Accepted Manuscript

Discovery of thieno[3,2-*c*]pyridin-4-amines as novel Bruton's tyrosine kinase (BTK) inhibitors

Xinge Zhao, Minhang Xin, Yazhou Wang, Wei Huang, Qiu Jin, Feng Tang, Gang Wu, Yong Zhao, Hua Xiang

PII:	\$0968-0896(15)00453-8
DOI:	http://dx.doi.org/10.1016/j.bmc.2015.05.043
Reference:	BMC 12339
To appear in:	Bioorganic & Medicinal Chemistry
Received Date:	31 March 2015
Revised Date:	21 May 2015
Accepted Date:	22 May 2015



Please cite this article as: Zhao, X., Xin, M., Wang, Y., Huang, W., Jin, Q., Tang, F., Wu, G., Zhao, Y., Xiang, H., Discovery of thieno[3,2-*c*]pyridin-4-amines as novel Bruton's tyrosine kinase (BTK) inhibitors, *Bioorganic & Medicinal Chemistry* (2015), doi: http://dx.doi.org/10.1016/j.bmc.2015.05.043

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

1	Discovery of thieno[3,2-c]pyridin-4-amines as novel Bruton's
2	tyrosine kinase (BTK) Inhibitors
3	
4	Xinge Zhao ^{a, b, †} , Minhang Xin ^{c, †} , Yazhou Wang ^b , Wei Huang ^d , Qiu Jin ^b , Feng Tang ^b ,
5	Gang Wu ^b , Yong Zhao ^b , Hua Xiang ^{a, *}
6	
7	^a Department of Medicinal Chemistry, School of Pharmacy, China Pharmaceutical University, No. 24,
8	Tongjiaxiang, Nanjing 210009, P.R. China
9	^b Jiangsu Simcere Pharmaceutical Co. Ltd., Jiangsu Key Laboratory of Molecular Targeted Antitumor
10	Drug Research, No 699-18, Xuan Wu District, Nanjing 210042, P.R. China
11	^c Department of Medicinal Chemistry, School of Pharmacy, Health Science Center, Xi' an Jiaotong
12	University, No 76, Yanta West Road, Xi ' an 710061, P.R.China
13	^d Key Laboratory of Pesticide and Chemical Biology, Ministry of Education, College of Chemistry,
14	Central China Normal University, Wuhan, 430079, P.R.China
15	
16	
17	
10	
10	
20	
21	
22	
23	
24	
25	\mathbf{G}^{-}
26	Footnotes
27	*Address correspondence to this author at the Department of Medicinal Chemistry, School of
28	Pharmacy, China Pharmaceutical University, No 24 Tongjiaxiang, Nanjing 210009, P.R. China; Tel:
29	+86-25-83271096; E-mail: <u>1020030692@cpu.edu.cn</u>
30	
31	[†] These authors contributed equally to this work.
32	

1	Abstract: A novel series of BTK inhibitors bearing thieno[3,2-c]pyridin-4-amine
2	framework as the core scaffold were designed, synthesized and well characterized. In this
3	paper, twenty one compounds displayed variant inhibitory activities against BTK in vitro,
4	and compound 14g showed the most potent inhibitory activity agianst BTK enzyme, with
5	the IC_{50} value of 12.8 nM. Moreover, compounds 14g displayed relatively good kinase
6	selectivity and was subsequently evaluated in vivo for profiling its PK properties. This
7	work identified the thieno[3,2-c]pyridin-4-amine derivatives as novel BTK inhibitors and
8	verified the value of thieno[3,2-c]pyridin-4-amine scaffold in drug design.
9	
10	
11	Keywords: BTK inhibitors, thieno[3,2-c]pyridin-4-amine, inhibitory activity, kinase
12	selectivity.
13	
14	
5	

1 **1. Introduction**

2 Bruton's tyrosine kinase (BTK) is a member of Tec family of cytoplasmic mammalian non-receptor tyrosine kinases. It is a crucial terminal kinase in the B cell receptor (BCR) 3 signaling pathway and essential for the development and activation of B cells. It has been 4 shown that aberrant activation of B cells plays a central role in the pathogenesis of B-cell 5 lymphomas and various autoimmune diseases. Therefore, inhibition of BTK has become 6 an attractive and potential therapeutic approach for the treatment of human diseases 7 associated with B cells lymphoproliferative disorders and autoimmune disorders, such as 8 hematological malignancies and rheumatoid arthritis $(RA)^{1-2}$. 9

