Badri, J.J. He, X. Zhang, S.R. Arumugam, H. Zhao, Ionic derivatives of betulinic acid exhibit antiviral activity against herpes simplex virus type-2 (HSV-2), but not HIV-1 reverse transcriptase, Bioorg. Med. Chem. Lett 25 (2015) 3168–3171.
- [15] H. Zhao, S.S. Holmes, G.A. Baker, S. Challa, H.S. Bose, Z. Song, lonic derivatives of betulinic acid as novel HIV-1 protease inhibitors, J. Enzyme Inhib. Med. Chem. 27 (2012) 715–721.
- [16] C. Suresh, H. Zhao, A. Gumbs, C.S. Chetty, H.S. Bose, New ionic derivatives of betulinic acid as highly potent anti-cancer agents, Bioorg. Med. Chem. Lett 22 (2012) 1734–1738.
- [17] P.T. Ho, K.Y. Ngu, An effective synthesis of N-(9-fluorenylmethyloxycarbonyl). alpha.-amino aldehydes from S-benzyl thioesters, J. Org. Chem. 58 (1993) 2313–2316.
- [18] P.C. Rodrigues, T. Roth, H.H. Fiebig, C. Unger, R. Mülhaupt, F. Kratz, Correlation of the acid-sensitivity of polyethylene glycol daunorubicin conjugates with their in vitro antiproliferative activity, Bioorg. Med. Chem. 14 (2006) 4110–4117.
- [19] A. Ammazzalorso, R. Amoroso, G. Bettoni, B. De Filippis, L. Giampietro, M. Pierini, M.L. Tricca, Asymmetric synthesis of (S)-ibuprofen by esterification with amides of (S)-lactic acid as chiral auxiliaries: experimental and theoretical results, Tetrahedron Lett. 43 (2002) 4325–4328.
- [20] T. Nitta, Y. Kuznetsov, A. McPherson, H. Fan, Murine leukemia virus glycosylated Gag (gPr80gag) facilitates interferon-sensitive virus release through lipid rafts, Proc. Natl. Acad. Sci. U. S. A. 107 (2010) 1190–1195.
- [21] T. Nitta, A. Hofacre, S. Hull, H. Fan, Identification and mutational analysis of a Rej response element in Jaagsiekte sheep retrovirus RNA, J. Virol. 83 (2009) 12499–12511.
- [22] N. Mueller-Lantzsch, H. Fan, Monospecific immunoprecipitation of murine leukemia virus polyribosomes: identification of p30 protein-specific messenger RNA, Cell 9 (1976) 579–588.
- [23] S.N. Rampersad, Multiple applications of Alamar Blue as an indicator of metabolic function and cellular health in cell viability bioassays, Sensors 12 (2012) 12347–12360.
- [24] T. Xu, Q. Pang, D. Zhou, A. Zhang, S. Luo, Y. Wang, X. Yan, Proteomic investigation into betulinic acid-induced apoptosis of human cervical cancer HeLa cells, PLoS One 9 (2014) e105768.
- [25] F. Li, R. Goila-Gaur, K. Salzwedel, N.R. Kilgore, M. Reddick, C. Matallana, A. Castillo, D. Zoumplis, D.E. Martin, J.M. Orenstein, G.P. Allaway, E.O. Freed, C.T. Wild, PA-457: a potent HIV inhibitor that disrupts core condensation by targeting a late step in Gag processing, Proc. Natl. Acad. Sci. U.S.A. 100 (2003) 13555–13560.
- [26] J. Zhou, C.H. Chen, C. Aiken, The sequence of the CA-SP1 junction accounts for the differential sensitivity of HIV-1 and SIV to the small molecule maturation inhibitor 3-O-{3',3'-dimethylsuccinyl}-betulinic acid,, Retrovirology [electronic resource] 1 (2004) 15.
- [27] T. Kanamoto, Y. Kashiwada, K. Kanbara, K. Gotoh, M. Yoshimori, T. Goto, K. Sano, H. Nakashima, Anti-human immunodeficiency virus activity of YK-FH312 (a betulinic acid derivative), a novel compound blocking viral maturation, Antimicrob. Agents Chemother. 45 (2001) 1225–1230.
- [28] A. Houari, P. Di Martino, Effect of chlorhexidine and benzalkonium chloride on bacterial biofilm formation, Lett. Appl. Microbiol. 45 (2007) 652–656.
- [29] M.A. Wainberg, B. Spira, G. Bleau, R. Thomas, Inactivation of human immunodeficiency virus type 1 in tissue culture fluid and in genital secretions by the spermicide benzalkonium chloride, J. Clin. Microbiol. 28 (1990) 156–158.
- [30] Y. Xu, Y. He, X. Li, C. Gao, L. Zhou, S. Sun, G. Pang, Antifungal effect of ophthalmic preservatives phenylmercuric nitrate and benzalkonium chloride on ocular pathogenic filamentous fungi, Diagn. Microbiol. Infect. Dis. 75 (2013) 64–67.