Ancillary ligand control of reactivity. Protonation at hydride vs. cyanide in trans-[FeH(CN)($R_2PCH_2CH_2PR_2$)₂] (R = Et, Ph, p-tolyl) and X-ray crystal structure determination of trans-[FeH(CNH)($R_2PCH_2CH_2PR_2$)₂]BF₄ (R = p-tolyl)

Patrick I. Amrhein, Samantha D. Drouin, Cameron E. Forde, Alan J. Lough and Robert H. Morris*

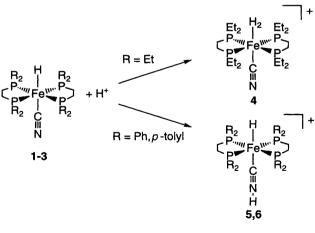
Department of Chemistry, University of Toronto, 80 St. George Street, Toronto, Ontario, Canada M5S 1A1

The ancillary ligands in *trans*-[FeH(CN)L₂] control whether protonation occurs at hydride to produce a dihydrogen complex or at cyanide to produce a hydrogen isocyanide complex.

In a situation where more than one product is possible in a reaction involving a transition-metal complex, judicious selection of the ancillary ligands and reaction conditions can direct the reaction pathway toward the desired product. Just such a system is observed in the protonation of *trans*-[FeH(CN)L₂] [L = depe (1), dppe (2), dtpe (3)].† The products of the protonation of 1–3 indicate that H⁺ adds to the Fe–H bond in 1 while it adds to the cyanide nitrogen in 2 and 3 (Scheme 1). The potential for protonation to occur at cyanide *vs.* hydride at an iron centre may exist in [NiFe]-hydrogenases where cyanide coordination on iron at the active site has been proposed.^{1,2} Tautomeric dihydrogen systems have been reported for [Os(H₂)(η^2 -S₂CH)-(CO)(PPrⁱ₃)₂]BF₄.³ and [Os(H₂)(η^2 -quinS)(CO)(PPh₃)₂]BF₄.⁴

Metathesis of chloride in *trans*-[FeH(Cl)L₂] (L = depe,⁵ dppe,⁶ dtpe⁷) for cyanide produces the complexes *trans*-[FeH(CN)L₂] [L = depe (1), dppe (2), dtpe (3)‡]. The hydridocyanide complexes 1–3 are characterized by ¹H and ³¹P NMR spectra and FABMS. The FABMS of 1 gives evidence that 1 is protonated to give 4 in the gas phase.

Protonation of *trans*-[FeH(CN)(depe)₂] **1** with 85% [Et₂OH]BF₄ in Et₂O under H₂ (1 atm) produces a yellow precipitate. The CD₂Cl₂ solution of this solid contains the dihydrogen compound *trans*-[Fe(H₂)(CN)(depe)₂]BF₄ **4**.§ This complex slowly decomposes to other species. In the ¹H NMR spectrum of this complex in CD₂Cl₂ a broad resonance is observed to high field (δ -14.05) due to the dihydrogen ligand. Measurement of the minimum spin-lattice relaxation time, *T*₁ = 15.2 ms (400 MHz, 229 K), allows the H–H separation to be



Scheme 1

calculated⁸ at 0.85 or 1.07 Å for a fast or slow spinning dihydrogen ligand, respectively.

The one-bond H–D coupling constant of 31.6 Hz was measured from the spectrum of the complex *trans*-[Fe(HD)(CN)(depe)₂]BF₄, **4'**, prepared by reacting **1** with [Ph₃PD]BF₄ in CD₂Cl₂. The coupling between the hydrogen and the four equivalent phosphorus nuclei is also observable in this system and a 1:1:1 triplet of quintets is observed, ${}^{2}J_{PH} =$ 5.8 Hz. By using an empirical correlation⁹ between ${}^{1}J_{HD}$ and d_{HH} a separation of 0.89 Å is calculated. This is consistent with the result of the T_1 determination if the rotational frequency of the dihydrogen ligand is much greater than the spectrometer frequency (400 MHz). Complex **4** represents the first iron dihydrogen complex of the form *trans*-[Fe(H₂)XL₂]⁺ to be prepared where X is not hydride.^{10,11} The related complex *trans*-[Ru(H₂)(CCPh)(dippe)₂]⁺ has been reported.¹² Addition of 1 equiv. of 85% [Et₂OH]BF₄ to *trans*-

