MedChemComm

CONCISE ARTICLE

Cite this: Med. Chem. Commun., 2014, 5, 1892

Structure-activity relationship studies of SETD8 inhibitors†‡

Angi Ma,^a Wenyu Yu,^d Yan Xiong,^a Kyle V. Butler,^a Peter J. Brown^d and Jian Jin^{*abc}

SETD8 (also known as SET8, PR-SET7, or KMT5A (lysine methyltransferase 5A)) is the only known lysine methyltransferase that catalyzes the monomethylation of histone H4 lysine 20 (H4K20). In addition to H4K20, SETD8 monomethylates non-histone substrates such as the tumor suppressor p53 and the proliferating cell nuclear antigen (PCNA). Because of its role in regulating diverse biological processes, SETD8 has been pursued as a potential therapeutic target. We recently reported the first substrate-competitive SETD8 inhibitor, UNC0379 (1), which is selective for SETD8 over 15 other methyltransferases. We characterized this inhibitor in a battery of biochemical and biophysical assays. Here we describe our comprehensive structure–activity relationship (SAR) studies of this chemical series. In addition to 2- and 4-substituents, we extensively explored 6- and 7-substituents of the quinazoline scaffold. These SAR studies led to the discovery of several new compounds, which displayed similar potencies as compound 1 and interesting SAR trends.

Received 21st July 2014 Accepted 16th September 2014

DOI: 10.1039/c4md00317a

www.rsc.org/medchemcomm

Introduction

Post-translational modifications (PTMs) of histones are critical in regulating gene expression and transcription.^{1–3} Among a myriad of PTMs, histone lysine methylation has been recognized as a major mechanism in chromatin regulation. Histone lysine methylation typically takes place at the N-terminal tails of core histone proteins (H3, H4, H2A, and H2B) and is catalyzed by protein lysine methyltransferases (PKMTs). It is worth noting that PKMTs also methylate many non-histone substrates.⁴

PKMTs have been increasingly recognized as a class of potential therapeutic targets by the medicinal chemistry and drug discovery community. Consequently, a number of selective inhibitors of PKMTs have been discovered during recent years.^{5–31} Related to PKMTs, protein arginine methyl-transferases (PRMTs) catalyze the methylation of arginine residues of histone and non-histone proteins.³² A number of selective inhibitors of PRMTs have also been reported.^{33–35}

SETD8 (also known as SET8, PR-SET7, or KMT5A (lysine methyltransferase 5A)), first characterized in 2002, is the only

known PKMT that catalyzes the monomethylation of histone H4 lysine 20 (H4K20).³⁶⁻³⁸ Monomethylation of H4K20 (H4K20me) and SETD8 has been implicated in DNA damage response and cell cycle progression.³⁸ In addition, SETD8 promotes epithelial-mesenchymal transition (EMT) by physically associating with TWIST, a master regulator of EMT.³⁹ SETD8 also monomethylates lysine 382 (K382) of the tumor suppressor p53 and lysine 248 (K248) of the proliferating cell nuclear antigen (PCNA) and plays a potential role in human carcinogenesis.^{40,41}

HEMISTRY

View Article Online

View Journal | View Issue

We recently reported UNC0379 (1) as the first substratecompetitive small-molecule inhibitor of SETD8 (Fig. 1).⁴² Compound 1 is active in multiple biochemical (*e.g.*, radioactive methyl transfer and microfluidic capillary electrophoresis) and biophysical (*e.g.*, isothermal titration calorimetry and surface plasmon resonance) assays, and importantly, selective for SETD8 over 15 other methyltransferases.⁴² The only other known selective inhibitor of SETD8 is a marine natural product nahuoic acid A, which is competitive with the co-factor *S*-adenosyl-L-methionine (SAM) (Fig. 1).²¹ In this article, we describe

[‡] Electronic supplementary information (ESI) available: Synthesis procedures, characterization data, and biochemical assays. See DOI: 10.1039/c4md00317a



Fig. 1 Structures of the known selective inhibitors of SETD8.

