

Conversion of 5-Hydroxymethylfurfural into 6-(Hydroxymethyl)pyridin-3-ol: A Pathway for the Formation of Pyridin-3-ols in Honey and Model Systems

Francisco J. Hidalgo, Cristina M. Lavado-Tena, and Rosario Zamora

J. Agric. Food Chem., **Just Accepted Manuscript** • DOI: 10.1021/acs.jafc.0c01679 • Publication Date (Web): 22 Apr 2020

Downloaded from pubs.acs.org on April 23, 2020

Just Accepted

“Just Accepted” manuscripts have been peer-reviewed and accepted for publication. They are posted online prior to technical editing, formatting for publication and author proofing. The American Chemical Society provides “Just Accepted” as a service to the research community to expedite the dissemination of scientific material as soon as possible after acceptance. “Just Accepted” manuscripts appear in full in PDF format accompanied by an HTML abstract. “Just Accepted” manuscripts have been fully peer reviewed, but should not be considered the official version of record. They are citable by the Digital Object Identifier (DOI®). “Just Accepted” is an optional service offered to authors. Therefore, the “Just Accepted” Web site may not include all articles that will be published in the journal. After a manuscript is technically edited and formatted, it will be removed from the “Just Accepted” Web site and published as an ASAP article. Note that technical editing may introduce minor changes to the manuscript text and/or graphics which could affect content, and all legal disclaimers and ethical guidelines that apply to the journal pertain. ACS cannot be held responsible for errors or consequences arising from the use of information contained in these “Just Accepted” manuscripts.

Conversion of 5-Hydroxymethylfurfural into 6-(Hydroxymethyl)pyridin-3-ol: A Pathway for the Formation of Pyridin-3-ols in Honey and Model Systems

Francisco J. Hidalgo, Cristina M. Lavado-Tena, and Rosario Zamora*

Instituto de la Grasa, Consejo Superior de Investigaciones Científicas, Carretera de Utrera km 1, Campus Universitario–Edificio 46, 41013-Seville, Spain

*Corresponding author: Rosario Zamora

Phone: +34954611550

Fax: +34954616790

e-mail: rzamora@ig.csic.es

1 ABSTRACT

2 The formation of 6-(hydroxymethyl)pyridin-3-ol by ring expansion of 5-
3 (hydroxymethyl)furfural (HMF) in the presence of ammonia-producing compounds was
4 studied to determine routes of formation of pyridin-3-ols in foods. 6-
5 (Hydroxymethyl)pyridin-3-ol was produced from HMF in model systems, mostly at
6 neutral pH values, as a function of reaction times and temperature, and with an activation
7 energy (E_a) of 74 ± 3 kJ/mol, which was higher than that of HMF disappearance (43 ± 4
8 kJ/mol). A reaction pathway is proposed, which is general for the formation of pyridin-
9 3-ols from 2-oxofurans. Thus, it explains the conversions of furfural into pyridin-3-ol and
10 of 2-acetylfuran into 2-methylpyridin-3-ol, which were also studied. When honey and
11 sugarcane honey were heated, they produced different pyridin-3-ols, although 6-
12 (hydroxymethyl)pyridin-3-ol was the pyridine-3-ol produced to the highest extent.
13 Obtained results suggest that formation of pyridin-3-ols in foods is unavoidable when 2-
14 oxofurans are submitted to thermal heating and ammonia (or ammonia-producing
15 compounds) is present.

16

17 **KEYWORDS:** *2-Acetylfuran; Carbonyl-amine reactions; Furfural; Honey; 5-*
18 *Hydroxymethylfurfural; Maillard reaction; Pyridin-3-ols; Reactive carbonyls*

19

20 INTRODUCTION

21 Maillard reaction is a major route for the formation of aromas as a consequence of
22 food processing.¹⁻³ The compounds responsible for these aromas have a great diversity
23 of structures. Thus, the formation of aldehydes, alcohols, thiols, sulfides, acids, lactones,
24 and a large variety of heterocyclic compounds, has been described.⁴ Among these last
25 compounds, the stale flavors of furan-2-carbaldehydes,⁵ or the roasted flavors of
26 pyrazines,⁶ for example, have been widely studied. However, some of the compounds
27 responsible for the produced aromas are not always stable and aroma changes are usually
28 observed with further processing or storage. Many of these changes are still very poorly
29 understood, which difficulties the prediction of the evolution of food aromas.

30 Among the different flavor-related substances produced in foods as a consequence of
31 Maillard reaction, 5-hydroxymethylfurfural (HMF) is an important compound commonly
32 produced to significant extents. Its flavor has been described as caramel, waxy, fatty,
33 musty, or cardboard. HMF is considered as an endogenous contaminant because of its
34 potential adverse effects,⁷ and it is also a quality indicator as its levels in foods provide
35 evidence of overheating during processing or inadequate storage conditions.⁸ In addition,
36 it has also been associated with color development in some foods.⁹ This compound has
37 been shown to be thermally unstable. Thus, its conversion upon heating into 5,5'-oxy-
38 dimethylene-*bis*(2-furaldehyde), 5-methylfurfural, 2,5-furandicarboxaldehyde, or 2-
39 acetyl-5-methylfuran, among others, has been described.^{10,11} In addition, diverse
40 chemical syntheses suggest that conversion of HMF into 6-(hydroxymethyl)pyridin-3-ol
41 is possible. To this respect, Müller *et al.*¹² showed that brief exposure to bromine in water-
42 methanol at 0 °C smoothly and effectively converts hydroxymethylfurfurylamines,
43 originated from either fructose or inuline sources, into 6-substituted pyridin-3-ols.
44 Furthermore, formation of pyridin-3-ols by ring expansion 2-acylfurans is well-known in

45 organic synthesis.^{13,14} Nevertheless, to the best of our knowledge, a relationship between
46 pyridin-3-ols, like 6-(hydroxymethyl)pyridin-3-ol, and 2-oxofurans, like HMF, has not
47 been established so far in foods. However, 6-(hydroxymethyl)pyridin-3-ol, like other
48 pyridin-3-ols, has been found in the hydrolysis of melanoidins¹⁵ or as a product of
49 Maillard reaction.¹⁶

50 Pyridines are typical food flavors, which are produced as a consequence of
51 processing.¹⁷ The formation pathways of alkylpyridines seem to be related to the
52 cyclizations and oligomerizations suffered by lipid-derived aldehydes in the presence of
53 ammonia-producing compounds.¹⁸ In addition, formation of alkylpyridiniums has been
54 described in roasted coffee by trigonelline decomposition.¹⁹ On the other hand, the
55 formation pathways of pyridin-3-ols are lesser known in spite of being pyridin-3-ol a
56 common flavor in coffee,²⁰ tea,²¹ or caramel colors,²² for example. In an attempt to
57 explore the role of 2-oxofurans in the appearance of pyridin-3-ols in foods, this study
58 investigates the formation of 6-(hydroxymethyl)pyridin-3-ol, pyridin-3-ol, and 2-
59 methylpyridin-3-ol in honeys and model systems.

60 MATERIALS AND METHODS

61 **Materials and chemicals.** As model food products, two kinds of honeys were studied.
62 One of them was a high quality multi-floral honey. The other one was a traditional plant
63 syrup, namely ‘miel de caña’ (sugarcane honey or sugarcane syrup). This syrup is
64 traditionally manufactured in Andalusia (Spain) from crushed sugarcane (*Saccharum*
65 *officinarum*) and, although produced only in limited quantities, it is highly appreciated by
66 consumers. For its preparation, the fresh cane juice is heated rapidly for decantation, then
67 filtered, and, finally, slowly heated until it looks like dark honey. A characterization of
68 this syrup was described previously.²³ Both products were purchased at local
69 supermarkets.

70 The tested pyridin-3-ols were pyridin-3-ol, 2-methylpyridin-3-ol, and 6-
71 (hydroxymethyl)pyridin-3-ol. As 2-oxofurans, HMF, furfural, and 2-acetylfuran were
72 employed. These and other chemicals employed in this study had the highest available
73 grade and were purchased from reliable commercial sources, including Sigma-Aldrich
74 (St. Louis, MO), Fluka (Buchs, Switzerland), and Alfa Aesar (Thermo Fischer GmbH,
75 Karlsruhe, Germany).

76 **Conversion of 2-Oxofurans into Pyridin-3-ols in Model Systems.** Mixtures of the
77 2-oxofuran (10 μmol dissolved in 20 μL of methanol) and an ammonia-producing
78 compound (30 μmol in 50 μL of water) were singly homogenized with 200 mg of 0.063–
79 0.200 mm silica gel (Macherey-Nagel, Düren, Germany), and 30 μL of 0.3 M buffer, pH
80 3–10. Samples were heated in closed test tubes for the indicated times (0–22 h) and
81 temperatures (100–180 $^{\circ}\text{C}$). The employed buffers were sodium citrate, pH 3–6, sodium
82 phosphate, pH 6–8, and sodium borate, pH 8–10. Times and temperatures employed in
83 each experience are indicated in the respective figure legends.

