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Structures and dynamics of the lowest excited triplet states and cation radicals of phenothiazine and 2-chlorophenothiazine: transient resonance Raman and absorption study¹

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Abstract

Transient resonance Raman and absorption spectra of the lowest excited triplet states T_1 and the cation radicals of phenothiazine and 2-chlorophenothiazine were measured. It was found that in the photoreaction of 2-chlorophenothiazine a transient exhibiting an absorption band at 556 nm was generated from the cation radical. The corresponding transient was not observed in the absorption spectrum of phenothiazine, a fact which suggests a possibility of phenothiazinyl radical generation for 2-chlorophenothiazine by photoinduced dechlorination. Vibrational assignments of the T_1 states and the cation radicals of the both compounds were made based on the frequency shifts on isotopic substitutions. Unusually large low-frequency shifts of the phenyl 8a and 8b modes were observed in the T_1 state but no appreciable shifts were detected in the cation radical, indicating that the phenyl rings are drastically weakened, and therefore, the phenyl C–C bonds are very much lengthened in the T_1 state. This implies that the excitation is strongly localized on the phenyl rings and the T_1 state has an $n-\pi^+$ character. © 1997 Elsevier Science B.V.

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1. Introduction

The photochemistry of phenothiazine and its derivatives (Fig. 1) has been extensively investigated [1-10] because of their pharmacological interest. Phenothiazine tranquilizers, particularly chlorpromazine (2-chloro-N-[3-dimethylamino-propyl]phenothiazine), have long been used for the treatment of psychotic disorders [11]. However, they are

known to cause both phototoxic and photoallergic reactions in the skin [12] and eyes [13,14] of patients receiving these drugs. While detailed mechanisms of the phototoxicity and photoallergy of these drugs are not known, it is obvious that photolytically generated excited states, radicals or the compounds derived from them play important roles in the mechanisms.

In this view, the photochemistry of phenothiazine and 2-chlorophenothiazine was investigated by means of time-resolved absorption and time-resolved resonance Raman spectroscopies in order to obtain information on the structures and dynamics of the excited states and radicals (neutral as well as cationic) which appear in the photochemical reactions of these

¹ Dedicated to Professor Kozo Kuchitsu on the occasion of his 70th birthday.

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compounds. Due to structural similarities, these two compounds are considered to be useful models for elucidating photochemical properties, particularly the structures and dynamics of the transients involved in the photochemical reactions of the clinically important drugs, i.e. promazine and chlorpromazine.

2. Experimental

The experimental arrangement of our time-resolved Raman spectroscopy system has been described previously [15]. Briefly, an excimer laser (Lambda Physik LPX120i) was used as a light source for pumping, and an excimer-laser-pumped dye laser (FL3002E) was used as a light source for Raman probing. The energy of the excimer laser used for the pumping was about 100 mJ pulse⁻¹ at the laser head and that of the dye laser used for Raman probing was 15-10 mJ pulse⁻¹ at the laser head depending on the wavelength. Both lasers (pulse width, 15-20 ns) were used at the repetition rate of 20 Hz. Time-resolved absorption spectra were measured by a laser flash photolysis system constructed in our laboratory which consists of an excimer laser (Lambda Physik LPX120i) for pumping, a xenon lamp for white light, and a 30 cm spectrometer equipped with a gated multichannel detector.

Phenothiazine was purchased from Kanto Chemical Co. Ltd and 2-chlorophenothiazine was synthesized

by the reaction of 3-chlorodiphenylamine with sulfur in the presence of a small amount (1 wt.% of diphenylamine) of iodine [16]. Both compounds were purified by recrystallization from ethanol. Although a small amount of a by-product, 4-chlorophenothiazine, was also produced in the above reaction, it was successfully removed by recrystallization. ³⁴S substituted phenothiazine and 2-chorophenothiazine were synthesized by the same reaction of ³⁴S (Isotec Inc., 90.46 at.%) with diphenylamine for the former and with 3-chlorodiphenylamine for the latter. Diphenylamine and 3-chlorodiphenylamine were purchased from Tokyo Kasei Organic Chemicals. N-D substituted phenothiazine was obtained by an exchange reaction with D₂O: stirring a mixture of CCl₄ solution of phenothiazine and D₂O under N₂ atmosphere, renewing D₂O several times, and finally removing CCl₄ by evaporation.