During recent years, a number of research groups are eager to pursue small molecular 10 BTK inhibitors, and many BTK-targeting agents have progressed to clinical stages. 11 Among these, ibrutinib (1, PCI-32765, ImbruvicaTM) is the most successful BTK 12 inhibitor, which was developed by Pharmacyclics and approved by US FDA for the 13 treatment of mantle cell lymphoma (MCL) and chronic lymphocytic leukemia (CLL) in 14 15 the past 2 years. And now, it is also ongoing in the clinical evaluation for other indications³. In addition, several BTK inhibitors including spebrutinib (2, CC-292, 16 AVL-292), ONO-4059 (3) and ACP-196 are currently moved into the phase II clinical 17 evaluation, and several molecules including MSC-2364447, PRN1008, BGB-3111 and 18 19 HM-71224 are quickly advanced into phase I clinical trials, although most of their structures are still not disclosed ⁴⁻⁶. Except that, there are some agents including 20 CGI-1746, GDC-0834, RN-486 (4) reported in discovery stage ⁷⁻¹⁵. Recently, we have 21 also reported our medicinal chemistry work on the discovery of BTK inhibitors, which 22 are structurally similar to RN-486, such as compounds 5 and 6^{16-17} . In general, the 23 reported small molecular BTK inhibitors can be classified into two classes (irreversible 24 inhibitors and reversible inhibitors) by their binding modes with the BTK catalytic 25 26 domains. Irreversible BTK inhibitors contain a Michael addition receptor moiety in the structures which can form a covalent bind with the conserved cysteine-481 residue of 27 BTK enzyme to achieve strong binding ¹. The agents including ibrutinib, spebrutinib and 28 ONO-4059 belong to this class. It is believed that irreversible BTK inhibitors can achieve 29 30 significant clinical benefit for treating hematological malignancies. For example,

1 ibrutinib shows significant progression free survival benefit and overall survival benefit in both MCL and CLL clinical trials. Nevertheless, it is thought reversible BTK inhibitors 2 3 might be more appropriate for treating RA. RA is different from hematological 4 malignancies, and is non-life-threatening disease. However, the irreversible inhibitors 5 might give some unexpected off-target toxicity because of the potential of covalently binding to non-target proteins and achieving the drug-protein conjugate. Thus, the 6 development of reversible BTK inhibitors would be more suitable to fulfill the treatment 7 need for non-life-threatening RA disease ¹⁸⁻¹⁹. Among the reported reversible BTK 8 9 inhibitors, 5-phenylpyrazinone-based or 5-phenylpyridinone-based inhibitors, such as 10 GDC-0834 and RN-486, demonstrated high BTK inhibitory potency and excellent selectivity. However, GDC-0834 was suspended in phase I because of poor 11 pharmacokinetic properties, and RN486 was not any progressed in the past 3 years ⁴. 12 13 Thus, there is still eagerly need to develop new classes of reversible BTK inhibitors.

14 (Insert Figure 1 and title for Figure 1 here)

15

Ibrutinib is a selective irreversible BTK inhibitor, showing very high potency against 16 BTK (IC₅₀ = 0.5 nM). Ibrutinib can irreversibly bind to BTK because of the acrylamide 17 moiety which can form a covalent bind with the conserved cysteine-481 residue of BTK. 18 19 In fact, ibrutinib was structurally optimized from the precursor compound PCI-29732, which was reported to be another potent BTK inhibitor ($IC_{50} = 8.2$ nM). Due to lack of 20 Michael addition receptor moiety, PCI-29732 is regard to non-covalently bind to BTK 21 enzyme. PCI-29732 is chemically similar to compound B43. The publicly available 22 23 crystal structure of B43 with BTK (PDB 3GEN) disclosed the binding mode of this type of reversible BTK inhibitors ²⁰⁻²¹. Generally, the backbone of 4-amino pyrrolopyrimidine 24 occupies the ATP binding pocket and makes several important interactions with the hinge. 25 The 4-amine directly forms two H-bonds with the gatekeeper Thr-474 and the backbone 26 27 Glu-475. The N-3 nitrogen of the pyrimidine participates in an H-bond with Met-477, and the phenoxyphenyl group enters a hydrophobic pocket and forms a face-to-edge 28 29 π -stacking interaction with Phe-540. These interactions above are critical for BTK 30 activity. Bioisosteric replacement is a highly attractive method in the drug design and

1 discovery, and we have used this strategy to discover several novel chemical series in hedgehog inhibitors programs²². Thieno[3,2-c]pyridines are very valuable heterocyclic 2 scaffolds which are abundantly described as central structural motifs in the design of 3 kinase inhibitors ²³. Based on the deeply understanding of the binding mode of B43 with 4 BTK enzyme and the principles of bioisosterism, we designed a novel series of BTK 5 inhibitors containing thieno[3,2-c]pyridin-4-amine scaffold. In the new designed series, 6 7 thieno[3,2-c]pyridin-4-amine group are expected to occupy the ATP binding pocket and keep the key hydrogen bonds, and the C3-phenoxyphenyl and its bioisosteric groups can 8 form the π -stacking interaction. Meanwhile, we tried to introduce alkyl and aryl groups 9 10 on the C7-position of thieno[3,2-c]pyridines to explore the SAR of the thieno[3,2-c]pyrid-4-amine scaffold. To the best of our knowledge, there is the first time 11 12 to describe the SARs on this position. Herein, we report our recent effort on the synthesis 13 and SARs of this series of BTK inhibitors.

14 (Insert Figure 2 and title for Figure 2 here)

15

16 **2.** Chemistry

As summarized in Table 1, the thieno[3,2-c]pyridin-4-amine derivatives (**12a-h**, **14a-i**, **19a-b and 24a-b**) were designed and synthesized. The synthetic routes for all target compounds are illustrated in Schemes 1–2.

