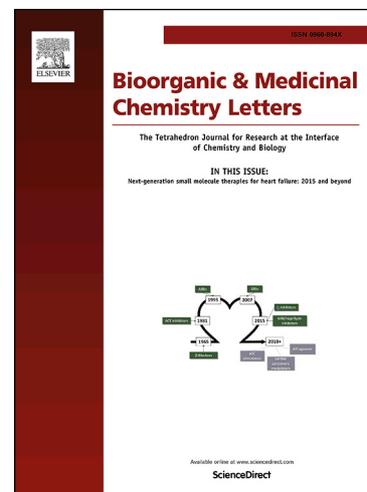


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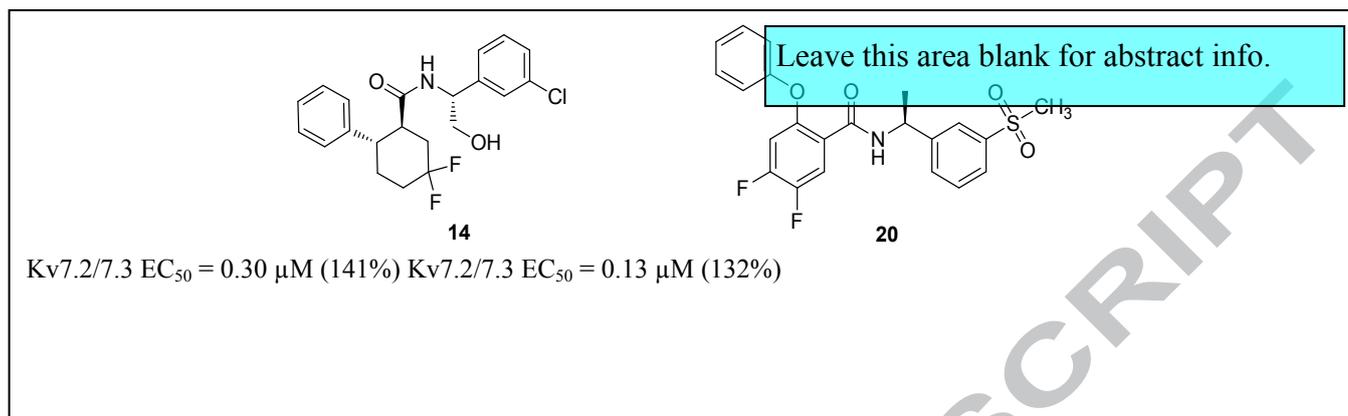
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## Novel K<sub>V</sub>7 ion channel openers for the treatment of epilepsy and implications for detrusor tissue contraction

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## ABSTRACT

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Neuronal voltage-gated potassium channels, K<sub>V</sub>7s, are the molecular mediators of the M current and regulate membrane excitability in the central and peripheral neuronal systems. Herein, we report novel small molecule K<sub>V</sub>7 openers that demonstrate anti-seizure activities in electroshock and pentylenetetrazol-induced seizure models without influencing Rotarod readouts in mice. The anti-seizure activity was determined to be proportional to the unbound concentration in the

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brain.  $K_v7$  channels are also expressed in the bladder smooth muscle (detrusor) and activation of these channels may cause localized undesired effects. Therefore, the impact of individual  $K_v7$  isoforms was investigated in human detrusor tissue using a panel of  $K_v7$  openers with distinct activity profiles among  $K_v7$  isoforms. *KCNQ4* and *KCNQ5* mRNA were highly expressed in detrusor tissue, yet a compound that has significantly reduced activity on homomeric  $K_v7.4$  did not reduce detrusor contraction. This may suggest that the homomeric  $K_v7.4$  channel plays a less significant role in bladder contraction and further investigation is needed.