[FeH(CN)(dppe)₂] 2 or trans-[FeH(CN)(dtpe)₂] 3 in Et₂O under H_2 (1 atm) results in yellow precipitates which in CD_2Cl_2 solution contain the coordinated hydrogen isocyanide comtrans-[FeH(CNH)(dppe)₂]BF₄ plexes 5,¶ or trans-[FeH(CNH)(dtpe)₂]BF₄ 6, respectively. A quintet resonance is observed to high field for the hydride ligands in the ¹H NMR spectrum of each compound. A broad doublet resonance with ${}^{2}J_{HC}$ = 98 Hz for the hydrogen isocyanide ligand is observed in the proton-coupled ${}^{13}C$ NMR spectrum of *trans*the $[FeH(^{13}CNH)(dppe)_2]BF_4$ 5'. The magnitude of $^2J_{HC}$ confirms the assignment of the ligand as CNH as a much larger one-bond coupling would be anticipated for an NCH ligand.13

In order to unequivocally determine the coordination mode of the hydrogen isocyanide ligand a single-crystal X-ray diffraction study of 6 was performed.** The coordination sphere about the iron (Fig. 1) is occupied by a square plane of P atoms and a C atom from the CNH ligand trans to the hydride ligand which was located and refined with an isotropic thermal parameter. The molecule has twofold crystallographic symmetry. The BF_4^- appears to be positioned in order to hydrogen bond with the NH, but the hydrogen was not located due to disorder. The F...N separation of 2.9 Å is less than the sum of the van der Waals radii of 3.1 Å. The coordination mode was determined to be Fe-CNH based on an analysis of the thermal ellipsoids and comparison with the other known hydrogen isocyanide structures. A search of the Cambridge Structural Database¹⁴ revealed that the M-C separations in hydrogen isocyanide complexes^{15,16} are ≤ 2.0 Å while the M–N separations in hydrogen cyanide^{17–21} (formonitrile) complexes are >2.3 Å. The Fe–C separation in 6 is 1.842(6) Å and the HFeCN unit is linear as required by symmetry.

The change in the site of protonation in these complexes can be explained by considering the differences in acidities between the hydrogen isocyanide complex **5** and the hypothetical complex *trans*-[FeH(CNH)(depe)₂]⁺ **7** ($\Delta p K_a^{CNH}$), and between the dihydrogen complex **4** and the hypothetical complex *trans*-[Fe(H₂)(CN)(dppe)₂]⁺ **8** ($\Delta p K_a^{H2}$). The change in the

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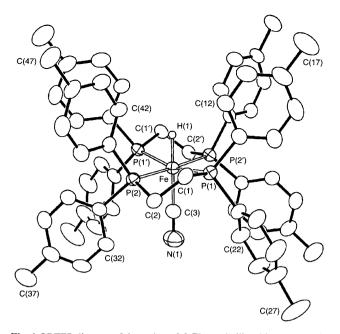


Fig. 1 ORTEP diagram of the cation of 6. Thermal ellipsoids represent the 50% probability surface. Selected bond lengths (Å) and angles (°): Fe–H(1) 1.39(6), Fe–C(3) 1.842(6), C(3)–N(1) 1.183(8), Fe–P(1) 2.2557(10), Fe–P(2) 2.335(9), H(1)–Fe–C(3) 180.0, Fe–C(3)–N(1) 180.0.

acidity of the dihydrogen ligand by changing ancillary ligands from depe to dppe is predicted to be 7 pK_a units $[\Delta pK_a^{H2} = pK_a^{H2}(4) - pK_a^{H2}(8)]$.^{††} The effect is large because the acidic proton in 4 or 8 is located three bonds away from the site where the change of substituent on phosphine occurs.²² The change in acidities of the hydrogen isocyanide ligands in 5 or 7 is expected to be much smaller because the acidic proton is five bonds removed from the substituent change. The change of ancillary ligand from depe to dppe lowers the pK_a of the hypothetical dihydrogen compound 8 below that of the hydrogen isocyanide complex 5.

