Reactivity, Structures, and NMR Spectroscopy of Half-Sandwich Pentamethylcyclopentadienyl Rhodium Amido Complexes Relevant to Transfer Hydrogenation

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Received October 15, 2008

The reactivity, structures, and NMR spectroscopy of a series of compounds relevant to asymmetric transfer hydrogenation, of general formula Cp*RhCl(S,S-4-RC₆H₄SO₂NCHPhCHPhNH₂) (Cp* = η^{5} - C_5Me_5 , S,S-2a R = Me, S,S-2b R = tBu, S,S-2c R = F), have been studied. ¹H/¹⁵N HMQC NMR spectra of 2a-2c were recorded with ¹⁵N in natural abundance making use of the coupling to the C₅Me₅ protons; the coupling constants J_{RhN} were ca. 15 and 20 Hz for the amino and amido nitrogens, respectively. 1 H/ 103 Rh NMR spectra of 2a and 2c were recorded similarly. The chloride ligand in S,S-2a has been shown to be very labile; reaction with CO afforded the cationic complex [Cp*Rh(CO){C(O)N- $(T_s)CHPhCHPhNH_2$][Cl] (3a) as a mixture of diastereomers, S,S,R_{Rh} and S,S,S_{Rh}, with opposite chirality at Rh; reaction with tBuNC gave [Cp*Rh(CNtBu)(S,S-TsCHPhCHPhNH₂)][PF₆] (4a), and reactions with LiBr and KI gave Cp*RhBr(S,S-TsNCHPhCHPhNH₂) (6a) and Cp*RhI(S,S-TsNCHPhCHPhNH₂) (7a), respectively. Complexes S,S-2c, S,S-6a, and S,S-7a have been characterized by X-ray crystallography; the amino N-Rh bond is significantly shorter than the amido N-Rh bond in all cases (difference Δr 0.058(2), 0.085(6), and 0.046(4) Å, respectively), and the complexes all possess intramolecular NH \cdots X (X = Cl, Br, I) hydrogen bonds. The reaction of **2a** with formic acid results in complete displacement of the chelating ligand and the formation of dinuclear $[{Cp*Rh}_2(\mu-H)(\mu-HCO_2)][BPh_4]$ (5a), which was also characterized crystallographically. Reaction of 2c with AgOTf resulted in the formation of $[Cp*Rh(OH_2)(4-FC_6H_4SO_2NCHPhCHPhNH_2)][OTf]([8c+H_2O][OTf]) or [Cp*Rh(4-FC_6H_4SO_2NCHPhCHPhNH_2)]_{2^-}$ $[OTf]_2$ ([8c]₂[OTf]₂) depending upon the conditions employed. [8c \cdot H₂O][OTf] was characterized by X-ray crystallography, and the structure showed that both CHPh centers in the ligand had been racemized, converting the S,S isomer of the starting material into a mixture of both R,R and S,S isomers.

Introduction

Asymmetric transfer hydrogenation catalyzed by halfsandwich complexes with monotosylated diamine ligands was described first by Noyori et al.^{1,2} The mechanism for the transfer hydrogenation of CO double bonds with iPrOH/KOH as the hydrogen donor has been described as outer-sphere metal-ligand bifunctional catalysis;³ a schematic for the $[(p-cymene)RuCl_2]_2/$ TsDPEN (TsDPEN = N-tosyl 1,2-diphenylethylenediamine) system is shown in Figure 1. First, [(p-cymene)RuCl₂]₂ and TsDPEN react to form the catalyst precursor (step 1). The precursor then reacts with base (KOH) to give a 16-electron species (step 2), which is hydrogenated by *i*PrOH (the hydrogen donor), leading to an 18-electron hydride complex (step 3). The latter transfers its hydride to the carbonyl carbon of the substrate and a proton from its amino nitrogen to the oxygen of the substrate, to give the product alcohol and re-form the 16-electron species (step 4). Step 4 is proposed to transfer hydrogen via a



Figure 1. Proposed catalytic cycle for outer-sphere metal–ligand bifunctional catalysis.³

six-membered, pericyclic transition state, without coordination of the substrate to Ru.

Catalytic transfer hydrogenation of C=N bonds using HCO₂H and its salts as the hydrogen donor^{4–12} was first reported by Noyori and co-workers in 1996.^{4,13} Due to its operational simplicity and wide substrate scope, it has become the method of choice for the synthesis of many chiral amines.

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Noyori and co-workers have isolated and characterized all three ruthenium complexes in Figure 1 by X-ray crystallography and have shown that they all reduce acetophenone with comparable activity and selectivity.³ Following this, attempts were made to study the behavior of the analogous Cp*M(III) $(M = Rh \text{ or } Ir, Cp^* = \eta^5 \cdot C_5 Me_5)$ systems;^{14–17} however, these systems are generally more active and the analogous intermediates have proved harder to isolate (the rhodium and iridium systems are also described in patents belonging to Piramal Healthcare).¹⁸⁻²⁰ Tani et al. showed that [Cp*RhCl₂]₂ and TsDPEN react with NaOH to give the 16-electron species Cp*Rh(TsNCHPhCHPhNH),¹⁴ but the rhodium hydride complex proved elusive. For iridium, a bis-amido 16-electron species was obtained as described above for rhodium; the same product was obtained by reaction of the chloride precursor with aqueous KOH in CH_2Cl_2 . The hydride $Cp*Ir(H)(TsNCHPhCHPhNH_2)$ was then prepared by reaction of this 16-electron complex with iPrOH. The reverse reaction of the hydride complex with acetone gave back the bis-amido complex.¹⁴

Ikariya and co-workers reported the synthesis of the related complexes Cp*RhCl(TsNC₆H₁₀NH₂) (where C₆H₁₀ is 1,2disubstituted cyclohexane), Cp*RhCl(TsNCHPhCHPhNH₂), Cp*IrCl(TsNC₆H₁₀NH₂), and Cp*IrCl(TsNCHPhCHPhNH₂), which also act as catalyst precursors for the asymmetric transfer hydrogenation of acetophenones.^{15,16} They showed that the reaction of Cp*IrCl(TsNC₆H₁₀NH₂) with 1 equiv of NaOH in *i*PrOH gave quantitative conversion to the 18-electron hydride $Cp*Ir(H)(TsNC_6H_{10}NH_2)$, which then reacted with acetone to give the bis-amido complex Cp*Ir(TsNC₆H₁₀NH). Recently, Heiden and Rauchfuss exploited the interconversion of Cp* $Ir(TsNCHPhCHPhNH), [Cp*Ir(TsNCHPhCHPhNH_2)]^+, and$ Cp*Ir(H)(TsNCHPhCHPhNH₂), to demonstrate that the hydride complex acts as a catalyst for the conversion of dihydrogen and dioxygen to water.²¹ They also showed that the 16-electron cation [Cp*Ir(TsNCHPhCHPhNH₂)]⁺ reacts with Lewis bases to form simple adducts $[Cp*Ir(TsNCHPhCHPhNH_2)L]^+$ (L = MeCN, NH₃, PAr₃, and CO).²²

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In spite of the successful synthesis of these 16-electron and hydride species, there has still been considerable debate about their role in the transfer hydrogenation of imines. Evidence from isotope labeling supports a monohydride mechanism in which the hydride and the proton undergoing transfer remain distinct for Rh, Ir, and Ru catalysts.^{23–25} On the other hand, it has been much harder to define when the outer-sphere (Noyori mechanism) occurs and when a more conventional inner-sphere mechanism occurs in which the substrate binds directly to the metal.11,26-33

A better understanding of the chemistry of complexes such as Cp*RhCl(TsNCHPhCHPhNH2) (2a) will surely aid our understanding of the catalytic systems, since 2a is generated in situ in catalytic asymmetric transfer hydrogenation reactions (the process is named CATHy, catalytic asymmetric transfer hydrogenation).¹⁸⁻²⁰ It also catalyzes the asymmetric Michael addition reaction^{34,35} and reacts with sodium phenoxide in dichloromethane, albeit very slowly and in low yield, to give a cyclometalated product.36

Our aim was to study the reactivity of the catalyst 2a with a focus on the identification of complexes relevant to transfer hydrogenation. Because of the poor solubility of the complexes derived from *N*-tosyl-1,2-diphenylethylenediamine (TsDPEN) (1a), we synthesized two additional ligand precursors, 4-tBu-C₆H₄SO₂DPEN) (1b) and 4-FC₆H₄SO₂DPEN (1c). The resulting metal complexes were more soluble than those of 1a, and the F atom in **1c** provided an additional NMR handle. We describe the reactions with CO, tBuNC, and AgOTf leading to cationic complexes. We show that the reaction of 2a with HCO₂H displaces the chelating chiral ligand completely. We also include the metathesis reactions that allow replacement of chloride with bromide and iodide. Throughout this work, we place emphasis on structural features including intramolecular hydrogen bonding (as noted previously)^{3,5,15} that we investigate through X-ray crystallography and NMR spectroscopy. We have also recorded ¹⁵N and ¹⁰³Rh NMR spectra as additional probes.

Results and Discussion

Synthesis and Structure of Cp*RhCl(4-RC₆H₄SO₂NCHPh-CHPhNH₂), 2a-2c. The N-aromatic sulfonyl-1,2-diphenylethylenediamine ligands were synthesized by the method of White et al. (Scheme 1).³⁷ Both the R,R and S,S isomers of **1b** and **1c** were prepared, but we show the S,S isomers in the schemes.

The ligands 1a-1c were complexed to rhodium by reaction with [Cp*RhCl₂]₂ to yield Cp*RhCl(4-RC₆H₄SO₂NCHPhCHPh- NH_2 [2a (R = Me), 2b (R = tBu), and 2c (R = F)]. Complex

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2a has been described previously.^{5,15,17,19} Complexes 2a-2c were prepared by treatment of [Cp*RhCl₂]₂ with 1a-1c and NEt₃ in CH₂Cl₂ and were crystallized as red-orange needles. The most interesting features of the ¹H NMR spectra of 2a-2care the chemical shift changes for the NH₂ protons that were observed when the solvent was changed. For 2a, the NH protons are observed at δ 3.46 and 3.88 in CD₂Cl₂, but are shifted to δ 4.42 and 3.60 in CD₃CN (Figure 2). This is consistent with intramolecular hydrogen bonding in CD₂Cl₂, a weak H-bond acceptor, and a more important role for intermolecular hydrogen bonding in CD₃CN, a stronger hydrogen bond acceptor. The acidity of the NH protons was confirmed when exchange with deuterium was observed in CD₃OD (Figure 2). The coupling constants of the CHPh-CHPh protons (H^5 and H^{10}), at 11–12 Hz, are diagnostic of three-bond couplings between diaxial, vicinal protons.38-40 This feature indicates that the phenyl groups are equatorial in the five-membered Rh-N-C-C-N ring, in agreement with the X-ray structure (see below).

Complex *S*,*S*-**2c** was characterized by X-ray crystal analysis (Figure 3). Table 1 compares its structure to those of 2a,⁵ the related ruthenium complex, [(*p*-cymene)RuCl(TsNCHPhCH-PhNH₂)],³ and bromide and iodide analogues described below. The amine hydrogen atoms were located in a difference map and refined isotropically, thereby demonstrating the presence of the intramolecular NH···Cl hydrogen bond. Further discussion of the structure is postponed until the other halide complexes have been described.

It has recently been suggested that [(arene)Ru(monotosylamidoamine)] complexes form dimers in solution through hydrogen bonding between the NH protons of one molecule





Figure 3. Molecular structure of *S*,*S*-**2c** (thermal ellipsoids shown at 50% probability). Hydrogen atoms are omitted except for those of the amino group. The dotted line shows the hydrogen bond.