3. Results and discussion

3.1. Time-resolved absorption spectra

Time-resolved absorption spectra of phenothiazine (PTH) in deoxygenated methanol reveal that two transients are involved in the photoreaction of this compound (Fig. 2(a)): a transient exhibiting a strong absorption band at 455 nm with a lifetime of about



Fig. 2. Transient absorption spectra of phenothiazine: (a) in deoxygenated methanol measured at 100, 300, 500, 700 ns, 1 and 3 μ s after excitation with UV (308 nm) light; (b) in aerated methanol measured at 100, 200, 500 ns, 5 and 100 μ s; (c) in oxygenated methanol measured at 100 ns. Concentration is 1.0×10^{-3} mol dm⁻³.

800 ns and another transient having a very weak peak at 517 nm with a much longer lifetime. In aerated methanol solutions an additional transient exhibiting a band at 387 nm and having a long lifetime appears with a concomitant intensity decrease of the 455 nm band as shown in Fig. 2(b). In oxygen-saturated methanol solutions the 455 nm band disappears completely and the 387 nm band becomes more intense (Fig. 2(c)). These observations indicate that the transient exhibiting the 455 nm band is the T₁ state ³PTH^{*} and the 387 nm band may be attributable to a compound generated from ³PTH^{*} and O₂. The band at 517 nm and a shoulder around 435 nm may be assigned to the cation radical.

The above results are in good accord with the previous reports in the literature. Henry and Kasha [4] observed a band at 467 nm in both EPA and 3-methylpentane solutions at 77 K and assigned the band to ³PTH^{*}. Iwaoka et al. [5] assigned the band at 465 nm observed by flash photolysis in ethanol solutions to ³PTH^{*}. They also observed a band at about 385 nm in aerated ethanol solutions and assigned it to a charge-transfer complex PTH-O₂ generated through ³PTH^{*}. Shine and Mach [2] observed bands at 437 and 515 nm in aqueous acetic acid solutions irradiated with ultraviolet light and assigned them to the cation radical PTH⁺. Alkaitis et al. [6] observed two bands at 460 and 520 nm and assigned the former to ³PTH^{*} and the latter to PTH⁺⁻ which they considered to be generated through monophotonic ionization process.

Time-resolved absorption spectra of 2-chlorophenothiazine (CPTH) are a little more involved. At 200 ns after irradiation with UV (308 nm) light three bands are observed at 438, 458 and 517 nm. These bands decrease in intensity with time while a new band appears at 556 nm as shown in Fig. 3. Corresponding with the spectra of phenothiazine the band at 458 nm can be assigned to the T_1 state ³CPTH^{*}, and the bands at 517 and 438 nm can be attributed to the cation radical CPTH⁺. We have confirmed that the 458 nm band was quenched by oxygen. The new band at 556 nm appears to be generated from the cation radical because the rise time of this band is approximately the same as the decay time of the 517 nm band, and besides, there exists an isosbestic point



Fig. 3. Transient absorption spectra of 2-chlorophenothiazine in deoxygenated methanol: measured at 200, 400 ns. 1 and 10 μ s after UV (308 nm) excitation. Concentration is 5.0×10^{-4} mol dm⁻³.

between this band and the 517 nm band; the 458 nm band decays much faster. It is interesting to note that the 517 nm band is much more intense, and therefore, the yield of the cation radical is much larger in 2-chlorophenothiazine than in phenothiazine. This may suggest that the ionization potential of CPTH is much lower than that of PTH.

The assignment of the 556 nm band could not be determined unambiguously in the present investigation. We call the transient exhibiting this band "X". Since the transient X is not involved in the photoreaction of phenothiazine, its generation must be related to the chlorine substituent. Therefore, it seems probable that the 556 nm band arises from the phenothiazinyl neutral radical generated by dechlorination with UV light. Transient absorption spectra of 2-chlorophenothiazine were not reported previously.