Compounds **12a–h** and **14a-i** were synthesized according to Scheme **1**. Commercially 20 3-bromothieno[3,2-c]pyridin-4-amine 9 21 available was treated with (4-phenoxyphenyl)boronic acid under palladium catalysis to give the Suzuki coupling 22 23 product 10 in 96 % yield. Iodination of 10 using N-iodosuccinimide afforded the key intermediate 11 in 70 % yield. Then treating 11 with various boronic acid reagents under 24 25 the Suzuki coupling condition provided the target compounds 12a-h as well as the 26 intermediate 13 in 35-90 % yields. Deprotection of 13 with trifluoroacetic acid provided the target compound 14a in 92 % yield, which was subsequently treated with various 27 halides under alkaline condition to produce target compound 14b-i in 40-95 % yields. 28

29 (Insert Scheme 1 and legend for Scheme 1 here)

1 Compounds 19a-b and 24a-b were prepared according to Scheme 2. Condensation of the commercially available boronobenzoic acid (15a-b) and aniline gave 16a-b, which 2 were subsequently reacted with 3-bromothieno[3,2-c]pyridin-4-amine 9 using Suzuki 3 coupling protocols to generate 17a-b. Treating 17a-b with N-iodosuccinimide provided 4 the iodides 18a-b, followed by Suzuki coupling reaction to afford the target compounds 5 6 **19a-b.** Likewise, the similar synthetic routes were employed to synthesize compounds 7 **24a-b.** Starting from the commercially available 4-aminophenylboronic acid pinacol ester (20) and m-chlorobenzoyl chloride, the desired condensation product 21 was obtained, 8 9 which was followed by Suzuki coupling reaction, iodination and again Suzuki coupling 10 reaction, leading to the target compounds **24a-b**.

- 11
- 12
- 13 **3. Result and discussion**

14 3.1. In vitro enzymatic assay of the new compounds against BTK kinase

(Insert Scheme 2 and legend for Scheme 2 here)

15 The newly synthesized thieno[3,2-c]pyridin-4-amine derivatives (12a-h, 14a-i, 19a-b

16 and 24a-b) were evaluated for their abilities to inhibit the BTK enzymatic activity. The

17 vitro IC_{50} values are summarized in Table 1.

18 (Insert Table 1 and title for Table 1 here)

19

Among the thieno[3,2-c]pyridin-4-amine series, when the six-membered ring aryl 20 21 group were introduced at the C7-position, such as phenyl (12a), 4-pyridyl (12b), 22 4-morpholinyl substituted phenyl (12c), 4-phenoxy substituted phenyl (12d), and 23 6-methoxynaphthalenyl (12e), the resulting compounds almost lost inhibitory effect against 24 BTK ($IC_{50} > 1000 \text{ nM}$). However, the five-membered ring aryl substituted compounds 12f 25 and 14a displayed weak potency against BTK. C7-furan-3-yl alogue 12f showed an IC_{50} 26 value of 767.5 nM, while C7-1H-pyrazol-4-yl alogue 14a was weaker than 12f, with an 27 IC_{50} value of 1052 nM. The introduction of three-membered ring of cyclopropyl (12g) or linear alkyl of n-butyl (12h) in the C7-position resulted in BTK inhibitory activity lost 28 $(IC_{50} > 10000 \text{ nM}, \text{ both})$. This initial SAR trend appeared that five-membered aryl 29 30 substituted at C7-position was a preference. Subsequently, a survey of several substituents

1 at the N1-position of pyrazolyl was investigated. Interestingly, Compounds 14b bearing 2 N1-methyl group showed an IC₅₀ value of 107.4 nM, which was 10 fold more potent than no-substituted pyrazolyl **14a**, while N1-methyl was changed into bulky alkyl groups such 3 4 as ethyl (14c), n-butyl (14d), benzyl (14e) or methoxypropyl (14f) afforded reduced BTK inhibitory activity, compared to 14b. However, unexpectedly, compound 14g bearing a 5 6 2-(methylamino)-2-oxoethyl at N1-position of pyrazolyl exhibited a higher potency 7 against BTK, with an IC_{50} value of 12.8 nM. Compared with these, 2-(methylamino)-2-oxoethyl (14g) produced increased activity likely due to the amide or 8 9 the carbonyl group. An attempt of replacement of 2-(methylamino)-2-oxoethyl with dimethylaminosulfuryl (14h) and acetyl (14i) led to reduced potency, which appeared 10 11 that the position of the amide or the carbonyl group was critical for BTK inhibitory activity. 12 N-(2-(methylamino)-2-oxoethyl)-1H-pyrazol-1-yl moiety at the C7-position may provide 13 an extra interaction with BTK residue, thereby showing the higher inhibitory activity. 14 Based on the best two substituents at the C7-position, the C3-position modification was attempted to carry out. The replacement of the oxygen linker of C3-phenoxyphenyl with 15 16 an amide afforded compounds 19a and 19b, as well as 24a and 24b. However, 17 unfortunately, these compounds showed reduced inhibitory activities. This finding suggested that the π -electron properties of the subsutituents on the C3-position of 18 19 thieno[3,2-c]pyrid-4-amine scaffold significantly affected the BTK inhibitory activity.