84 At the end of the heating period, samples were cooled and 1 mL of methanol and 20
85 μL of the internal standard solution (13 mg of 1-octadecanol in 25 mL of methanol) were
86 added. Suspensions were stirred for 1 min and centrifuged for 5 min at 2000 g . The
87 supernatant was collected, evaporated to dryness, and the residue was successively treated
88 with 200 μL of *N,O*-bis(trimethylsilyl)trifluoroacetamide (BSTFA), heated for 30 min at
89 60 $^{\circ}\text{C}$, and studied by gas chromatography coupled to mass spectrometry (GC-MS).

90 The 2-oxofurans tested were HMF, furfural, and 2-acetylfuran. The assayed ammonia-
91 producing compounds were ammonia and ammonium chloride.

92 **Formation of Pyridin-3-ols in Honeys.** Honeys (1 g) were heated in closed test tubes
93 for 0–10 h at either 60 or 100 $^{\circ}\text{C}$. After cooling, 100 μL of water was added and the

94 mixture was stirred. Then, 4 mL of acetonitrile and 20 μ L of the internal standard solution
95 (13 mg of 1-octadecanol in 25 mL of methanol) were added. Mixtures were successively
96 stirred for 1 min, sonicated for 10 min, stirred for 1 min, and the acetonitrile layer was
97 decanted and taken to dryness. The residue was treated with 300 μ L of *N,O*-
98 *bis*(trimethylsilyl)trifluoroacetamide (BSTFA), heated for 30 min at 60 $^{\circ}$ C, and studied
99 by gas chromatography coupled to mass spectrometry (GC-MS).

100 **GC-MS Analyses.** GC-MS analyses were conducted with an Agilent 7820A gas
101 chromatograph coupled with an Agilent 5977 mass selective detector (quadrupole type)
102 using a fused-silica HP-5MS UI capillary column (30 m length, 0.25 mm inner diameter,
103 0.25 μ m coating thickness) from Agilent. One microliter of sample was injected in the
104 pulsed splitless mode. Working conditions were as follows: carrier gas, helium (1 mL/min
105 at constant flow); injector, 250 $^{\circ}$ C; transfer line to mass selective detector, 280 $^{\circ}$ C;
106 electron ionization (EI), 70 eV; ion source temperature, 230 $^{\circ}$ C; and mass range, 28-550
107 amu. Oven temperature conditions were from 80 $^{\circ}$ C (1 min) to 140 $^{\circ}$ C at 20 $^{\circ}$ C/min, then
108 to 300 $^{\circ}$ C at 50 $^{\circ}$ C/min, and finally held at 300 $^{\circ}$ C for 4 min.

109 **Identification of 6-(Hydroxymethyl)pyridin-3-ol, Pyridin-3-ol, 2-Methylpyridin-**
110 **3-ol and HMF.** Identification of the three pyridin-3-ols and HMF was carried out by
111 comparison of retention indexes and mass spectra, and by co-elution with authentic
112 standards, which were derivatized analogously. Mass spectra of the assayed compounds
113 as well as that of the internal standard are collected in the Supporting Information. The
114 following ions (M^{+} – methyl) were employed for quantitation purposes: m/z 254 for 6-
115 (hydroxymethyl)pyridin-3-ol, m/z 152 for pyridin-3-ol, m/z 166 for 2-methylpyridin-3-ol,
116 m/z 183 for HMF, and m/z 327 for the IS.

117 **Quantitation of 6-(Hydroxymethyl)pyridin-3-ol, Pyridin-3-ol, 2-Methylpyridin-**
118 **3-ol and HMF.** Quantitation of the three pyridin-3-ols and HMF in model systems and

119 honeys was carried out by preparing standard curves of those compounds in the
120 corresponding assayed systems and following the whole procedures described above
121 (without heating). For each curve, six different concentration levels of the three pyridin-
122 3-ols and HMF were used. Pyridin-3-ols and HMF contents were directly proportional to
123 the corresponding compound/internal standard area ratios ($r > 0.99$, $p < 0.001$). The
124 coefficients of variation at the different assayed concentrations were $< 10\%$.

125 **Statistical Analysis.** All data given are mean \pm standard deviation (SD) of at least
126 three independent experiments. Analysis of variance was employed to compare the
127 different groups. When significant F values were obtained, group differences were
128 evaluated by the Tukey test.²⁴ These studies were conducted using Origin version 7.0
129 (OriginLab Corp., Northampton, MA, USA). The significance level is $p < 0.05$ unless
130 otherwise indicated.

131 RESULTS AND DISCUSSION

132 **Effect of Reaction Conditions on the Conversion of HMF into 6-**
133 **(Hydroxymethyl)pyridin-3-ol.** When HMF was heated in the presence of either
134 ammonia or ammonium chloride, the formation of 6-(hydroxymethyl)pyridin-3-ol was
135 observed. The conversion yield depended on the reaction conditions, including reaction
136 pH, reactant concentrations, time and temperature.

137 Figure 1 shows the effect of pH on both the formation of 6-(hydroxymethyl)pyridin-
138 3-ol and the disappearance of HMF. As observed, HMF disappearance and 6-
139 (hydroxymethyl)pyridin-3-ol formation increased when the pH increased from 3 to 7. The
140 maximum amount of 6-(hydroxymethyl)pyridin-3-ol was produced at pH 6–8 with
141 sodium phosphate buffer. At higher pH values and in the presence of borate buffer, the
142 amount of the produced 6-(hydroxymethyl)pyridin-3-ol was much lower. For some

143 unknown reason, HMF disappearance also occurred at pH 8–10 in the presence of sodium
144 borate buffer, but this disappearance did not produce the pyridine to a high extent.
145 Between pH 3 and pH 8 the formation of 6-(hydroxymethyl)pyridin-3-ol and the
146 disappearance of HMF were correlated ($r = 0.96$, $p = 0.002$). The conversion yield of the
147 furan into the pyridine depended on the pH and ranged from 0.4% at pH 3 to 1.3% at pH
148 7–8. Although there was not a significant difference among the amounts of 6-
149 (hydroxymethyl)pyridin-3-ol produced at either pH 7 or pH 8, pH 8 was selected for the
150 rest of this study because the minimum amount of HMF was recovered at this pH.

151 6-(Hydroxymethyl)pyridin-3-ol formation also depended on the HMF/ammonia ratio.
152 Figure 2 shows the effect of increasing amounts of HMF on the formation of 6-
153 (hydroxymethyl)pyridin-3-ol in the presence of ammonium chloride. The maximum
154 amount of the pyridine was achieved when 10 μmol of HMF were heated in the presence
155 of 30 μmol of ammonium chloride. Nevertheless, the conversion yield was higher when
156 smaller amounts of HMF were employed. Thus, conversion yield was 3% with 2.5 μmol
157 of HMF, and this yield decreased when higher amounts of HMF were employed. Most of
158 this study was carried out with 10 μmol of HMF because the highest amount of 6-
159 (hydroxymethyl)pyridin-3-ol was obtained by using these conditions.

160 Conversion yield also depended on the concentration of the ammonia-producing
161 compound. Figure 3 shows the effect of increasing amounts of ammonia and ammonium
162 chloride on the formation of 6-(hydroxymethyl)pyridin-3-ol. The amount of this pyridine
163 increased linearly ($r > 0.9993$, $p < 0.0007$) between 0 and 20 μmol of the amino compound
164 and it remained constant when more than 30 μmol of the amino compound was added.
165 The behavior was inverse for the HMF recovered. It decreased linearly ($r = 0.996$, $p <$
166 0.004) between 0 and 20 μmol of the amino compound and it remained constant when

167 more than 30 μmol of the amino compound was added. In fact, formation of 6-
168 (hydroxymethyl)pyridin-3-ol and disappearance of HMF were correlated ($r = 0.98$, $p <$
169 0.0001). Moreover, the behavior of the two amino compounds assayed (ammonia and
170 ammonium chloride) was very similar. Both of them produced analogous disappearance
171 of HMF, although the conversion yield for 6-(hydroxymethyl)pyridin-3-ol seemed to be
172 slightly higher when ammonium chloride was employed. For that reason, ammonium
173 chloride was employed as ammonia-producing compound in most of the experiments
174 carried out in this study. In addition, 30 μmol of ammonium chloride was selected because
175 this was the minimum amount of the ammonia-producing compound that produced the
176 pyridine to a higher extent when starting from 10 μmol of HMF.