Table 1.	Selected Bon	l Lengths (Å) and	Angles	(deg)	for 2	a, 2c,	6a,
		7a, and	l Ru					

bond	$\frac{2\mathbf{a}^a}{(\mathbf{X} = \mathbf{Cl})}$	2c (X = Cl)	$6a \\ (X = Br)$	7a (X = I)	$\frac{\mathbf{R}\mathbf{u}^{b}}{(\mathbf{X}=\mathbf{C}\mathbf{l})}$
$ \frac{M-X}{M-N(1)} \\ M-N(2) \\ \Delta r(M-N)^{d} \\ N(1)-M-N(2) \\ N(1)-M-X \\ N(2)-M-X \\ X \cdots N(1) \\ X \cdots N(1) \\ X \cdots N(1) $	2.421(5) 2.103(6) 2.184(6) 0.081(8) 77.5(5) 84.1(2) 94.0(2) 3.032(7)	2.4175(5) 2.1069(17) 2.1649(17) 0.058(2) 78.70(6) 83.36(5) 91.81(5) 3.0183(19) 2.55(2)	$\begin{array}{c} 2.4968(7)\\ 2.101(4)\\ 2.186(4)\\ 0.085(6)\\ 77.27(15)\\ 84.52(12)\\ 94.65(10)\\ 3.106(4)\\ 2.61^c\end{array}$	$\begin{array}{c} 2.7107(3)\\ 2.122(3)\\ 2.168(2)\\ 0.046(4)\\ 78.76(10)\\ 84.01(8)\\ 93.75(6)\\ 3.263(3)\\ 2.78(3)\end{array}$	$\begin{array}{c} 2.435(4)\\ 2.116(9)\\ 2.145(8)\\ 0.029(12)\\ 79.5(3)\\ 81.1(2)\\ 86.4(2)\\ 2.968(10)\\ 2.57^c \end{array}$
$X \cdots H = N(1)$		111.7(17)	123^{c}	115(2)	104^{c}

^{*a*} Ref 5, H not located. ^{*b*} Ref 3 $\mathbf{Ru} = [(p-cymene)RuCl(TsNCHPhCH-PhNH₂)]$. ^{*c*} No esd available since hydrogen location could not be refined, or was not refined. ^{*d*} $\Delta r(M-N)$ is the difference between the two Rh–N distances.

and a sulfonyl oxygen of another, or between the NH protons of one molecule and the chloride of a second.⁴¹ This has been demonstrated by PGSE NMR experiments and calculations, and also by ESI mass spectrometry. An ESI mass spectrum of **2a** (in CH₃OH) showed a peak at m/z = 603 for the monomer [M - Cl]⁺ and also at m/z = 1241 for [M₂ - Cl]⁺, suggesting that both monomer and dimer may be present in solution.

We also wished to determine ¹⁵N NMR data for **2a** as a basis for further study of catalytic intermediates. [¹⁵N]-TsDPEN was synthesized as a mixture of *R*,*R* and *S*,*S* isomers according to the method of Corey et al. (Scheme 2) with ca. 40% ¹⁵N.⁴² It was not necessary to resolve the racemic mixture because the ¹H NMR spectra of the associated metal complexes are identical. The DPEN ligand was tosylated to give ¹⁵N-labeled TsDPEN (**1a**-¹⁵N) in 46% yield. The FAB mass spectrum for **1a**-¹⁵N

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a 40% 15N-labeled NH4OAc was used, and a full range of isotopomers were formed; only the S,S isomer is shown but a mixture of S,S and R,R isomers was formed.



Figure 4. ¹H-¹⁵N HMQC spectrum of 2a-¹⁵N (CD₂Cl₂, 300 K) showing couplings between the two nitrogen nuclei and the C5Me5 protons. $\delta({}^{1}\text{H})$ horizontal, $\delta({}^{15}\text{N})$ vertical.

compared well with the calculated spectrum. ¹⁵N-Labeled 2a (designated 2a-¹⁵N) was synthesized from 1a-¹⁵N, as described above.

The ¹H NMR spectrum (500 MHz, CD_2Cl_2 , 300K) of **2a-**¹⁵N displays coupling of both NH protons to ¹⁵N. In the ¹⁵N-enriched complex, H^a (numbering in Figure 2) generates two apparent triplet resonances with $J_{\rm NH} = 82$ Hz on either side of the peak of the ¹⁴N isotopomer. Proton H^b generates two doublets with $J_{\rm NH} = 72$ Hz on either side of the doublet of the ¹⁴N isotopomer. A ¹H-¹⁵N HMQC spectrum showed a multiplet at δ -263.6 assigned as the amino nitrogen with coupling to both NH protons, H^5 , H^{10} , and the $C_5(CH_3)_5$ protons. A second multiplet at δ -210.6 with coupling to H⁵, H¹⁰, and the C₅(CH₃)₅ protons was assigned as the amido nitrogen. A ¹H-¹⁵N HMQC spectrum (50.7 MHz, CD₂Cl₂, 300 K) was recorded and showed Rh coupled doublets at δ -263.6 and -210.6 with $J_{\text{RhN}} = 14$ and 18 Hz, respectively (Figure 4). The observation of coupling between the TsDPEN ligand nitrogens and the protons of the Cp* ligand led us to try recording natural abundance ¹H/¹⁵N HMQC spectra of complexes 2a-2c. This was found to be surprisingly straightforward. Complex **2b** had peaks at δ –263.7 and -209.5 with $J_{\rm RhN} = 17$ and 20 Hz, respectively, while complex 2c showed ¹⁵N peaks at δ –263.8 and –210.2 with $J_{\rm RhN} = 16$ and 21 Hz, respectively.





Figure 5. ¹³C-¹³C COSY (125 MHz, CD₂Cl₂, 300 K) experiment, showing couplings between the terminal CO and acyl CO in the two diastereomers of ¹³C-labeled **3a**.

¹⁰³Rh NMR chemical shifts were also recorded via a ¹H-¹⁰³Rh HMQC method making use of the coupling to the C₅Me₅ protons (data in Experimental Section).

Reactivity of Cp*RhCl(TsNCHPhCHPhNH₂), 2a-2c.1. Reactions with Carbon Monoxide, tert-Butylisonitrile, and Formic Acid. Rhodium complexes are known to catalyze both the decomposition of HCO₂H⁴³ and the water-gas shift reaction,⁴⁴ thereby providing a potential source of CO. The reactivity of the catalytic species toward CO is therefore of interest when using HCO₂H for transfer hydrogenation. CO (3 atm) was admitted to a solution of S_1S_2a in CD_2Cl_2 , and the reaction was followed by ¹H NMR spectroscopy. Over a period of 5 days, the Cp* resonance of S,S-2a at δ 1.80 decreased in intensity and was replaced by two new Cp* proton signals at δ 1.68 and 1.83 with approximately equal intensities, indicating complete conversion to products. Over an extended period, the system evolves further and Cp*Rh(CO)₂ is formed as the major product, as indicated by the associated ¹H and ¹³C NMR spectra and IR spectrum. Analysis of the first stage of reaction suggested that the initial products were two diastereoisomers S, S, R_{Rh} and S,S,S_{Rh} of [Cp*Rh(CO){C(O)N(Ts)CHPhCHPhNH₂}][Cl] (**3a**) with the same ligand chirality, but with opposite chirality at Rh. The ¹³C NMR spectrum of the two diastereomers of 3a shows resonances at δ 181.6 (d, $J_{\rm RhC}$ = 27 Hz) and 181.7 (d, $J_{\rm RhC} = 27$ Hz) assigned to OCN and signals at δ 188.1 (d, $J_{\rm RhC}$ = 77 Hz) and 188.2 (d, J_{RhC} = 77 Hz) for the CO ligands. A $^{13}C^{-13}C$ COSY spectrum of **3a** (synthesized by reaction of **2a** with 1 atm ¹³CO and 2 atm ¹²CO) showed couplings between RhCO at 188.1 and the OCN at δ 181.6, along with couplings between the CO at δ 188.2 and the OCN at δ 181.7 (Figure 5), consistent with the presence of two diastereomers. The ¹H NMR spectrum (500 MHz, CD₂Cl₂, 300 K) of **3a** showed two peaks for Cp* (δ 1.68 and 1.83), two for the tosyl methyl group (δ 2.28 and 2.31), for the CH protons of the backbone, and for the NH protons. All these observations are consistent with the formation of a pair of diastereoisomers.

Complex 3a may form via substitution of chloride by CO, followed by migration of the amide bond of the chelating ligand onto CO, which creates a coordinatively unsaturated, planar, 16-electron, carbamoyl complex. CO could then bind from the front or the back, forming two diastereomers of 3a with the same ligand chirality, but with opposite chirality at Rh (Scheme

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Scheme 3. Proposed Mechanism for the Reaction of 2a with CO to Form 5a



3). Both ESI and FAB mass spectrometry revealed peaks at m/z = 603 for $[M - 2CO]^+$; the parent molecular ion was not detected. Insertion of CO into mononuclear amido and amidinato complexes has been observed previously to form carbamoyl complexes.^{45–48} Recently Rauchfuss et al. have reported a similar loss of chirality in related complexes, $[Cp*Ir-(L)(TsNCHPhCHPhNH_2]^+$ (L = PPh₃, P(4-FC₆H₄)₃, P(4-MeOC₆H₄)₃, or CO).²²

On release of the pressure of CO from the NMR tube (before complete conversion to Cp*Rh(CO)₂), 3a converted back to 2a sufficiently slowly to allow IR spectra to be recorded. The IR spectrum showed a peak at 2056 cm⁻¹ consistent with a terminal CO ligand and a peak at 1600 cm^{-1} assigned to the acyl group of the bidentate ligand for **3a**. When ¹³C-enriched CO was used, the peaks for the ¹³C isotopomers were observed at 2003 cm⁻¹ (2012 cm⁻¹ calcd, based on the change in reduced mass) and 1562 cm⁻¹ (1566 cm⁻¹ calcd). The reaction between **2a** and CO may be accelerated by the presence of methanol. When complex 2a was dissolved in CD₃OD with 3 atm of CO, the color changed immediately from orange to yellow. The NMR spectrum of the product solution was consistent with conversion to the two diastereoisomers of 3a, as described for the reaction in CD₂Cl₂. The ¹H NMR spectrum (500 MHz, CD₃OD, 300 K) of **3a** showed two peaks for Cp* (δ 1.68 and 1.82), two for the tosyl methyl group (δ 2.25 and 2.31), a multiplet consistent with the CH protons of the ligand backbone at δ 4.48–4.52, and another at δ 5.03–5.08. Notably, the two isomers were formed initially with a 4:1 ratio (δ 1.68 dominant), but equalized in intensity after about an hour. This experiment therefore also suggests reversible CO coordination leading to thermodynamic equilibrium.