20

21 3.2. Kinase selectivity assay

22 From the BTK inhibition results above, compound 14b and 14g were picked out as the 23 representative BTK inhibitors among the thieno[3,2-c]pyridin-4-amine series, and they 24 were further evaluated for their inhibitory activities against 4 kinds of our in-house RTKs. 25 The data were summarized in Table 2. Compared to BTK enzyme, both compounds only showed moderate inhibitory activities against KDR kinase, and moreover, compound 14g 26 27 displayed 10-fold more selective against BTK than KDR. This result indicated that **14g** was a relatively selective BTK inhibitor, and deserving further identification against other 28 kinase profiles. 29

30 (Insert Table 2 and title for Table 2 here)

1

2 3.3. In vivo pharmacokinetic (PK) profiles

3 Compound 14g stood out as the most potent BTK inhibitor in this 4 thieno[3,2-c]pyridin-4-amine series, with an IC_{50} value of 12.8 nM. Therefore, it was further investigated by profiling its pharmacokinetic (PK) properties in vivo. Table 35 illustrated the PK profiles of compound 14g in SD rats, by iv (2.5 mg/kg) and po (10 6 7 mg/kg) administration. Following intravenous injection with 2.5 mg/kg in SD rats, 14g showed a satisfactory exposure (AUC = 1459.1 hr*ng/mL) and volume of distribution 8 (Vz = 900 mL/kg), but a short half-time ($T_{1/2}$ = 0.36 h). The short elimination phase half 9 life was probably due to its large systemic clearance (Cl = 1716.4 mL/hr/kg). 10 Subsequently, compound 14g was further evaluated by oral administration of 10 mg/kg 11 12 dose. It was found the drug was also quickly eliminated, with large systemic clearance of 8670.1 mL/hr/kg and short half time ($T_{1/2} = 1.01$ h), although the area-under-curve (AUC 13 14 = 1365.7 hr*ng/mL) and the oral bioavailability ($\mathbf{F} = 23.4 \%$) are acceptable in this dose. The PK properties of compound **14g** suggested that some groups in this structure maybe 15 16 easily be metabolized and eliminated, thereby exhibiting a short half-life. Thus, in the 17 next structural modification, efforts should be made for reducing the metabolic effect and optimizing the PK characteristics, meanwhile, increasing the in vitro biological activities. 18 (Insert Table 3 and title for Table 3 here) 19

20

21 3.4. Docking study

an effort 22 In to elucidate the binding mode for this kind of 23 thieno[3,2-c]pyridin-4-amines, We performed the molecular docking of compound 14g 24 with BTK enzyme (PDB code 3GEN). The CDOCKER program of the Discovery Studio 25 2.5 software package was used, and the binding model of compound 14g with BTK enzyme was shown in Figure 3. It was found that the entire structure of 14g was 26 27 favorably located in the BTK pocket. The 4-amino group of thieno[3,2-c]pyridine fragment of 14g formed two hydrogen bonds with Thr-474 and Glu-475, respectively. 28 29 Furthermore, the N atom in the 5-position of thieno[3,2-c]pyridine generated another 30 hydrogen bonding interaction with the backbone of Met-477 in the BTK kinase hinge

1 region. These three hydrogen bonds were very critical for maintaining the BTK inhibitory 2 activity. Meanwhile, the N' atom in the 2-position of pyrazolyl group substituted at 3 C7-position of thieno[3,2-c]pyridin-4-amine formed a hydrogen bond with the water-36, which was similar to **B43** with BTK. Interestingly, it was noticeable that the carbonyl of 4 5 2-(methylamino)-2-oxoethyl group in 1-position of pyrazolyl could afford a hydrogen bond interaction with Cys-481, which might contribute to the higher BTK inhibition of 6 7 compound **14g** in the series. Overall, the docking results afforded some understandings 8 on the observed BTK inhibitory activities for the thieno[3,2-c]pyridin-4-amines. 9 (Insert Figure 3 and title for Figure 3 here)

10

11 **4.** Conclusion

In this paper, we have developed a novel series of BTK inhibitors bearing 12 thieno[3,2-c]pyridin-4-amine framework as the core scaffold. Twenty one compounds 13 were design, synthized, and evaluated. These compounds displayed variant inhibitory 14 15 activities against BTK in vitro, and compound 14g gave the highest potency with the IC₅₀ value of 12.8 nM. Moreover, compounds 14g displayed relatively good kinase selectivity. 16 However, in vivo evaluation showed that compound 14g had poor PK properties. Thus, 17 further structural modification is deserved to be made for improving the PK 18 19 characteristics, as well as increasing the in vitro biological activities.

20

21 **5. Experimental**

22 5.1 Chemistry

All chemical reagents were purchased from commercial vendors and used without further purification unless noted especially. The melting points for the compounds were performed on a Melt-Temp II apparatus and uncorrected. ¹H NMR spectra (400 MHz) and ¹³C NMR (100 MHz) spectra data were recorded in CDCl₃ or DMSO- d_6 on a Bruker BioSpin AG (Ultrashield Plus AV 400) spectrometer. MS data were recorded at an Agilent-6120 quadrupole LC/MS (ESI) while HRMS were recorded at a Water Q-Tof micro mass spectrometer. The HPLC study for the compounds was verified using a

mixture of solvent (methanol/water or acetonitrile/water) at the flow rate of 2 mL/min
and peak detection at 254 nm under UV. Column chromatography was carried out on
silica gel (200–300 mesh) purchased from Qindao Ocean Chemical Company of China.
Thin-layer chromatography (TLC) analyses were carried out on silica gel GF254.
Ibrutinib was synthesized according to the reference literature.