3.2. Transient resonance Raman spectra

The Raman spectrum of the ground state S_0 of phenothiazine in acetone and resonance Raman spectra of ³PTH^{*} and PTH⁺⁻ in methanol are compared in Fig. 4. One feature of interest in these spectra is that the phenyl ring stretch modes 8a and 8b (Wilson vibration number [17]) observable at 1603 and 1575 cm⁻¹ in the S_0 spectrum are both very drastically down-shifted to 1507 cm⁻¹ in the T₄ state spectrum, while they do not exhibit appreciable frequency-shifts in the spectrum of the cation radical. Since the low-frequency shifts of the 8a and 8b modes (phenyl ring stretch modes) on



Fig. 4. The Raman spectrum of phenothiazine in the S₀ state and transient resonance Raman spectra of ³PTH^{*} and PTH⁺: (a) nearly saturated solution of S₀ in acetone. Probe wavelength: 581 nm; (b) ³PTH^{*} in deoxygenated methanol measured at 150 ns after the pumping with 308 nm light. Probe wavelength: 456 nm. Concentration: 2.0×10^{-3} mol dm⁻³; (c) PTH⁺ in oxygenated methanol measured at 100 ns after the pumping with 308 nm light. Probe wavelength: 517 nm. Concentration: 2.0×10^{-3} mol dm⁻³. The asterisk (*) denotes solvent bands.



Fig. 5. The Raman spectra of 2-chlorophenothiazine in the S₀ state and transient resonance Raman spectra of ⁵CPTH^{*} and CPTH^{*}: (a) nearly saturated solution of S₀ in acetone. Probe wavelength: 1064 nm: (b) ³CPTH^{*} in deoxygenated methanol measured at 150 ns after the pumping with 308 nm light. Probe wavelength: 456 nm. Concentration: 2.0×10^{-3} mol dm ⁻³: (c) CPTH^{*} in oxygenated methanol measured at 100 ns after the pumping with 308 nm light. Probe wavelength: 517 nm. Concentration: 2.0×10^{-3} mol dm ⁻³. The asterisk (*) denotes solvent bands.

going from the S_0 state to T_1 state are usually around $20-30 \text{ cm}^{-1}$ [18,19], the down-shift of 96 cm⁻¹ in phenothiazine is unusually large. This suggests that the excitation is strongly localized on the phenyl rings and that the phenothiazine phenyl rings are drastically weakened in the T_1 state. This point will be discussed later.

The Raman spectrum of the S_0 state of 2-chlorophenothiazine (CPTH) in acetone and resonance Raman spectra of ³CPTH^{*} and CPTH⁺ in methanol are shown in Fig. 5. Like phenothiazine the phenyl 8a and 8b modes at 1596 and 1567 cm⁻¹ are also drastically down-shifted to around 1510 cm⁻¹ in the spectrum of the T₁ state, whereas they do not exhibit appreciable shifting in the spectrum of the cation radical. The localized excitation on the phenyl rings is considered to be operative also in this compound. Comparison of the spectra of phenothiazine and 2chlorophenothiazine reveals that in the spectra of phenothiazine the bands corresponding to the bands of 2-chlorophenothiazine at 585 cm⁻¹ of the S₀ state, at 538 cm⁻¹ of the T₁ state, and at 553 cm⁻¹ of the cation radical are not observed. These bands can be assigned to the C–Cl stretching mode of CPTH, ³CPTH^{*} and CPTH⁺, respectively.

In order to obtain information on the structural changes on-going from the S_0 state to the T_1 state and to the cation radical, it is necessary to make more detailed vibrational assignments of their Raman bands. For this purpose we have synthesized isotopically substituted analogues, viz. ³⁴S substituted analogue (PTH-³⁴S) and N-D



Fig. 6. Comparison of Raman spectra of the S₀ states of phenothiazine and its isotopically substituted analogues: (a) PTH normal species in acetone (nearly saturated solution) measured with 581 nm light; (b) $PTH-^{34}S$ in acetone (nearly saturated solution) measured with 581 nm light; (c) $PTH-d_1$ in methanol- d_1 (saturated solution) measured with 532 nm light. The asterisk (*) denotes solvent bands.

substituted analogue (PTH- d_1), and measured the Raman or resonance Raman spectra of their S₀ and T₁ states and cation radicals.

