

# Synthesis and evaluation of unsymmetrical heterocyclic thioureas as potent $\beta$ -glucuronidase inhibitors

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**Abstract** Thiourea analogs **1–20** were synthesized and evaluated for their in vitro  $\beta$ -glucuronidase inhibitory potential. The compounds **9** ( $0.86 \pm 0.01 \mu\text{M}$ ), **6** ( $1.24 \pm 0.01 \mu\text{M}$ ), **16** ( $1.64 \pm 0.02 \mu\text{M}$ ) and **15** ( $2.12 \pm 0.02 \mu\text{M}$ ) showed potent activity. Other analogs **1–5**, **7**, **8**, **10**, **11**, **13**, **17**, **20** showed better activity than standard drug D-saccharic acid 1,4-lactone ( $47.34 \pm 0.21 \mu\text{M}$ ) ranging 4.36–34.4  $\mu\text{M}$ . All synthetic compounds were characterized by different spectroscopic methods. This study has identified a new class of potent inhibitors  $\beta$ -glucuronidase.

**Keywords** Thioureas ·  $\beta$ -Glucuronidase inhibition · Heterocyclic

## Introduction

Urea derivatives have broad spectrum of biological activities as colchicine-binding antagonist, (Fortin *et al.*, 2007, 2009) and CXCR3 antagonist, (Watson *et al.*, 2008) anti-HIV, antiviral, antibacterial, analgesic and HDL-elevating properties (Sivan *et al.*, 2013; Venkatachalam *et al.*, 2004; Holešová *et al.*, 2014; Flieger *et al.*, 2012; Bloom *et al.*, 2004; Seth *et al.*, 2004; Lee *et al.*, 2003). The urea analogs are found to be very selective, and they are well suited to interact with the viral proteases and showed no interaction with the human proteases. Since, after first report of these inhibitors, various derivatives have rapidly been synthesized, class of cyclic compounds may soon become a feasible choice to the currently available antiretroviral agents. The substituted cyclic urea shown below has shown an increase in antiretroviral potency (Bäckbro *et al.*, 1997; Sham *et al.*, 1996a; Hultén *et al.*, 1997; Ala *et al.*, 1998).

Cyclic urea derivatives are also cytotoxic on a large number of cancer cell lines and remain active on most chemo-resistant cells (Fortin *et al.*, 2007; Liu *et al.*, 2014). Chalcon reacts with isocyanate, thioisocyanate and amines to produce important industrial products belonging to urea and thiourea which possess a series of biological activities including herbicidal activities, inhibition of nitric oxide, antimicrobial, anti-HIV, antiviral, analgesic and HDL-elevating properties (Yang *et al.*, 2012). Derivatives of 1-phenyl-3-{4-[(2*E*)-3-phenylprop-2-enoyl]-phenyl}-urea (**2**) exhibited anti-inflammatory and antimalarial activities (Santos *et al.*, 2008).

Classes of heterocyclic ureas, the azacyclic ureas, are the potent inhibitors of HIV-1 protease; inhibition of HIV protease is an important approach for the intervention of HIV infection but possesses very low bioavailability (Chrusciel and Strohbach, 2004; Sham *et al.*, 1996b).

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*N*-Acyl azide ureas **3** are excellent inhibitors of viral protease and have shown good antiviral activities against resistant strains (Zhao *et al.*, 2005).

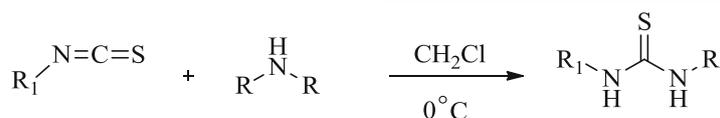
$\beta$ -Glucuronidase is an enzyme which catalyzes the glucuronosyl-*O*-bonds cleavage (Sperker *et al.*, 1997). This is also up-regulated in different pathological conditions, such as infection of urinary tract (Bank and Bailine, 1965; Roberts *et al.*, 1967; Ronald *et al.*, 1971; Kallet and Lapco, 1967), renal disease (Gonick *et al.*, 1973), rejection of transplantation (Schapiro *et al.*, 1968), epilepsy (Plum 1967), larynx and breast cancer (Boyland *et al.*, 1957). Besides, this enzyme is also involved in inflammatory joint diseases, like rheumatoid arthritis (Caygill and Pitkeathly, 1966), and some hepatic diseases, and AIDS is also reported due to over-expression of this enzyme. Literature report showed that the bacterial  $\beta$ -glucuronidase inhibitor leads to a decrease in carcinogen-induced colonic tumors (Reddy, 1976). The current study is focused on the discovery of  $\beta$ -glucuronidase inhibitors of pharmacological importance. Some synthetic  $\beta$ -glucuronidase inhibitors reported in recent past (Jamil *et al.*, 2014; Khan *et al.*, 2014a, b, c; Cheng *et al.*, 2015; Ahmad *et al.*, 2012).

Our research group is continuously working on the chemistry and biology of new compounds (Aziz *et al.*, 2014; Musharraf *et al.*, 2012; Anouar *et al.*, 2013), sulfur-containing compounds (Khan *et al.*, 2008, 2009, 2010, 2011) and hydrazones (Khan *et al.*, 2012; Taha *et al.*, 2013, 2014).

## Results and discussion

### Chemistry

We previously reported that unsymmetrical urea derivatives are excellent antiglycating agent; therefore, we further explored this class for discovering other biological uses. The synthetic procedure which was adopted was quite simple and required very short time to complete. Urea derivatives **1–20** were synthesized from different amines treating them with isocyanate (2 mmol). The crude products were recrystallized with ethanol afforded the pure products (Scheme 1).



In continuation of our work on enzyme inhibition (Rahim *et al.*, 2015; Taha *et al.*, 2015a; Taha *et al.*, 2015b; Khan *et al.*, 2014d), the compounds **1–20** were evaluated for in vitro  $\beta$ -glucuronidase inhibitory potential (Table 1).

The compounds **9**, **6**, **15** and **16** showed  $0.86 \pm 0.01$ ,  $1.24 \pm 0.01$ ,  $2.12 \pm 0.02$  and  $1.64 \pm 0.02$   $\mu\text{M}$  activity far better than standard drug D-saccharic acid 1,4-lactone  $47.34 \pm 0.21$   $\mu\text{M}$ , respectively. 3-Chlorophenyl-2-methylpiperazinethiourea (**9**) showed potent activity that may be due to 3-chlorophenyl ring which can be proved by comparing the activity of compound **5** ( $28.12 \pm 0.35$   $\mu\text{M}$ ) having phenyl ring with no substitution on 2-methylpiperazinethiourea. The second most active compound also has 3-chlorophenyl ring and 4-phenylpiperazinethiourea; when 3-chlorophenyl is replaced with phenyl as in compound **4**, the activity decreased almost two times. The compounds **15** and **16** have quinoline ring and phenyl and 3-chlorophenyl, respectively; the activity of 3-chlorophenyl is slightly better than phenyl substituted. Compounds **3** and **17** have morpholine and phenyl and 3-chlorophenyl thiourea, respectively; it was observed that both compounds have almost same inhibition potential.