Recently, Ikariya et al. showed that the related cyclometalated iridium chloride complexes react with $AgBF_4$ and CO to form metal carbonyl cations with ν_{CO} very close to the values that we report for the terminal CO.⁴⁹ Rauchfuss and co-workers have reported the complexes [Cp*Ir(CO)(TsCH₂CH₂NH₂)][BF₄], [Cp*Ir(CO)(Ts-1,2-C₆H₁₀NH₂)][BF₄], and [Cp*Ir(CO)(TsCH-PhCHPhNH₂)][BF₄] with ν_{CO} of 2059, 2059, and 2064 cm⁻¹.^{21,22}

Reaction of **2a** with *t*BuNC in CH₃OH gives an immediate color change to yellow; extraction into toluene followed by

Scheme 4. Reactions of 2a with tBuNC, HCO₂H, LiBr, and



addition of KPF₆ yields [Cp*Rh(CN*t*Bu)(TsNCHPhCHPh-NH₂)][PF₆] (**4a**) as a yellow powder (Scheme 4). The complex was crystallized from THF/hexane to give **4a** as yellow needles. The IR spectrum shows $\nu_{\rm CN}$ at 2112 cm⁻¹ (KBr disk). The ¹³C NMR spectrum shows a resonance for *t*BuNC at δ 134.8 as a doublet of multiplets with $J_{\rm RhC} = 69$ Hz. These data are consistent with those for Cp*RhCl₂(CNMe), which exhibits a doublet of triplets at δ 136.3 with $J_{\rm RhC} = 69$ Hz and $J_{\rm NC} = 20$ Hz.⁵⁰

The reaction of **2a** with HCO₂H (10 equiv) in methanol was performed to establish how **2a** reacts with a single component of the system for catalytic transfer hydrogenation. Upon addition of formic acid, the color of the solution turned immediately from orange to red, and a red powder was isolated by adding 2 equiv of NaBPh₄ to a solution of the metal complex in methanol. ESI mass spectrometry (M⁺ = 557) was consistent with formation of the triply bridged dimer [{Cp*Rh}₂(μ -H)(μ -Cl)-(μ -HCO₂)][BPh₄] ([**5a**][**BPh₄**]), and this assignment was con-

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Figure 6. Molecular structure of the cation in $[5a][BPh_4] \cdot OEt_2$ (thermal ellipsoids shown at 50% probability). Hydrogen atoms are omitted except for hydride. Selected bond lengths [Å] and angles [deg]: Rh(1) ··· Rh(2) 2.8251(3), Rh(1)-O(2) 2.1044(18), Rh(2)-O(1) 2.117(2), Rh(1)-H(1) 1.73(3), Rh(2)-H(1) 1.77(3), Rh(1)-Cl(1) 2.4290(7), Rh(2)-Cl(1) 2.4201(7); Rh(1)-Cl(1)-Rh(2) 71.27(2), Rh(1)-O(2)-C(21) 123.79(18), Rh(2)-O(1)-C(21) 123.25(18), O(1)-C(21)-O(2) 128.3(3).

firmed by X-ray analysis using a crystal grown from acetone/ Et₂O at -20 °C (Scheme 4). Related triply bridged cationic dimers of general formula [{Cp*Rh}₂(μ -H)(μ -OCOR)₂][PF₆] have been reported by Maitlis et al.⁵¹

The crystal structure of [5a][BPh4] · OEt2 (Figure 6) reveals Rh(1)-O(2) and Rh(2)-O(1) distances of 2.1044(18) and 2.117(2) Å, respectively, longer than those in $[{Cp*Rh}_2(\mu-H)_2 (\mu$ -CO₂CH₃)]⁺ (2.071 and 2.070 Å).⁵² The hydride in the cation 5a was readily located on a difference map. Rh(1)-H(1) and Rh(2)-H(1) distances of 1.73(3) and 1.77(3) Å, respectively, are comparable to those in $[{Cp*Rh}_2(\mu-H)_2(\mu-CO_2CH_3)]^+$ (1.73) Å). The geometry of the bridge is different from that of its analogues. The Rh ··· Rh distance for the cation 5a of 2.8251(3) Å may be compared with 2.680 Å in $[{Cp*Rh}_2(\mu-H)_2 (\mu$ -CO₂CH₃)]^{+.52} The average Rh(1)-Cl distance of 2.425(1) Å is shorter than that in $[{Cp*Rh}_2(\mu-Cl)_3]^+$ (2.457 Å),⁵³ while the Rh(1)-Cl(1)-Rh(2) angle of $71.27(2)^{\circ}$ for **5a** is much more acute than that for $[{Cp*Rh}_2(\mu-Cl)_3]^+$ (81.23° average). The O(1)-C(21)-O(2) angle of $128.3(3)^{\circ}$ is wider than that of $[{Cp*Rh}_2(\mu-H)_2(\mu-CO_2CH_3)]^+$ (124.80°).

The ¹H NMR spectrum (500 MHz, (CD₃)₂CO, 300 K) of **[5a][BPh₄]** showed a triplet at δ -8.67 (J_{RhH} = 27 Hz) for the bridging hydride, a singlet at δ 1.83 for the Cp* protons, and a triplet at δ 7.41 (J_{RhH} = 4 Hz) for the bridging formate proton.

The isolation of [5a][BPh₄] raises the question of whether cation 5a could be formed under catalytic conditions when the formic acid is buffered by triethylamine. As an experimental test, we investigated the reduction of d_6 -acetone by d_{15} triethylamine/formic acid catalyzed by 2a in CD₃CN solution. The molar ratio of acetone substrate to catalyst was 20:1. The reaction was carried out both at 250 K and at 300 K with the same result. Examination of the hydride region of the ¹H NMR spectrum showed one hydride species only with a triplet at δ -8.76 (J = 27 Hz), consistent with significant formation of the hydride-bridged cation 5a. The same species was observed with CD₃OD and CD₂Cl₂ as solvents.

The surprising observation of 5a under catalytic conditions raises the question of whether reaction of the catalyst with



Figure 7. ¹H NMR (C*H*Ph and NH₂ region) spectra of 2a (top), 6a (middle), and 7a (bottom) 500 MHz, CD₃CN, 300 K.

formic acid is a significant deactivation pathway. We compared the catalytic activities of **2a** and **[5a][BPh4]** for the transfer hydrogenation of the imine 6,7-dimethoxy-1-methyl-3,4-dihydroisoquinoline. Under conditions when **2a** gave excellent activity and ee's of 90%,^{20,54} complex **[5a][BPh4]** showed very poor activity. However, the catalytic system of **{5a + 1a}** gave activity and selectivity comparable to that of **2a**. Thus we conclude that reaction of **2a** with formic acid to form **5a** may occur to some extent under catalytic conditions despite the presence of NEt₃. Nevertheless, catalytic activity is not necessarily blocked because the presence of free chiral ligand may allow recovery of activity.

2. Reactions with Halide Salts. The addition of LiBr or KI to a solution of S,S-2a in methanol resulted in quantitative conversion to the corresponding bromide Cp*RhBr(S,S-TsNCHPhCHPhNH₂) (S,S-6a) or iodide Cp*RhI(S,S-TsNCHPhCHPhNH₂) (S,S-7a), respectively (Scheme 4). The addition of iodide salts is accompanied by a color change from pale orange to dark red. Both complexes were characterized by ¹H and ¹³C NMR spectroscopy, FAB mass spectrometry, and X-ray crystallography. The ¹H NMR spectra show most variation between 2a, 6a, and 7a in the proton chemical shifts of the ligand backbone and NH₂ groups, as illustrated in Figure 7.

Complex *S*,*S*-**6a** was recrystallized by slow diffusion of hexane into a concentrated solution in THF. Crystals of *S*,*S*-**7a** \cdot CH₃OH were obtained by slow evaporation of a solution in CH₃OH. Selected bond lengths, angles, and interatomic distances are compared to those for **2c** and literature values for **2a** and for **Ru** ((*p*-cymene)RuCl(TsNCHPhCHPhNH₂)) in Table 1. The

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Figure 8. Molecular structures of S, S-6a (above) and S, S- $7a \cdot CH_3OH$ (below) (thermal ellipsoids shown at 50% probability). Hydrogen atoms are omitted except for the amino group and methanol OH. The hydrogen bonds are shown as dotted lines.

amine hydrogen atoms were located by difference maps; their positions were refined successfully for 7a but their positions were constrained in the refinement of **6a**. The structures of **2a**, 2c, 6a, and 7a are very similar and also close to that of Ru.³ The most important features are (a) that the Rh-N(1) (NH_2) bond is significantly shorter than the Rh-N(2) (NS) bond and (b) that each complex possesses an intramolecular $NH \cdots X$ hydrogen bond. The phenyl groups lie in equatorial positions on the five-membered Rh-N-C-C-N ring, as in related structures and in agreement with the evidence provided by the ¹H NMR data (see above).²² The Rh-NH₂ bond length is invariably shorter than the Rh-NS bond length, but the differences $\Delta r(M-N)$ are larger for **2a** and **6a** (0.081(8) and 0.085(6) Å) than for **2c** and **7a** (0.058(2) and 0.046(4) Å). The changes arise from an increased Rh-N(1) and decreased Rh-N(2) distance in 7a and a decreased Rh-N(2) distance in **2c.** The esd on $\Delta r(M-N)$ for the ruthenium complex is too large to compare to the values for the other complexes with confidence. The N····X and NH····X distances are consistent with an NH····X hydrogen bond in each complex (X = Cl, Br, I). The NH····X distances are shorter than the sum of the van der Waals radii by 0.40, 0.44, and 0.40 Å in 2c, 6a, and 7a, respectively (van der Waals radii of Cl 1.75, Br 1.85, I 1.98, and H 1.20 Å).^{55,56} Complex 7a crystallizes as the methanol solvate and exhibits a further hydrogen bond between the methanol oxygen and the NH hydrogen that does not interact with iodine (Figure 8). In 2c, there are also short S=O(1) ··· HC(30)



Figure 9. Molecular structure and unit cell for $[8c-H_2O][OTf]$ (thermal ellipsoids shown at 50% probability). Hydrogen atoms and anions are omitted for clarity. Selected bond lengths [Å] and angles [deg]: Rh(1)-N(1) 2.1328(19), Rh(1)-O(3) 2.1631(18), Rh(1)-N(2) 2.141(2); N(1)-Rh(1)-N(2) 77.61(8), N(1)-Rh(1)-O(3) 85.24(7), N(2)-Rh(1)-O(3) 90.08(7).

and $S=O(1)\cdots HC(12)$ contacts with $O\cdots C$ of 2.881(3) and 2.899(2) Å, respectively. The intramolecular hydrogen bonds have been noted in previous structures, but the striking difference in the Rh–N distances has not been emphasized, perhaps because the esd's on the older structures were larger than on those reported here.^{3,5,15}

3. Halide Abstraction. In an attempt to provide a route to a crystalline analogue of the CO complex 3a, the reaction of S_1S_2c with AgOTf (OTf = CF₃SO₃) in CH₃OH was carried out. The chloride abstraction succeeded although the CO reaction proved unsuccessful; the addition of AgOTf to a solution of 2c in methanol led to an immediate color change from orange to magenta and precipitation of AgCl. After filtering off the insoluble salt and removing the solvent under reduced pressure, a purple solid was obtained. This solid was identified as [Cp*Rh(4-FC₆H₄SO₂NCHPhCHPhNH₂)][OTf] ([8c][OTf]). A sample was dissolved in CD₃CN for NMR analysis, resulting in a yellow solution, suggesting CD₃CN coordination to Rh. Attempts to locate the coordinated CH₃CN by running a ¹H NMR spectrum in CH₃CN using solvent suppression were unsuccessful, although exchange could be occurring on the NMR time scale. The coordination of solvent was supported by an IR band at 2239 cm⁻¹ consistent with the CN stretch of coordinated CH₃CN.⁵⁷

Complex [8c][OTf] was characterized by ¹H, ¹³C, and ¹⁹F NMR spectroscopy in CD₃CN. The ¹H NMR spectrum displayed peaks at δ 4.50 and 3.95 for the NH₂ protons, a resonance at δ 1.83 for Cp*, and signals at δ 4.15 and δ 3.76 for the CHPh protons. The ¹³C NMR resonances of the sulfonyl aromatic ring carbons all show couplings to ¹⁹F, as for **2c**. A diffuse reflectance UV-vis spectrum of [8c][OTf] showed $\lambda_{max} = 520$ nm. The ESI mass spectrum in CH₃OH showed the expected peak at m/z = 607 for [M]⁺, without coordinated solvent.