Raman spectra of phenothiazine and its isotopically substituted analogues in the S_0 state, i.e. PTH, PTH-³⁴S and PTH-d₁ are compared in Fig. 6. It is seen that only one band exhibits an appreciable shifting on each of the ³⁴S and N-D substitutions: the band at 434 cm⁻¹ of PTH is down-shifted to 430 cm⁻¹ in the spectrum of PTH-³⁴S and the band at 1242 cm⁻¹ of PTH is down-shifted to 1238 cm⁻¹ in the spectrum of PTH-d₁. The 434 cm⁻¹ band can be assigned to a vibrational mode involving the largest contribution of the C-S symmetric stretch (for simplicity hereafter we call this mode "C-S symmetric stretch") and the 1242 cm⁻¹ band is assignable to a mode in which the contribution of the C-N symmetric stretch is the largest (hereafter "C–N symmetric stretch"). The frequency 434 cm⁻¹ appears to be considerably lower than the usual C–S stretching frequencies which are expected to lie around 580–750 cm⁻¹ [20]. The lower frequency of the C–S symmetric stretch of PTH is considered to be ascribed to the mixing with skeletal deformation of the ring. Although in this figure the PTH spectrum measured in acetone is compared with the PTH-d₁ spectrum measured in methanol-d₁, this comparison is justified because the frequency difference between the spectrum of PTH in acetone and that in methanol is negligibly small.

Resonance Raman spectra of phenothiazine and its isotopically substituted analogues in the T₁ state, i.e. ³PTH^{*}, ³PTH^{*}-³⁴S and ³PTH^{*}-d₁ are shown in Fig. 7. A strong band at 466 cm⁻¹ of the S₀ state is



Fig. 7. Transient resonance Raman spectra of the T₁ state of phenothiazine and its isotopically substituted analogues in deoxygenated methanol: (a) ³PTH^{*} normal species; (b) ³PTH^{*}- 34 S; (c) ³PTH^{*} - d₁. Pump wavelength: 308 nm; probe wavelength: 456 nm. Concentration: 2.0 × 10⁻³ mol dm⁻³. (For the measurement of ³PTH^{*} - d₁, methanol-d₁ was used to avoid the exchange of N–D deuterium with methanol O–H hydrogen.)

down-shifted to 460 cm⁻¹ on the ³⁴S substitution. The 466 cm⁻¹ band can be assigned to the C–S symmetric stretch. The band at 930 cm⁻¹ is also down-shifted to 917 cm⁻¹ on the ³⁴S substitution. This band is assignable to the first overtone of the 466 cm⁻¹ band. This assignment is supported by the observation that the frequency decrease 13 cm⁻¹ of the 930 cm⁻¹ band is about twice as large compared to the frequency decrease 6 cm⁻¹ of the 466 cm⁻¹ band. The band at 1219 cm⁻¹ of the normal species ³PTH^{*} is down-shifted to 1215 cm⁻¹ on deuteration of the N–H group. The 1219 cm⁻¹ band can be assigned to the C–N symmetric stretch.

Resonance Raman spectra of 2-chlorophenothiazine and its ${}^{34}S$ substituted analogue in the T₁ state, i.e. ${}^{3}CPTH^{*}$ and ${}^{3}CPTH^{*}-{}^{34}S$ are shown in Fig. 8. The bands at 454 and 904 cm⁻¹ of the normal species ${}^{3}\text{CPTH}^{*}$ are down-shifted to 449 and 895 cm⁻¹, respectively on the ${}^{34}\text{S}$ substitution. The 454 cm⁻¹ band can be assigned to the C–S symmetric stretch and the 904 cm⁻¹ band to the first overtone of the 454 cm⁻¹ band.

Resonance Raman spectra of the cation radicals of phenothiazine and its isotopically substituted analogues, i.e. PTH⁺, PTH⁺⁺–³⁴S and PTH⁺⁺-d₊ are shown in Fig. 9. It is seen that the bands at 473 and 946 cm⁻¹ of the normal species PTH⁺⁺ are down-shifted to 466 and 932 cm⁻¹, respectively on the ⁻³⁴S substitution. The 7 cm⁻¹ magnitude of the down shift on the ⁻³⁴S substitution indicates that the 473 cm⁻¹ is assignable to the C–S symmetric stretch. The 946 cm⁻¹ band exhibiting the down shift of 14 cm⁻¹ is considered



Fig. 8. Transient resonance Raman spectra of the T_1 state of 2-chlorophenothiazine and its ³⁴S substituted analogue in deoxygenated methanol: (a) ³CPTH^{*} normal species; (b) ³CPTH-³⁴S. Pump wavelength: 308 nm and probe wavelength is 456 nm. Concentration: 2.0×10^{-3} mol dm⁻³.

to arise from the first overtone of the 473 cm⁻¹ band. We note that the C–S symmetric stretch of PTH⁺ is quite harmonic. The band at 1262 cm⁻¹ of PTH⁺ is down-shifted to 1257 cm⁻¹ on the deuteration of the N–H group. This band is assignable to the C–N symmetric stretch.