The compounds **2**, **7**, **10**, **11** and **13** having piperidine ring with different substitutions showed good activities. The activity pattern is common, and 3-chlorophenylthiourea showed better activity than phenylthiourea. Phenyl-4-methylpiperazinethiourea (**1**) showed less activity as compared to 3-chlorophenyl-4-ethylpiperazinethiourea (**8**) which may be due to 3-chlorophenyl. The compound **20** showed fewer activities than compounds **1** and **8** because 4-acyl group replaces ethyl group. Compounds **12**, **14**, **18** and **19** showed less than 50 % inhibition and so considered as inactive. Hence, we have discovered new potent derivatives of thiourea for  $\beta$ -glucuronidase inhibition. Further study on this compound could produce new molecules for the  $\beta$ -glucuronidase inhibition.

## Experimental

### Materials and methods

NMR experiments were performed on Avance Bruker AM 300 and 500 MHz machines. CHN analyses were carried out on a Carlo Erba Strumentazione-Mod-1106, Italy. Electron impact mass spectra (EI MS) were recorded on a

Finnigan MAT-311A (Germany) mass spectrometer. Thin-layer chromatography (TLC) was monitored on pre-coated silica gel aluminum plates (Kieselgel 60, 254, E. Merck, Germany). Chromatograms were visualized by iodine vapors or UV at 254 and 365 nm.

Entry No	R <sub>1</sub>	R	Yield (%)	Entry No	R <sub>1</sub>	R	Yield (%)
1			85	11			80
2			74	12			82
3			80	13			77
4			70	14			85
5			82	15			78
6			75	16			88
7			83	17			90
8			78	18			84
9			80	19			83
10			76	20			84

Scheme 1 Synthesis of heterocyclic thioureas 1–20

**Table 1** In vitro  $\beta$ -glucuronidase activity of compounds **1–20**

Compounds	IC <sub>50</sub> ( $\mu\text{M} \pm \text{SEM}^{\text{a}}$ )	Compounds	IC <sub>50</sub> ( $\mu\text{M} \pm \text{SEM}^{\text{a}}$ )
<b>1</b>	20.50 $\pm$ 0.26	<b>11</b>	31.40 $\pm$ 0.54
<b>2</b>	25.40 $\pm$ 0.30	<b>12</b>	NA <sup>b</sup>
<b>3</b>	8.34 $\pm$ 0.15	<b>13</b>	11.21 $\pm$ 0.26
<b>4</b>	4.36 $\pm$ 0.08	<b>14</b>	NA <sup>b</sup>
<b>5</b>	28.12 $\pm$ 0.35	<b>15</b>	2.12 $\pm$ 0.02
<b>6</b>	1.24 $\pm$ 0.01	<b>16</b>	1.64 $\pm$ 0.02
<b>7</b>	18.46 $\pm$ 0.24	<b>17</b>	8.42 $\pm$ 0.36
<b>8</b>	15.4 $\pm$ 0.46	<b>18</b>	NA <sup>b</sup>
<b>9</b>	0.86 $\pm$ 0.01	<b>19</b>	NA <sup>b</sup>
<b>10</b>	34.4 $\pm$ 0.42	<b>20</b>	47.34 $\pm$ 0.21
D-Saccharic acid 1,4-lactone <sup>c</sup>	48.4 $\pm$ 1.25 $\mu\text{M}$	–	–

<sup>a</sup> SEM is the standard error of the mean

<sup>b</sup> NA Not active

<sup>c</sup> D-saccharic acid 1,4-lactone, standard inhibitor for  $\beta$ -glucuronidase

### General procedure for the synthesis of compounds **1–20**

Thiourea derivatives **1–20** were synthesized from different heterocyclic amines by treating with isocyanate. Heterocyclic amines (2 mmol) dissolved in dichloromethane (10 mL), and temperature was maintained at 0 °C; then isocyanate (2 mmol) was added drop-wise with constant stirring. The formation of white solid after 1–1.5 h confirmed the completion of reaction. The resultant solid products **1–20** were filtered and washed with hexane. Recrystallization with ethanol afforded the yield between 60 and 85 %.

### Biological assays

$\beta$ -Glucuronidase (E.C. 3.2.1.31 from bovine liver, G-0251) and *p*-nitrophenyl- $\beta$ -D-glucuronide (N-1627) were purchased from Sigma Chemical Co. (St. Louis, MO, USA).

### Assay for $\beta$ -D-glucuronidase

$\beta$ -D-Glucuronidase inhibition was determined by measuring the absorbance of the *p*-nitrophenol which is produced from the substrate at 405 nm. The total reaction volume was 250  $\mu\text{L}$ . The reaction mixture contains 5  $\mu\text{L}$  of test compound solution, 185  $\mu\text{L}$  of 0.1 M acetate buffer and 10  $\mu\text{L}$  of enzyme, and it was incubated at 37 °C for 30 min. The plates were read on a multiplate reader at 405 nm after the addition of 50  $\mu\text{L}$  of 0.4 mM *p*-nitrophenyl- $\beta$ -D-glucuronide. All assays were performed in triplicate.

*4-Methyl-N-phenylpiperazine-1-carbothioamide (1)* Yield: 85 %; <sup>1</sup>H NMR: (500 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  7.90 (s, 1H, NH), 7.54 (dd, 2H, *J* = 6.4, *J* = 1.7 Hz, H-2'/H-6'), 7.23 (t, 2H, *J* = 7.8 Hz, H-3', H-5'), 6.98 (t, 1H, *J* = 8.4 Hz, H-4'), 3.52 (t, 4H, *J* = 10.1 Hz, 2-CH<sub>2</sub>/6-CH<sub>2</sub>), 2.33 (t, 4H, *J* = 10.0 Hz, 3-CH<sub>2</sub>/5-CH<sub>2</sub>), 2.50 (s, CH<sub>3</sub>); <sup>13</sup>C NMR (75 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  181.7 (C=S, C), 138.4 (C, C-1'), 129.1 (CH, C-3'), 129.1 (CH, C-5'), 128.3 (CH, C-4'), 126.6 (CH, C-2'), 126.6 (CH, C-6'), 56.7 (CH<sub>2</sub>, C-2), 56.7 (CH<sub>2</sub>, C-6), 51.6 (CH<sub>2</sub>, C-3), 51.6 (CH<sub>2</sub>, C-5), 46.5 (CH<sub>3</sub>, C); EI MS: *m/z* (rel. abund%) 235.

*N-Phenylpiperidine-1-carbothioamide (2)* Yield: 74 %; <sup>1</sup>H NMR: (500 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  8.42 (s, 1H, NH), 7.46 (d, 2H, *J* = 8.0 Hz, H-2'/H-6'), 7.23 (t, 2H, *J* 8.0 Hz, H-3'/H-5'), 6.93 (t, 1H, *J* = 7.0 Hz, H-4'), 3.44 (t, 2H, *J* = 10.0 Hz, 2-CH<sub>2</sub>), 2.52 (t, 2H, *J* = 11 Hz, 6-CH<sub>2</sub>), 1.58 (m, 6H, 3-CH<sub>2</sub>/4-CH<sub>2</sub>/5-CH<sub>2</sub>); <sup>13</sup>C NMR (75 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  187.3 (C=S, C), 138.4 (C, C-1'), 129.1 (CH, C-3'), 129.1 (CH, C-5'), 128.5 (CH, C-4'), 126.4 (CH, C-2'), 126.4 (CH, C-6'), 54.5 (CH<sub>2</sub>, C-2), 54.5 (CH<sub>2</sub>, C-6), 25.7 (CH<sub>2</sub>, C-3), 25.7 (CH<sub>2</sub>, C-5), 24.9 (CH<sub>2</sub>, C-4); EI MS: *m/z* (rel. abund%) 220.