3-(4-Phenoxyphenyl)thieno[3,2-c]pyridin-4-amine solution of 6 (10). А 3-bromothieno[3,2-c]pyridin-4-amine (3 g, 13 mmol), Pd(PPh₃)₄ (1.5 g, 1.3 mmol) and 7 8 (4-phenoxyphenyl)boronic acid (3.06 g, 14.3 mmol) in toluene (20 mL) was degassed 9 with nitrogen for 5 min followed by addition of EtOH (4 mL), H₂O (2 mL) and Na₂CO₃ (3.5 g, 32.5 mmol) under continuous flow of nitrogen. The reaction mixture was stirred at 10 90 °C for 2 h. The reaction mixture was cooled, filtered through celite, diluted with water 11 12 (45 mL), and extracted with $(3 \times 60 \text{ mL})$ ethyl acetate. The combined organic layers were 13 dried over sodium sulfate and were concentrated in vacuo, the crude product was purified 14 on a silica gel column using (20-80 % ethyl acetate/hexanes) as eluent to afford 10 (4.0 g, 96 %) as a white solid. MS m/z 319.1 [M+H]⁺; ¹H NMR (400 MHz, DMSO- d_6) δ ppm 15 7.84-7.82 (d, J=5.60 Hz, 1H), 7.59 (s, 1H), 7.48-7.43 (m, 4H), 7.27-7.26 (d, J=5.60 Hz, 16 1H), 7.22-7.18 (t, J=14.4 Hz, 1H), 7.14-7.10 (t, J=15.6 Hz, 4H), 5.43 (brs, 2H). 17

7-Iodo-3-(4-phenoxyphenyl)thieno[3,2-c]pyridin-4-amine (11). A solution of 18 19 3-(4-phenoxyphenyl)thieno[3,2-c]pyridin-4-amine (2.5)7.9 mmol) g, and N-Iodosuccinimide (2.14 g, 9.5 mmol) in DMF (10 mL) was stirred at r t for 2 h. After 20 completion of reaction, the reaction mixture was diluted with the mixed ice and water, the 21 22 precipitated solid was filtered off and the crude product was purified on a silica gel 23 column using (20-80 % ethyl acetate/ petroleum ether) as eluent to afford 11 (2.4 g, 70.0 %) as a yellow solid. MS m/z 445.0 $[M+H]^+$; ¹H NMR (400 MHz, DMSO- d_6) δ ppm 24 8.02 (s, 1H), 7.57 (s, 1H), 7.48-7.42 (m, 4H), 7.22-7.18 (t, J=14.8 Hz, 1H), 7.14-7.10 (t, 25 26 J=15.2 Hz, 4H), 5.61 (brs, 2H).

3-(4-Phenoxyphenyl)-7-phenylthieno[3,2-c]pyridin-4-amine (12a). A solution of
7-iodo-3-(4-phenoxyphenyl)thieno[3,2-c]pyridin-4-amine (100 mg, 0.225 mmol),
Pd(PPh₃)₄ (26 mg, 0.0225 mmol) and phenylboronic acid (33 mg, 0.27 mmol) in
Ethylene glycol dimethyl ether (10 mL) was degassed with nitrogen for 5 min followed

1 by addition of H₂O (2 mL) and Na₂CO₃ (72 mg, 0.675 mmol) under continuous flow of nitrogen. The reaction mixture was stirred at 80 °C for 3 h. The reaction mixture was 2 3 cooled, filtered through celite, diluted with water (45 mL), and extracted with $(3 \times 20 \text{ mL})$ ethyl acetate. The combined organic layers were dried over sodium sulfate and were 4 5 concentrated in vacuo, the crude product was purified on a silica gel column using (20-50 % ethyl acetate/ petroleum ether) as eluent to afford 12a (71 mg, 80.1 %) as a 6 white solid. Mp:172-173.4 °C; MS m/z 395.1 $[M+H]^+$; ¹H NMR (400 MHz, CDCl₃) δ 7 ppm 7.99 (s, 1H), 7.66 (s, 1H), 7.52-7.39 (m, 9H) 7.20-7.18 (t, J=14.8 Hz, 1H), 7.13-7.09 8 9 (t, J=15.2 Hz, 4H), 4.86 (brs, 2H); HPLC 95.4 %; HRMS (ESI) m/z calcd for C₂₅H₁₈N₂OS [M+H]⁺. 395.1140, found 395.1218. 10

3-(4-Phenoxyphenyl)-7-(pyridin-4-yl)thieno[3,2-c]pyridin-4-amine (12b). 12b was
synthesized by the general method described above compound 12a (27 mg, 66.3 %). Mp:
230.2-231.8 °C; MS m/z 396.1 [M+H]⁺; ¹H NMR (400 MHz, CDCl₃) δ ppm 8.72-8.71 (d,
J=5.56 Hz, 2H), 8.04 (s, 1H), 7.63-7.62 (d, J=5.52 Hz, 2H), 7.45-7.39 (dd, J=8.44 Hz,
J=7.64 Hz, 4H), 7.20-7.19 (t, J=4.44 Hz, 2H), 7.12-7.10 (d, J=8.56 Hz, 4H), 4.87 (brs,
2H); HPLC 99.4 %; HRMS(ESI) m/z calcd for C₂₄H₁₈N₃OS [M+H]⁺. 396.1092, found
396.1172.