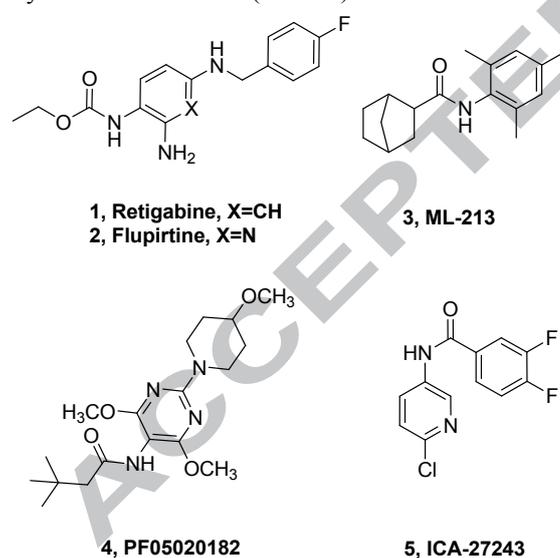
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Neuronal voltage-gated potassium channels composed of the following heterodimeric  $K_v7$  isoforms,  $K_v7.2$ , 7.3, 7.4, and 7.5, are coded by *KCNQ2-5*, respectively.  $K_v7$  ion channels are the molecular mediators of the M current and regulate membrane excitability in the central and peripheral neuronal systems.<sup>1</sup> Genetic mutations in *KCNQ3* and *KCNQ4*, are associated with excitability disorders including multiple forms of epilepsy and deafness.<sup>2, 3</sup> Therefore,  $K_v7$  channel opening is a potential therapeutic approach for aberrant neuroexcitation conditions.<sup>4</sup>

Previously reported small molecule  $K_v7$ -openers suppress neuronal activity at sub-micromolar concentrations *in vitro*. Suppression of neuronal activities with  $K_v7$ -openers translate to functional endpoints in a wide range of pre-clinical models.<sup>5</sup> In human clinical studies, Retigabine (**1**) and Flupirtine (**2**) (Figure 1) that act on pan- $K_v7.2-7.5$  channels demonstrated clinical efficacy in partial epilepsy<sup>6</sup> and multiple forms of pain,<sup>7</sup> respectively. However, these molecules have undesired effects including of discolouration to the skin, nails, lips and ocular tissues (Retigabine)<sup>8</sup> and hepatotoxicity (Flupirtine)<sup>9, 10</sup> that are suspected as  $K_v7$ -unrelated actions. Furthermore, the frequency of urinary adverse events such as urinary retention was increased in patients receiving Retigabine compared with placebo in clinical studies.<sup>11</sup> Urinary adverse events were suspected as target-related because some *KCNQ* isoforms are expressed in human urinary bladder smooth muscle<sup>12</sup> and pan- $K_v7$  openers affected bladder contractibility *ex vivo* in human<sup>12</sup> and guinea pig.<sup>13</sup>

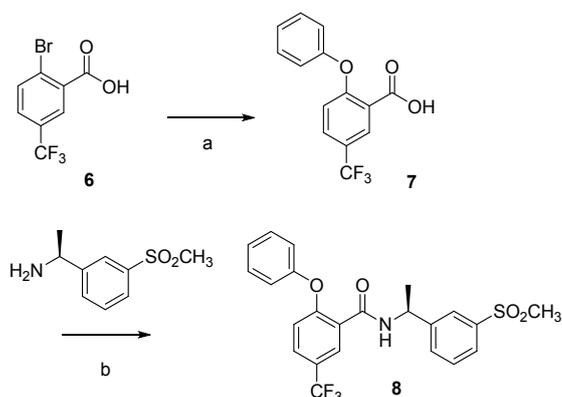
While  $K_v7$  channel openers have demonstrated clinical efficacy, improved therapeutic agents are desired. Discovery efforts leading to chemically distinct  $K_v7.2/7.3$  channel opener series (**1-5**, Figure 1) have been described in the literature<sup>5, 14-16</sup> and, except for the ICA-27243 (**5**) series of compounds, all have been reported to activate human pan- $K_v7$ -mediated currents. In this study, we aimed to discover novel  $K_v7$  isoform-selective openers for the treatment of epilepsy and to investigate the implications of  $K_v7$  isoform activities in human urinary bladder contraction.