^aDepartment of Structural and Chemical Biology, Icahn School of Medicine at Mount Sinai, New York, NY, 10029, USA. E-mail: jian.jin@mssm.edu; Tel: +1-212-659-8699 ^bDepartment of Oncological Sciences, Icahn School of Medicine at Mount Sinai, New York, NY, 10029, USA

^cDepartment of Pharmacology and Systems Therapeutics, Icahn School of Medicine at Mount Sinai, New York, NY, 10029, USA

^dStructural Genomics Consortium, University of Toronto, Toronto, Ontario, M5G 1L7, Canada

[†] This article is part of the MedChemComm "Epigenetics" themed issue.

View Article Online MedChemComm

our comprehensive structure–activity relationship (SAR) studies of the quinazoline template represented by compound **1**, which resulted in the discovery of interesting SAR trends and novel analogs with similar potencies as compound **1**.

Results and discussion

Our strategy for studying the SAR of the UNC0379 series was to extensively explore the 2-, 4-, 6-, and 7-substituents (Fig. 2). We previously reported the initial SAR results of the 2- and 4-substituents.⁴² In these new studies, we investigated not only additional 2- and 4-substituents, but also 6- and 7-substituents, the two regions that have not been previously explored for SETD8.

We first explored the 4-amino moiety of the quinazoline scaffold. Compounds **1–15** (Scheme 1 and Table 1) were synthesized from commercially available 2,4-dichloro-6,7-dimethoxyquinazoline and corresponding amines in good yields. As previously reported,⁶ we displaced the 4-chloro group with the first set of amines at room temperature. The 2-chloro group was substituted by the second set of amines under microwave heating conditions, yielding the desired 2,4-diamino-6,7-dimethoxyquinazolines (Scheme 1).

The ring size of the terminal cyclic amino group did not have a significant impact on SETD8 potency (Table 1). Pyrrolidine (compound 1), piperidine (compound 2), and azepane (compound 3) resulted in similar potencies. The replacement of the cyclic amino group with an acyclic amino group such as the dimethyl amine group (compound 4) did not lead to a significant potency change either. However, increasing the length of the side chain from 5 to 6 carbons (compound 4 versus compound 5) resulted in a decrease in potency. We previously reported that decreasing the length of the side chain also led to a decrease in potency.42 Interestingly, replacing the dimethyl amine group with the primary amino group slightly decreased the potency (compound 5 versus compound 6). We next explored various conformation-constrained analogs and found that these compounds (7-10) were not as potent as compound 1. In addition to exploring the length of the side chain, we also attempted to replace the straight 5-carbon chain with several amide-containing linkers and found that amide 11 was significantly less potent than compound 1 and amide 12 was completely inactive. We previously reported that compound 13 was weakly active against SETD8.42 Consistent with the results



Fig. 2 The four highlighted regions were explored for studying the SAR of the UNC0379 series.



Scheme 1 Typical synthesis of 2,4-diamino-6,7-dimethoxyquinazolines. Reagents and conditions: (a) R^1 amines, THF, *N*,*N*-diisopropylethylamine, room temperature; (b) R^2 amines, *n*-BuOH, DIPEA, microwave, heat.

of amide **12**, amides **14** and **15** did not display any activity against SETD8. Taken together, these results suggest that: (1) the terminal pyrrolidine group can be modified without potency loss and (2) the 5-carbon linker is optimal. In addition, we previously demonstrated that the basicity of the pyrrolidine nitrogen was important for maintaining potency for SETD8.⁴²

We next investigated various 2-substituents at the quinazoline core (Scheme 2 and Table 2). Synthesis of compounds **16– 18** was described previously.⁴² Compounds **20–25** were prepared according to the synthesis route illustrated in Scheme 1. Buchwald–Hartwig amination reaction conditions⁴³ were used to synthesize compounds **19** and **26–32** from the known intermediate 2-chloro-6,7-dimethoxy-*N*-(5-(pyrrolidin-1-yl)pentyl)quinazolin-4-amine⁴² and commercially available amines (Scheme 2). Compounds **19** and **26–32** could not be generated using standard nucleophilic conditions (Scheme 1) in good yields. A Suzuki coupling reaction⁴⁴ was used to prepare compounds **33– 36** from the same intermediate and commercially available aromatic boronic acids (Scheme 2).