18**7-(4-Morpholinophenyl)-3-(4-phenoxyphenyl)thieno[3,2-c]pyridin-4-amine**(12c).19**12c** was synthesized by the general method described above compound **12a** (28 mg,2067.1 %). Mp: 207.7-209.4 °C; MS m/z 480.2 [M+H]⁺; ¹H NMR (400 MHz, DMSO- d_6) δ 21ppm 7.85 (s, 1H), 7.62 (s, 1H), 7.53-7.43 (m, 7H), 7.22-7.21 (t, *J*=8.54 Hz, 1H),227.15-7.07 (m, 5H), 5.45 (brs, 2H), 3.78-3.76 (t, *J*=8.00 Hz, 4H), 3.19-3.17 (t, *J*=8.00 Hz,234H); HPLC 95.4 %; HRMS (ESI) m/z calcd for C₂₉H₂₅N₃O₂S [M+H]⁺. 480.1740, found24480.1756.

3,7-Bis(4-phenoxyphenyl)thieno[3,2-c]pyridin-4-amine (12d). 12d was synthesized
by the general method described above compound 12a (22 mg, 76.1 %). Mp: 185.5-187.3
°C; MS m/z 487.2 [M+H]⁺; ¹H NMR (400 MHz, CDCl₃) δ ppm 7.92 (s, 1H), 7.63-7.61 (d,
J=8.04 Hz, 2H), 7.46-7.43 (d, J=8.34 Hz, 2H), 7.42-7.36 (dd, J=8.00 Hz, J=8.00 Hz,4H),
7.20-7.17 (t, J=13.20 Hz, 2H), 7.15-7.10 (m, 8H); HPLC 95.4 %; HRMS (ESI) m/z calcd

30 for $C_{31}H_{22}N_2O_2S$ [M+H]⁺. 487.1478, found 487.1482.

1 7-(6-Methoxynaphthalen-2-yl)-3-(4-phenoxyphenyl)thieno[3,2-c]pyridin-4-amine (12e). 12e was synthesized by the general method described above compound 12a (32 mg, 2 72.1 %). Mp: 205.1-207.2 °C; MS m/z 475.2 $[M+H]^+$; ¹H NMR (400 MHz, CDCl₃) δ 3 ppm 8.06-8.05 (d, J=3.4 Hz, 2H), 7.87-7.85 (d, J=8.04 Hz, 1H), 7.82-7.80 (t, J=8.41 Hz, 4 1H), 7.77-7.75 (d, J=8.00 Hz, 1H), 7.47-7.45 (d, J=8.20 Hz, 2H), 7.42-7.39 (t, J=12.18 5 Hz, 2H), 7.21-7.16 (dd, J=6.04 Hz, J=4.84 Hz, 4H), 7.12-7.10 (dd, J=2.32 Hz, J=2.46 Hz, 6 4H), 4.75 (brs, 2H), 3.96 (s, 3H); HPLC 97.7 %; HRMS (ESI) m/z calcd for 7 8 $C_{30}H_{22}N_2O_2S$ [M+H]⁺. 485.1474, found 485.1485. 9 7-(Furan-2-yl)-3-(4-phenoxyphenyl)thieno[3,2-c]pyridin-4-amine (12f). 12f was synthesized by the general method described above compound **12a** (38 mg, 63.0 %). Mp: 10 163.4-165.1 °C; MS m/z 385.1 [M+H]⁺; ¹H NMR (400 MHz, CDCl₃) δ ppm 8.05 (s, 1H), 11 12 7.89 (s, 1H), 7.56 (s, 1H), 7.44-7.38 (dd, J=8.04 Hz, J=8.46 Hz, 4H), 7.21-7.18 (t, 13 J=12.04 Hz, 2H), 7.11-7.08 (dd, J=3.20 Hz, J=4.00 Hz, 4H), 6.83 (s, 1H), 4.74 (brs, 2H); HPLC 97.5 %; HRMS (ESI) m/z calcd for C₂₃H₁₆N₂O₂S [M+H]⁺. 385.1005, found 14 385.1018. 15

16**7-Cyclopropyl-3-(4-phenoxyphenyl)thieno[3,2-c]pyridin-4-amine** (12g). 12g was17synthesized by the general method described above compound 12a (21 mg, 42.0 %). Mp:18180.5-182.4 °C; MS m/z 359.1 [M+H]⁺; ¹H NMR (400 MHz, CDCl₃) δ ppm 7.69 (s, 1H),197.40 (s, 4H), 7.15 (s, 2H), 7.09 (s, 4H), 4.66 (brs, 2H), 1.97 (s, 1H), 0.97 (s, 2H), 0.75 (s,202H). HPLC 95.8 %; HRMS (ESI) m/z calcd for $C_{22}H_{18}N_2OS$ [M+H]⁺. 359.1212, found21359.1211.