In conclusion the site of protonation of trans-[FeH(CN)L₂] can be directed to hydride (producing a dihydrogen ligand) or to cyanide (producing a hydrogen isocyanide ligand) by controlling the nature of the ancillary ligands.

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Footnotes

+ Abbreviations used: quinS = 8-quinolinethiolate, depe = 1,2-bis(diethylphosphino)ethane, dppe = 1,2-bis(diphenylphosphino)ethane, dtpe = 1,2-bis(di-*p*-tolylphosphino)ethane, dippe = 1,2-bis(diisopropylphosphino)ethane.

‡ The characterisation of these complexes will be reported elsewhere.

§ trans-[Fe(H₂)(CN)(depe)₂]BF₄ 4. Addition of 1 equiv. of acid {85% [Et₂OH]BF₄ in Et₂O or [Ph₃PH]BF₄ in CD₂Cl₂} to 1 produced 4 as revealed by NMR. ¹H NMR (CD₂Cl₂): δ –14.05 [br s, FeH₂, *T*₁ (min, 400 MHz) 15.2 ms]. trans-[Fe(HD)(CN)(depe)₂]BF₄ 4'. 1 equiv. of [Ph₃PD]BF₄ was added to 1 in CD₂Cl₂. ¹H NMR (CD₂Cl₂): δ –14.08 (1 : 1 : 1 t of qut, FeHD, ¹J_{DH} 31.6 Hz, ²J_{PH} 5.8 Hz). trans-[Fe(H₂)(¹³CN)(depe)₂]BF₄ 4". Preparation is as above for 4 except [FeH(¹³CN)(depe)₂] was employed. ³¹P[⁴H]-NMR (CD₂Cl₂): δ 77.7 (d, ²J_{CP} 17 Hz). ¹³C NMR (CD₂Cl₂): δ 145.6 (m, FeCN.

¶ trans-[FeH(CNH)(dppe)₂]BF₄ **5**. Protonation of **2** (100 mg, 0.11 mmol) in Et₂O (8 ml) under H₂ with 1.1 equiv. of 85% [Et₂OH]BF₄ (20 μ l, 0.12 mmol) in diethyl ether produced **5**. ¹H NMR (CD₂Cl₂): $\delta -10.7$ (qnt, FeH, ²J_{PH} 49 Hz). FABMS *m*/*z* 880 (calc. 880). trans-[FeH(CNH)(dppe)₂]BF₄ **5**'. Preparation is as for **5** except [FeH(¹³CN)(dppe)₂] was used. ³¹P{¹H} NMR (CD₂Cl₂): δ 88.0 (d, ²J_{CP} 16 Hz). ¹³C NMR (CD₂Cl₂): δ 179.4 (br d, ²J_{HC} 98 Hz, CNH).

|| *trans*-[FeH(CNH)(dtpe)₂]BF₄ **6**. Protonation of **3** with 1 equiv. of 85% [Et₂OH]BF₄ in Et₂O under H₂ produced **6**. ³¹P{¹H} NMR (CD₂Cl₂): δ 86.5 (s). ¹H NMR (CD₂Cl₂): δ −10.78 (qnt, FeH, ²*J*_{PH} 47 Hz). FABMS *m*/*z* 992 (calc. 992).

** Complex 6 crystallises in the orthorhombic space group *Pbcn* with unitcell parameters a = 25.700(4), b = 12.453(2), c = 19.151(3) Å, Z = 4. Atomic coordinates, bond lengths and angles, and thermal parameters have been deposited at the Cambridge Crystallographic Data Centre (CCDC). See Information for Authors, Issue No. 1. Any request to the CCDC for this material should quote the full literature citation and the reference number 182/134.