As shown in Table 2, replacing the pyrrolidine (compound 1) with either piperidine (compound 16) or azepane (compound 17) resulted in a significant loss of potency, suggesting that a larger group is disfavored. On the other hand, the replacement of the pyrrolidine (compound 1) with the dimethyl amine group (compound 18) did not lead to a significant potency change. These results were reported previously.⁴² Interestingly, the 3,3difluoroazetidine (compound 19) resulted in a large loss of potency, possibly due to the increase of the compound's polarity. We attempted to synthesize a simple azetidine analog, but found that the target molecule was unstable and could not be isolated. Adding a methyl substituent to the pyrrolidine (compound 20) led to about two-fold drop in potency. We previously reported that compound 21 displayed some activity for SETD8.42 Based on this result, we attempted to introduce a pyrrolidine with a 2-5-carbon linker and were disappointed to find that these compounds (22-25) did not exhibit any activity for SETD8. We also explored unsubstituted and substituted Nmethylanilines and anilines. As shown in Table 2, unsubstituted N-methylaniline (compound 26) and N-methylaniline with either an electron-donating group (methoxy, compound 27) or an electron-withdrawing group (nitro, compound 28) at the para-position were inactive. Similarly, unsubstituted aniline (compound 29) and anilines with either an electron-donating group (compound 30) or an electron-withdrawing group (compounds 31 and 32) at the para-position did not display any activity. Finally, we attempted to replace the pyrrolidine with a phenyl or substituted phenyl group containing an

Table 1 SAR of the 4-amino moiety

MeO

MeO

 HN^{R^1}

Compound



SETD8 IC_{50}^{a} (μM)



 a IC_{50} determination experiments were performed in duplicate. b IC_{50} value was reported previously. 42



Scheme 2 Synthesis of compounds 19 and 26–36. Reagents and conditions: (a) amines, $Pd(OAc)_2$, (+)-BINAP, Cs_2CO_3 , THF, microwave, heat; (b) aromatic boronic acid, $Pd(PPh_3)_4$, K_2CO_3 , dioxane–water, microwave, heat. Ar, aromatic ring.

electron-donating or an electron-withdrawing group at the *para*position, but found that none of these compounds (**33–36**) were active against SETD8. Taken together, these results suggest that the SAR at the 2-substituent is very tight. The vast majority of the modifications we made led to a significant or complete loss of potency for SETD8.

We also extensively explored the 6- and 7-substituents (Scheme 3 and Table 3), which were not studied previously. Compounds **37–45** were prepared from different 6,7-substituted 2,4-dichloroquinazolines (synthesis of these intermediates is detailed in the ESI‡) according to the synthesis route illustrated in Scheme 1. Synthesis of compounds **46–49** is outlined in Scheme 3. Briefly, debenzylation of compound **45** *via* hydrogenation produced compound **46**. Nucleophilic substitution reactions between phenol **46** and various alkyl bromides afforded compounds **47–50**.

We found that the removal of both methoxy groups (compound **37**) or the 6-methoxy group (compound **38**) completely abolished activity (Table 3). Interestingly, the removal of the 7-methoxy group (compound **39**) led to about 7-



Table 2 (Contd.)





 a IC_{50} determination experiments were performed in duplicate. b IC_{50} value was reported previously. 42

fold loss in potency, but it retained some activity against SETD8. These results suggest that the 6-methoxy group may be more important for maintaining SETD8 potency compared with the 7-methoxy group.