A crystal suitable for X-ray analysis was grown by vapor diffusion of hexane into a solution of [8c][OTf] in CH₃OH/ toluene (1:1). The structure obtained was a derivative of [8c][OTf] with a water molecule coordinated to the rhodium center, [8c-H₂O][OTf] (Figure 9). The most significant difference in the structure of [8c-H₂O][OTf] from those of 2a and 2c is a change from a chiral to an achiral space group. Complex [8c-H₂O][OTf] crystallized in the achiral space group $P\bar{1}$, whereas those used to solve structures of 2a-2c were the chiral space groups $P2_1$ or $P2_12_12_1$. The hydrogen atoms of the coordinated water and amine were located in a difference map

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and refined isotropically. Figure 9 shows that the two molecules in the cell have opposite chirality at both Rh and ligand, and it appears that the ligand has racemized. At the molecular level, there are also important changes: the Rh–NH₂ distance in [**8c**-H₂O][OTf] is 0.026(3) Å longer than in 2c, while the Rh–NTs distance is 0.024(3) Å shorter, with the result that Δr (Rh–N) is insignificant. The Rh–O bond in [**8c**-H₂O][OTf] of 2.1631(18) Å is comparable to the mean value in [Cp*Rh(OH₂)₃][OTf]₂ (2.156 Å)⁵⁸ and that in [Cp*Rh(PMe₃)(OH₂)(C₆F₅CF₂)][BF₄] (2.164(7) Å).⁵⁹ The amine hydrogen atoms engage in hydrogen bonding to the oxygen atoms of the triflate counterion, while the water hydrogen atoms form hydrogen bonds with the fluorine and oxygen atoms of the counterion and with the sulfonyl oxygens.

The crystal structure leaves the question open whether the majority of the complex has racemized or whether the result depends on the selection of crystals. Further evidence for racemization of the TsDPEN ligand was obtained by measuring the optical rotation of a solution of S_1S-2c in CHCl₃, reacting this with AgOTf to form [8c][OTf], and measuring the optical rotation of the resulting solution at constant concentration. The solution of complex S,S-2c had an optical rotation of -1.5° , whereas the solution of the product [8c-H₂O][OTf] had an optical rotation of -0.1° . Furthermore, subsequent conversion of [8c-H₂O][OTf] back to 2c (via addition of Bu₄NCl) resulted in an optical rotation of -0.6° , well short of the initial optical rotation. This significant reduction of optical activity for the product solution is evidence that a substantial proportion of the ligand in the complex has racemized and the effect is not confined to a few crystals.

A second crop of crystals of [8c][OTf] was grown as described above, but under anhydrous conditions, and proved to have a different structure. In the absence of water or a suitable coordinating solvent, the cationic 16-electron complex dimerized. Although the structure is of poor quality, the principal features are important. An oxygen of the sulfonyl group of one molecule has filled the vacant site at that of the other and vice versa to give the dimer [Cp*Rh(4-FC₆H₄SO₂NCHPhCHPh-NH₂)]₂[OTf]₂ ([8c]₂[OTf]₂, structure below). As with [8c-H₂O][OTf], the structure of [8c]₂[OTf]₂ was solved with the achiral $P\bar{1}$ space group and opposite enantiomers are paired in the unit cell. For comparison, $[Cp*Rh(R^1COCHCOR^2)][PF_6]$ has been shown to exist as a dimer by coordination of the central acetylacetonate carbon of one complex to Rh in another and vice versa.⁶⁰ An ESI mass spectrum of [8c][OTf] showed a significant peak at m/z = 1363 for $[M_2(SO_3CF_3)]^+$, providing some evidence for the existence of the dimer in solution as well as in the solid state.



The racemization of the ligand in $[8c-H_2O][OTf]$ is of importance in asymmetric transfer hydrogenation. A possible mechanism for this racemization of the ligand is outlined (Scheme 5): step 1 is the reversible dissociation of coordinated

Scheme 5. Possible Mechanism for the Racemization of the FsNCHPhCHPhNH₂ Ligand in $[8c-S]^+$ (S, = solvent, Fs = 4-FC₆H₄SO₂)



solvent; step 2 is a β -elimination; step 3 is an imine—enamine tautomerism, which scrambles the chirality; steps 4 and 5 are the reverse of steps 1 and 2, so both enantiomers are as likely to be formed and a racemic mixture ensues. The *meso* isomer is likely to be much higher in energy and disfavored. This mechanism remains unproven, but it provides a plausible explanation of the observations.

The reactivity of **[8c][OTf]** toward two-electron donors was studied as a route to new cationic Rh complexes; although the reaction with CO was unsuccessful, an adduct was formed by reaction with *t*BuNC. Upon addition of 1 equiv of *t*BuNC to a solution of **8c** in CH₃OH, an immediate color change from magenta to yellow was observed. Workup by removing the solvent under reduced pressure, extraction into CH₃CN, and removal of CH₃CN under vacuum gave a yellow solid, which has been characterized by ¹H, ¹³C, and ¹⁹F NMR spectroscopy and ESI mass spectrometry and was found to be [Cp*Rh(CN*t*Bu)-(4-FC₆H₄SO₂NCHPhCHPhNH₂)][OTf] (**4c**).

Conclusions

A group of 18-electron half-sandwich rhodium complexes, of the type $[Cp*Rh(X)(4-RC_6H_4SO_2NCHPhCHPhNH_2)]$ (X = Cl, Br, I; R = Me, *t*Bu, F), of significance in asymmetric transfer hydrogenation catalysis^{5-7,10,15,17,20} has been isolated. The complexes have been characterized by multinuclear NMR spectroscopy and by X-ray crystallography. The NMR spectra show significant solvent dependence of the ¹H chemical shift of the NH₂ protons characteristic of NH ···· X hydrogen bonding. The ¹H–¹⁵N HMQC NMR spectra show that the chemical shifts of the inequivalent nitrogen atoms are separated by ca. 50 ppm; moreover, the resonances exhibit significant coupling to the C₅Me₅ protons and to the rhodium nucleus. These methods offer scope for wider application for characterization of the nitrogen environment in solution. Notable features of the X-ray structure are the significantly longer Rh-N bonds to the amido nitrogen than the amino nitrogen and the presence of an intramolecular

Table 2. Crystallographic Data for 2c, [5a][BPh4] · OEt2, 6a, 7a, and [8c · H2O][OTf]

	2c	$[5a][BPh_4] \cdot OEt_2$	6a	7a	[8c-H ₂ O][OTf]
formula	C30H33ClFN2O2RhS	C49H62BClO3Rh2	C31H36BrN2O2RhS	C32H40IN2O3RhS	$C_{34.5}H_{38.5}F_4N_2O_6RhS_2$
M_{\perp}	643.00	951.07	683.50	762.53	820.20
a/Å	7.8845(4)	11.7528(10)	8.3927(4)	8.4842(5)	12.286(5)
b/Å	11.3176(5)	13.6373(12)	16.1471(8)	9.1856(6)	12.454(5)
c/Å	31.7839(14)	14.2877(12)	21.6575(11)	20.6402(13)	13.377(5)
α/deg		106.264(2)			107.266(7)
β/deg		92.571(2)		97.4630(10)	113.603(7)
γ/deg		92.506(2)			97.270(7)
V/Å ³	2836.2(2)	2192.3(3)	2935.0(3)	1594.92(17)	1719.2(12)
T/K	100(2)	110(2)	110(2)	110(2)	110(2)
space group	$P2_{1}2_{1}2_{1}$	$P\overline{1}$	$P2_{1}2_{1}2_{1}$	$P2_1$	$P\overline{1}$
Ζ	4	2	4	2	2
μ (Mo K α)/mm ⁻¹	0.807	0.854	2.045	1.603	0.688
reflns measd	42 801	22 323	32 890	21 931	23 383
reflns indep	8180	10 753	8364	7863	8538
$R_{\rm int}$	0.0289	0.0221	0.0803	0.0244	0.0267
final $R[I > 2\sigma(I)]$	$R_1 = 0.0254$	$R_1 = 0.0351$	$R_1 = 0.0429$	$R_1 = 0.0248$	$R_1 = 0.0336$
	$wR_2 = 0.0625$	$wR_2 = 0.0847$	$wR_2 = 0.1177$	$wR_2 = 0.0585$	$wR_2 = 0.0848$
final R (all data)	$R_1 = 0.0268$	$R_1 = 0.0440$	$R_1 = 0.0548$	$R_1 = 0.0272$	$R_1 = 0.0380$
-	$wR_2 = 0.0672$	$wR_2 = 0.0889$	$wR_2 = 0.1212$	$wR_2 = 0.0663$	$wR_2 = 0.0869$
CCDC number	703218	703216	703215	703214	703217

NH····X hydrogen bond that may prepare the complex for proton abstraction.^{3,5,15}

Direct reaction of Cp*Rh(Cl)(S,S-TsNCHPhCHPhNH₂) with CO results in the reversible formation of [Cp*Rh(CO)- $\{C(O)N(Ts)CHPhCHPhNH_2\}$ [Cl] as two diastereomers, S,S,R_{Rh} and S,S,S_{Rh}, with different configurations at rhodium. This species has been identified by $2D^{-13}C^{-13}C$ NMR spectroscopy with ¹³CO labeling and by IR spectroscopy. The growth of the second isomer may be monitored during reaction in d_4 -methanol as solvent. Further reaction with CO leads to the complete displacement of the nitrogen ligands. The nitrogen ligands are also completely displaced by reaction of 2a with formic acid, yielding dinuclear [{Cp*Rh}₂(μ -H)(μ -Cl)(μ -HCO₂)][BPh₄], [5a]-[BPh₄], closely related to carboxylate complexes observed previously.⁵¹ Cation **5a** is also detected under catalytic conditions for reduction of d_6 -acetone with d_{15} -NEt₃ and formic acid; that is, the buffering effect of triethylamine is not sufficient to prevent some reaction. Although formation of complex [5a][B- Ph_4 represents a deactivation path for the catalyst, catalytic activity can be recovered by reaction of [5a][BPh₄] with the ligand 1a.

Reaction of Cp*RhCl(S,S-4-FC₆H₄SO₂NCHPhCHPhNH₂) with AgOTf generates a purple 16-electron complex that exists as the dimer [Cp*Rh(4-FC₆H₄SO₂NCHPhCHPhNH₂)]₂[OTf]₂ in the crystalline state. In the presence of adventitious water, it is isolated as a salt of the aqua cation [Cp*Rh(4-FC₆H₄SO₂-NCHPhCHPhNH₂)(OH₂)][OTf]. The notable feature of this reaction is that it results in racemization of both CHPh centers (i.e., formation of a mixture of *R*,*R* and *S*,*S* from initial *S*,*S* material). Thus halide abstraction in this manner would result in loss of enantioselectivity of the catalyst.

Experimental Section

General Procedure. Unless otherwise stated, synthetic work was carried out in air with untreated solvents. Commercially available reagents were obtained from the following sources: [Cp*RhCl₂]₂, *R*,*R* and *S*,*S*-DPEN, *R*,*R* and *S*,*S*-TsDPEN, 6,7-dimethoxy-3,4-dihydroisoquinoline (donated by Piramal Healthcare); RhCl₃ · *x*H₂O (Precious Metals Online); hexamethyl(dewar)benzene, benzil, 4-flu-

orobenzenesulfonyl chloride, 4-*tert*-butylbenzenesulfonyl chloride, triethylamine, lithium, ammonium-¹⁵N acetate (98 atom % ¹⁵N), formic acid (Aldrich); tosyl chloride (Alfa Aesar); d₁₅-triethylamine from Cambridge Isotope Laboratories. Ar, N₂, CO, and H₂ gases (BOC) and were used as received without further purification. [Cp*RhCl₂]₂ was also made as previously reported.⁶¹ When necessary, dry solvents (Fisher Scientific AR or HPLC grade) were prepared by refluxing over Mg/I₂ (CH₃OH), CaH₂ (CH₃CN, CH₂Cl₂), or sodium/benzophenone (THF) and were degassed and, along with air- and moisture-sensitive materials, stored under a dry Ar atmosphere. Manipulations of these compounds were performed using standard Schlenk, high-vacuum, and glovebox techniques. Deuterated solvents (Aldrich) were dried using CaH₂ (CD₃OH, CD₃CN, and CD₂Cl₂) and degassed by three freeze–pump–thaw cycles.