*N-Phenylmorpholine-4-carbothioamide (3)* Yield: 80 %; <sup>1</sup>H NMR: (500 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  7.38 (dd, 2H, *J* = 8.0, *J* = 2.0, Hz, H-2'/H-6') 7.29 (t, 2H, *J* = 8.0 Hz, H-3'/H-5'), 7.07 (t, 1H, *J* = 8.0 Hz, H-4'), 3.75 (t, 4H, *J* = 10.4 Hz, 3-CH<sub>2</sub>, 5-CH<sub>2</sub>), 3.48 (t, 4H, *J* = 10.3 Hz, 2-CH<sub>2</sub>/6-CH<sub>2</sub>); <sup>13</sup>C NMR (75 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  181.9 (C=S, C), 138.4 (C, C-1'), 129.1 (CH, C-3'), 129.1 (CH, C-5'), 128.5 (CH, C-4'), 126.4 (CH, C-2'), 126.4 (CH, C-6'), 66.3 (CH<sub>2</sub>, C-3), 66.3 (CH<sub>2</sub>, C-5), 50.1 (CH<sub>2</sub>, C-2), 50.1 (CH<sub>2</sub>, C-6); EI MS: *m/z* (rel. abund%) 222.

*N*-4-Diphenylpiperazine-1-carbothioamide (**4**) Yield: 70 %;  $^1\text{H}$  NMR: (500 MHz, DMSO- $d_6$ ):  $\delta$  7.38, (dd, 2H,  $J = 8.0$ ,  $J = 2.0$  Hz, H-2'/H-6'), 7.31 (m, 4H, H-3'/H-5'/H-2''/H-6''), 7.06 (m, 3H, H-3''/H-5''/H-4'), 6.91 (t, 1H,  $J = 8.0$  Hz, H-4''), 3.72 (t, 4H,  $J = 10.0$  Hz, 2-CH<sub>2</sub>/6-CH<sub>2</sub>), 3.24 (t, 4H,  $J = 10.1$  Hz, 3-CH<sub>2</sub>,5-CH<sub>2</sub>);  $^{13}\text{C}$  NMR (75 MHz, DMSO- $d_6$ ):  $\delta$  181.7 (C=S, C), 149.5 (C, C-1'), 138.4 (C, C-1'), 129.7 (CH, C-3'), 129.7 (CH, C-5'), 128.9 (CH, C-3'), 128.9 (CH, C-5'), 128.5 (CH, C-4'), 126.4 (CH, C-2'), 126.4 (CH, C-6'), 121.8 (CH, C-4'), 114.4 (CH, C-2'), 114.4 (CH, C-6'), 56.8 (CH<sub>2</sub>, C-2), 56.8 (CH<sub>2</sub>, C-6), 53.6 (CH<sub>2</sub>, C-3), 53.6 (CH<sub>2</sub>, C-5); EI MS:  $m/z$  (rel. abund%) 297.

3-Methyl-*N*-phenyl-4-(phenylcarbamothioyl)piperazine-1-carboxamide (**5**) Yield: 82 %;  $^1\text{H}$  NMR: (500 MHz, DMSO- $d_6$ ):  $\delta$  8.56 (s, 1H, NH), 8.52 (s, 1H, NH), 7.50 (m, 4H, H-2'/H-6'/H-2''/H-6''), 7.29 (m, 4H, H-3'/H-5'/H-3''/H-5''), 7.01-6.94 (m, 2H, H-4'/H-4''), 4.45 (t, 1H,  $J = 6.5$  Hz, 3a-CH), 4.16 (d, 1H,  $J = 11$  Hz, 3b-CH), 3.97 (t, 4H,  $J = 11$  Hz, 6-CH<sub>2</sub>), 3.18 (t, 1H,  $J = 11$  Hz, 5-CH<sub>2</sub>), 2.99 (m, 1H, 2-CH), 1.11 (d, 3H,  $J = 7.2$  Hz, CH<sub>3</sub>);  $^{13}\text{C}$  NMR (75 MHz, DMSO- $d_6$ ):  $\delta$  181.3 (C=S, C), 138.6 (C, C-1'), 129.1 (CH, C-3'), 129.1 (CH, C-5'), 128.5 (CH, C-4'), 126.6 (CH, C-2'), 126.6 (CH, C-6'), 66.6 (CH, C-2), 57.5 (CH<sub>2</sub>, C-3), 56.9 (CH<sub>2</sub>, C-6), 47.8 (CH<sub>2</sub>, C-5), 18.5 (CH<sub>3</sub>, C); EI MS:  $m/z$  (rel. abund%) 354.

*N*-(3-Chlorophenyl)-4-phenylpiperazine-1-carbothioamide (**6**) Yield: 75 %;  $^1\text{H}$  NMR: (300 MHz, DMSO- $d_6$ ):  $\delta$  8.24 (s, 1H, NH), 7.86 (d, 1H,  $J = 2.0$  Hz, H-2'), 7.49 (dt, 1H,  $J = 8.0$ ,  $J = 2.0$  Hz, H-6'), 7.32 (d, 1H,  $J = 2.0$  Hz, H-4'), 7.29-7.22 (m, 2H, H-2''/H-6''), 7.04-6.98 (m, 3H, H-4''/H-6''/H-6'), 6.90 (t, 1H,  $J = 7.0$  Hz, H-4''), 3.74 (t, 4H,  $J = 10.0$  Hz, H-3/H-5), 3.24 (t, 4H,  $J = 10.0$  Hz, H-2/H-6);  $^{13}\text{C}$  NMR (75 MHz, DMSO- $d_6$ ):  $\delta$  181.7 (C=S, C), 149.4 (C, C-1'), 138.4 (C, C-1'), 134.5 (C, C-3'), 130.3 (CH, C-5'), 129.5 (CH, C-3'), 129.5 (CH, C-5'), 128.2 (CH, C-4'), 126.8 (CH, C-2'), 124.5 (CH, C-6'), 121.8 (CH, C-4'), 114.4 (CH, C-2'), 114.4 (CH, C-6'), 56.8 (CH<sub>2</sub>, C-2), 56.8 (CH<sub>2</sub>, C-6), 53.4 (CH<sub>2</sub>, C-3), 53.5 (CH<sub>2</sub>, C-4); EI MS:  $m/z$  (rel. abund%) 331.

*N*-(3-Chlorophenyl)piperidine-1-carbothioamide (**7**) Yield: 83 %;  $^1\text{H}$  NMR: (500 MHz, DMSO- $d_6$ ):  $\delta$  7.50 (t, 1H,  $J = 4.0$  Hz, H-2'), 7.28 (dt, 1H,  $J = 8.5$ ,  $J = 2.0$  Hz, H-4'), 7.28 (t, 1H,  $J = 8.0$  Hz, H-5'), 6.99 (dt, 1H,  $J_6' = 8.0$  -  $J = 2.0$  Hz, H-6'), 3.49 (t, 4H,  $J = 10.5$  Hz, H-2,6), 1.68-1.63 (m, 2H, H-4), 1.60-1.58 (m, 4H, H-3,5);  $^{13}\text{C}$  NMR (75 MHz, DMSO- $d_6$ ):  $\delta$  187.1 (C=S, C), 138.6 (C, C-1'), 134.5 (C, C-3'), 130.6 (CH, C-5'), 128.2 (CH, C-4'), 126.8 (CH, C-2'), 124.7 (CH, C-6'), 54.3 (CH<sub>2</sub>, C-2), 54.3 (CH<sub>2</sub>, C-6), 25.5 (CH<sub>2</sub>, C-3), 25.5 (CH<sub>2</sub>, C-5), 24.7 (CH<sub>2</sub>, C-4); EI MS:  $m/z$  (rel. abund%) 254.