A high throughput screen of the GSK compound collection using the heterodimeric  $K_v7.2/7.3$  channel (See Supplemental Information for methods of all biological experiments) revealed two similar, but structurally distinct lead series, Benzamide series (Table 1) and Cyclohexamide series (Table 2).



**Figure 1.** Representative  $K_v7.2/7.3$  openers

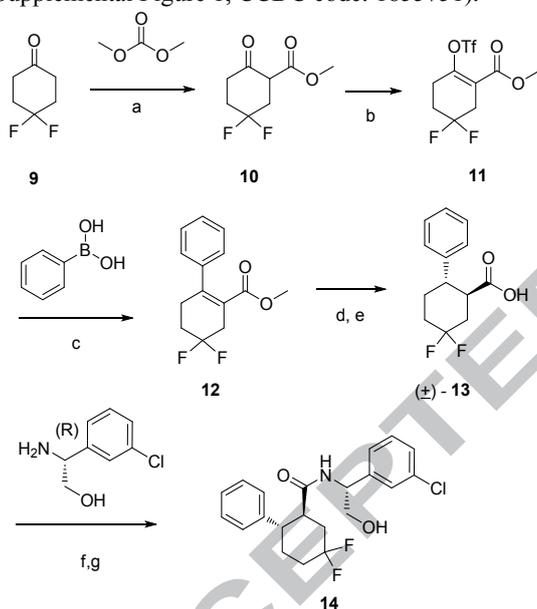
The synthesis of the benzamide series is depicted in scheme 1. Ortho-bromo benzoic acids **6** were coupled with phenolic reagents via an  $S_NAr$  reaction to provide biaryl ethers **7**. Subsequent amide coupling with chiral benzyl amines gave benzamide products (Table 1).



**Scheme 1.** Synthesis of benzamide derivatives.

Reagents and conditions: (a) Phenol, CuI, K<sub>2</sub>CO<sub>3</sub>, DMF, 130 °C, 16h; (b) HOBt, EDC, DIPEA, DMF, 60 °C, 2h

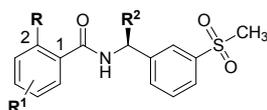
Cyclohexamide compounds (Scheme 2) were synthesized starting from 4,4-difluoro cyclohexanone (**9**) and dimethyl carbonate to afford ketoester **10**. Converting **10** to the corresponding enol triflate **11** and subsequent cross-coupling with phenylboronic acid gave the unsaturated aryl ester **12**. Hydrogenation followed by saponification under basic conditions provided the racemic *trans*-cyclohexyl carboxylic acid **13**. Amide coupling with appropriately substituted chiral benzyl amines yielded K<sub>v</sub>7.2/7.3 openers which were separated into pure diastereomers using chiral HPLC (Table 2). Only one enantiomer in the cyclohexamide series was determined to be consistently active in the K<sub>v</sub>7.2/7.3 primary assay and this absolute stereochemistry was confirmed by an X-ray crystal structure of compound **14** (Supplemental Figure 1, CCDC code: 1855751).



**Scheme 2.** Synthesis of aryl amide analogs.

Reagents and conditions: (a) NaH, DMF, 0 °C - RT, 16h; (b) Comins' reagent, NaH, THF, 0 °C - RT, 16h; (c) Pd(dppf)Cl<sub>2</sub>·CH<sub>2</sub>Cl<sub>2</sub>, Na<sub>2</sub>CO<sub>3</sub>, Toluene, H<sub>2</sub>O, EtOH, 90 °C, 16h; (d) H<sub>2</sub>, Pd/C, MeOH, RT 16h; (e) NaOEt, EtOH, 90 °C; (f) HATU, DIPEA, THF, RT, 16h; (g) Chiral prep HPLC.