Resonance Raman spectra of the cation radicals of 2-chlorophenothiazine and its ${}^{34}S$ substituted analogue, i.e. CPTH⁺ and CPTH⁺ - ${}^{34}S$, are shown in Fig. 10. The band at 465 cm⁻¹ is down-shifted to 459 cm⁻¹ on the ${}^{34}S$ substitution. This band can be assigned to the C-S symmetric stretch. The first overtone bands of the 465 and 459 cm⁻¹ bands are not clearly detectable due to overlapping by nearby bands.

3.3. Structures and dynamics of the T_i state and the cation radical

We have shown by time-resolved resonance Raman spectroscopy that in both phenothiazine and 2-chlorophenothiazine the 8a and 8b modes (phenyl ring stretch mode) of the phenyl rings are drastically down-shifted in the T_1 state, whereas they do not exhibit appreciable frequency shifts in the cation radical. In contrast the C–S symmetric stretch is upshifted considerably both in the T_1 state and cation radical. These observations imply that (1) the phenyl rings of both PTH and CPTH are drastically weakened and therefore the C–C bonds of the phenyl rings are lengthened in the T_1 state, but these structural changes do not occur in the cation radical, and (2) the C–S bonds of PTH and CPTH are strengthened and therefore the C–S bonds are shortened both in the T_1 state and in the cation radical.

Since the T_1 state can be represented by the electron configuration in which an electron is elevated from the HOMO (highest occupied molecular orbital) to the LUMO (lowest unoccupied molecular orbital), while the electronic state of the cation radical is most adequately expressed by the electron configuration in which an electron is removed from the HOMO as depicted in Fig. 11, the weakening of the phenyl rings only in the T_1 state and strengthening of the C-S bonds both in the T_1 and in the cation radical indicate that the LUMO is an anti-bonding π -orbital strongly localized on the phenyl rings, and the



Fig. 9. Transient resonance Raman spectra of the cation radical of phenothiazine and its isotopically substituted analogues in methanol: (a) PTH⁺ normal species; (b) PTH⁺ $-^{34}$ S; (c) PTH⁺-d₁. Pump wavelength: 308 nm and probe wavelength is 517 nm. Concentration: 2.0×10^{-3} mol dm⁻³. (For the measurement of PTH⁺-d₁ methanol-d₁ was used to avoid the exchange of N–D deuterium with methanol O–H hydrogen.)

HOMO is a non-bonding orbital strongly localized on the S atom. This implies that the T₁ state is of $n-\pi^*$ character.

The generation of the transient X in the photoreaction of 2-chlorophenothiazine is of interest. Since the corresponding transient is not generated in the photoreaction of phenothiazine, the production of the transient X must be closely related to the chlorine substituent. Chlorpromazine, a clinically important drug for psychotic disorders, has a chlorine substituent at the same position (2-position) of the phenothiazine skeleton and is known to be an order of magnitude more phototoxic as well as photoallergic than promazine which has no chlorine substituent. Therefore, it is not wholly improbable that the corresponding transient X of chlorpromazine is one of the prime cause of phototoxicity and photoallergy of chlorpromazine. The identity of the transient X is not clarified as yet, but it seems probable that it is the phenothiazinyl neutral radical generated by photoinduced dechlorination.

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Fig. 10. Transient resonance Raman spectra of the cation radical of 2-chlorophenothiazine and its 34 S substituted analogue in methanol: (a) CPTH⁺ normal species; (b) CPTH⁺– 34 S. Pump wavelength: 308 nm and probe wavelength is 517 nm. Concentration is 2.0×10^{-3} mol dm⁻³. The asterisk (*) denotes solvent bands.



Fig. 11. Electronic configurations of the T₁ state and cation radical.

performed using the apparatus at the Materials Characterization Central Laboratory, Waseda University.

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