*N*-(3-Chlorophenyl)-4-ethylpiperazine-1-carbothioamide (**8**) Yield: 78 %;  $^1\text{H}$  NMR: (500 MHz, DMSO- $d_6$ ):  $\delta$  7.52, (t, 1H,  $J = 2.0$  Hz, H-2'), 7.28 (ddd, 1H,  $J = 8.0$ ,  $J = 2.0$ ,  $J = 2.0$  Hz, H-4'), 7.27 (t, 1H,  $J = 8.5$  Hz, H-5'), 7.04 (ddd, 1H,  $J = 8.0$ ,  $J = 2.0$ ,  $J = 2.0$  Hz, H-4'), 3.56 (t, 4H,  $J = 10.0$  Hz, H-2/H-6), 2.56-2.52 (m, 6H, 2xH-3/H-5/CH<sub>2</sub>), 1.20 (d, 3H,  $J = 8.0$  Hz, CH<sub>3</sub>);  $^{13}\text{C}$  NMR (75 MHz, DMSO- $d_6$ ):  $\delta$  181.7 (C=S, C), 138.6 (C, C-1'), 134.5 (C, C-3'), 130.3 (CH, C-5'), 128.2 (CH, C-4'), 126.8 (CH, C-2'), 124.5 (CH, C-6'), 57.2 (CH<sub>2</sub>, C-2), 57.2 (CH<sub>2</sub>, C-6), 56.6 (CH<sub>2</sub>, C-3), 56.6 (CH<sub>2</sub>, C-5), 49.7 (CH<sub>2</sub>, C), 13.4 (CH<sub>3</sub>, C); EI MS:  $m/z$  (rel. abund%) 283.

*N*<sup>1</sup>, *N*<sup>4</sup>-Bis(3-chlorophenyl)-2-methyl-1,4-piperazinedicarboxamide (**9**) Yield: 80 %;  $^1\text{H}$  NMR: (500 MHz, DMSO- $d_6$ ):  $\delta$  8.71 (s, 1H, NH), 8.62 (s, 1H, NH), 7.63 (s, 2H, H-2'/H-2''), 7.40 (d, 2H,  $J = 8.0$  Hz, H-5'/H-5''), 7.24 (dt, 2H,  $J = 8.0$ ,  $J = 2.0$  Hz, H-4'/H-4''), 7.02 (dd, 2H,  $J = 2.0$ ,  $J = 2.0$ , H-6'/H-6''), 4.39 (s, 1H, H-3a), 4.06 (d, 1H,  $J = 12.0$  Hz, H-3b), 3.97 (t, 2H,  $J = 13.0$  Hz, H-5), 3.17 (t, 2H,  $J = 13.0$  Hz, H-6), 2.98-2.91 (m, 1H, H-2);  $^{13}\text{C}$  NMR (75 MHz, DMSO- $d_6$ ):  $\delta$  181.3 (C=S, C), 138.4 (C, C-1'), 134.5 (C, C-3'), 130.3 (CH, C-5'), 128.2 (CH, C-4'), 126.8 (CH, C-2'), 124.7 (CH, C-6'), 66.6 (CH, C-2), 57.7 (CH<sub>2</sub>, C-3), 56.7 (CH<sub>2</sub>, C-6), 47.8 (CH<sub>2</sub>, C-5), 18.2 (CH<sub>3</sub>, C); EI MS:  $m/z$  (rel. abund%) 422.

3-Methyl-*N*-phenylpiperidine-1-carbothioamide (**10**) Yield: 76 %;  $^1\text{H}$  NMR: (500 MHz, DMSO- $d_6$ ):  $\delta$  8.36 (s, 1H, NH), 7.46 (dd, 2H,  $J = 8.0$ ,  $J = 1.5$  Hz, H-2'/H-6'), 7.32 (t, 2H,  $J = 8.0$ ,  $J = 8.0$  Hz, H-3'/H-5'), 6.99 (t, 1H,  $J = 8.0$  Hz, H-4'), 4.08 (t, 2H,  $J = 10.5$  Hz, 6-CH<sub>2</sub>), 2.72 (dd, 1H,  $J = 6.0$ ,  $J = 8.0$  Hz, H-2b), 2.52 (dd, 1H,  $J = 7.0$ ,  $J = 6.5$  Hz, H-2a), 1.78-1.67 (m, 2H, H-4b/H-4a), 1.59-1.38 (m, 2H, 5-CH<sub>2</sub>), 1.16-1.05 (m, 1H, H-3), 0.91 (d, 3H,  $J = 6.6$  Hz, CH<sub>3</sub>);  $^{13}\text{C}$  NMR (75 MHz, DMSO- $d_6$ ):  $\delta$  187.3 (C=S, C), 138.4 (C, C-1'), 128.9 (CH, C-3'), 128.9 (CH, C-5'), 128.3 (CH, C-4'), 126.4 (CH, C-2'), 126.4 (CH, C-6'), 60.3 (CH<sub>2</sub>, C-2), 54.6 (CH<sub>2</sub>, C-6), 32.7 (CH<sub>2</sub>, C-4), 28.8 (CH, C-3), 23.2 (CH<sub>2</sub>, C-5), 18.6 (CH<sub>3</sub>, C); EI MS:  $m/z$  (rel. abund%) 234.

4-Methyl-*N*-phenylpiperidine-1-carbothioamide (**11**) Yield: 80 %;  $^1\text{H}$  NMR: (500 MHz, DMSO- $d_6$ ):  $\delta$  8.42 (s, 1H, NH), 7.46 (dd, 2H,  $J = 8.0$ ,  $J = 2.0$  Hz, H-2'/H-6') 7.31 (t, 2H,  $J = 8.0$  Hz, H-3'/H-5'), 7.06 (t, 1H,  $J = 7.2$  Hz, H-4'), 4.07 (d, 2H,  $J = 13.0$  Hz, 2-CH<sub>2</sub>), 2.79 (t, 2H,  $J = 13.0$  Hz, 6-CH<sub>2</sub>) 1.66-1.58 (m, 3H, 4-CH/3-CH<sub>2</sub>), 1.15-1.08 (m, 2H, 5-CH<sub>2</sub>) 1.02 (d,  $J = 13.0$  Hz, CH<sub>3</sub>);  $^{13}\text{C}$  NMR (75 MHz, DMSO- $d_6$ ):  $\delta$  187.1 (C=S, C), 138.4 (C, C-1'), 129.1 (CH, C-3'), 129.1 (CH, C-5'), 128.3 (CH, C-4'), 126.4 (CH, C-2'), 126.4 (CH, C-6'), 51.8 (CH<sub>2</sub>, C-2), 51.8 (CH<sub>2</sub>, C-6), 34.2 (CH<sub>2</sub>, C-3), 34.2 (CH<sub>2</sub>, C-5), 32.2 (CH, C-4), 20.3 (CH<sub>3</sub>, C); EI MS:  $m/z$  (rel. abund%) 234.