177 Finally, time and temperature also played a major role on the conversion of HMF into
178 6-(hydroxymethyl)pyridin-3-ol. Figure 4 shows the time-courses for the formation of 6-
179 (hydroxymethyl)pyridin-3-ol at 100–160 $^{\circ}\text{C}$. As observed, the amount of the pyridine
180 increased linearly ($r = 0.91$, $p < 0.04$) as a function of heating time. Furthermore,
181 formation rates, obtained from the slopes of the lines of the best fit, increased with
182 temperature. In addition, there was a small lag period, which was higher when
183 temperature was lower. Reaction rates for this formation were employed in an Arrhenius
184 plot to obtain the activation energy (E_a) for the formation of 6-(hydroxymethyl)pyridin-
185 3-ol from HMF. The obtained plot is shown in Figure 5. The activation energy was
186 obtained from the slope of the line of best fit and resulted to be 74 ± 3 kJ/mol.

187 Similarly, time-courses for the disappearance of HMF at 100–160 $^{\circ}\text{C}$ are shown in
188 Figure 6. Recovered HMF decreased linearly ($r > 0.99$, $p < 0.04$) as a function of heating
189 times and disappearance rates increased with temperature. Disappearance rates were
190 employed in an Arrhenius plot for determining the E_a for HMF disappearance. It was 43

191 ± 4 kJ/mol. This value was lower than that found above for 6-(hydroxymethyl)pyridin-3-
192 ol formation. This explains that HMF is easily decomposed, as observed previously,¹⁰ and
193 it is only partially converted into 6-(hydroxymethyl)pyridin-3-ol.

194 **Conversion of 2-Oxofurans into Pyridin-3-ols.** The conversion of HMF into the
195 corresponding pyridin-3-ol is not exclusive for this furan. To confirm that other 2-
196 oxofurans can suffer analogous transformations, the conversion of other two products of
197 the Maillard reaction (furfural²⁵ and 2-acetylfuran²⁶) into the corresponding pyridin-3-ols
198 (pyridin-3-ol and 2-methylpyridin-3-ol, respectively) was studied. Figure 7 shows the
199 formation of the corresponding pyridin-3-ols after 3 and 22 h at 100 °C. The figure also
200 shows the formation of 6-(hydroxymethyl)pyridin-3-ol by heating of HMF under the
201 same reaction conditions for comparison purposes. As expected, the corresponding
202 pyridin-3-ols were always produced, although the conversion yield depended on the
203 employed 2-oxofuran. Thus, the highest yield was observed for 6-
204 (hydroxymethyl)pyridin-3-ol. This pyridine was produced with a yield of 0.5% after 3 h
205 at 100 °C, and the yield increased to 1.3% after 22 h at 100 °C. The yields of the other
206 two pyridines were similar among them and were lower than those of 6-
207 (hydroxymethyl)pyridin-3-ol. The pyridine that was produced to a higher extent was
208 pyridin-3-ol (0.03% and 0.11% after 3 and 22 h, respectively). This pyridine has been
209 found in different foods that had suffered Maillard reaction.^{16–18} The yield for 2-
210 methylpyridin-3-ol was 0.01% and 0.06% after 3 and 22 h, respectively.

211 **Proposed Pathway for the Conversion of 2-Oxofurans into Pyridin-3-ols.** Above
212 described results suggest that, in the presence of ammonia and moderate temperatures, 2-
213 oxofurans are decomposed and the formation of pyridin-3-ols is produced. A pathway
214 that explains this transformation is shown in Figure 8. This reaction pathway is based on
215 that previously published by Gruber.¹⁴ Thus, in the presence of ammonia, the

216 corresponding imine should be produced in a first step. Then, the addition of a second
217 molecule of ammonia would produce the opening of the ring with the formation of a
218 compound derived from the dicarbonyl precursor of the furan. This attack is consequence
219 of the polarization of the C–O bond in the furan ring because of the difference of
220 electronegativities between both atoms.²⁷ Finally, cyclization of this intermediate with
221 the exit of a molecule of ammonia would produce the corresponding pyridine derivative.

222 This reaction only requires the presence of an oxo group at position 2 of the furan ring.
223 This means that it should be produced with any other 2-oxofuran that might be present.
224 This explains, for example, the presence of both 6-methylpyridin-3-ol and 5-
225 methylfurfural in coffee aroma.²⁸ In addition, it could also be expected to be produced in
226 analogous 2-oxoheterocycles. For example, 2-acetylthiophene is a well-known Maillard
227 reaction product.²⁹ According to the proposed reaction pathway, it should be converted
228 to a certain extent into 2-methylpyridin-3-thiol. To the best of our knowledge, this flavor
229 compound has not been yet identified in foods. However, other Maillard reaction product,
230 2-thiophenecarboxaldehyde, which is present in coffee³⁰ or pork broth,³¹ for example,
231 should produce pyridin-3-thiol. This last pyridine has been identified in soy sauce, which
232 is also a source of 2-acetylthiophene.³²

233 Although the presence of the oxo group is a requisite for the reaction, reaction yield
234 also depended on the existence of substituents in the carbon at position 5 of the furan ring.
235 The hydroxyl group of the hydroxymethyl substituent at position 5 of HMF has a negative
236 inductive effect. This effect should contribute to increase the electron deficiency of
237 carbon at position 5 of the furan ring, therefore converting this atom into a better
238 electrophile. For this reason, HMF is converted more easily into 6-
239 (hydroxymethyl)pyridin-3-ol than furfural into pyridin-3-ol. This suggests that, although
240 pyridin-3-ol is more frequently described than 6-(hydroxymethyl)pyridin-3-ol in most

241 foods, this last pyridine is likely to be present to a higher extent, but it has mostly gone
242 unnoticed until now. To confirm this last hypothesis, and to test the formation of pyridin-
243 3-ols in food products, the effect of thermal heating on commercial honeys was studied.
244 Obtained results will be discussed in the next subsection.

245 **Formation of Pyridin-3-ols in Honeys.** Two different types of honeys were studied.
246 One of them was a high quality honey, which had not suffered any previous thermal
247 overheating. The other one was a sugarcane honey. As described in the Materials and
248 Methods section, the production of this last honey requires thermal treatment that is likely
249 to promote Maillard reaction to a high extent.

250 Figure 9 shows the effect of heating on the formation of both HMF and 6-
251 (hydroxymethyl)pyridin-3-ol in the studied honey. As expected, because of its high
252 quality, very small amounts of HMF and 6-(hydroxymethyl)pyridin-3-ol were detected in
253 this honey at the initial step. However, when the honey was heated, both compounds were
254 produced to a significant extent. On the other hand, formation of pyridin-3-ol was not
255 observed, and only trace amounts of 2-methylpyridin-3-ol were detected when the honey
256 was heated at 100 °C for long heating periods. This indicated that, similarly to that
257 observed in model systems, 6-(hydroxymethyl)pyridin-3-ol is produced to a higher extent
258 than other pyridines. In addition, HMF and 6-(hydroxymethyl)pyridin-3-ol formation
259 time-courses were different. Thus, HMF increased exponentially as a function of heating
260 time, as observed previously.³³ On the other hand, 6-(hydroxymethyl)pyridin-3-ol
261 concentration increased linearly ($r = 0.94$, $p < 0.0002$). This difference of time-courses
262 does not necessarily mean that 6-(hydroxymethyl)pyridin-3-ol is not produced by ring
263 expansion of HMF. As observed in the studied model systems (Figure 2), increased
264 amounts of HMF do not always produce increasing amounts of 6-
265 (hydroxymethyl)pyridin-3-ol. This conversion requires the presence of ammonia (or other

266 ammonia-producing compound) that can be the limiting factor in this conversion. In
267 addition, and analogously to that observed in model systems, HMF disappearance should
268 produce other compounds in addition to 6-(hydroxymethyl)pyridin-3-ol.

269 Results obtained with the sugarcane honey studied were somewhat different. Because
270 this food product suffers a heating process during its production, both HMF and 6-
271 (hydroxymethyl)pyridin-3-ol were already present in the initial sample. In addition, both
272 of them increased linearly ($r > 0.93$, $p < 0.0008$) when the sugarcane honey was heated.
273 In fact, there was a correlation between them at both 60 °C ($r > 0.97$, $p = 0.005$) and 100
274 °C ($r > 0.97$, $p < 0.0001$). Surprisingly, 6-(hydroxymethyl)pyridin-3-ol/HMF ratio was
275 higher at 60 °C (~0.27) than at 100 °C (~0.034). This suggests that disappearance of HMF
276 at low temperature mainly produces the pyridine. However, higher temperatures favor the
277 transformation of HMF into other compounds.