We next investigated several 6-substituents while holding the 7-methoxy group constant and found that the replacement of



Scheme 3 Synthesis of compounds 46-49. Reagents and conditions: (a) H₂, Pd/C, EtOH, room temperature; (b) RBr, K₂CO₃, DMF, room temperature.

Table 3 SAR of the 6- and 7-substituted groups



 a IC_{50} determination experiments were performed in duplicate. b IC_{50} value was reported previously. 42

the 6-methoxy group with the 6-ethoxy group did not result in a significant change in potency (compound **1** *versus* compound **40**). On the other hand, the 6-isopropoxy group (compound **41**) and the 6-chloro group (compound **42**) led to about 8-fold and 6-fold loss in potency, respectively, suggesting that a larger group or a less electron-donating group is disfavored at this position.

Lastly, we explored various 7-substituents while holding the 6-methoxy group constant. Slightly increasing the size of the 7methoxy group to the 7-ethoxy group (compound **43**) did not

significantly change the potency. On the other hand, the larger 7-isopropoxy group (compound 44) and the 7-benzyloxy group (compound 45) led to more than 6-fold and 17-fold potency drops, respectively. Interestingly, the 7-hydroxy group (compound 46) completely abolished activity against SETD8. We next studied whether a linear chain could be tolerated at the 7-position. We were pleased to find that compound 47, which contains the 7-methoxyethyloxy group, was as potent as compound 1. However, the 7-methoxypropoxy group (compound 48) and the 7-hydroxypropoxy group (compound 49) led to a significant decrease in potency. Interestingly, compound 50, which contains the 7-formylaminoethyloxy group, retained the same potency as compounds 1 and 47. Taken together, these results suggest that the 7-position is amenable to modifications and there may be an opportunity to create more potent inhibitors of SETD8 by further exploring this region.

Conclusions

We comprehensively studied the SAR of the quinazoline scaffold represented by UNC0379 (compound 1), which was recently discovered as the first substrate-competitive inhibitor of SETD8. We found a number of interesting SAR trends. They include: (1) at the 4-position, the terminal pyrrolidine group can be modified without potency loss and the 5-carbon linker is optimal; (2) at the 2-position, modifications are generally not tolerated and pyrrolidine and dimethylamino groups are optimal; (3) at the 6position, the methoxy and ethoxy groups are preferred and a larger group or a less electron-donating group is disfavored; and (4) at the 7-position, modifications can be well tolerated and further exploration of this region may result in more potent SETD8 inhibitors. These SAR studies also led to the discovery of several novel compounds (3, 40, 43, 47 and 50), which exhibited similar potencies as compound 1. During the revision of this paper, several novel inhibitors of SETD8 have been reported.45

Acknowledgements

The research described here was supported by the grant R01GM103893 from the U.S. National Institute of General Medical Sciences of the National Institutes of Health. The Structural Genomics Consortium is a registered charity (number 1097737) that receives funds from the Canada Foundation for Innovation, Eli Lilly Canada, GlaxoSmithKline, the Ontario Ministry of Economic Development and Innovation, the Novartis Research Foundation, Pfizer, AbbVie, Takeda, Janssen, Boehringer Ingelheim, Bayer and the Wellcome Trust.

References

- 1 R. A. Copeland, M. E. Solomon and V. M. Richon, *Nat. Rev. Drug Discovery*, 2009, **8**, 724–732.
- 2 C. H. Arrowsmith, C. Bountra, P. V. Fish, K. Lee and M. Schapira, *Nat. Rev. Drug Discovery*, 2012, **11**, 384–400.
- 3 K. Helin and D. Dhanak, Nature, 2013, 502, 480-488.
- 4 S. G. Clarke, Trends Biochem. Sci., 2013, 38, 243-252.