*N*-(3-Chlorophenyl)-4-(2-methoxyphenyl)piperazine-1-carbothioamide (**12**) Yield: 82 %;  $^1\text{H}$  NMR: (500 MHz, DMSO- $d_6$ ):  $\delta$  7.54, (t, 1H,  $J = 2$  Hz, H-2'), 7.31 (ddd, 1H,  $J = 1.6$ ,  $J = 8.0$  Hz, H-4'), 7.28 (t, 1H,  $J = 8.0$  Hz, H-5'), 7.07-6.97 (m, 4H, H-3''/H-4''/H-5''/H-6''), 6.96 (dt, 1H,  $J = 8.4$ ,  $J = 2.3$  Hz, H-6'), 3.89 (s, 3H, OCH<sub>3</sub>), 3.71 (t, 4H,  $J = 10.0$  Hz, 2-CH<sub>2</sub>/6-CH<sub>2</sub>), 3.08 (t, 4H,  $J = 10.0$  Hz, 3-CH<sub>2</sub>/5-CH<sub>2</sub>);  $^{13}\text{C}$  NMR (75 MHz, DMSO- $d_6$ ):  $\delta$  181.7 (C=S, C), 162.3 (C, C-1''), 141.2 (C, C-2''), 138.4 (C, C-1'), 134.5 (C, C-3'), 130.3 (CH, C-5'), 128.2 (CH, C-4'), 126.8 (CH, C-2'), 124.5 (CH, C-6'), 123.1 (CH, C-6''), 122.0 (CH, C-4''), 121.8 (CH, C-5''), 113.4 (CH, C-3''), 56.6 (CH<sub>2</sub>, C-2), 56.6 (CH<sub>2</sub>, C-6), 55.7 (OCH<sub>3</sub>, C), 53.7 (CH<sub>2</sub>, C-3), 53.7 (CH<sub>2</sub>, C-5); EI MS:  $m/z$  (rel. abund%) 361.

*N*-(3-Chlorophenyl)-2,6-dimethylpiperidine-1-carbothioamide (**13**) Yield: 77 %;  $^1\text{H}$  NMR: (500 MHz, DMSO- $d_6$ ):  $\delta$  8.42 (s, 1H, NH), 7.72, (t, 1H,  $J = 2.0$  Hz, H-2'), 7.47 (dd, 1H,  $J = 8.0$  Hz, H-4'), 7.28 (t, 1H,  $J = 8.0$  Hz, H-5'), 7.07 (dt, 1H,  $J = 8.0$ ,  $J = 2$  Hz, H-6'), 7.28 (t,  $J = 8.0$  Hz, H-5') 6.99 (dt, 1H,  $J = 7.5$ ,  $J = 2.0$  Hz, H-4'), 4.36-4.30 (m, 2H, 4-CH<sub>2</sub>), 1.58-1.54 (m, 1H, 2-CH), 1.51-1.46 (m, 4H, 3-CH<sub>2</sub>/5-CH<sub>2</sub>), 1.42-1.37 (m, 1H, 6CH), 1.14 (d, 3H,  $J = 7$  Hz, CH<sub>3</sub>), 1.04 (d, 3H,  $J = 7$  Hz, CH<sub>3</sub>);  $^{13}\text{C}$  NMR (75 MHz, DMSO- $d_6$ ):  $\delta$  187.1 (C=S, C), 138.4 (C, C-1'), 134.5 (C, C-3'), 130.3 (CH, C-5'), 128.2 (CH, C-4'), 126.8 (CH, C-2'), 124.5 (CH, C-6'), 64.2 (CH, C-2), 64.2 (CH, C-6), 30.0 (CH<sub>2</sub>, C-3), 30.0 (CH<sub>2</sub>, C-5), 20.2 (CH<sub>3</sub>, C), 20.2 (CH<sub>3</sub>, C), 14.0 (CH<sub>2</sub>, C-4); EI MS:  $m/z$  (rel. abund%) 284.

*N*-(3-Chlorophenyl)-4-(4-methoxyphenyl)piperazine-1-carbothioamide (**14**) Yield: 85 %;  $^1\text{H}$  NMR: (500 MHz, DMSO- $d_6$ ):  $\delta$  8.79, (s, 1H, H-NH), 7.68 (t, 1H,  $J = 2.0$  Hz, H-2'), 7.43 (dd, 1H,  $J = 8.0$ ,  $J = 2.0$  Hz, H-4'), 7.25 (t, 1H,  $J = 8.0$  Hz, H-5'), 7.02 (d, 1H,  $J = 2.0$  Hz, H-2'), 7.01 (d, 2H,  $J = 9$  Hz, H-2''/H-6'') 6.87 (d, 2H,  $J = 6.0$  Hz, H-3''/H-5''), 3.69 (s, 3H, CH<sub>3</sub>), 3.60 (t, 4H,  $J = 9.5$  Hz, 3-CH<sub>2</sub>/5-CH<sub>2</sub>), 3.06 (t, 4H,  $J = 2$ -CH<sub>2</sub>/6-CH<sub>2</sub>);  $^{13}\text{C}$  NMR (75 MHz, DMSO- $d_6$ ):  $\delta$  181.7 (C=S, C), 152.9 (C, C-4''), 146.2 (C, C-1''), 138.4 (C, C-1'), 134.5 (C, C-3'), 130.3 (CH, C-5'), 128.2 (CH, C-4'), 126.8 (CH, C-2'), 124.5 (CH, C-6'), 115.2 (CH, C-2''), 115.2 (CH, C-6''), 115.1 (CH, C-3''), 115.1 (CH, C-5''), 56.6 (CH<sub>2</sub>, C-2), 56.6 (CH<sub>2</sub>, C-6), 55.7 (OCH<sub>3</sub>, C), 53.4 (CH<sub>2</sub>, C-3), 53.4 (CH<sub>2</sub>, C-5); EI MS:  $m/z$  (rel. abund%) 361.

*I*-Phenyl-3-(quinolin-8-yl)thiourea (**15**) Yield: 78 %;  $^1\text{H}$  NMR: (500 MHz, Methanol- $d_4$ ):  $\delta$  8.82 (dd, 1H,  $J = 2.0$ ,  $J = 5.0$  Hz, H-2), 8.52 (dd, 1H,  $J = 7.0$ ,  $J = 2.0$  Hz, H-4), 8.29 (dd, 1H,  $J = 8.0$ ,  $J = 2.0$  Hz, H-7), 7.54-7.48 (m, 5H, H-3, H-5/H-6, H-2'/H-6'), 7.33 (t, 2H,  $J = 7.6$  Hz, H-3/H-5), 7.06 (t, 1H,  $J = 7.6$  Hz, H-4');  $^{13}\text{C}$  NMR

(75 MHz, DMSO- $d_6$ ):  $\delta$  179.8 (C=S, C), 148.5 (CH, C-2), 138.4 (C, C-1'), 137.1 (C, C-10), 136.6 (CH, C-4), 133.7 (C, C-9), 129.1 (C, C-5), 129.1 (CH, C-3'), 129.1 (CH, C-5'), 128.3 (CH, C-4'), 127.4 (CH, C-7), 126.6 (CH, C-2'), 126.6 (CH, C-6'), 121.3 (CH, C-3), 116.6 (CH, C-8), 113.7 (CH, C-6); EI MS:  $m/z$  (rel. abund%) 279.