In the benzamide series, thioether screening lead **15** (Table 1) was metabolically unstable in pooled human liver microsomes (HLM, >3 mL/min/g liver) as were phenyl ether analogs lacking electron withdrawing groups (EWGs) at R<sub>1</sub> (not shown). Alkyl substituents at R<sub>2</sub> were tolerated, however, unsubstituted phenyl ethers provided the highest level of K<sub>v</sub>7.2/7.3 activation. Functionality at R<sub>2</sub> was limited to methyl or hydroxymethyl with the specific stereochemistry shown. The methyl sulfone at the 3-position of the benzylic ring remained unchanged as it provided the best combination of solubility and K<sub>v</sub>7.2/7.3 activity.



**Table 1.** Benzamide series potencies (μM)

Cmpd R R<sup>1</sup> R<sup>2</sup> EC<sub>50</sub><sup>a</sup> Sol.(μM)<sup>b</sup>

**15** SPh 5-CF<sub>3</sub> H 0.20 (175) 34

**16** OPh 5-CF<sub>3</sub> H 0.16 (245) 23

**8** OPh 5-CF<sub>3</sub> CH<sub>3</sub> 0.04 (134) 57

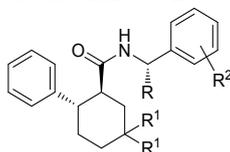
**17** OPh 5-CF<sub>3</sub> CH<sub>2</sub>OH 0.08 (77) 11

18 OPh 5-SF<sub>5</sub> H 0.08 (185) 7  
 19 OPh 5-SF<sub>5</sub> CH<sub>2</sub>OH 0.02 (101) 121  
 20 OPh 4-F,5-F CH<sub>3</sub> 0.13 (132) 145

<sup>a</sup>measured using IWQ (% of Maximal response to ML213)

<sup>b</sup>kinetic aqueous solubility measured using CAD or CLND

Initially, poor aqueous solubility was observed in the cyclohexamide series. This issue was resolved by the addition of a hydroxymethyl group at position R. Again, the stereospecificity at this position was limited to that shown in Table 2. Stabilities under HLMs were compromised without EWGs on the cyclohexyl ring. Gem-difluoro substitution on the cyclohexyl ring provided stability and avoided the introduction of an additional chiral center. The phenyl group adjacent to the carboxamide was ideally left unsubstituted much like the phenyl group in the benzamide series. Lipophilic substituents at the meta-position of the benzyl ring improved target activity without sacrificing significant solubility. Methylsulfone substitution was not well-tolerated on the benzyl ring in this series.



**Table 2.** Cyclohexamide series potencies ( $\mu\text{M}$ )

Cmpd R R<sup>1</sup> R<sup>2</sup> EC<sub>50</sub><sup>a</sup> Sol.( $\mu\text{M}$ )<sup>b</sup>

21 CH<sub>3</sub> H H 0.25(115) 57

22 CH<sub>2</sub>OH H H 0.79(165) 455

23 CH<sub>2</sub>OH F H 1.3(108) 332

24 CH<sub>3</sub> F H 0.20(140) 25

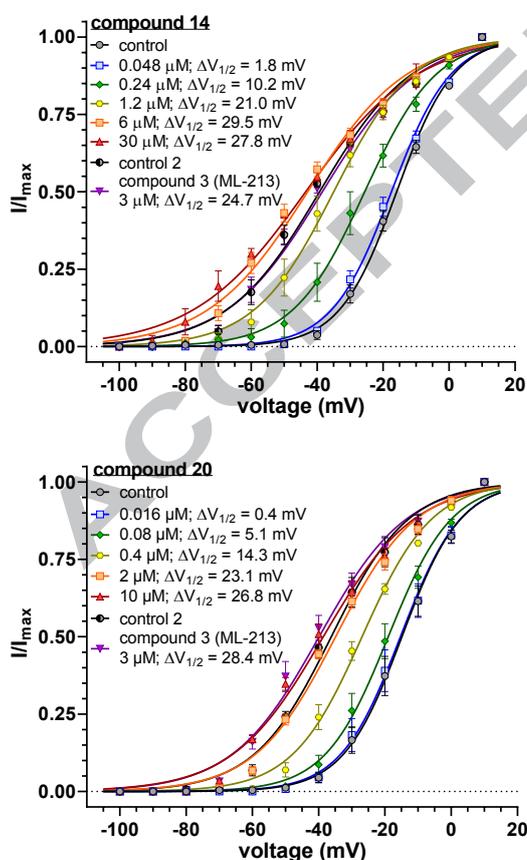
14 CH<sub>2</sub>OH F 3-Cl 0.30(141) 318