- 5 S. Kubicek, R. J. O'Sullivan, E. M. August, E. R. Hickey, Q. Zhang, M. L. Teodoro, S. Rea, K. Mechtler, J. A. Kowalski, C. A. Homon, T. A. Kelly and T. Jenuwein, *Mol. Cell*, 2007, 25, 473-481.
- 6 F. Liu, X. Chen, A. Allali-Hassani, A. M. Quinn, G. A. Wasney,
 A. Dong, D. Barsyte, I. Kozieradzki, G. Senisterra, I. Chau,
 A. Siarheyeva, D. B. Kireev, A. Jadhav, J. M. Herold,
 S. V. Frye, C. H. Arrowsmith, P. J. Brown, A. Simeonov,
 M. Vedadi and J. Jin, *J. Med. Chem.*, 2009, 52, 7950–7953.
- 7 Y. Chang, T. Ganesh, J. R. Horton, A. Spannhoff, J. Liu, A. Sun, X. Zhang, M. T. Bedford, Y. Shinkai, J. P. Snyder and X. Cheng, *J. Mol. Biol.*, 2010, 400, 1–7.
- 8 F. Liu, X. Chen, A. Allali-Hassani, A. M. Quinn, T. J. Wigle, G. A. Wasney, A. Dong, G. Senisterra, I. Chau, A. Siarheyeva, J. L. Norris, D. B. Kireev, A. Jadhav, J. M. Herold, W. P. Janzen, C. H. Arrowsmith, S. V. Frye, P. J. Brown, A. Simeonov, M. Vedadi and J. Jin, *J. Med. Chem.*, 2010, **53**, 5844–5857.
- M. Vedadi, D. Barsyte-Lovejoy, F. Liu, S. Rival-Gervier, A. Allali-Hassani, V. Labrie, T. J. Wigle, P. A. DiMaggio, G. A. Wasney, A. Siarheyeva, A. Dong, W. Tempel, S.-C. Wang, X. Chen, I. Chau, T. Mangano, X.-P. Huang, C. D. Simpson, S. G. Pattenden, J. L. Norris, D. B. Kireev, A. Tripathy, A. Edwards, B. L. Roth, W. P. Janzen, B. A. Garcia, A. Petronis, J. Ellis, P. J. Brown, S. V. Frye, C. H. Arrowsmith and J. Jin, *Nat. Chem. Biol.*, 2011, 7, 566–574.
- 10 F. Liu, D. Barsyte-Lovejoy, A. Allali-Hassani, Y. He, J. M. Herold, X. Chen, C. M. Yates, S. V. Frye, P. J. Brown, J. Huang, M. Vedadi, C. H. Arrowsmith and J. Jin, *J. Med. Chem.*, 2011, 54, 6139–6150.
- A. D. Ferguson, N. A. Larsen, T. Howard, H. Pollard, I. Green, C. Grande, T. Cheung, R. Garcia-Arenas, S. Cowen, J. Wu, R. Godin, H. Chen and N. Keen, *Structure*, 2011, 19, 1262–1273.
- 12 S. R. Daigle, E. J. Olhava, C. A. Therkelsen, C. R. Majer, C. J. Sneeringer, J. Song, L. D. Johnston, M. P. Scott, J. J. Smith, Y. Xiao, L. Jin, K. W. Kuntz, R. Chesworth, M. P. Moyer, K. M. Bernt, J. C. Tseng, A. L. Kung, S. A. Armstrong, R. A. Copeland, V. M. Richon and R. M. Pollock, *Cancer Cell*, 2011, **20**, 53–65.
- 13 Y. Yao, P. Chen, J. Diao, G. Cheng, L. Deng, J. L. Anglin,
 B. V. V. Prasad and Y. Song, *J. Am. Chem. Soc.*, 2011, 133, 16746–16749.
- 14 Y. Yuan, Q. Wang, J. Paulk, S. Kubicek, M. M. Kemp, D. J. Adams, A. F. Shamji, B. K. Wagner and S. L. Schreiber, ACS Chem. Biol., 2012, 7, 1152–1157.
- S. K. Knutson, T. J. Wigle, N. M. Warholic, C. J. Sneeringer, C. J. Allain, C. R. Klaus, J. D. Sacks, A. Raimondi, C. R. Majer, J. Song, M. P. Scott, L. Jin, J. J. Smith, E. J. Olhava, R. Chesworth, M. P. Moyer, V. M. Richon, R. A. Copeland, H. Keilhack, R. M. Pollock and K. W. Kuntz, *Nat. Chem. Biol.*, 2012, **8**, 890–896.
- 16 M. T. McCabe, H. M. Ott, G. Ganji, S. Korenchuk, C. Thompson, G. S. Van Aller, Y. Liu, A. P. Graves, A. D. Iii, E. Diaz, L. V. Lafrance, M. Mellinger, C. Duquenne, X. Tian, R. G. Kruger, C. F. McHugh, M. Brandt, W. H. Miller, D. Dhanak, S. K. Verma, P. J. Tummino and C. L. Creasy, *Nature*, 2012, **492**, 108–112.