*I*-(3-Chlorophenyl)-3-(quinolin-8-yl)thiourea (**16**) Yield: 88 %;  $^1\text{H}$  NMR: (500 MHz, Methanol- $d_4$ ):  $\delta$  8.80 (dd, 1H,  $J = 2.0$ ,  $J = 4.5$  Hz, H-2), 8.52 (dd, 1H,  $J = 6.0$ ,  $J = 2.5$  Hz, H-3), 8.29 (dd, 1H,  $J = 8.4$ ,  $J = 1.7$  Hz, H-7), 7.74 (t, 1H,  $J = 2.0$  Hz, H-2'), 7.57-7.49 (m, 3H, H-4/H-5/H-6), 7.38 (ddd, 1H,  $J = 8.4$ ,  $J = 2.0$ ,  $J = 1.6$  Hz, H-4'), 7.25 (t, 1H,  $J = 8.0$  Hz, H-5'), 7.04 (ddd, 1H,  $J = 7.0$ ,  $J = 2.0$ ,  $J = 1.6$  Hz, H-6);  $^{13}\text{C}$  NMR (75 MHz, DMSO- $d_6$ ):  $\delta$  179.8 (C=S, C), 148.5 (CH, C-2), 138.4 (C, C-1'), 137.1 (C, C-10), 136.6 (CH, C-4), 134.7 (C, C-3'), 133.9 (C, C-9), 130.5 (CH, C-5'), 129.1 (C, C-5), 128.2 (CH, C-4'), 127.2 (CH, C-7), 126.8 (CH, C-2'), 124.5 (CH, C-6'), 121.1 (CH, C-3), 116.4 (CH, C-8), 113.7 (CH, C-6); EI MS:  $m/z$  (rel. abund%) 313.

*N*-(3-chlorophenyl)morpholine-4-carbothioamide (**17**) Yield: 90 %;  $^1\text{H}$  NMR: (500 MHz, DMSO- $d_6$ ):  $\delta$  8.71 (s, 1H, NH) 7.66 (t, 1H,  $J = 2.0$  Hz, H-2), 7.41 (dt, 1H,  $J = 8.0$ ,  $J = 2.0$  Hz, H-4), 7.29 (t, 1H,  $J = 8.0$  Hz, H-5), 7.02 (dd, 1H,  $J = 7.8$ ,  $J = 2.0$  Hz, H-6), 3.62 (t, 4H,  $J = 5.0$  Hz, 2'-CH<sub>2</sub>/6'-CH<sub>2</sub>), 3.52 (t, 4H,  $J = 5.0$  Hz, 3'-CH<sub>2</sub>, 5'-CH<sub>2</sub>);  $^{13}\text{C}$  NMR (75 MHz, DMSO- $d_6$ ):  $\delta$  181.7 (C=S, C), 138.4 (C, C-1'), 134.5 (C, C-3'), 130.3 (CH, C-5'), 128.2 (CH, C-4'), 126.8 (CH, C-2'), 124.5 (CH, C-6'), 66.3 (CH<sub>2</sub>, C-3), 66.3 (CH<sub>2</sub>, C-5), 50.1 (CH<sub>2</sub>, C-2), 50.1 (CH<sub>2</sub>, C-6); EI MS:  $m/z$  (rel. abund%) 256.

*N*-Phenyl-4-(pyridin-2-yl)piperazine-1-carbothioamide (**18**) Yield: 84 %;  $^1\text{H}$  NMR: (500 MHz, DMSO- $d_6$ ):  $\delta$  8.15 (dd, 1H,  $J = 1.5$ ,  $J = 5.0$  Hz, H-6''), 7.62 (ddd, 1H,  $J = 5.0$ ,  $J = 5.0$ ,  $J = 1.5$  Hz, H-5''), 7.39 (d, 2H,  $J = 8.5$  Hz, H-2'/H-6'), 7.25 (t, 2H,  $J = 8.5$  Hz, H-3'/H-5'), 7.06 (t, 1H,  $J = 7.6$  Hz, H-4'), 6.88 (d, 1H,  $J = 8.4$  Hz, H-3''), 6.72 (dd, 1H,  $J = 8.0$ ,  $J = 1.5$  Hz, H-4''), 3.62 (t, 4H,  $J = 9.2$  Hz, 2-CH<sub>2</sub>/6-CH<sub>2</sub>), 3.59 (t, 4H,  $J = 9.2$  Hz, 3-CH<sub>2</sub>/5-CH<sub>2</sub>);  $^{13}\text{C}$  NMR (75 MHz, DMSO- $d_6$ ):  $\delta$  181.9 (C=S, C), 158.2 (C, C-2''), 148.0 (CH, C-6''), 138.6 (C, C-1'), 138.4 (CH, C-4''), 128.9 (CH, C-3'), 128.9 (CH, C-5'), 128.3 (CH, C-4'), 126.4 (CH, C-2'), 126.4 (CH, C-6'), 117.8 (CH, C-5''), 106.3 (CH, C-3''), 56.9 (CH<sub>2</sub>, C-2), 56.9 (CH<sub>2</sub>, C-6), 46.2 (CH<sub>2</sub>, C-3), 46.2 (CH<sub>2</sub>, C-5); EI MS:  $m/z$  (rel. abund%) 298.

*N*-(3-Chlorophenyl)-4-(pyridin-2-yl)piperazine-1-carbothioamide (**19**) Yield: 83 %;  $^1\text{H}$  NMR: (500 MHz, DMSO- $d_6$ ):  $\delta$  8.12 (dd, 1H,  $J = 5.0$ ,  $J = 1.5$  Hz, H-6''), 7.62 (ddd, 1H,  $J = 5.0$ ,  $J = 5.0$ ,  $J = 1.5$  Hz, H-5''), 7.55 (t, 1H,  $J = 4.5$  Hz, H-2'), 7.29 (dd, 1H,  $J = 7.0$ ,  $J = 2.0$  Hz,

H-4'), 7.26 (t, 1H,  $J = 8.0$  Hz, H-5'), 7.02 (ddd 1H,  $J = 8.0$ ,  $J = 1.8$ ,  $J = 2.0$  Hz, H-6'), 6.86 (d, 1H,  $J = 8.8$  Hz, H-3''), 6.74 (dd, 1H,  $J = 7.0$ ,  $J = 5.0$  Hz, H-4''), 3.69 (t, 4H,  $J = 10.0$  Hz, 2-CH<sub>2</sub>/6-CH<sub>2</sub>), 3.56 (t, 4H,  $J = 10.0$  Hz, 3-CH<sub>2</sub>/5-CH<sub>2</sub>); <sup>13</sup>C NMR (75 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  181.7 (C=S, C), 158.2 (C, C-2''), 148.0 (CH, C-6''), 138.4 (C, C-1'), 138.2 (CH, C-4''), 134.5 (C, C-3'), 130.3 (CH, C-5'), 128.2 (CH, C-4'), 126.8 (CH, C-2'), 124.5 (CH, C-6'), 117.8 (CH, C-5''), 106.1 (CH, C-3''), 56.6 (CH<sub>2</sub>, C-2), 56.6 (CH<sub>2</sub>, C-6), 46.2 (CH<sub>2</sub>, C-3), 46.2 (CH<sub>2</sub>, C-5); EI MS: *m/z* (rel. abund%) 332.