278 Differently to the studied honey, the studied sugarcane honey also contained the other
279 analyzed pyridines (pyridin-3-ol and 2-methylpyridin-3-ol) at the initial step. In addition,
280 concentrations of both pyridines increased linearly ($r > 0.90$, $p < 0.002$) as a function of
281 heating. However, and analogously to the observed in the studied model systems and the
282 studied honey, the concentrations of both pyridines were always lower than that of 6-
283 (hydroxymethyl)pyridin-3-ol.

284 Obtained results suggest that, analogously to that observed in honey, sugarcane honey,
285 and model systems, conversion of 2-oxofurans into pyridin-3-ols is unavoidable when 2-
286 oxofurans are submitted to thermal heating and ammonia (or an ammonia-producing
287 compound) is present. This suggests that, although not frequently found in foods, pyridin-
288 3-ols should be common components of foods in which Maillard reaction has occurred,
289 even in those foods submitted to soft heating or stored for a limited time period.

290 Obtained results also show that 6-(hydroxymethyl)pyridin-3-ol is an additional,
291 previously unknown in foods, transformation product of HMF. Some authors have shown
292 doubts about the usefulness of using HMF as a quality indicator of inadequately processed
293 foods because of its instability,⁹ although in many food systems, after a short lag time, a
294 constant increase in HMF concentration is observed, which indicates a faster formation
295 than degradation. Nevertheless, because aromaticity of pyridines is usually higher than
296 that of furans,³⁵ 6-(hydroxymethyl)pyridin-3-ol is expected to be more stable than HMF.
297 Therefore, although reaction yield of 6-(hydroxymethyl)pyridin-3-ol is much lower than
298 that of HMF, the potential use of 6-(hydroxymethyl)pyridin-3-ol as an alternative marker
299 of thermal stress in some foods might be suggested.

300 Finally, a last consequence of the described reaction is related to the observed changes
301 in the antioxidant activity of honeys submitted to thermal heating.^{34,35} As described
302 above, 6-(hydroxymethyl)pyridin-3-ol increases upon heating, and pyridin-3-ols have
303 been shown to be efficient chain-breaking antioxidants.³⁶ The potential contribution of 6-
304 (hydroxymethyl)pyridin-3-ol to the observed increase of antioxidant activity in honey
305 might be hypothesized.

306 **ASSOCIATED CONTENT**

307 **Supporting Information**

308 The Supporting Information is available free of charge at

309 Mass spectra of trimethylsilyl derivatives of pyridin-3-ol, 2-methylpyridin-3-ol, 6-
310 (hydroxymethyl)pyridin-3-ol, 5-hydroxymethylfurfural (HMF), and 1-octadecanol (PDF)

311 **AUTHOR INFORMATION**

312 **Corresponding author**

313 *Telephone: +34 954 611 550. Fax: +34 954 616 790. E-mail: rzamora@ig.csic.es.

314 **Funding**

315 This study was supported by the Ministerio de Ciencia, Innovación y Universidades
316 (MCIU) from Spain, the Agencia Estatal de Investigación (AEI) from Spain, and the
317 Fondo Europeo de Desarrollo Regional (FEDER) from the European Union (Project
318 RTI2018-096632-B-100).

319 **Notes**

320 The authors declare no competing financial interest.

321

322 **REFERENCES**

- 323 (1) McGorin R. J. Key aroma compounds in oats and oat cereals. *J. Agric. Food Chem.*
324 **2019**, *67*, 13778–13789.
- 325 (2) Poisson, L.; Schaerer, A.; Spreng, S.; Mestdagh, F.; Blank, I.; Davidek, T.
326 Generation of α -diketones and 4-hydroxy-2,5-dimethyl-3(2H)-furanone upon
327 coffee roasting—Impact of roast degree on reaction pathways. *J. Agric. Food Chem.*
328 **2019**, *67*, 13829–13839.
- 329 (3) Brehm, L.; Frank, O.; Junger, M.; Wimmer, M.; Ranner, J., Hofmann, T. Novel
330 taste-enhancing 4-amino-2-methyl-5-heteroalkylpyridines formed from thiamine
331 by Maillard-type reactions. *J. Agric. Food Chem.* **2019**, *67*, 13986–13997.
- 332 (4) Starowicz, M.; Zielinski, H. How Maillard reaction influences sensorial properties
333 (color, flavor and texture) of food products? *Food Rev. Int.* **2019**, *35*, 707–725.
- 334 (5) Vrzal, T.; Sterba, K.; Jurková, M.; Olsovska, J. The usage of a reflectometric
335 method for 5-(hydroxymethyl)furan-2-carbaldehyde determination as a stale flavor
336 sensor for beer. *Food Packag. Shelf Life* **2019**, *19*, 1–6.
- 337 (6) Liu, C. J.; Yang, Q.; Linforth, R.; Fisk, I. D.; Yang, N. Modifying Robusta coffee
338 aroma by green bean chemical pre-treatment. *Food Chem.* **2019**, *272*, 251–257.
- 339 (7) Zeng, R.; Zhang, G.; Zheng, J.; Zhou, H.; Wang, Y.; Huang, C.; Hu, W.; Ou, S.
340 Formation and identification of two hydroxymethylfurfural-glycine adducts and
341 their cytotoxicity and absorption in Caco-2 cells. *J. Agric. Food Chem.* **2020**, *68*,
342 384–389.
- 343 (8) Lee, C.-H.; Chen, K.-T.; Lin, J.-A.; Chen, Y.-T.; Chen, Y.-A.; Wu, J.-T.; Hsieh,
344 C.-W. Recent advances in processing technology to reduce 5-
345 hydroxymethylfurfural in foods. *Trends Food Sci. Technol.* **2019**, *93*, 271–280.

- 346 (9) Francisquini, J. D.; Rocha, J.; Martins, E.; Stephani, R.; da Silva, P. H. F.; Renhe,
347 I. R. T.; Perrone, I. T.; de Carvalho, A. F. 5-Hydroxymethylfurfural formation and
348 color change in lactose-hydrolyzed Dulce de leche. *J. Dairy Res.* **2019**, 477–482.
- 349 (10) Chambel, P., Oliveira, M. B., Andrade, P. B., Fernandes, J. O., Seabra, R. M.,
350 Ferreira, M. A. Identification of 5,5'-oxy-dimethylene-bis(2-furaldehyde) by
351 thermal decomposition of 5-hydroxymethyl-2-furfuraldehyde. *Food Chem.* **1998**,
352 *63*, 473–477.
- 353 (11) Nikolov, P. Y.; Yayalayan, V. A. Thermal decomposition of 5-(hydroxymethyl)-2-
354 furaldehyde (HMF) and its further transformations in the presence of glycine. *J.*
355 *Agric. Food Chem.* **2011**, *59*, 10104–10113.
- 356 (12) Müller, C.; Diehl, V.; Lichtenthaler, F. W. Building blocks from sugars. Part 23.
357 Hydrophilic-3-pyridinols from fructose and isomaltulose. *Tetrahedron* **1998**, *54*,
358 10703–10712.
- 359 (13) Leditschke, H. die Synthese von 3-Oxyl-2-aryl-pyridinen aus Phenyl- α -furyl-
360 ketonen. *Chem. Ber* **1952**, *85*, 202–204.
- 361 (14) Gruber, W. Synthesis of 3-hydroxy-2-alkylpyridines. *Can. J. Chem.* **1953**, *31*, 564–
362 568.
- 363 (15) Tsuchida, H.; Komoto, M.; Kato, H.; Fujimaki, M. Isolation and identification of
364 two β -hydroxy-pyridine derivatives from the nondialyzable melanoidin-
365 hydrolyzate. *Agric. Biol. Chem.* **1973**, *37*, 403–409.
- 366 (16) Olsson, K.; Pernemalm, P.-A.; Theander, O. Formation of aromatic compounds
367 from carbohydrates. VII. Reaction of D-glucose and glycine in slightly acidic,
368 aqueous solution. *Acta Chem. Scand. B* **1978**, *32*, 249–256.
- 369 (17) Maga, J. A. Pyridines in foods. *J. Agric. Food Chem.* **1981**, *29*, 895–898.