- 17 S. K. Verma, X. Tian, L. V. LaFrance, C. Duquenne, D. P. Suarez, K. A. Newlander, S. P. Romeril, J. L. Burgess, S. W. Grant, J. A. Brackley, A. P. Graves, D. A. Scherzer, A. Shu, C. Thompson, H. M. Ott, G. S. V. Aller, C. A. Machutta, E. Diaz, Y. Jiang, N. W. Johnson, S. D. Knight, R. G. Kruger, M. T. McCabe, D. Dhanak, P. J. Tummino, C. L. Creasy and W. H. Miller, *ACS Med. Chem. Lett.*, 2012, 3, 1091–1096.
- 18 W. Zheng, G. Ibáñez, H. Wu, G. Blum, H. Zeng, A. Dong, F. Li, T. Hajian, A. Allali-Hassani, M. F. Amaya, A. Siarheyeva, W. Yu, P. J. Brown, M. Schapira, M. Vedadi, J. Min and M. Luo, *J. Am. Chem. Soc.*, 2012, **134**, 18004– 18014.
- 19 W. Qi, H. Chan, L. Teng, L. Li, S. Chuai, R. Zhang, J. Zeng, M. Li, H. Fan, Y. Lin, J. Gu, O. Ardayfio, J.-H. Zhang, X. Yan, J. Fang, Y. Mi, M. Zhang, T. Zhou, G. Feng, Z. Chen, G. Li, T. Yang, K. Zhao, X. Liu, Z. Yu, C. X. Lu, P. Atadja and E. Li, *Proc. Natl. Acad. Sci. U. S. A.*, 2012, 109, 21360–21365.
- 20 W. Yu, E. J. Chory, A. K. Wernimont, W. Tempel, A. Scopton,
 A. Federation, J. J. Marineau, J. Qi, D. Barsyte-Lovejoy, J. Yi,
 R. Marcellus, R. E. Iacob, J. R. Engen, C. Griffin, A. Aman,
 E. Wienholds, F. Li, J. Pineda, G. Estiu, T. Shatseva,
 T. Hajian, R. Al-awar, J. E. Dick, M. Vedadi, P. J. Brown,
 C. H. Arrowsmith, J. E. Bradner and M. Schapira, *Nat. Commun.*, 2012, 3, 1288.
- 21 D. E. Williams, D. S. Dalisay, F. Li, J. Amphlett, W. Maneerat, M. A. Chavez, Y. A. Wang, T. Matainaho, W. Yu, P. J. Brown, C. H. Arrowsmith, M. Vedadi and R. J. Andersen, *Org. Lett.*, 2013, 15, 414–417.
- 22 J. L. Anglin, L. Deng, Y. Yao, G. Cai, Z. Liu, H. Jiang, G. Cheng, P. Chen, S. Dong and Y. Song, *J. Med. Chem.*, 2012, 55, 8066–8074.
- 23 K. D. Konze, A. Ma, F. Li, D. Barsyte-Lovejoy, T. Parton, C. J. Macnevin, F. Liu, C. Gao, X. P. Huang, E. Kuznetsova, M. Rougie, A. Jiang, S. G. Pattenden, J. L. Norris, L. I. James, B. L. Roth, P. J. Brown, S. V. Frye, C. H. Arrowsmith, K. M. Hahn, G. G. Wang, M. Vedadi and J. Jin, ACS Chem. Biol., 2013, 8, 1324–1334.
- 24 S. K. Knutson, N. M. Warholic, T. J. Wigle, C. R. Klaus, C. J. Allain, A. Raimondi, M. Porter Scott, R. Chesworth, M. P. Moyer, R. A. Copeland, V. M. Richon, R. M. Pollock, K. W. Kuntz and H. Keilhack, *Proc. Natl. Acad. Sci. U. S. A.*, 2013, **110**, 7922–7927.
- 25 W. Beguelin, R. Popovic, M. Teater, Y. Jiang, K. L. Bunting, M. Rosen, H. Shen, S. N. Yang, L. Wang, T. Ezponda, E. Martinez-Garcia, H. Zhang, Y. Zheng, S. K. Verma, M. T. McCabe, H. M. Ott, G. S. Van Aller, R. G. Kruger, Y. Liu, C. F. McHugh, D. W. Scott, Y. R. Chung, N. Kelleher, R. Shaknovich, C. L. Creasy, R. D. Gascoyne, K. K. Wong, L. Cerchietti, R. L. Levine, O. Abdel-Wahab, J. D. Licht, O. Elemento and A. M. Melnick, *Cancer Cell*, 2013, 23, 677–692.
- 26 F. Liu, D. Barsyte-Lovejoy, F. Li, Y. Xiong, V. Korboukh,
 X. P. Huang, A. Allali-Hassani, W. P. Janzen, B. L. Roth,
 S. V. Frye, C. H. Arrowsmith, P. J. Brown, M. Vedadi and
 J. Jin, *J. Med. Chem.*, 2013, 56, 8931–8942.