Unless otherwise stated, DPEN, TsDPEN, $4-FC_6H_4SO_2DPEN$, $4-tBuC_6H_4SO_2DPEN$, and all metal complexes have been synthesized and used as single diastereomers, with either the *R*,*R* or *S*,*S* configuration at the ligand, which, when coordinated to the metal, enforces the opposite chirality at Rh; the metal complex of the *S*,*S* ligand will have the *R* configuration at the metal center and *vice versa*. When referring to DPEN and related ligands, the specific chirality is not always mentioned, as both have been used interchangeably.

Most NMR spectra were recorded on Bruker AMX500, DRX400, or AMX300 spectrometers. The chemical shift values are reported relative to the residual protons of the deuterated solvent, according to the standard Bruker list. Numbering of hydrogen and carbon atoms refers to Figure 2. $^{1}H^{-103}$ Rh HMQC spectra were run on a Bruker Avance II 700 MHz spectrometer calibrated against [Cp*RhCl₂]₂ at δ 2303.⁶² $^{1}H^{-15}$ N HMQC spectra were referenced to pyridine as δ 0. FAB mass spectra were recorded on a VG Auto-Spec, and ESI mass spectra were recorded on a Thermo Electron LCQ Classic. Infrared spectra were recorded on a Unicam RS 10000E FTIR instrument. Polarimetry measurements were made in a Jasco DIP-370 digital polarimeter using a 2 mL cell and CHCl₃ solvent. Microanalysis was carried out by Elemental Microanalysis Ltd., Okehampton, Devon.

Synthesis of 4-RC₆H₄SO₂DPEN Ligands.³⁷ DPEN was dissolved in CH₂Cl₂ in a two-necked, 500 mL round-bottom flask. The solution was cooled to 0 °C, NEt₃ was added, and the mixture was stirred magnetically. The appropriate sulfonyl chloride was

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dissolved in CH₂Cl₂ and added to a dropping funnel, which was placed in the side neck of the round-bottom flask. The sulfonyl chloride solution was added to the stirred DPEN/NEt₃ solution dropwise over an hour, and the solution was warmed to room temperature and stirred overnight. It was washed successively with water (80 mL), NaHCO₃ solution (80 mL), water (80 mL), and brine (80 mL) and then dried over MgSO₄; the solvent was removed under reduced pressure to give a white solid, which was crystallized from EtOAc/hexane.

4-tBuC₆H₄SO₂DPEN (1b). 1b was synthesized as described above from DPEN (2.10 g, 9.9 mmol) and NEt₃ (2.10 mL, 14.9 mmol) in CH₂Cl₂ (150 mL) and 4-tert-butylbenzenesulfonyl chloride (2.00 g, 8.7 mmol) in CH₂Cl₂ (50 mL). Yield: 1.72 g (48%). ¹H NMR (500 MHz, CD₂Cl₂, 300 K, numbering as in Figure 2): δ 1.29 (9H, s, (CH₃)₃C), 4.14 (1H, d, $J_{\rm HH} = 5$ Hz, H⁵ or H¹⁰), 4.35 (1H, d, $J_{\rm HH} = 5$ Hz, H⁵ or H¹⁰), 7.12–7.15 (10H, m, Ph), 7.21 (2H, d, $J_{\rm HH} = 8$ Hz, H²), 7.33 (2H, d, $J_{\rm HH} = 8$ Hz, H³). ¹³C NMR (125 MHz, CD₂Cl₂, 300 K): δ 31.36 (s, (CH₃)₃C), 35.36 (s, (CH₃)₃C), 61.0 (s, C⁵ or C¹⁰), 63.86 (s, C⁵ or C¹⁰), 126.10 (s, C³ or C⁴), 127.15 (s, C³ or C⁴), 127.17 (s, Ph), 127.60 (s, Ph), 127.81 (s, Ph), 128.70 (s, Ph), 128.83 (s, Ph), 129.99 (s, Ph), 137.60 (s, C¹), 140.15 (s, C⁶ or C¹¹), 142.30 (s, C⁶ or C¹¹), 156.08 (s, C⁴). MS (FAB, *m/z*): 410 ([MH⁺]). IR (KBr/cm⁻¹): 3356 (*v*_{NH}), 3298 (*v*_{NH}), 3291 (br, $\nu_{\rm NH}$). Anal. Calcd for C₂₄H₂₈N₂O₂S: C, 70.55; H, 6.91; N, 6.86. Found: C, 70.84; H, 7.08; N 6.87.

4-FC₆H₄SO₂DPEN (1c). 1c was synthesized as described above from DPEN (2.46 g, 11.6 mmol) and NEt₃ (2.40 mL, 17.4 mmol) in CH₂Cl₂ (150 mL) and 4-fluorobenzenesulfonyl chloride (2.00 g, 10.4 mmol) in CH₂Cl₂ (50 mL). Yield: 2.30 g (60%). ¹H NMR (500 MHz, CD₂Cl₂, 300 K, numbering as in Figure 2): δ 4.17 (1H, d, $J_{\rm HH} = 5$ Hz, H⁵ or H¹⁰), 4.38 (1H, d, $J_{\rm HH} = 5$ Hz, H⁵ or H¹⁰), 6.86 (2 H, apparent t, $J_{\rm HH} = 9$ Hz, $J_{\rm HF} = 9$ Hz, H^2), 7.17–7.20 (10H, m, Ph), 7.41 (2H, dd, $J_{\rm HH} = 9$, $J_{\rm HF} = 5$ Hz, H³). ¹³C NMR (125 MHz, CD₂Cl₂, 300 K): δ 60.86 (s, C⁵ or C¹⁰), 63.84 (s, C⁵ or C^{10}), 116.18 (d, $J_{FC} = 23$ Hz, C^2), 127.06 (s, Ph), 127.55 (s, Ph), 128.00 (s, Ph), 128.04 (s, Ph), 128.84 (s, Ph), 128.94 (s, Ph), 129.96 (d, $J_{FC} = 9$ Hz, C²), 136.91 (d, $J_{FC} = 4$ Hz, C⁴), 140.09 (s, C⁶ or C¹¹), 142.19 (s, C⁶ or C¹¹), 165.09 (d, $J_{FC} = 251$ Hz, C¹). ¹⁹F NMR (480 MHz, CD₂Cl₂, 300 K): δ -107.54 (s). MS (FAB, *m/z*): 371 $([MH]^+)$. IR (KBr/cm⁻¹): 3350 (ν_{NH}), 3338 (ν_{NH}), 3291 (ν_{NH}). Anal. Calcd for C₂₀H₁₉FN₂O₂S: C, 64.85; H, 5.17; N, 7.56. Found: C, 64.72; H, 5.16; N, 7.49

¹⁵N-Labeled DPEN.⁴² A mixture of benzil (3.31 g, 15.8 mmol), cyclohexanone (1.63 mL,15.8 mmol), ammonium acetate (7.5 g ¹⁴N and 5.24 g ¹⁵N), and acetic acid (30 mL) was refluxed for 1 h. The cooled mixture was slowly poured into vigorously stirred water (50 mL) and stirred for an additional 2 h, then left overnight. The crystals formed were collected by filtration, washed with water (4 × 10 mL), and dried in vacuo to give 5-spirocyclohexyl-2,3diphenyisoimidazole (A) (4.41 g, 97%), which was used in the next step without further purification. To a magnetically stirred solution of A (4.41 g, 15.3 mmol) in THF (20 mL) and NH₃ (25 mL) at -78 °C was added Li (601 mg, 85.9 mmol). Stirring was continued for 2 h and the reaction monitored by TLC, at which time all of the starting material had been consumed. Ammonium chloride (4.0 g) was added at -78 °C and the reaction mixture warmed very slowly to evaporate the NH₃. When the temperature of the reaction mixture reached 0 °C, water (20 mL) and Et₂O (20 mL) were added, the mixture was shaken, and the phases were separated. The aqueous phase was washed with Et_2O (3 \times 20 mL), and the combined organic extracts were washed with brine (20 mL), dried over Na₂SO₄, filtered, and concentrated in vacuo to a volume of 15 mL. The resulting solution was cooled to 0 °C, 2 M HCl (15 mL) was added with vigorous stirring, and the solution was warmed to room temperature and stirred for 1 h. Water (30 mL) and CH₂Cl₂ (30 mL) were added, the mixture was shaken, and the phases were separated. The organic phase was washed with water (10 mL), and the combined aqueous phases were washed with CH₂Cl₂ (20 mL). The aqueous phase was treated with 2 M NaOH (20 mL) and extracted with CH₂Cl₂ (5 × 10 mL). The combined organic phases were dried over Na₂SO₄, filtered, and concentrated *in vacuo* to give ¹⁵N-labeled DPEN (1.36 g, 44%). ¹H NMR (500 MHz, CDCl₃, 300 K): δ 1.98 (4H, s), 4.09 (2H, s), 7.17–7.33 (10H, m). MS (FAB, *m/z*): 213 ([MH]⁺), 214 (singly labeled [MH]⁺), 215 (doubly labeled [MH]⁺).

¹⁵N-Labeled TsDPEN (1a-¹⁵N). 1a-¹⁵N was synthesized as described above from ¹⁵N-labeled DPEN (178 mg, 0.84 mmol), NEt₃ (150 μL, 1.1 mmol) in CH₂Cl₂ (30 mL), and tosyl chloride (122 mg, 0.64 mmol) in CH₂Cl₂ (15 mL). Yield: 109 mg (46%). ¹H NMR (500 MHz, CDCl₃, 300 K): δ 2.33 (3H, s, CH₃), 4.16 (1H, d, $J_{HH} = 6$ Hz, CHNH₂), 4.39 (1H, d, $J_{HH} = 6$ Hz, CHNTs), 6.98 (2H, d, $J_{HH} = 8$ Hz, H⁴), 7.12–7.18 (10H, m, Ph) Ar, 7.32 (2H, d, $J_{HH} = 8$ Hz, H²). MS (FAB, *m/z*, relative intensity): 367 (63, [MH]^{+ 14}N/¹⁴N/¹²C/³²S isotopomer), 368 (100, [MH]⁺¹⁴N/¹⁴N/¹²C/³²S, ¹⁵N/¹⁵N/¹²C/³²S isotopomers), 370 (37, [MH]^{+ 14}N/¹⁵N/¹²C/³⁴S, ¹⁵N/¹⁵N/¹³C/³²S isotopomers), 371 (18, [MH]^{+ 15}N/¹²C/³⁴S isotopomer).

General Synthesis for Cp*RhCl(4-RC₆H₄SO₂NCHPhCHPh-NH₂).¹⁷ [Cp*RhCl₂]₂ and 4-RC₆H₄SO₂DPEN were dissolved in CH₂Cl₂, NEt₃ was added dropwise with stirring, and the resulting solution was stirred for 1 h. The solution was washed with water, dried over MgSO₄, and crystallized from CH₂Cl₂/hexane as orange needles.