- 370 (18) Zamora, R.; Lavado-Tena, C. M.; Hidalgo, F. J. Oligomerization of reactive
371 carbonyls in the presence of ammonia-producing compounds: A route for the
372 production of pyridines in foods. *Food Chem.* **2020**, *304*, 125284.
- 373 (19) Stadler, R. H.; Varga, N.; Milo, C.; Schilter, B.; Vera, F. A.; Welti, D. H.
374 Alkylpyridiniums. 2. Isolation and quantification in roasted and ground coffees. *J.*
375 *Agric. Food Chem.* **2002**, *50*, 1200–1206.
- 376 (20) Moon, J.-K.; Shibamoto, T. Role of roasting conditions in the profile of volatile
377 flavor chemicals formed from coffee beans. *J. Agric. Food Chem.* **2009**, *57*, 5823–
378 5831.
- 379 (21) Kuo, P. C.; Lay, Y. Y.; Chen, Y. J.; Yang, W. H.; Tzen, J. T. Changes in volatile
380 compounds upon aging and drying in oolong tea production. *J. Sci. Food Agric.*
381 **2011**, *91*, 293–301.
- 382 (22) Myers, D. V.; Howell, J. C. Characterization and specifications of caramel colours:
383 an overview. *Food Chem. Toxicol.* **1992**, *30*, 359–363.
- 384 (23) Ruiz-Matute, A. I.; Soria, A. C.; Sanz, M. L.; Martínez-Castro, I. Characterization
385 of traditional Spanish edible plant syrups based on carbohydrate GC-MS analysis.
386 *J. Food Comp. Anal.* **2010**, *23*, 260–263.
- 387 (24) Snedecor, G. W.; Cochran, W. G. *Statistical Methods*, 7th ed.; Iowa State
388 University Press: Ames, IA, 1980.
- 389 (25) Lu, S. Y.; Cui, H. P.; Zhan, H.; Hayat, K.; Jia, C. S. Hussain, S.; Tahir, M. U.;
390 Zhang, X. M.; Ho, C. T. Timely addition of glutathione for its interaction with
391 deoxypentosone to inhibit the aqueous Maillard reaction and browning of
392 glycyglycine-arabinose system. *J. Agric. Food Chem.* **2019**, *67*, 6586–6593.

- 393 (26) Kanzler, C.; Schestkova, H.; Haase, P. T.; Kroh, L. W. Formation of reactive
394 intermediates, color, and antioxidant activity in the Maillard reaction of maltose in
395 comparison to D-glucose. *J. Agric. Food Chem.* **2017**, *65*, 8957–8965.
- 396 (27) Adam, W.; Grimison, A. Sigma-polarization in 5-membered heterocyclic ring
397 systems. *Theoret. Chim. Acta* **1967**, *7*, 342–351.
- 398 (28) Dong, W.; Hu, R.; Long, Y.; Li, H.; Zhang, Y.; Zhu, K.; Chu, Z. Comparative
399 evaluation of the volatile profiles and taste properties of roasted coffee beans as
400 affected by drying method and detected by electronic nose, electronic tongue, and
401 HS-SPME-GC-MS. *Food Chem.* **2019**, *272*, 723–731.
- 402 (29) Begum, N.; Raza, A.; Song, H. L.; Zhang, Y.; Zhang, L.; Liu, P.. Effect of thermal
403 treatment on aroma generation from bovine bone marrow extract during enzymatic
404 hydrolysis. *J. Food Process. Preserv.* **2019**, *43*, e14105.
- 405 (30) Pua, A.; Lau, H.; Liu, S. Q.; Tan, L. P.; Goh, R. M. V.; Lassabliere, B.; Leong, K.-
406 C.; Sun, J.; Cornuz, M.; Yu, B.. Improved detection of key odourants in Arabica
407 coffee using gas chromatography-olfactometry in combination with low energy
408 electron ionisation gas chromatography-quadrupole time-of-flight mass
409 spectrometry. *Food Chem.* **2020**, *302*, 125370.
- 410 (31) Wang, Y.; Song, H.; Zhang, Y.; Tang, J.; Yu, D. Determination of aroma
411 compounds in pork broth produced by different processing methods. *Flavour Frag.*
412 *J.* **2016**, *31*, 319–328.
- 413 (32) Kim, H.-J.; Lee, E.-J.; Shin, O.-S.; Ji, W.-D; Choi, M.-R.; Kim, J.-K. Volatile
414 components in the soy sauce manufactured by bacillus species and fused yeast. *J.*
415 *Microbiol. Biotechnol.* **1996**, *6*, 194–201.
- 416 (33) Chua, L. S. The extent of hydroxymethylfurfural formation in honey by heating
417 temperature and duration. *Lett. Org. Chem.* **2018**, *15*, 233–240.

- 418 (34) Braghini, F.; Biluca, F. C.; Gonzaga, L. V.; Kracik, A. S.; Vieira, C. R. W.; Vitali,
419 L.; Micke, G. A.; Costa, A. C. O.; Fett, R. Impact of short-term thermal treatment
420 on stingless bee honey (Meliponinae): Quality, phenolic compounds and
421 antioxidant capacity. *J. Food Process. Preserv.* **2019**, *43*, 13954.
- 422 (35) Molaveisi, M. ; Beigbabaie, A. ; Akbari, E.; Noghabi, M. S. ; Mohamadi, M.
423 Kinetics of temperature effect on antioxidant activity, phenolic compounds and
424 color of Iranian jujube honey. *Heliyon* **2019**, *5*, 01129.
- 425 (36) Kumar, S.; Johansson, H.; Kanda, T.; Engman, L.; Mueller, T.; Jonsson, M.;
426 Pedulli, G. F.; Petrucci, S.; Valgimigli, L. Catalytic chain-breaking pyridinol
427 antioxidants. *Org. Lett.* **2008**, *10*, 4895–4898.
- 428
- 429

FIGURE CAPTIONS

Figure 1. Effect of pH on the formation of 6-(hydroxymethyl)pyridin-3-ol (circles) and on the disappearance of 5-hydroxymethylfurfural (HMF, up pointing triangles). HMF (10 μmol) was heated in the presence of ammonium chloride (30 μmol) and 30 μL 0.3 M buffer for 22 h at 100 $^{\circ}\text{C}$. Three buffers were assayed: sodium citrate buffer (open symbols), sodium phosphate buffer (half-closed symbols), and sodium borate buffer (closed symbols). Data are given in nmol of the produced 6-(hydroxymethyl)pyridin-3-ol per μmol of the amount of HMF added to the reaction mixture.

Figure 2. Effect of 5-hydroxymethylfurfural (HMF, Δ) concentration on the formation of 6-(hydroxymethyl)pyridin-3-ol (\circ). HMF was heated in the presence of ammonium chloride (30 μmol) and 0.3 M sodium phosphate buffer, pH 8, for 22 h at 100 $^{\circ}\text{C}$.

Figure 3. Effect of amine compound concentration on the formation of 6-(hydroxymethyl)pyridin-3-ol (\circ and \diamond) and on the disappearance of 5-hydroxymethylfurfural (HMF, Δ and ∇). HMF (10 μmol) was heated in the presence of the amine compound and 0.3 M buffer for 22 h at 100 $^{\circ}\text{C}$. Two amine compounds were assayed: ammonia (\diamond and Δ) and ammonium chloride (\circ and ∇). Data are given in nmol of the produced 6-(hydroxymethyl)pyridin-3-ol per μmol of the amount of HMF added to the reaction mixture.

Figure 4. Effect of time and temperature on the formation of 6-(hydroxymethyl)pyridin-3-ol. 5-Hydroxymethylfurfural (HMF, 10 μmol) was heated in the presence of ammonium chloride (30 μmol) and 0.3 M sodium phosphate buffer, pH 8. The assayed temperatures were: 100 (∇), 120 (Δ), 140 (\circ), and 160 (\square) $^{\circ}\text{C}$. Data are given in nmol of the

produced 6-(hydroxymethyl)pyridin-3-ol per μmol of the amount of HMF added to the reaction mixture.

Figure 5. Arrhenius plot obtained for the formation of 6-(hydroxymethyl)pyridin-3-ol (\circ) and the disappearance of 5-hydroxymethylfurfural (HMF, \square). HMF ($10 \mu\text{mol}$) was heated in the presence of ammonium chloride ($30 \mu\text{mol}$) and 0.3 M sodium phosphate buffer, pH 8, at the temperatures indicated in Figures 4 and 6.

Figure 6. Effect of time and temperature on the disappearance of 5-hydroxymethylfurfural (HMF). HMF ($10 \mu\text{mol}$) was heated in the presence of ammonium chloride ($30 \mu\text{mol}$) and 0.3 M sodium phosphate buffer, pH 8. The assayed temperatures were: 100 (∇), 120 (\triangle), 140 (\circ), and 160 (\square) $^{\circ}\text{C}$.

Figure 7. Formation of pyridin-3-ols [6-(hydroxymethyl)pyridin-3-ol (stripped bars), pyridin-3-ol (open bars), and 2-methylpyridin-3-ol (crosshatched bars)] from 2-oxofurans [5-hydroxymethylfurfural (HMF), furfural, and 2-acetylfuran, respectively]. The 2-oxofuran ($10 \mu\text{mol}$) was heated in the presence of ammonium chloride ($30 \mu\text{mol}$) and 0.3 M sodium phosphate buffer, pH 8, at $100 \text{ }^{\circ}\text{C}$ for the indicated times. Data are given in nmol of the produced pyridin-3-ol per μmol of the amount of 2-oxofuran added to the reaction mixture.