25 CH<sub>2</sub>OH F 3-CF<sub>3</sub> 0.08(96) 119

<sup>a</sup>measured using IWQ (% of Maximal response to ML213)

<sup>b</sup>kinetic aqueous solubility measured using CAD or CLND

The selection criteria for progressing compounds to *in vivo* studies were minimally: aqueous kinetic solubilities of  $>100 \mu\text{M}$ , K<sub>v</sub>7.2/7.3 channel opening EC<sub>50</sub>  $\leq 0.30 \mu\text{M}$  (IonWorks™ Quattro), and HLM clearance values under 3 mL/min/g liver. All compounds had P-glycoprotein (P-gp) ratios of less than 2. Some compounds elicited complex curves wherein the currents measured were actually depressed as compared with vehicle controls at more depolarized potentials (i.e., generally higher than -10 mV). These compounds were not advanced to *in vivo* studies. Examples of IV curves of compounds in QPatch (see supplemental section) that met with our progression criteria are indicated in Figure 2.



**Figure 2.** Examples of IV curve

Prior to *in vivo* testing in rodents, activities of representative compounds from both chemical series **14** and **20** were measured in primary cultured rat cortical neurons using a multiple-electrode array system (see supplemental section). Both

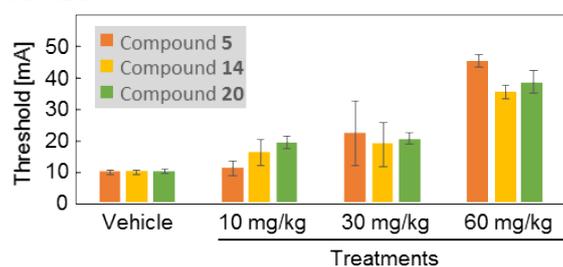
compounds inhibited spontaneous neuronal firing activities at similar concentrations to those observed with cell lines over-expressing  $K_V7$  isoforms.  $IC_{50}$  values of 0.28 and 0.11  $\mu M$  were determined for compounds **14** and **20**, respectively.

Compounds **14** and **20** were also found to be sufficiently brain penetrant (Table 3) to progress to maximum electroshock-threshold (MES-T, Figure 3A) and Rotarod (Figure 3B) studies that measure anti-seizure effects and CNS tolerability, respectively. Dose-dependent increases in seizure threshold with no meaningful effects in the Rotarod model were similar to the results observed with the experimental control ICA-27243 (Compound **5** in Figure 3A and B). Similar results were obtained using the pentylenetetrazol (PTZ)-induced seizure model (Supplemental Figure 2). In addition, pharmacodynamic (PD) effects observed in both the PTZ and MES-T mouse models correlated well with the predicted target engagement across compounds at all dose levels tested (Supplemental Figure 3).