7-Butyl-3-(4-phenoxyphenyl)thieno[3,2-c]pyridin-4-amine 22 (12h). 12h was 23 synthesized by the general method described above compound **12a** (21 mg, 42.0 %). Mp: 147.5-148.6 °C; MS m/z 375.2 [M+H]⁺; ¹H NMR (400 MHz, CDCl₃) δ ppm 7.72 (s, 1H), 24 7.43-7.37 (dd, J=8.04 Hz, J=7.82 Hz, 4H), 719-7.15 (t, J=15.60 Hz, 1H), 7.12 (s, 1H), 25 711-7.07 (t, J=15.80 Hz, 4H), 4.56(br s, 2H), 719-7.15 (t, J=15.60 Hz, 1H), 2.78-2.75(t, 26 J=12.00 Hz, 2H), 1.77-1.70 (m, 2H), 1.45-1.38 (m, 2H), 0.98-0.95 (q, J=12.40 Hz, 3H); 27 HPLC 95.4 %; HRMS (ESI) m/z calcd for $C_{23}H_{22}N_2OS [M+H]^+$. 375.1526, found 28 375.1538. 29

30 Tert-butyl 4-(4-amino-3-(4-phenoxyphenyl)thieno[3,2-c]pyridin-7-yl)-1H-pyrazole

-1-carboxylate (13). A solution of 11 (150 mg, 0.337 mmol), 1-Boc-pyrazole-4-boronic
acid pinacol ester (100 mg, 0.337 mmol) and Pd(dppf)Cl₂·CH₂Cl₂ (27 mg, 0.0337 mmol)
in 1,4-dioxane (10 mL) was degassed with nitrogen for 5 min followed by addition
Na₂CO₃ (107 mg, 2M in water) under continuous flow of nitrogen. The reaction mixture
was stirred at 80 °C for 1 h, the catalyst was removed by filtration through Celite and the
filtrate concentrated to a residue. The residue was used in the next reaction without
further purification. MS m/z 485.1 [M+H]⁺.

3-(4-Phenoxyphenyl)-7-(1H-pyrazol-4-yl)thieno[3,2-c]pyridin-4-amine (14a). 8 Α mixture of the crude product 13 (100 mg, 0.21 mmol) was stirred in CF₃COOH (3 mL) at 9 10 r t for 2 h, after completion of reaction, the reaction mixture was diluted with the mixed ice and water, and then 1M NaOH solution was added to alkalify to pH 7-8, the 11 12 precipitated solid was filtered, and concentrated under reduced pressure. The crude product was purified on a silica gel column using (5-10 % CH₃OH/ DCM) as eluent to 13 afford **14a** (59 mg, 75 %) as a gray solid. Mp: 232.3-234.2 °C; MS m/z 385.1 [M+H]⁺. ¹H 14 NMR (400 MHz, DMSO-*d*₆) δ ppm 13.09 (brs, 1H), 8.15 (brs, 1H), 8.06 (s, 1H), 7.95 (br 15 16 s, 1H), 7.54 (s, 1H), 7.48-7.43 (dd, J=7.20 Hz, J=8.20 Hz, 4H), 7.22-7.18 (t, J=6.80 Hz, 1H), 7.14-7.12 (dd, J=2.80 Hz, J=2.80 Hz, 4H), 5.42 (brs, 2H); HPLC 99.7 %; HRMS 17 (ESI) m/z calcd for $C_{22}H_{16}N_4OS [M+H]^+$.385.1117, found 385.1128. 18

19

7-(1-Methyl-1H-pyrazol-4-yl)-3-(4-phenoxyphenyl)thieno[3,2-c]pyridin-4-amine

(14b). A suspension of 14a (100 mg, 0.26 mmol), CH₃I (55 mg, 0.39 mmol) and K₂CO₃ 20 (71.7 mg, 0.52 mmol) in DMF (5 mL) was stirred for 12 h at rt. After completion of 21 reaction, the reaction mixture was diluted with the mixed ice and water, the precipitated 22 23 solid was filtered off and the crude product was purified on a silica gel column using 24 (20-50%) ethyl acetate/ petroleum ether) as eluent to afford 14b (19mg, 42.0%) as a white solid. Mp: 201-203 °C; MS m/z 456.2 $[M+H]^+$; ¹H NMR (400 MHz, DMSO- d_6) 25 δppm 8.17 (s, 1H), 8.07 (br s, 2H), 7.93 (s, 1H), 7.56 (s, 1H), 7.51-7.43 (dd, J=8.00 Hz, 26 J=8.20 Hz, 4H), 7.22-7.19 (t, J=6.40 Hz, 1H), 7.14-7.12 (dd, J=4.40 Hz, J=4.00 Hz, 4H), 27 5.44 (brs, 2H), 4.86 (s, 2H), 2.65 (s, 1H); HPLC 98.6%; HRMS(ESI) m/z calcd for 28 $C_{25}H_{21}N_5O_2S$ [M+H]⁺. 456.1488, found 456.1501. 29