Figure 8. Proposed pathway for the formation of pyridine-3-ols from the corresponding 2-oxofurans. For furfural and pyridin-3-ol, $R_1 = \text{H}$ and $R_2 = \text{H}$. For 2-acetylfuran and 2-methylpyridin-3-ol, $R_1 = \text{H}$ and $R_2 = \text{CH}_3$. For 5-hydroxymethylfurfural (HMF) and 6-(hydroxymethyl)pyridin-3-ol, $R_1 = \text{CH}_2\text{OH}$ and $R_2 = \text{H}$.

Figure 9. Effect of heating time on the formation of 6-(hydroxymethyl)pyridin-3-ol (○) and 5-hydroxymethylfurfural (HMF, △) in honey heated at, A, 60, and B, 100 °C. Honey (1 g) was heated in closed test tubes for the indicated times and temperatures.

Figure 10. Effect of heating time on the formation of 6-(hydroxymethyl)pyridin-3-ol (○), 5-hydroxymethylfurfural (HMF, △), pyridin-3-ol (◁), and 2-methylpyridin-3-ol (▷) in sugarcane honey heated at, A, 60, and B, 100 °C. Sugarcane honey (1 g) was heated in closed test tubes for the indicated times and temperatures.

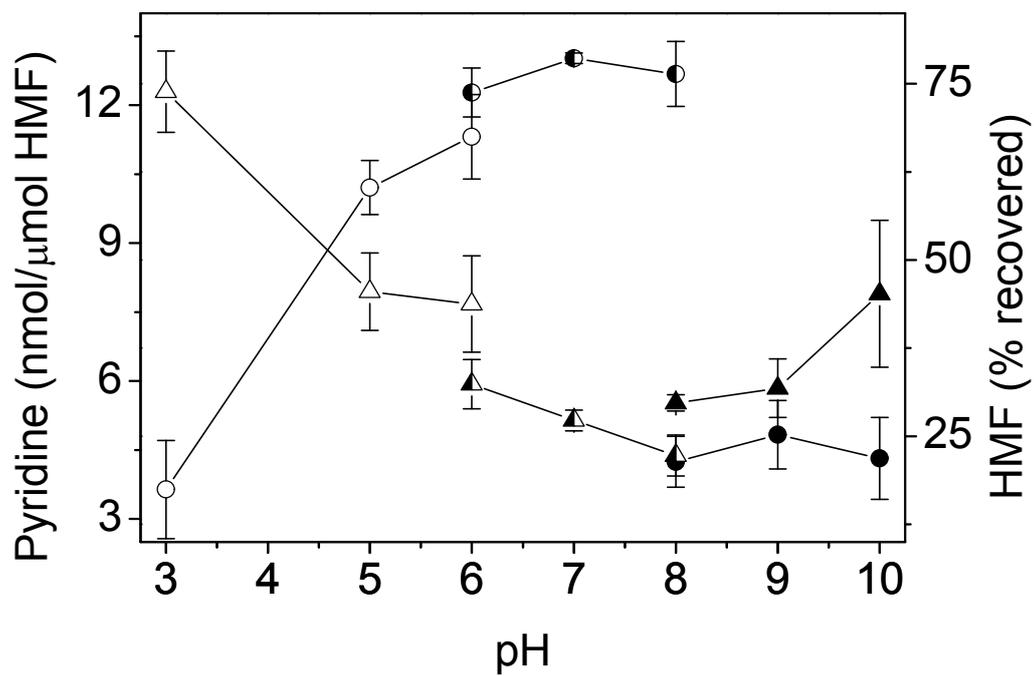


Figure 1

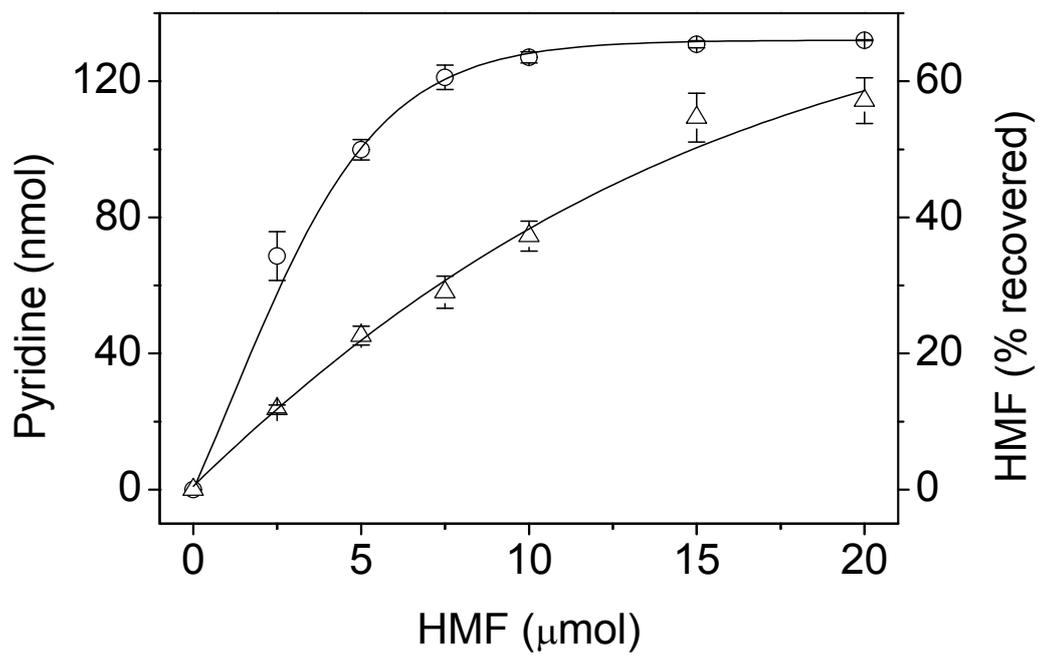


Figure 2

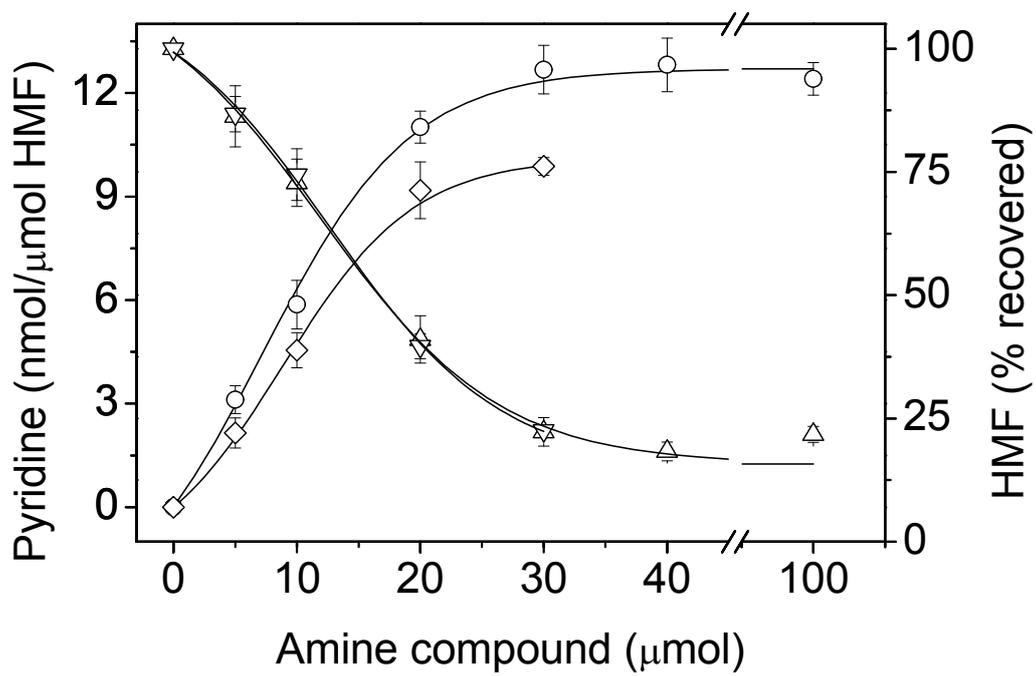


Figure 3

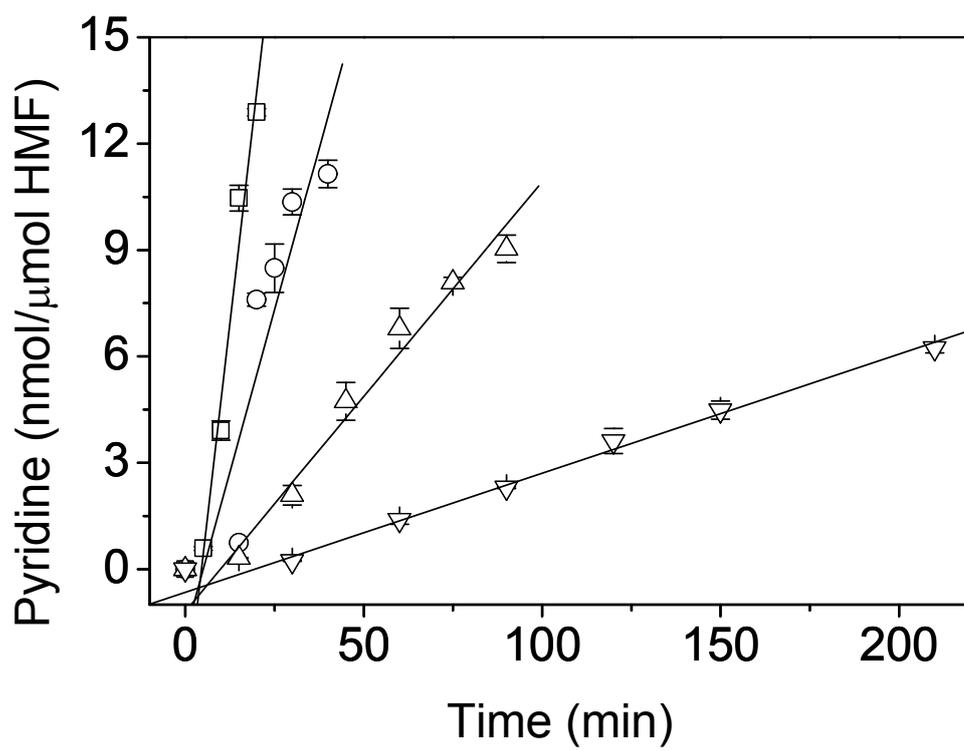


Figure 4

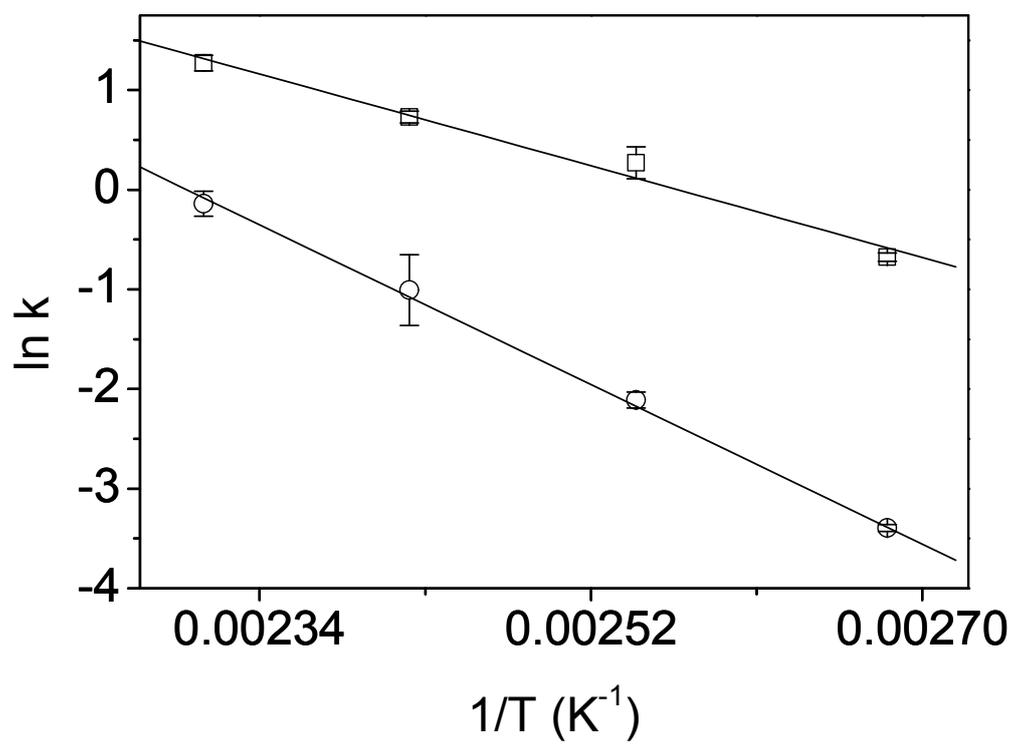


Figure 5

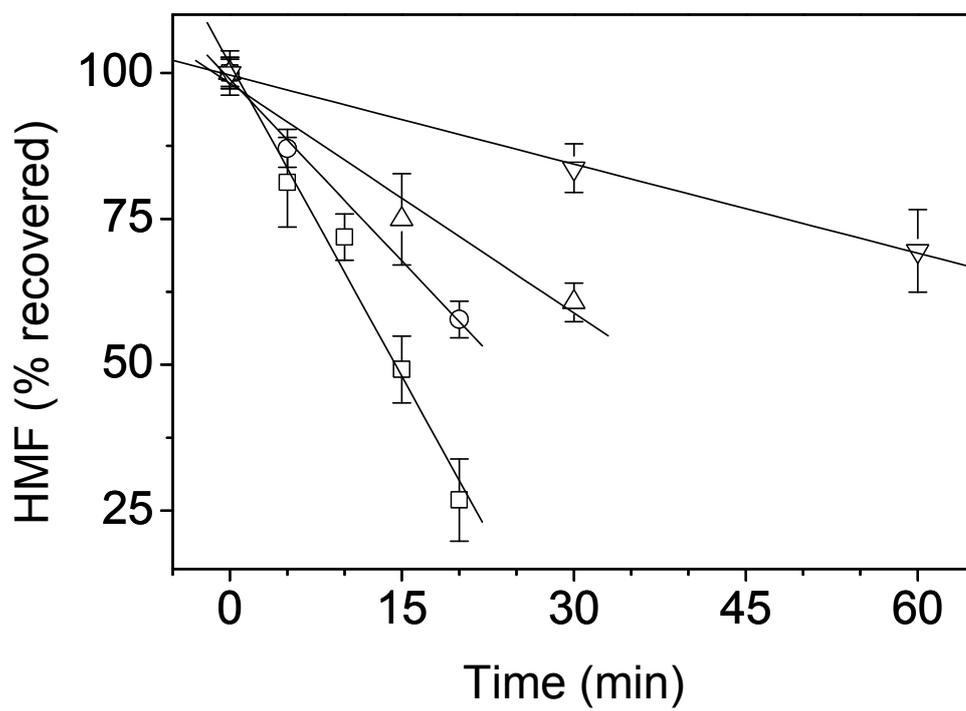


Figure 6

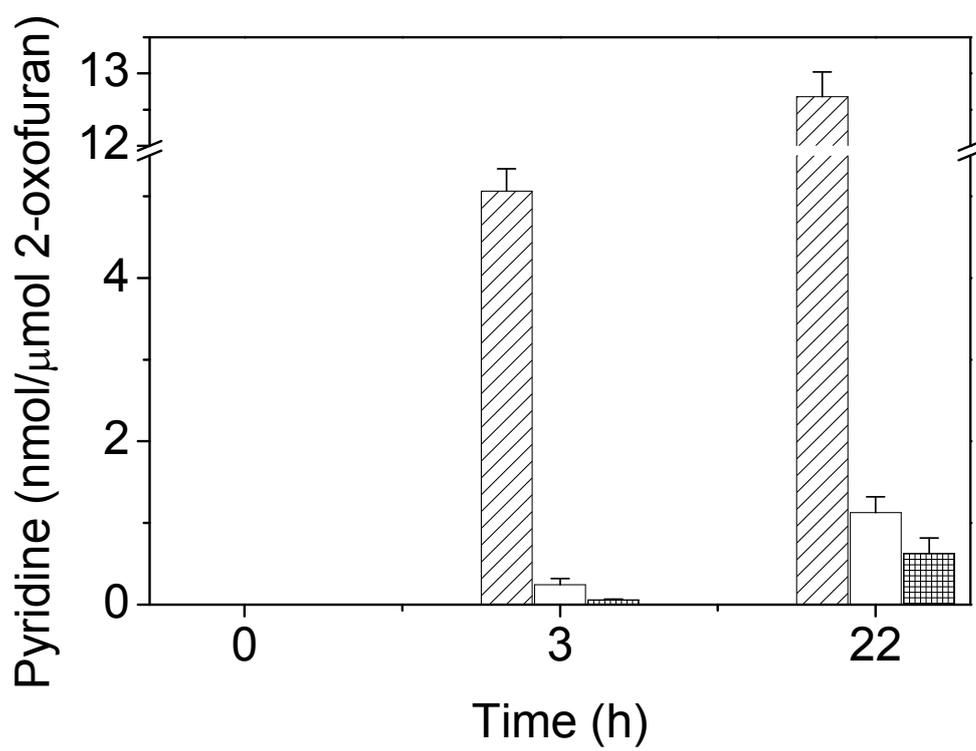
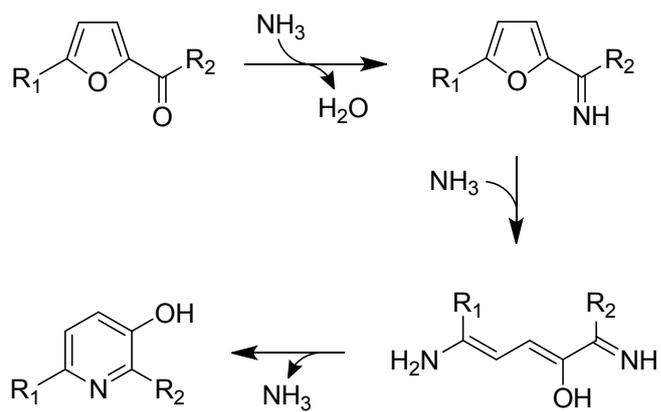