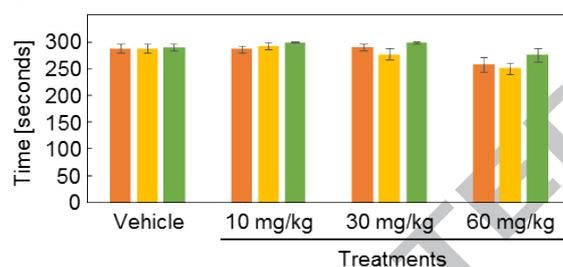
**Table 3.** Pharmacokinetic parameters for selected  $K_V7.2/7.3$  channel openers following intravenous dosing in mice

Cmpd	Cl <sub>b</sub>	DNAUC <sub>0-t</sub>	Br/Bl (mL/min/kg)	(( $\mu g$ .h/mL) ratio /mg/kg)
<b>14</b>	48 ± 2.1	0.51 ± 0.5	0.8	
<b>20</b>	31 ± 2.6	0.33 ± 0.5	1.3	

#### A. MES-T



#### B. Rotarod



**Figure 3. MES-T seizure and Rotarod studies**

After determining that compounds from these two series could achieve efficacy in seizure models, we sought to understand if the compounds demonstrated isoform selectivity amongst the  $K_V7.2-7.5$  channels. Compounds meeting the previously described selection criteria for activity and solubility were cross-screened against the heterodimeric  $K_V7.3/7.5$  channel using the IonWorks™ Barracuda platform (Molecular Devices). All compounds demonstrated similar levels of activities in both  $K_V7.2/7.3$  and  $K_V7.3/7.5$  assays (Table 4). To further scrutinize the binding site interactions, select compounds were tested in a cell line that has point mutations ( $K_V7.2W236L$  and  $K_V7.3W265L$ ) in the pore domain (Table 5). No compounds from these two series showed measurable activity up to 10  $\mu M$  in this assay, suggesting that these compounds bind in the  $K_V7.2/7.3$  pore domain. The activity of Icaegen compound ICA-27243 (**5**) was unaffected in this mutant  $K_V7.2/7.3$  assay, supporting reports of compound binding in the voltage sensor domain.<sup>17</sup>

**Table 4.** Activities in human  $K_V7.2-7.5$  channels ( $\mu M$ )<sup>a</sup>

Cmpd	$K_V7.2/7.3$	$K_V7.3/7.5$	$K_V7.4$
<b>14</b>	0.10	0.36	0.10
<b>19</b>	0.06	0.08	>10
<b>20</b>	0.20	0.20	0.16
<b>24</b>	0.40	0.40	4.0
<b>25</b>	0.05	0.25	0.02
<b>ML-213</b>	0.04	0.50	1.6
<b>ICA-27243</b>	0.46	>10	>10

<sup>a</sup>activities determined by SyncroPatch platform.

**Table 5.** Activities in human wildtype and mutant  $K_V7.2/7.3$  channels ( $\mu M$ )<sup>a</sup>

Cmpd	WT $K_V7.2/7.3$	$K_V7.2W236L/$	$K_V7.3W265L$
<b>15</b>	0.02	>30	
<b>16</b>	0.05	>30	
<b>21</b>	0.10	>30	
<b>ML-213</b>	0.04	>11	
<b>ICA-27243</b>	0.46	0.43-1.81	