**4-Acetyl-N-phenylpiperazine-1-carbothioamide (20)** Yield: 84 %; <sup>1</sup>H NMR: (500 MHz, Methanol-*d*<sub>4</sub>):  $\delta$  7.37 (dd, 2H,  $J = 7.6$ ,  $J = 1.8$ , Hz, H-2'/H-6'), 7.32 (t, 2H,  $J = 8.5$  Hz, H-3'/H-5'), 7.07 (t, 1H,  $J = 8.5$  Hz, H-4'), 3.67 (t, 4H,  $J = 10.0$  Hz, 2-CH<sub>2</sub>/6-CH<sub>2</sub>), 3.56 (t, 4H,  $J = 10.0$  Hz, 3-CH<sub>2</sub>/5-CH<sub>2</sub>), 2.12 (s, 3H, CH<sub>3</sub>); <sup>13</sup>C NMR (75 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  181.9 (C=S, C), 168.7 (C=O, C), 138.4 (C, C-1'), 129.1 (CH, C-3'), 129.1 (CH, C-5'), 128.5 (CH, C-4'), 126.4 (CH, C-2'), 126.4 (CH, C-6'), 56.7 (CH<sub>2</sub>, C-2), 56.7 (CH<sub>2</sub>, C-6), 49.4 (CH<sub>2</sub>, C-3), 49.4 (CH<sub>2</sub>, C-5), 21.2 (CH<sub>3</sub>, C); EI MS: *m/z* (rel. abund%), 263.

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## References

- Ahmad S, Hughes MA, Yeh LA, Scott JE (2012) Potential repurposing of known drugs as potent bacterial  $\beta$ -glucuronidase inhibitors. *J Biomol Screen* 17:957–965
- Ala JP, DeLoskey RJ, Huston EE, Jadhav PK, Lam PYS, Eyermann CJ, Hodge CN, Schadt MC, Lewandowski FA, Weber PC, McCabe DD, Duke JL, Chang CH (1998) Molecular recognition of cyclic urea HIV-1 protease inhibitors. *J Biol Chem* 273:12325–12331
- Anouar EH, Raweh S, Bayach I, Taha M, Baharudin MS, Meo FD, Hasan MH, Adam A, Ismail NH, Weber JF, Trouillas P (2013) Antioxidant properties of phenolic Schiff bases: structure-activity relationship and mechanism of action. *J Comput Aided Mol Des* 27:951–964
- Aziz AN, Taha M, Ismail NH, Anouar EH, Yousuf S, Jamil W, Awang K, Ahmat N, Khan KM, Kashif SM (2014) Synthesis, crystal structure, DFT studies and evaluation of the antioxidant activity of 3,4-Dimethoxybenzamine schiff bases. *Molecules* 19:8414–8433
- Bäckbro K, Löwgren S, Österlund K, Atepo J, Unge T (1997) Unexpected binding mode of a cyclic sulfamide HIV-1 protease inhibitor. *J Med Chem* 40:898–902
- Bank N, Bailine SH (1965) Urinary  $\beta$ -glucuronidase activity in patients with urinary-tract infection. *N Engl J Med* 272:70–75
- Bloom JD, Dushin RG, Curran KJ, Donahue F, Norton EB, Terefenko E, Jonas TR, Ross AA, Feld B, Lang SA, Grandi D (2004) *Bioorgan Med Chem* 14:3401–3406
- Boyland E, Gasson JE, Williams DC (1957) Enzyme activity in relation to cancer; the urinary  $\beta$ -glucuronidase activity of patients suffering from malignant disease. *Br J Cancer* 11:120–129
- Caygill JC, Pitkeathly DA (1966) A study of  $\beta$ -acetylglucosaminase and acid phosphatase in pathological joint fluids. *Ann Rheum Dis* 25:137–144
- Cheng TC, Chuang KH, Roffler SR, Cheng KW, Leu YL, Chuang CH, Huang CC, Kao CH, Hsieh YC, Chang LS, Cheng TL, Chen CS (2015) Discovery of specific inhibitors for intestinal *E. coli*  $\beta$ -Glucuronidase through in silico virtual screening. *Sci World J* 2015:740815. doi:10.1155/2015/740815
- Chrusciel RA, Strohbach JW (2004) Non-peptidic HIV protease inhibitors. *Curr Top Med Chem* 4:1097–1114
- Flieger J, Żelazko AC, Rządowska M, Szacoń E, Matusiuk D (2012) Usefulness of reversed-phase HPLC enriched with room temperature imidazolium based ionic liquids for lipophilicity determination of the newly synthesized analgesic active urea derivatives. *J Pharm Biomed Anal* 66:58–67
- Fortin JS, Lacroix J, Desjardins M (2007) *N*-Phenyl-*N'*-(2-chloroethyl)urea analogs of combretastatin A-4: is the *N*-phenyl-*N'*-(2-chloroethyl)urea pharmacophore mimicking the trimethoxy phenyl moiety. *Bioorgan Med Chem* 15:4456–4469
- Fortin S, Wei L, Moreau E, Labrie P, Petitclerc É, Kotra LP, Gaudreault RC (2009) Mechanism of action of *N*-phenyl-*N'*-(2-chloroethyl)ureas in the colchicine-binding site at the interface between  $\alpha$ - and  $\beta$ -tubulin. *Bioorgan Med Chem* 17:3690–3697
- Gonick HC, Kramer HJ, Schapiro AE (1973) Urinary  $\beta$ -glucuronidase activity in renal disease. *Arch Intern Med* 132:63–69
- Holešová S, Valášková M, Hlaváč D, Madejová J, Samlíková M, Tokarský J, Pazdziora E (2014) Antibacterial kaolinite/urea/chlorhexidine nanocomposites: experiment and molecular modeling. *Appl Surf Sci* 305:783–791
- Hultén J, Bonham NM, Nilroth U, Hansson T, Zuccarello G, Bouzide A, Åqvist J, Classon B, Danielson HA, Karlén Kvarnstrom I, Samuelsson B, Hallberg A (1997) Cyclic HIV-1 protease inhibitors derived from mannitol: synthesis, inhibitory potencies, and computational predictions of binding affinities. *J Med Chem* 40:885–889
- Jamil W, Perveen S, Shah SAA, Taha M, Ismail NH, Perveen S, Ambreen N, Khan KM, Choudhary MI (2014) Phenoxyacetylhydrazide schiff bases:  $\beta$ -Glucuronidase inhibitors. *Molecules* 19:8788–8802
- Kallet HA, Lapco L (1967) Urine  $\beta$ -glucuronidase activity in urinary tract disease. *J Urol* 97:352–356
- Khan KM, Ali M, Taha M, Perveen S, Choudhary MI, Voelter W (2008) An expedient and selective approach towards disulfides using sodium bromate/sodium hydrogen sulfite reagent. *Lett Org Chem* 5:432–434
- Khan KM, Taha M, Ali M, Perveen S (2009) A mild and alternative approach towards symmetrical disulfides using H<sub>3</sub>IO<sub>5</sub>/NaHSO<sub>3</sub> combination. *Lett Org Chem* 6:319–320
- Khan KM, Taha M, Rahim F, Ali M, Jamil W, Perveen S, Choudhary MI (2010) An improved method for the synthesis of disulfides by periodic acid and sodium hydrogen sulfite in water. *Lett Org Chem* 7:244
- Khan KM, Taha M, Naz F, Khan M, Rahim F, Samreen Perveen S, Choudhary MI (2011) Synthesis and in vitro leishmanicidal activity of disulfide derivatives. *Med Chem* 7:704–710
- Khan KM, Taha M, Naz F, Ali S, Perveen S, Choudhary MI (2012) Acylhydrazide schiff bases: DPPH radical and superoxide anion scavengers. *Med Chem* 8:705–710
- Khan KM, Naz F, Taha M, Khan A, Perveen S, Choudhary MI, Voelter W (2014a) Synthesis and in vitro urease inhibitory activity of *N*, *N'*-disubstituted thioureas. *Eur J Med Chem* 74:314–323