- 27 R. F. Sweis, M. Pliushchev, P. J. Brown, J. Guo, F. L. Li, D. Maag, A. M. Petros, N. B. Soni, C. Tse, M. Vedadi, M. R. Michaelides, G. G. Chiang and W. N. Pappano, ACS Med. Chem. Lett., 2014, 5, 205–209.
- 28 S. Garapaty-Rao, C. Nasveschuk, A. Gagnon, E. Y. Chan, P. Sandy, J. Busby, S. Balasubramanian, R. Campbell, F. Zhao, L. Bergeron, J. E. Audia, B. K. Albrecht, J. C. Harmange, R. Cummings and P. Trojer, *Chem. Biol.*, 2013, **20**, 1329–1339.
- 29 S. R. Daigle, E. J. Olhava, C. A. Therkelsen, A. Basavapathruni, L. Jin, P. A. Boriack-Sjodin, C. J. Allain, C. R. Klaus, A. Raimondi, M. P. Scott, N. J. Waters, R. Chesworth, M. P. Moyer, R. A. Copeland, V. M. Richon and R. M. Pollock, *Blood*, 2013, **122**, 1017–1025.
- 30 C. G. Nasveschuk, A. Gagnon, S. Garapaty-Rao,
 S. Balasubramanian, R. Campbell, C. Lee, F. Zhao,
 L. Bergeron, R. Cummings, P. Trojer, J. E. Audia,
 B. K. Albrecht and J.-C. P. Harmange, ACS Med. Chem. Lett., 2014, 5, 378–383.
- 31 Y. He, I. Korboukh, J. Jin and J. Huang, *Acta Biochim. Biophys. Sin.*, 2012, **44**, 70–79.
- 32 M. T. Bedford and S. Richard, Mol. Cell, 2005, 18, 263-272.
- 33 A. Siarheyeva, G. Senisterra, A. Allali-Hassani, A. Dong,
 E. Dobrovetsky, G. A. Wasney, I. Chau, R. Marcellus,
 T. Hajian, F. Liu, I. Korboukh, D. Smil, Y. Bolshan, J. Min,
 H. Wu, H. Zeng, P. Loppnau, G. Poda, C. Griffin, A. Aman,
 P. J. Brown, J. Jin, R. Al-awar, C. H. Arrowsmith,
 M. Schapira and M. Vedadi, *Structure*, 2012, 20, 1425–1435.
- 34 F. Liu, F. Li, A. Ma, E. Dobrovetsky, A. Dong, C. Gao, I. Korboukh, J. Liu, D. Smil, P. J. Brown, S. V. Frye, C. H. Arrowsmith, M. Schapira, M. Vedadi and J. Jin, *J. Med. Chem.*, 2013, 56, 2110–2124.