^{††} It is assumed that the complexes *trans*-[Fe(H₂)(CN)L₂]⁺ require the same energy of hydrogen abstraction to produce *trans*- [FeH(CN)L₂]⁺ and H·.²³ The difference in reduction potential of *trans*-[FeH(CN)L₂]⁺ for L = depe vs. dppe is expected to be $\Delta E_{1/2} = -0.4$ V.^{23,24} Therefore $\Delta pK_a = -16.9 \ \Delta E_{1/2} = 7.^{23}$

References

- 1 K. A. Bagley, C. J. van Garderen, M. Chen, E. C. Duin, S. P. Albracht and W. H. Woodruff, *Biochemistry*, 1994, **31**, 9229.
- 2 K. A. Bagley, E. C. Duin, W. Roseboom, S. P. J. Albracht and W. H. Woodruff, *Biochemistry*, 1995, 34, 5527.
- 3 M. J. Albéniz, M. L. Buil, M. A. Esteruelas, A. M. López, L. A. Oro and B. Zeier, *Organometallics*, 1994, 13, 3746.
- 4 M. Schlaf and R. H. Morris, J. Chem. Soc., Chem. Commun., 1995, 625.
- 5 J. Chatt and R. G. Hayter, J. Chem. Soc., 1961, 5507.
- 6 P. Giannoccaro and A. Sacco, Inorg. Synth., 1977, 17, 69
- 7 E. P. Cappellani, S. D. Drouin, G. Jia, P. A. Maltby, R. H. Morris and C. T. Schweitzer, J. Am. Chem. Soc., 1994, 116, 3375.
- 8 K. A. Earl, G. Jia, P. A. Maltby and R. H. Morris, J. Am. Chem. Soc., 1991, 113, 3027.
- 9 P. A. Maltby, M. Schlaf, M. Steinbeck, A. J. Lough, R. H. Morris, W. T. Klooster, T. F. Koetzle and R. C. Srivastava, J. Am. Chem. Soc., 1996, 118, in the press.
- 10 D. M. Heinekey and W. J. Oldham, Chem. Rev., 1993, 93, 913.
- 11 P. G. Jessop and R. H. Morris, Coord. Chem. Rev., 1992, 121, 155.
- 12 M. Jimenez-Tenorio, M. C. Puerta and P. Valerga, J. Chem. Soc., Chem. Commun., 1993, 1750.
- 13 G. J. Schrobilgen, J. Chem. Soc., Chem. Commun., 1988, 863.
- 14 F. H. Allen, O. Kennard and R. Taylor, Chem. Des. Autom. News, 1993, 8, 31.
- 15 E. Bar, J. Fuchs, D. Rieger, F. Aguilar-Parrilla, H.-H. Limbach and W. P. Fehlhammer, Angew. Chem., Int. Ed. Engl., 1991, 30, 88.
- 16 D. Rieger, E. Hahn and W. P. Fehlhammer, J. Chem. Soc., Chem. Commun., 1990, 285.
- 17 G. Constant, J.-C. Daran, Y. Jeannin and R. Morancho, J. Coord. Chem., 1973, 2, 203.
- 18 C. Chavant, G. Constant, Y. Jeannin and R. Morancho, Acta Crystallogr., Sect. B, 1975, 31, 1823.
- 19 J. P. Smit, W. Purcell, A. Roodt and J. G. Leipoldt, J. Chem. Soc., Chem. Commun., 1993, 1388.
- 20 P. G. Jones, H. W. Roesky and J. Schimkowiak, J. Chem. Soc., Chem. Commun., 1988, 730.
- 21 G. Constant, J. C. Daran and Y. Jeannin, Acta Crystallogr., Sect. B, 1971, 27, 2388.
- 22 D. D. Perrin, B. Dempsey and E. P. Serjeant, *pKa Prediction for Organic Acids and Bases*, Chapman and Hall, London, 1981.
- 23 R. H. Morris, Inorg. Chem., 1992, 31, 1471.
- 24 A. B. P. Lever, Inorg. Chem., 1990, 29, 1271

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