Cp*RhCl(TsNCHPhCHPhNH₂) (2a). [Cp*RhCl₂]₂ (2.0 g, 3.2 mmol), TsDPEN (2.4 g, 6.5 mmol), and NEt₃ (1.3 g, 12.9 mmol) were reacted in CH₂Cl₂ (120 mL), to give 2a (4.2 g, 93%). ¹H NMR (500 MHz, CDCl₃, 300 K, numbering as in Figure 2): δ 1.80 (15H, s, C₅Me₅), 2.21 (3H, s, Me), 3.46 (1H, br d, $J_{\rm HH} \approx 11$ Hz, NH^b), 3.70 (1H, ddd, $J_{\rm HH} = 3$, 12, 14 Hz, H¹⁰), 3.84 (1H, br apparent t, $J_{\rm HH} \approx 12$, 12 Hz, NH^a), 3.95 (1H, d, $J_{\rm HH} = 11$ Hz, H⁵), 6.64 $(2H, d, J_{HH} = 7 \text{ Hz}, H^7 \text{ or } H^8), 6.76 (2H, t, J_{HH} = 7 \text{ Hz}, H^7 \text{ or } H^8),$ 6.80-6.85 (5H, m, H³, H⁹, and H¹²), 7.09-7.14 (3H, m, H¹³ and H¹⁴), 7.29 (2H, d, H²). ¹³C NMR (125 MHz, CD₂Cl₂, 300 K): δ 9.97 (s, C₅Me₅) 21.40 (s, Me), 69.91 (s, C⁵), 72.05 (s, C¹⁰), 94.92 (d, $J_{RhC} = 8$ Hz, C_5Me_5), 126.83 (s, C³, C⁹, or C¹³), 127.54 (s, C⁷) or C⁸), 127.59 (s, C³, C⁹, or C¹³), 128.27 (s, C³, C⁹, or C¹³), 128.49 (s, C²), 128.78 (s, C¹³ or C¹⁴), 128.97 (s, C¹³ or C¹⁴), 129.34 (s, C⁷ or C⁸), 139.79 (s, C¹), 140.06 (s, C⁶ or C¹¹), 140.29 (s, C⁶ or C¹¹), 142.27 (s, C⁴). ¹⁵N NMR (50.7 MHz, CD₂Cl₂, 300 K): δ -263.6 $(d, J_{RhN} = 14 \text{ Hz}, \text{NH}_2), -210.6 (d, J_{RhN} = 18 \text{ Hz}, \text{NTs}).$ ¹⁰³Rh-¹H HMQC (22.18 MHz, CDCl₃, 300 K): δ 2550.5. MS (FAB, *m/z*): 639 ([MH]⁺). IR (KBr/ cm⁻¹): 3281 ($\nu_{\rm NH}$), 3209 ($\nu_{\rm NH}$).

¹H NMR (500 MHz, CD₃CN, 300 K): δ 1.80 (15H, s, C₅Me₅), 2.24 (3H, s, Me), 3.60 (1H, br apparent t, $J_{\text{HH}} \approx 12$, 12 Hz, NH^a), 3.78 (1H, ddd, $J_{\text{HH}} = 3$, 11, 14 Hz, H¹⁰), 3.91 (1H, d, $J_{\text{HH}} = 11$ Hz, H⁵), 4.41 (1H, br d, $J_{\text{HH}} \approx 9$ Hz, NH^b), 6.68 (2H, d, $J_{\text{HH}} = 8$ Hz, H⁷ or H⁸), 6.77 (2H, t, $J_{\text{HH}} = 7$ Hz, H⁷ or H⁸), 6.83–6.88 (5H, m, H³, H⁹, and H¹²), 7.11–7.14 (3H, m, H¹³ and H¹⁴), 7.32 (2H, d $J_{\text{HH}} = 8$ Hz, H²).

Cp*RhCl(4-*t***BuC₆H₄SO₂NCHPhCHPhNH₂) (2b).** [Cp*RhCl₂]₂ (0.76 g, 1.23 mmol), 4-*t*BuC₆H₄SO₂DPEN (1.0 g, 2.45 mmol), and NEt₃ (0.68 mL, 4.9 mmol) were reacted in CH₂Cl₂ (50 mL) to give **2b** (1.0 g, 60%). Crystals suitable for X-ray analysis were grown by slow diffusion of hexane into a concentrated solution of **2b** in CH₂Cl₂. ¹H NMR (500 MHz, CD₂Cl₂, 300 K, numbering as in Figure 2): δ 1.21 (9H, s, CMe₃), 1.80 (15H, s, C₅Me₅), 3.47 (1H, br d, *J*_{HH} \approx 10 Hz, NH^b), 3.69 (1H, ddd, *J*_{HH} = 3, 12, 14 Hz, H¹⁰), 3.90 (1H, br apparent t, *J*_{HH} \approx 11, 11 Hz, NH^a), 4.07 (1H, d, *J*_{HH} = 11 Hz, H⁵), 6.59 (2H, d, *J*_{HH} = 7 Hz, H⁷), 6.66 (2H, t, *J*_{HH} = 8 Hz, H⁸), 6.71–6.74 (1H, m, H⁹), 6.85–6.87 (1H, m, H¹²), 6.96 (2H, d, *J*_{HH} = 9 Hz, H³). ¹³C (125 MHz, CD₂Cl₂, 300 K): δ 9.96

(s, C₅Me₅), 31.40 (s, *CMe*₃), 69.75 (s, C⁵), 71.90 (s, C¹⁰), 99.48 (d, $J_{\rm RhC} = 9$ Hz, C_5 Me₅), 124.60 (s, C²), 126.96 (s, C⁹), 127.53 (s, C⁸), 127.84 (s, C¹²), 127.86 (s, C³), 128.76 (s, C¹³ or C¹⁴), 128.96 (s, C¹³ or C¹⁴), 129.35 (s, C⁷), 139.72 (s, C⁶ or C¹¹), 140.12 (s, C⁶ or C¹¹), 143.02 (s, C⁴), 152.53 (s, C¹). ¹⁵N NMR (50.7 MHz, CD₂Cl₂, 300 K): δ –263.7 (d, $J_{\rm RhN} = 17$ Hz, NH₂), –208.6 (d, $J_{\rm RhN} = 20$ Hz, NTs). MS (FAB, *m*/*z*): 681 ([MH⁺]). IR (KBr/cm⁻¹): 3287 ($\nu_{\rm NH}$), 3233 ($\nu_{\rm NH}$). Anal. Calcd for C₃₄H₄₂ClN₂O₂RhS: C, 59.95; H, 6.22; N, 4.11. Found: C, 59.70; H, 6.25; N, 4.16.

¹H NMR (500 MHz, CD₃CN, 300 K): δ 1.24 (9H, s, CMe₃), 1.80 (15H, s, C₅Me₅), 3.64 (1H, br apparent t, $J_{\text{HH}} = 11$, 11 Hz, NH^b), 3.78 (1H, ddd, $J_{\text{HH}} = 3$, 12, 14 Hz, H¹⁰), 4.00 (1H, d, $J_{\text{HH}} =$ 11 Hz, H⁵), 4.42 (1H, br d, $J_{\text{HH}} = 9$ Hz, NH^a), 6.63 (2H, d, $J_{\text{HH}} =$ 7 Hz, H⁷), 6.68 (2H, d, $J_{\text{HH}} = 7$ Hz, H⁸), 6.73–6.76 (1H, m), 6.90–6.92 (2H, m), 7.04 (2H, d, $J_{\text{HH}} = 9$ Hz), 7.12–7.13 (3H, m), 7.27 (2H, d $J_{\text{HH}} = 9$ Hz).

Cp*RhCl(4-FC₆H₄SO₂NCHPhCHPhNH₂) (2c). [Cp*RhCl₂]₂ (0.83 g, 1.35 mmol), 4-FC₆H₄SO₂DPEN (1.0 g, 2.7 mmol), and NEt₃ (0.75 mL,5.4 mmol) were reacted in CH₂Cl₂ (50 mL) to give **2c** (1.35 g, 78%). Crystals suitable for X-ray analysis were grown by slow diffusion of hexane into a concentrated solution of 2c in CH₂Cl₂. ¹H NMR (500 MHz, CDCl₃, 300 K): δ 1.80 (15H, s, C_5Me_5), 3.54 (1H, br d, $J_{HH} \approx 10$ Hz, NH^b), 3.70 (1H, ddd, $J_{HH} =$ 2, 12, 14 Hz, H¹⁰), 3.84 (1H, br apparent t, $J_{\rm HH} \approx 11$, 11 Hz, NH^a), $3.96 (1H, d, J_{HH} = 11 \text{ Hz}, \text{H}^5), 6.62 (2H, d, J_{HH} = 9 \text{ Hz}, \text{H}^7 \text{ or } \text{H}^8),$ 6.65-6.68 (2H, m, H⁷ or H⁸), 6.78 (2H, apparent t, $J_{\rm HH} = 8, 8$ Hz, $\rm H^2),~6.82{-}6.85$ (3H, m, $\rm H^9$ and $\rm H^{12}),~7.10$ (3H, m, $\rm H^{13}$ and $\rm H^{14}),$ 7.40 (2H, dd, $J_{\rm HH} = 8$ Hz, $J_{\rm HF} = 5$ Hz, H³). ¹³C (125 MHz, CD₂Cl₂, 300 K): δ 9.96 (s, C₅Me₅), 69.85 (s, C⁵), 71.87 (s, C¹⁰), 94.92 (d, $J_{\text{RhC}} = 8$ Hz, C_5 Me₅), 113.9 (C^{2,d}, $J_{\text{FC}} = 22$ Hz), 127.09 (s, C⁹ or C^{12} , 127.55 (s, C⁹ or C¹²), 127.66 (s, C⁷ or C⁸), 128.79 (s, C¹³ or C¹⁴), 128.96 (s, C¹³ or C¹⁴), 129.32 (s, C⁷ or C⁸), 130.79 (d, $J_{FC} = 6$ Hz, C³), 139.87 (s, C⁵ or C¹⁰), 139.94 (s, C⁵ or C¹⁰), 141.70 (s, C⁴), 162.42 (d, $J_{FC} = 6$ Hz, C³), 139.87 (s, C⁵ or C¹⁰), 139.94 (s, C⁵ or C¹⁰), 141.70 (s, C⁴), 162.42 (d, $J_{FC} = 6$ Hz, C³), 139.87 (s, C⁵ or C¹⁰), 139.94 (s, C⁵ or C¹⁰), 141.70 (s, C⁴), 162.42 (d, $J_{FC} = 6$ Hz, C³), 139.87 (s, C⁵ or C¹⁰), 139.94 (s, C⁵ or C¹⁰), 141.70 (s, C⁴), 162.42 (d, $J_{FC} = 6$ Hz, C³), 139.87 (s, C⁵ or C¹⁰), 139.94 (s, C⁵ or C¹⁰), 141.70 (s, C⁴), 162.42 (d, $J_{FC} = 6$ Hz, C³), 162.42 (d, J_{FC} = 6 Hz, C³), 1 C⁴), 163.43 (d, $J_{FC} = 250$ Hz,C¹). ¹⁹F NMR (480 MHz, CD₂Cl₂, 300 K): $\delta -112.76$ (s). ¹H-¹⁵N HMQC NMR (50.7 MHz, CD₂Cl₂, 300 K): δ –263.8 (d, $J_{\rm RhN}$ = 16 Hz, NH2), –210.2 (d, $J_{\rm RhN}$ = 21 Hz, NTos). ¹H $^{-103}$ Rh HMQC (22.18 MHz, CDCl₃, 300 K): δ 2036.5. MS (FAB, m/z): 643 ([MH]⁺). IR (KBr/cm⁻¹): 3292 ($\nu_{\rm NH}$), 3234 (*v*_{NH}). Anal. Calcd for C₃₀H₃₃ClFN₂O₂RhS: C, 56.04; H, 5.17, N, 4.36. Found: C, 55.70; H, 5.39; N 4.39.