- Khan KM, Rahim F, Wadood A, Taha M, Khan M, Naureen S, Ambreen N, Hussain S, Perveen S, Choudhary MI (2014b) Evaluation of bisindole as potent  $\beta$ -Glucuronidase inhibitors: synthesis and in silico based studies. *Bioorgan Med Chem Lett* 24:1825–1829
- Khan KM, Saad SM, Shaikh NN, Hussain S, Fakhri MI, Perveen S, Taha M, Choudhary MI (2014c) Synthesis and  $\beta$ -glucuronidase inhibitory activity of 2-arylquinazolin-4(3H)-ones. *Bioorgan Med Chem* 22:3449–3454
- Khan KM, Ambreen N, Taha M, Halim SA, Zaheer-ul-Haq Naureen S, Rasheed S, Perveen S, Ali S, Choudhary MI (2014d) Structure-based design, synthesis and biological evaluation of  $\beta$ -Glucuronidase inhibitors. *J Comput Aided Mol Des* 28: 577–585
- Lee J, Kang M, Shin M, Kim JM, Kang SU, Lim JO, Choi HK (2003) *N*-(3-acyloxy-2-benzylpropyl)-*N'*-[4-(methylsulfonylamino)benzyl]thiourea analogs: novel potent and high affinity antagonists and partial antagonists of the vanilloid receptor. *J Med Chem* 46:3116–3126
- Liu Z, Wang Y, Lin H, Zuo D, Wang L, Zhao Y, Gong P (2014) Design, synthesis and biological evaluation of novel thieno[3,2-d]pyrimidine derivatives containing diaryl urea moiety as potent antitumor agents. *Eur J Med Chem* 85:215–227
- Musharraf SG, Bibi A, Shahid N, Najam-ul-Haq M, Khan M, Taha M, Mughal UR, Khan KM (2012) Acylhydrazide and isatin schiff bases as alternate UV laser desorption ionization (LDI) matrices for low molecular weight (LMW) peptides analysis. *Am J Anal Chem* 3:779–789
- Plum CM (1967)  $\beta$ -glucuronidase activity in serum, cerebrospinal fluid and urine in normal subjects and in neurological and mental patients. *Enzymol Biol Clin* 8:97–112
- Rahim F, Ullah K, Ullah H, Wadood A, Taha M, Rehman AU, Uddin I, Ashraf M, Shaikat A, Rehman W, Hussain S, Khan KM (2015) Triazinoindole analogs as potent inhibitors of  $\alpha$ -glucosidase: synthesis, biological evaluation and molecular docking studies. *Bioorg Chem* 58:81–87
- Reddy BS (1976) Dietary factors and cancer of the large bowel. *Semin Oncol* 3:351–359
- Roberts AP, Frampton J, Karim SM, Beard RW (1967) Estimation of  $\beta$ -glucuronidase activity in urinary-tract infection. *N Engl J Med* 276:1468–1470
- Ronald AR, Silverblatt F, Clark H, Cutler RE, Turck M (1971) Failure of urinary  $\beta$ -glucuronidase activity to localize the site of urinary tract infection. *Appl Environ Microbiol* 21:990–992
- Santos LD, Lima LA, Cechinel-Filho V, Corrêa R, Buzzi FC, Nunes RJ (2008) Synthesis of new 1-phenyl-3-{4-[(2*E*)-3-phenylprop-1-enyl]phenyl}-thiourea and urea derivatives with antinociceptive activity. *Bioorgan Med Chem* 16:8526–8534
- Schapiro A, Paul W, Gonick H (1968) Urinary  $\beta$ -glucuronidase in urologic diseases of the kidneys. *J Urol* 100:146–157
- Seth PP, Ranken R, Robinson DE, Osgood SA, Risen LM, Rodgers EL, Migawa MT, Jefferson EA, Swayze EE (2004) Aryl urea analogs with broad-spectrum antibacterial activity. *Bioorgan Med Chem* 14:5569–5572
- Sham HL, Zhao C, Marsh KC, Betebenner DA (1996a) Novel azacyclic ureas that are potent inhibitors of HIV-1 protease. *J Biochem Biophys Res Commun* 225:436–440
- Sham HL, Zhao C, Stewart KD, Betebenner DA, Lin S (1996b) A novel, picomolar inhibitor of human immunodeficiency virus type 1 protease. *J Med Chem* 39:392–397
- Sivan SK, Vangala R, Manga V (2013) Molecular docking guided structure based design of symmetrical *N, N'*-disubstituted urea/thiourea as HIV-1 gp120–CD4 binding inhibitors. *Bioorgan Med Chem* 21:4591–4599
- Sperker B, Backman JT, Kromer K (1997) The role of  $\beta$ -glucuronidase in drug disposition and drug targeting in humans. *Clin Pharm* 33:18–31
- Taha M, Ismail NH, Jamil W, Yousuf S, Jaafar FM, Ali MI, Kashif SM, Hussain E (2013) Synthesis, evaluation of antioxidant activity and crystal structure of 2,4-Dimethylbenzoylhydrazones. *Molecules* 18:10912–10929
- Taha M, Naz H, Rasheed S, Ismail NH, Rahman AA, Yousuf S, Choudhary MI (2014) Synthesis of 4-Methoxybenzoylhydrazones and evaluation of their antiglycation activity. *Molecules* 19:1286–1301
- Taha M, Ismail NH, Lalani S, Fatmi MQ, Atiahab Siddiqui S, Khan KM, Imran S, Choudhary MI (2015a) Synthesis of novel inhibitors of  $\alpha$ -glucosidase based on the benzothiazole skeleton containing benzohydrazide moiety and their molecular docking studies. *Eur J Med Chem* 92:387–400
- Taha M, Ismail NH, Baharudin MS, Lalani S, Mehboob S, Khan KM, Yousuf S, Siddiqui S, Rahim F, Choudhary MI (2015b) Synthesis crystal structure of 2-methoxybenzoylhydrazones and evaluation of their  $\alpha$ -glucosidase and urease inhibition potential. *Med Chem Res* 24:1310–1324
- Venkatachalam TK, Mao C, Uckun FM (2004) Effect of stereochemistry on the anti-HIV activity of chiral thiourea compounds. *Bioorgan Med Chem* 12:4275–4284
- Watson RJ, Allen DR, Birch HL, Chapman GA, Gayle A, Knight LA, Oliver K, Owen DA, Thomas EJ, Tremayne N, Williams SC (2008) Development of CXCR3 antagonists. Part 3: tropenyl and homotrophenyl-piperidine urea derivatives. *Bioorgan Med Chem Lett* 18:147–151
- Yang W, Liu H, Li M, Wang F, Zhou W, Fan J (2012) Synthesis, structures and antibacterial activities of benzoylthiourea derivatives and their complexes with cobalt. *J Inorg Biochem* 116: 97–105
- Zhao C, Sham HL, Sun M, Stoll VS, Stewart KD, Lin S, Mo H (2005) Synthesis and activity of *N*-acyl azacyclic urea HIV-1 protease inhibitors. *Bioorgan Med Chem Lett* 15:549–555