Figure 7

**Figure 8**

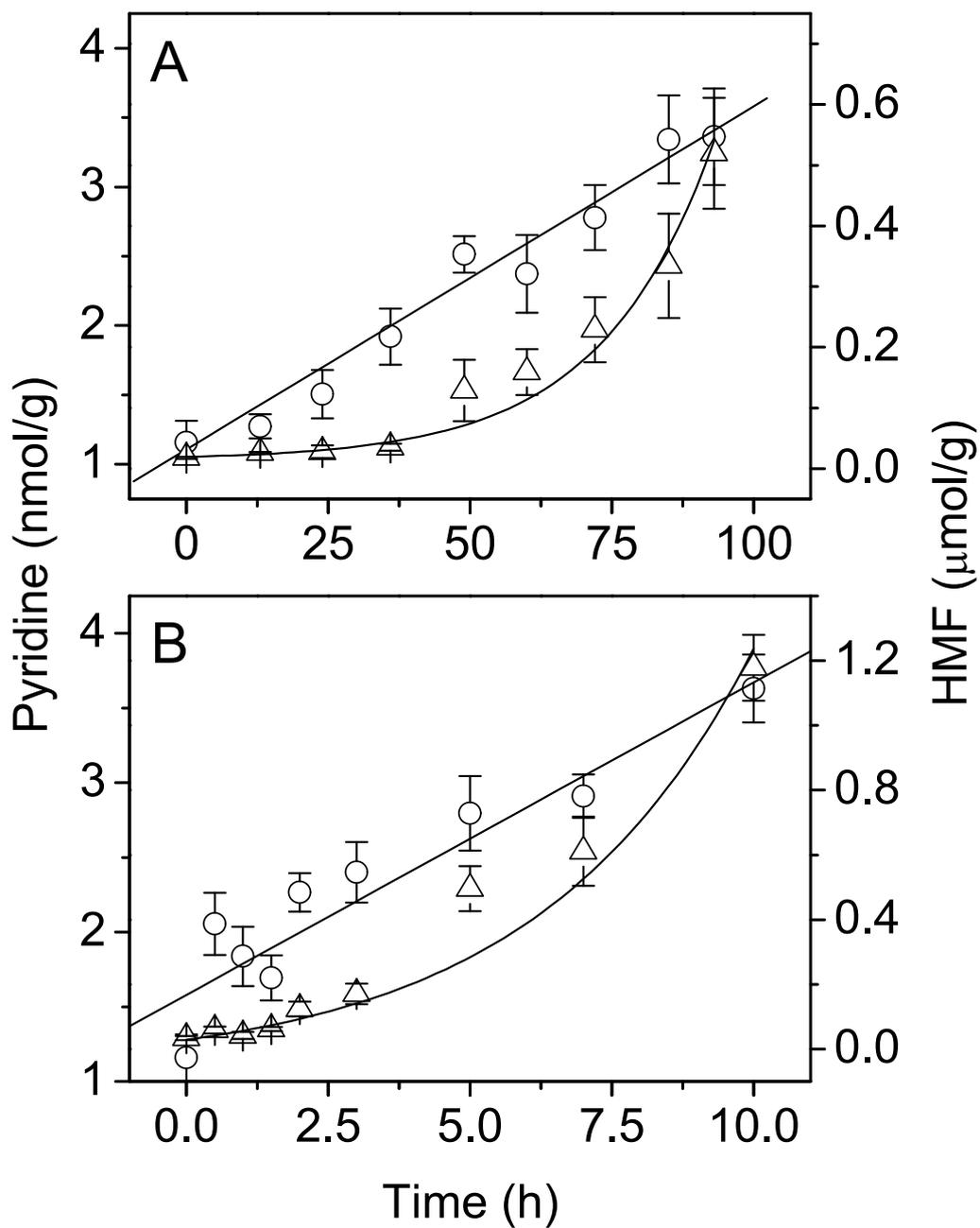


Figure 9

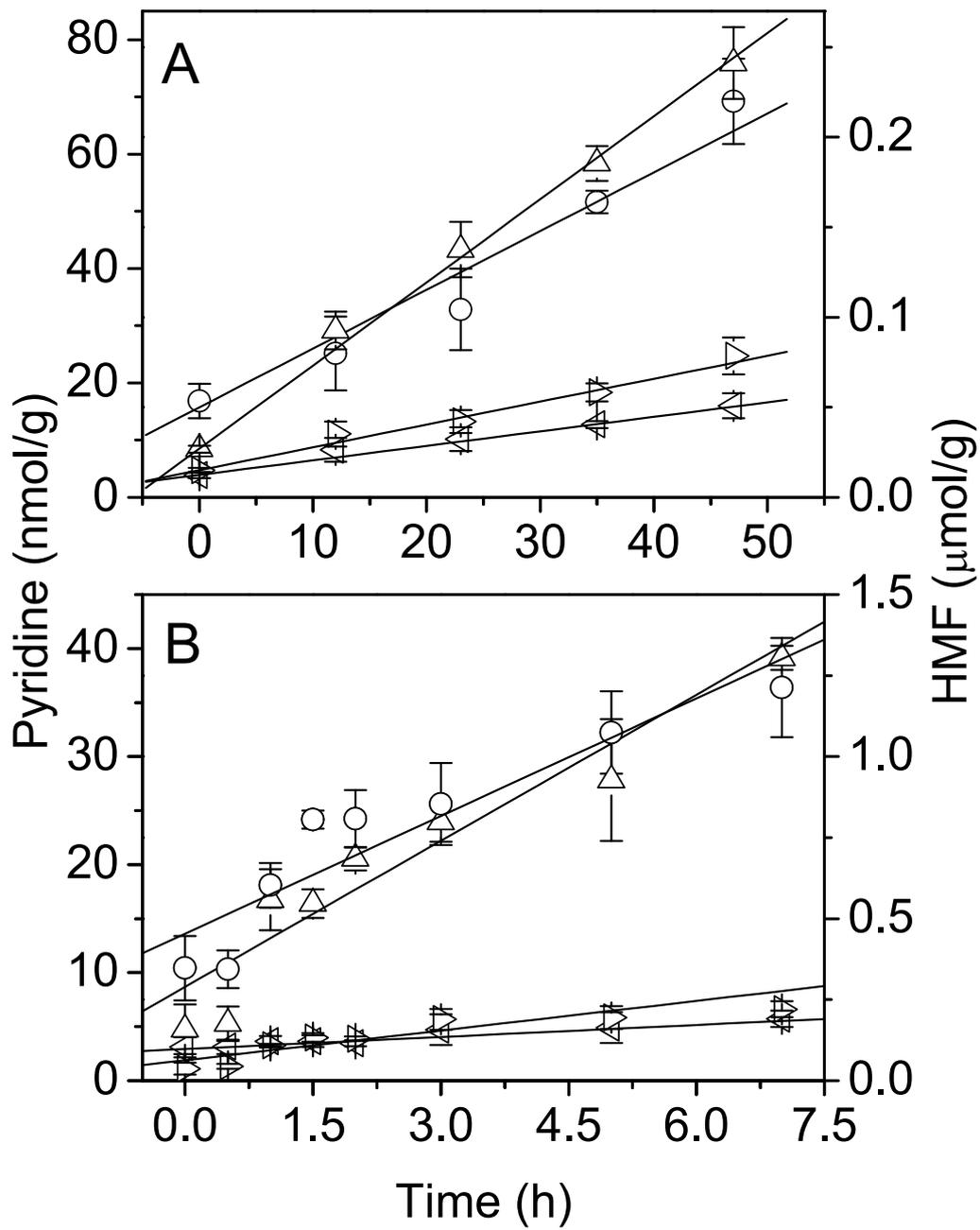


Figure 10

GRAPHIC FOR TABLE OF CONTENTS