¹H NMR (500 MHz, CD₃CN, 300 K): δ 1.80 (15H, s, C₅Me₅), 3.61 (1H, br apparent t, $J_{\rm HH} \approx 11$, 11 Hz, NH^b), 3.80 (1H, ddd, $J_{\rm HH} = 3$, 12, 14 Hz, H¹⁰), 3.94 (1H, d, $J_{\rm HH} = 11$ Hz, H⁵), 4.44 (1H, br d, $J_{\rm HH} \approx 8$ Hz, NH^a), 6.69 (2H, d, $J_{\rm HH} = 7$ Hz), 6.75–6.79 (4H, m), 6.83–6.85 (1H, m), 6.89–6.91 (2H, m) 7.12–7.14 (3H, m), 7.44 (2H, dd $J_{\rm HH} = 9$ Hz, $J_{\rm FH} = 5$ Hz).

¹⁵N-Labeled Cp*RhCl(TsNCHPhCHPhNH₂) (2a-¹⁵N). [Cp*Rh-Cl₂]₂ (36.0 mg, 58 μmol), TsDPEN (41.0 mg, 112 μmol), and NEt₃ (33 μL, 237 μmol) were reacted in CH₂Cl₂ (5 mL), to give ¹⁵N-enriched **2a** (54 mg, 71%). ¹H NMR (500 MHz, CDCl₃, 300 K): δ 1.80 (15H, s, C₅Me₅), 2.21 (3H, s, Me), 3.48 (1H, dd, J_{NH} = 72 Hz, J_{HH} = 10 Hz), 3.70 (1H, ddd, J_{HH} = 14, 12, 3 Hz), 3.86 (1H, d of apparent t, J_{NH} = 82 Hz, J_{HH} = 11, 11 Hz), 3.96 (1H, d, J_{HH} = 11 Hz), 6.63 (2H, d, J_{HH} = 7 Hz), 6.75 (2H, t, J_{HH} = 7 Hz), 6.80-6.85 (5H, m), 7.06-7.14 (3H, m), 7.28 (2H, d, J_{HH} = 7 Hz). ¹⁵N{¹H} NMR (50.7 MHz, CD₂Cl₂, 300 K): δ -263.6 (d, J_{RhN} = 14 Hz, NH₂), -210.6 (d, J_{RhN} = 18 Hz, NTs). MS (FAB, *m*/z): 639 ([MH⁺] ¹⁴N/¹⁴N isotopomer), 640 ([MH]^{+ 14}N/¹⁵N isotopomer), 641 ([MH]^{+ 15}N/¹⁵N isotopomer).

Synthesis of [Cp*Rh(CO)(CONTsCHPhCHPhNH₂)][Cl] (3a). 3a was prepared by adding CO (3 atm) to an NMR tube fitted with a Young's tap containing a degassed solution of **2a** in dry CD₂Cl₂, CD₃OD, or a mixture of CD₂Cl₂/CD₃OD (1%). ¹H NMR (500 MHz, CD₂Cl₂, 300 K): δ 1.68 (s, 15H, C₅Me₅), 1.83 (s, 15H, C₅Me₅), 2.28 (s, 3H, Me), 2.31 (s, 3H, Me), 4.49–4.51 (m, 2H, CH), 5.04–5.09 (m, 2H, CH), 6.02 (d, *J*_{HH} = 6 Hz, 1H, NH), 6.07 (d, *J*_{HH} = 6 Hz, 1H, NH), 6.82–7.40 (m, 28H, aromatic), 7.56 (m, 1H, NH), 7.78 (d, $J_{\rm HH} = 8$ Hz, 1H, NH). ¹³C NMR (75 MHz, CD₂Cl₂, 300 K): δ 9.06 (s, C_5Me_5), 9.30 (s, C_5Me_5), 21.16 (s, Me), 60.72 (s, CH), 60.84 (s, CH), 63.55 (s, CH), 64.10 (s, CH), 106.24 (d, $J_{\rm RhC} = 5$ Hz, C_5Me_5), 106.55 (d, $J_{\rm RhC} = 5$ Hz, C_5Me_5), 124.86 (s), 126.90 (s), 127.26 (s), 127.59 (s), 127.66 (s), 127.78 (s), 127.83 (s), 128.44 (s), 128.68 (s), 129.10 (s), 129.32 (s), 137.30 (s), 137.46 (s), 137.95 (s), 138.24 (s), 138.36 (s), 138.43 (s), 142.77 (s), 142.98 (s), 181.58 (d, $J_{\rm RhC} = 27$ Hz, NCO), 181.75 (d, $J_{\rm RhC} = 27$ Hz, NCO), 188.08 (d, $J_{\rm RhC} = 77$ Hz, CO), 188.14 (d, $J_{\rm RhC} = 77$ Hz, CO), 1³C NMR (125 MHz, CD₂Cl₂, 300 K): δ 181.6 (d, $J_{\rm RhC} = 77$ Hz, CO), 181.7 (d, $J_{\rm RhC} = 77$ Hz, CO), 188.1 (d, $J_{\rm RhC} = 77$ Hz, CO), 188.2 (d, $J_{\rm RhC} = 77$ Hz, CO), 188.1 (d, $J_{\rm RhC} = 77$ Hz, CO), 188.2 (d, $J_{\rm RhC} = 77$ Hz, CO), 188.1 (d, $J_{\rm RhC} = 77$ Hz, CO), 188.2 (d, $J_{\rm RhC} = 77$ Hz, CO). MS (FAB, m/z): 603 ([M – 2CO]⁺). IR (CH₂Cl₂/cm⁻¹): 2056 ($\nu_{\rm CO}$), 1600 ($\nu_{\rm CO}$).

[Cp*Rh(tBuNC)(TsNCHPhCHPhNH₂)][PF₆] (4a). 4a was prepared by addition of tBuNC (13 mg, 0.16 mmol) to a solution of 2a (100 mg, 0.16 mmol) in CH₃OH (5 mL), stirring for 1 h, removing the solvent under reduced pressure, and extracting into toluene. The addition of KPF₆ (59 mg, 0.32 mmol) to the toluene solution resulted in the precipitation of a yellow solid. The yellow solid was extracted into THF, filtered, and crystallized from THF/ hexane to give 4a (102 mg, 76%) as yellow needles. ¹H NMR (500 MHz, CD₃CN, 300 K): δ 1.70 (s, 9H, *t*Bu), 1.91 (s, 15H, C₅Me₅), 2.15 (s, 3H, Me), 3.75 (ddd, $J_{\rm HH} = 3$, 12, 14 Hz, 1H, H¹⁰), 4.09 (br d, $J_{\rm HH} \approx 11$ Hz, 1H, NH^a), 4.15 (d, $J_{\rm HH} = 11$ Hz, 1H, H⁵), 4.29 (br apparent t, $J_{\rm HH}\approx$ 11, 11 Hz, 1H, NH^b), 6.60 (d, $J_{\rm HH}=$ 7 Hz, 2H, Ph), 6.65 (t, $J_{\rm HH} = 8$ Hz, 2 H, Ph), 6.74 (d, $J_{\rm HH} = 8$ Hz, 2H, H²), 6.74-6.76 (m, 1H, H⁹, or H¹⁴), 7.03-7.04 (m, 2H, Ph), 7.12 (d, $J_{\rm HH} = 8$ Hz, 2H, H³), 7.18 (m, 3H, Ph). ¹³C NMR (125 MHz, CD₃CN, 300 K): δ 10.33 (s, C₅Me₅), 21.50 (s), 30.64 (s, CMe₃), 60.37 (s, CMe₃), 71.74 (s, C¹⁰), 73.84 (s, C⁵), 102.26 (d, $J_{RhC} = 7$ Hz, C₅Me₅), 127.60 (s, Ph), 128.52 (s, Ph), 128.77 (s, Ph), 129.03 (s, C²), 129.33 (s, C³), 129.62 (s, Ph), 130.00 (s, Ph), 134.8 (d, $J_{\text{RhC}} = 69$, CN), 139.69 (s, Ph), 140.11 (s, Ph), 141.40 (s, C¹ or C⁴), 141.91 (s, C¹ or C⁴). ³¹P NMR (202 MHz, CD₃CN, 300 K): δ -144.61 (sept, $J_{\rm FP} = 702$ Hz). MS (ESI, m/z): 686 ([M⁺]). IR (KBr/ cm⁻¹): 3336 ($\nu_{\rm NH}$), 3292 ($\nu_{\rm NH}$); 2212 ($\nu_{\rm CN}$). Anal. Calcd for C₃₆H₄₅F₆N₃O₂PRhS: C, 51.99; H, 5.42; N, 5.05. Found: C, 51.68; H, 5.48; N, 4.99.

[(Cp*Rh)₂(µ-H)(µ-Cl)(µ-HCO₂)][BPh₄] ([5a][BPh₄]). 2a (200 mg, 0.32 mmol) was dissolved in CH₃OH (10 mL) with the aid of sonication, and HCO₂H (150 mg, 3.26 mmol) was added via microsyringe. The solution turned from orange to red and was stirred for a further 15 min before stirring was stopped and NaBPh₄ (220 mg, 0.64 mmol) was added as a solid, and the flask was swirled to afford mixing, causing a red precipitate to settle from the solution upon standing. The solid was removed by filtration, dried under reduced pressure, and crystallized from acetone/Et₂O (1:1) at -20°C to give [5a][BPh₄] as red crystals (121 mg, 43%). Crystals suitable for X-ray analysis were grown from a solution of [5a][BPh₄] in (CH₃)₂CO/Et₂O (1:1) at -20 °C. ¹H NMR (500 MHz, (CD₃)₂CO, 300 K): δ -8.59 (t, J_{RhH} = 27.4 Hz, 1H, Rh-H-Rh), 1.94 (s, 30H, C₅Me₅), 6.77 (t, $J_{\rm HH} = 7.2$ Hz, 4H, BPh₄), 6.92 (t, $J_{\rm HH} = 7.5$ Hz, 8H, BPh₄), 7.33–7.35 (m, 8H, BPh₄), 7.46 (t, $J_{\rm HH}$ = 3.5 Hz, 1H, HCO_2). ¹³C NMR (125 MHz, $(CD_3)_2CO$, 300 K): δ 10.09 (s, C₅Me₅), 98.93-98.99 (m, C₅Me₅), 122.29 (s, BPh₄), $126.01 - 126.08 \text{ (q, } J_{BC} = 3 \text{ Hz, BPh}_4\text{), } 137.07 - 137.08 \text{ (m, BPh}_4\text{), }$ 164.40 - 165.58 (q, $J_{BC} = 49$ Hz, BPh₄), 173.84 (s, HCO₂). IR (KBr/ cm⁻¹): 1556 (ν_{CO}). MS (ESI, m/z): 557 ([M]⁺). Anal. Calcd for C45H52BClO2Rh2: C, 61.63; H, 5.98. Found: C, 61.21; H, 6.12.

Transfer Hydrogenation of d_6 -Acetone with d_{15} -NEt₃/Formic Acid Catalyzed by 2a. (*S*,*S*)-2a (4.4 mg, 6.8 μ mol) was dissolved in CD₃CN; d_6 -acetone (10 μ L, 0.14 mmol) was added to the tube via microsyringe followed by d_{15} -NEt₃ (35 μ L, 0.27 mmol). At each stage of addition, a ¹H NMR spectrum was recorded. The sample was cooled to 250 K, HCO₂H (26 μ L, 0.680 mmol) was added to the tube, and spectra were recorded every 10 min for 1 h. The experiment was repeated at 300 K.

Metathesis Reactions of 2a and 2c. $Cp*RhCl(4-RC_6H_4SO_2NC-HPhCHPhNH_2)$ was dissolved in the minimum volume of CH₃OH (often with the aid of sonication), the relevant reagent was added as a solid (LiBr, KI, AgOTf) or liquid (*t*BuNC), and the solution was stirred for 1 h or more. The solvent was removed under reduced pressure and the residue extracted into CH₃CN, filtered through Celite, and concentrated to dryness under vacuum. The product was recrystallized by layering a THF solution with hexane.