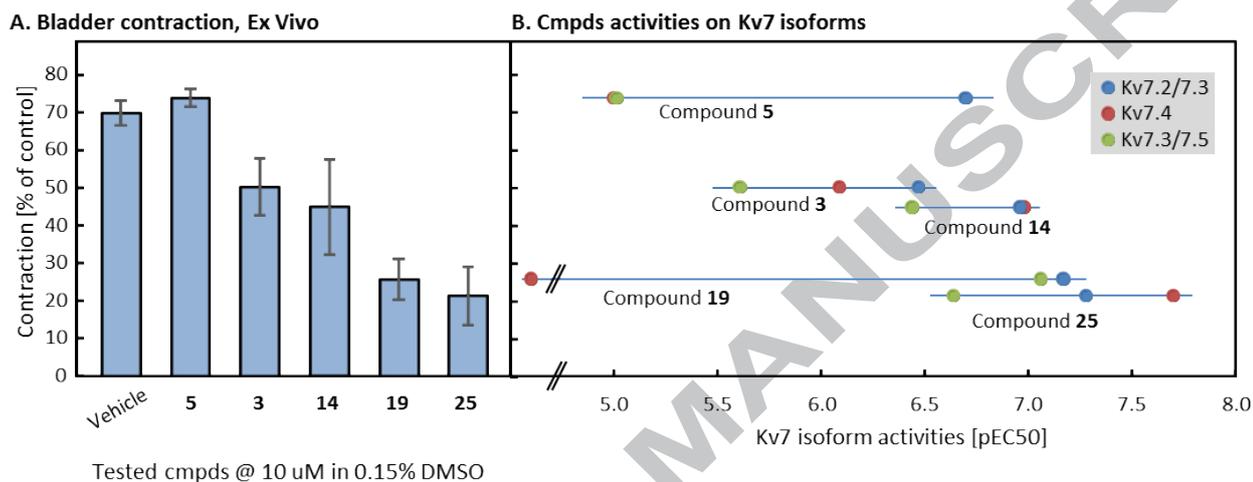
**Retigabine 0.32 >30**

<sup>a</sup>activities determined by <sup>86</sup>Rb efflux assay

It has been reported that K<sub>v</sub>7.4 is implicated in pain and urinary bladder contraction. Therefore, we measured the effects of compound **19**, which has significantly reduced activity on K<sub>v</sub>7.4, in a bladder contraction model.

Urinary adverse events were reported more frequently in patients receiving the pan-K<sub>v</sub>7 activator Retigabine (**1**), as compared with placebo, although most patients were able to continue with treatment.<sup>11</sup> *KCNQ* isoforms are expressed in human detrusor and K<sub>v</sub>7 openers were shown to influence bladder contraction.<sup>12, 18</sup> In this study, we have examined expression of *KCNQ* isoforms (Supplemental Figure 4) and effects on human bladder contraction *ex vivo* (Figure 4). We tested compounds that have diverse *KCNQ* isoform activity profiles to determine how different isoform profiles influence bladder contraction. While the inhibition of bladder contraction was clearly associated with K<sub>v</sub>7 activation, it wasn't clear how individual K<sub>v</sub>7.2-7.5 isoforms may contribute. Compound **19**, which has significantly reduced homomeric K<sub>v</sub>7.4 activity, did not reduce impact on bladder contraction, and this may suggest that the homomeric K<sub>v</sub>7.4 channel plays a less critical role in bladder contraction. Further studies are needed to conclude a role of K<sub>v</sub>7 isoforms in bladder contraction.

**Figure 4.** Effects of K<sub>v</sub>7 openers with differential activities on K<sub>v</sub>7.2/7.3, 7.4 and 7.3/7.5 isoforms on carbachol-induced contraction in human bladder.



In the present study, we have identified novel pan-K<sub>v</sub>7 openers, that are free from structural alerts. Representatives of novel K<sub>v</sub>7 openers demonstrated anti-seizure effects in a dose-dependent manner without meaningful changes in the Rotarod CNS tolerability model. We found that effects on bladder contraction depend on pan-K<sub>v</sub>7 activity, but perhaps not on homomeric K<sub>v</sub>7.4 activity. Further investigation is required to conclude the roles of individual K<sub>v</sub>7 isoforms in bladder contraction.

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### Supplementary Materials

X-Ray crystal structure of compound **24** (Supplemental Figure 1), synthetic procedures including yields, seizure protection in PTZ-induced seizure model (Supplemental Figure 2), methods and some data for electrophysiological and biological experiments, PK/PD analysis in MES-T and PTZ models (Supplemental Figure 3), and quantification of *KCNQ* isoforms mRNA expression in the bladder (Supplemental Figure 4), were included in Supplementary Section.

- Novel pan-Kv7 openers for the treatment of epilepsy are discussed
- Compounds from two series of Kv7 openers show efficacy in PTZ and MES-T seizure models
- Bladder contraction is associated with pan-KV7 activity, but perhaps not with homomeric Kv7.4 activity as reported

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