- 35 J. M. Yost, I. Korboukh, F. Liu, C. Gao and J. Jin, *Curr. Chem. Genomics*, 2011, 5, 72–84.
- 36 K. Nishioka, J. C. Rice, K. Sarma, H. Erdjument-Bromage, J. Werner, Y. Wang, S. Chuikov, P. Valenzuela, P. Tempst, R. Steward, J. T. Lis, C. D. Allis and D. Reinberg, *Mol. Cell*, 2002, 9, 1201–1213.
- 37 J. Fang, Q. Feng, C. S. Ketel, H. Wang, R. Cao, L. Xia, H. Erdjument-Bromage, P. Tempst, J. A. Simon and Y. Zhang, *Curr. Biol.*, 2002, **12**, 1086–1099.
- 38 D. B. Beck, H. Oda, S. S. Shen and D. Reinberg, *Genes Dev.*, 2012, **26**, 325–337.
- 39 F. Yang, L. Sun, Q. Li, X. Han, L. Lei, H. Zhang and Y. Shang, *EMBO J.*, 2012, **31**, 110–123.
- 40 X. Shi, I. Kachirskaia, H. Yamaguchi, L. E. West, H. Wen, E. W. Wang, S. Dutta, E. Appella and O. Gozani, *Mol. Cell*, 2007, 27, 636–646.
- 41 M. Takawa, H.-S. Cho, S. Hayami, G. Toyokawa, M. Kogure, Y. Yamane, Y. Iwai, K. Maejima, K. Ueda, A. Masuda, N. Dohmae, H. I. Field, T. Tsunoda, T. Kobayashi, T. Akasu, M. Sugiyama, S.-i. Ohnuma, Y. Atomi, B. A. J. Ponder, Y. Nakamura and R. Hamamoto, *Cancer Res.*, 2012, 72, 3217–3227.
- 42 A. Ma, W. Yu, F. Li, R. M. Bleich, J. M. Herold, K. V. Butler, J. L. Norris, V. Korboukh, A. Tripathy, W. P. Janzen, C. H. Arrowsmith, S. V. Frye, M. Vedadi, P. J. Brown and J. Jin, *J. Med. Chem.*, 2014, 57, 6822–6833.
- 43 J. P. Wolfe and S. L. Buchwald, *Org. Synth.*, 2004, **10**, 423–430.
- 44 N. Miyaura and A. Suzuki, Chem. Rev., 1995, 95, 2457-2483.
- 45 G. Blum, G. Ibáñez, X. Rao, D. Shum, C. Radu, H. Djaballah, J. C. Rice and M. Luo, *ACS Chem. Biol.*, 2014, DOI: 10.1021/ cb500515r.