Synthesis of Cp*RhBr(TsNCHPhCHPhNH₂) (6a). 6a was prepared as described above from 2a (150 mg, 0.23 mmol), LiBr (21 mg, 0.24 mmol), and CH₃OH (10 mL). After removal of CH₃CN the product was crystallized from THF/hexane to give 6a (140 mg, 89%) as red crystals. Crystals suitable for X-ray analysis were grown by slow diffusion of hexane into a concentrated solution of **6a** in THF. ¹H NMR (500 MHz, CD₃CN, 300 K, numbering as in Figure 2): δ 1.82 (s, 15H, C₅Me₅), 2.19 (s, 3H, Me), 3.75 (br apparent t, $J_{\rm HH} \approx 11$, 11 Hz, 1H, NH^a), 3.83 (ddd, $J_{\rm HH} = 3$, 12, 14 Hz, 1H, H¹⁰), 4.14 (d, $J_{\rm HH} = 11$ Hz, 1H, H⁵), 4.62 (br d, $J_{\rm HH} \approx 10$ Hz, NH^b), 6.65-6.71 (m, 4H, Ph), 6.76-6.77 (m, 1H, Ph), 6.80 (d, $J_{\rm HH} = 8$ Hz, 2H, H³), 7.05 (m, 2H, Ph), 7.12–7.13 (m, 3H, Ph), 7.23 (d, $J_{\rm HH} = 8$ Hz, 2H, H²). ¹³C NMR (125 MHz, CD₃CN, 300 K): δ 10.41 (s, C₅Me₅), 21.55 (s, Me), 70.39 (s, C¹⁰), 72.55 (s, C⁵), 95.96 (d, $J_{RhC} = 9$ Hz, C_5Me_5), 127.47 (s, Ph), 128.31 (s, Ph), 128.88 (s, C² and Ph), 129.13 (s, Ph), 129.20 (s, H²), 129.48 (s, Ph), 130.34 (s, Ph), 140.52 (s, C¹), 140.96 (s, Ph), 141.08 (s, Ph), 143.07 (s, C⁴). IR (KBr)/cm⁻¹): 3271 (ν_{NH}), 3210 (ν_{NH}). MS (FAB, *m*/z): 682 ([M⁺] ⁷⁹Br isotopomer), 684 ([M⁺] ⁸¹Br isotopomer).

Cp*RhI(TsNCHPhCHPhNH2) (7a). 7a was prepared as described above from 2a (250 mg, 0.34 mmol) and KI (57 mg, 0.34 mmol) in CH₃OH (12 mL). The product was recrystallized from THF/hexane to give 7a (174 mg, 70%) as dark red crystals. Crystals suitable for X-ray analysis were grown by slow evaporation of a solution of **7a** in CH₃OH. ¹H NMR (500 MHz, CD₃CN, 300 K): δ 1.92 (s, 15H, C₅Me₅), 2.21 (s, 3H, Me), 3.83 (m, 1H, H^a), 3.84 (m, 1H, H¹⁰), 4.01 (d, $J_{\rm HH} = 11$ Hz, 1H, H⁵), 4.35 (br d, $J_{\rm HH} \approx 9$ Hz, 1H, H^b), 6.66-6.69 (m, 4H, Ph), 6.74-6.76 (m, 1H, Ph), 6.80 (d, $J_{\rm HH} = 8$ Hz, H³), 6.92–6.94 (m, 2H, Ph), 7.12–7.13 (m, 3H, Ph), 7.21 (d, $J_{\rm HH} = 8$ Hz, 2H, H²). ¹³C NMR (125 MHz, CD₃CN, 300 K): δ 11.11 (s, C₅Me₅), 21.46 (s, Me), 71.29 (s, C¹⁰), 72.16 (s, C⁵), 96.12 (d, $J_{\text{RhC}} = 8$ Hz, C_5 Me₅), 127.37 (s, Ph), 128.06 (s, Ph), 128.67 (s, C²), 128.87 (s, C³), 129.06 (s, Ph), 129.51 (s, Ph), 130.58 (s, Ph), 140.19 (s, C¹), 140.33 (s, Ph), 141.22 (s, Ph), 144.46 (s, C⁴). IR (KBr/cm⁻¹): 3278 (ν_{NH}), 3233 (ν_{NH}). MS (FAB, *m/z*): 731 $([MH]^+)$. Anal. Calcd for C₃₁H₃₆IN₂O₂RhS: C, 50.97; H, 4.97; N, 3.83. Found: C, 50.45; H, 5.03; N, 3.61.

[Cp*Rh(4-FC₆H₄SO₂NCHPhCHPhNH₂)][CF₃SO₃], [8c][OTf] and [8c-H₂O][OTf]. [8c][OTf] was prepared as described above from 2c (500 mg, 0.78 mmol), AgCF₃SO₃ (200 mg, 0.78 mmol), and CH₃OH (40 mL). The product mixture was filtered through Celite before the solvent was removed under vacuum. Compound [8c][OTf] (470 mg, 80%) was obtained as a purple powder. Crystals suitable for X-ray analysis were grown by slow diffusion of hexane into a concentrated solution of [8c][OTf] in CH₃OH/toluene (1:1) but proved to be [8c-H₂O][OTf]. ¹H NMR (500 MHz, CD₃CN, 300 K): δ 1.83 (s, 15H, C₅Me₅), 3.75 (ddd, J_{HH} = 3, 12, 14 Hz, 1H, H¹⁰), 3.95 (apparent t, 11 Hz,1H NH^b), 4.15 (d, 11 Hz, 1H, H⁵), 4.50 (d, J_{HH} = 10 Hz, 1H, NH^a), 6.63–6.68 (m, 4H, Ph), 6.70 (apparent t, J_{HH} = 8 Hz, J_{FH} = 8 Hz, 2H, H²), 6.76–6.79 (m, 1H, Ph), 7.06–7.08 (m, 2H, Ph), 7.16–7.17 (m, 3H, Ph), 7.31 (dd, J_{HH} = 11 Hz, J_{FH} = 5 Hz, 2H, H³). ¹³C NMR (125 MHz, CD₃CN, 300 K): δ 9.71 (s, C₅*Me*₅), 70.65 (s, C¹⁰), 72.64 (s, C⁵), 98.08 (d, *J*_{RhC} = 8 Hz, *C*₅Me₅), 115.10 (d, *J*_{FC} = 22 Hz, C²), 122.09 (q, *J*_{FC} = 316 Hz, CF₃), 127.49 (s, Ph), 128.23 (s, Ph), 129.09 (s, Ph), 129.13 (s, Ph), 129.81 (s, Ph), 130.85 (d, *J*_{FC} = 10 Hz, C³), 139.59 (s, Ph ipso), 139.94 (s, Ph ipso), 142.09 (d, *J*_{FC} = 4 Hz, C⁴), 163.84 (d, *J*_{FC} = 243 Hz, C¹). ¹⁹F NMR (470 MHz, CD₃CN, 300 K): δ -79.2 (s, 3F, CF₃), -113.0 (s, 1F, FC₆H₄). MS (ESI, *m/z*): 607 ([M]⁺). IR (KBr/cm⁻¹): 3278 (*v*_{NH}), 3246 (*v*_{NH}). Anal. Calcd for [**8c**-H₂O][OTf], C₃₁H₃₅F₄N₂O₆RhS₂: C, 48.07; H, 4.55; N, 3.62. Found: C, 47.89; H, 4.51; N, 3.65.

[Cp*Rh(tBuNC)(4-FC₆H₄SO₂NCHPhCHPhNH₂)][CF₃SO₃] (4c). 4c was prepared from 8c by adding tBuNC (11 mg, 0.13 mmol) to a solution of 8c (100 mg, 0.13 mmol) in CH₃OH (3 mL), filtering, and removing the solvent under vacuum, to give 4c (46 mg, 55%) as a yellow powder. ¹H NMR (500 MHz, CD₃CN, 300 K): δ 1.70 (s, 9H, CMe₃), 1.91 (s, 15H, C₅Me₅), 3.75 (ddd, $J_{\rm HH} = 3$, 12, 14 Hz, 1H, H^{10}), 4.12–4.18 (m, 1H, NH^a), 4.15 (d, $J_{HH} = 11$ Hz, 1H, H⁵), 4.44 (br apparent t, $J_{\rm HH} \approx 12$, 12 Hz, 1H, nH^b), 6.63–6.68 (m, 4H, Ph), 6.70 (dd, $J_{\rm HH} = 8$ Hz, $J_{\rm FH} = 8$ Hz, 2H, H²), 6.76–6.79 (m, 1H, Ph), 7.04-7.07 (m, 2H, Ph), 7.17-7.18 (m, 3H, Ph), 7.26–7.28 (dd, $J_{\text{FH}} = 5$ Hz, $J_{\text{HH}} = 9$ Hz, 2H, H³). ¹³C NMR (125 MHz, CD₃CN, 300 K): δ 10.39 (s, C₅Me₅), 30.68 (s, CMe₃), 60.62 (s, CMe₃), 71.80 (s, C¹⁰), 73.76 (s, C⁵), 102.26 (d, $J_{RhC} = 7$ Hz, C_5 Me₅), 115.62 (d, $J_{FC} = 23$ Hz, C²), 122.81 (q, $J_{FC} = 335$ Hz, CF₃), 128.01 (s, Ph), 128.75 (s, Ph), 129.14 (s, Ph), 129.69 (s, Ph), 130.20 (s, Ph), 131.44 (d, $J_{FC} = 10$ Hz, C³), 139.61 (s, Ph), 140.09 (s, Ph), 141.37 (d, $J_{FC} = 4$ Hz, C⁴), 164.0 (d, $J_{FC} = 243$ Hz, C¹). ¹⁹F NMR (470 MHz, CD₃CN, 300 K): δ -79.23 (s, 3F, CF₃), -112.63 (s, 1F, FC₆H₄). MS (ESI, *m/z*): 690 ([M]⁺).

X-ray Crystallography. X-ray data were collected on a Bruker SMART Apex X-ray diffractometer for $\theta > 1.72^{\circ}$ up to $\theta <$ 28.1-29.99°. The crystals were cooled to 110 K using an Oxford Cryosystems Cryostream. Diffractometer control, data collection, and initial unit cell determination were performed using SMART (v5.625 Bruker-AXS). Frame integration and unit-cell refinement were carried out with SAINT+ (v6.22, Bruker AXS). Absorption corrections were applied by SADABS (v2.03, Sheldrick). The structures were solved by direct methods using SHELXS-97 and refined by full-matrix least-squares using SHELXL-97.63,64 Hydrogen atoms bound to carbon were placed using a riding model and included in the refinement at calculated positions. Hydrogen atoms attached to nitrogen in 2c, 6a, and 7a were located on difference maps after all non-hydrogen atoms had been located and were refined isotropically (2c, 7a). Crystal data for complexes 2c, [5a][BPh₄] • OEt₂, 6a, 7a, and 8c-H₂O are shown in Table 2.

Acknowledgment. We acknowledge the support of EPSRC and NPIL Ltd. We also appreciated discussions with Matthew Stirling, George Hodges, Anthony Haynes, and Richard Douthwaite. We thank David Taylor for recording the rhodium NMR spectra, and Richard Lindup for assistance with preparing the manuscript. We thank the reviewer for helpful criticism of the manuscript.

Supporting Information Available: Cif files for 2c, $[5a][BPh_4] \cdot OEt_2$, 6a, 7a, and $8c \cdot H_2O$. This material is available free of charge via the Internet at http://pubs.acs.org.

OM8009969

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