30 7-(1-Ethyl-1H-pyrazol-4-yl)-3-(4-phenoxyphenyl)thieno[3,2-c]pyridin-4-amine

1 (14c). 14c was synthesized by the general method described above compound 14b (90 mg, 87.1 %) as a white solid. Mp: 180.5-182.0 °C; MS m/z 399.2 $[M+H]^+$. ¹H NMR (400) 2 MHz, CDCl₃) δ ppm 7.91-7.90 (d, *J*=6.20 Hz, 1H), 7.76 (s, 1H), 7.65 (s, 1H), 7.43-7.38 3 (dd, J=8.40 Hz, J=7.80 Hz, 4H), 7.21-7.15 (dd, J=6.00 Hz, J=7.60 Hz, 2H), 7.12-7.07 (dd, 4 J=4.80 Hz, J=5.20 Hz, 4H), 5.20 (brs, 2H), 4.00 (s, 3H); HPLC 95.3 %; HRMS (ESI) m/z 5 6 calcd for C₂₃H₁₈N₄OS [M+H]⁺.399.1274, found 399.1276. 7-(1-Butyl-1H-pyrazol-4-yl)-3-(4-phenoxyphenyl)thieno[3,2-c]pyridin-4-amine 7 (14d). 14d was synthesized by the general method described above compound 14b (38.5 8 mg, 83.0 %). Mp: 160-162 °C; MS m/z 413.2 $[M+H]^+$; ¹H NMR (400 MHz, CDCl₃) δ 9 ppm 8.02 (s, 1H), 7.90 (s, 1H), 7.78 (s, 1H), 7.45-7.38 (dd, J=8.40 Hz, J=8.00 Hz, 4H), 10 7.18-7.16 (t, J=8.20 Hz, 2H), 7.11-7.08 (dd, J=4.20 Hz, J=4.40 Hz, 4H), 4.68 (brs, 2H), 11 12 4.28-4.27 (q, J=4.00 Hz, 2H), 1.60-1.56 (t, J=2.40 Hz, 3H); HPLC 99.0 %; HRMS (ESI) 13 m/z calcd for C₂₄H₂₀N₄OS [M+H]⁺.413.1430, found 413.1432. 14 7-(1-Benzyl-1H-pyrazol-4-yl)-3-(4-phenoxyphenyl)thieno[3,2-c]pyridin-4-amine (14e). 14e was synthesized by the general method described above compound 14b (18.3 15 mg, 57.2 %). Mp: 165-167 °C; MS m/z 441.2 $[M+H]^+$; ¹H NMR (400 MHz, CDCl₃) δ 16 ppm 8.00 (s, 1H), 7.89 (s, 1H), 7.76 (s, 1H), 7.44-7.38 (dd, J=8.40 Hz, J=8.00 Hz, 4H), 17 7.20-7.18 (t, J=8.20 Hz, 2H), 7.12-7.08 (dd, J=4.20 Hz, J=4.40 Hz, 4H), 4.87 (brs, 2H), 18 19 4.23-4.219(t, J=14.0 Hz, 2H), 1.93 (m, 2H), 1.41 (m, 2H), 1.00-0.97 (t, J=9.20 Hz, 3H); HPLC 95.5 %; HRMS (ESI) m/z calcd for C₂₆H₂₄N₄OS [M+H]⁺.441.1670, found 20 441.1673. 21 7-(1-(3-Methoxypropyl)-1H-pyrazol-4-yl)-3-(4-phenoxyphenyl)thieno[3,2-c]pyridi 22 **n-4-amine** (14f). 14f was synthesized by the general method described above compound 23 **14b** (21 mg, 86.5 %). Mp: 182-183.5 °C; MS m/z 475.2 [M+H]⁺; ¹H NMR (400 MHz, 24 25 CDCl₃) δppm 8.02 (s, 1H), 7.95 (s, 1H), 7.77 (s, 1H), 7.44-7.30(m, 9H), 7.19-7.15(m, 2H), 7.11-7.08 (dd, J=4.20 Hz, J=4.40 Hz, 4H), 5.41 (s, 2H), 4.67 (s, 2H); HPLC 95.6 %; 26

27 HRMS (ESI) m/z calcd for $C_{29}H_{22}N_4OS$ [M+H]⁺. 475.1587, found 475.1588.

28 2-(4-(4-Amino-3-(4-phenoxyphenyl)thieno[3,2-c]pyridin-7-yl)-1H-pyrazol-1-yl)-N-

29 methylacetamide (14g). 14g was synthesized by the general method described above

compound **14b** (23 mg, 65.2 %). Mp: 145-147 °C; MS m/z 457.2 [M+H]⁺; ¹H NMR (400 1 2 MHz, DMSO-*d*₆) δ ppm 8.17 (s, 1H), 8.05 (s, 1H), 7.91 (s, 1H), 7.56 (s, 1H), 7.50-7.43 3 (dd, J=8.80 Hz, J=8.40 Hz, 4H), 7.22-7.19 (t, J=12.4 Hz, 1H), 7.15-7.11 (dd, J=4.40 Hz, J=4.20 Hz, 4H), 4.86 (brs, 2H), 4.33 (t, 2H), 3.40 (t, 2H), 3.39 (s, 3H), 2.20 (t, 2H); 4 HPLC 96.6 %; HRMS (ESI) m/z calcd for C₂₆H₂₄N₄O₂S [M+H]⁺. 457.1692, found 5 6 457.1700. 4-(4-Amino-3-(4-phenoxyphenyl)thieno[3,2-c]pyridin-7-yl)-N,N-dimethyl-1H-pyra 7 **zole-1-sulfonamide** (14h). 14h was synthesized by the general method described above 8

compound 14b (18 mg, 88.5 %). Mp: 170-172 °C; MS m/z 492.2 [M+H]⁺; ¹H NMR (400 MHz, DMSO-*d*₆) δ ppm 8.61 (s, 1H), 8.43 (s, 1H), 8.22 (s, 1H), 7.63 (s, 1H), 7.51-7.43
(dd, *J*=8.80 Hz, *J*=8.40 Hz, 4H), 7.23-7.19 (t, *J*=12.4 Hz, 1H), 7.15-7.13 (dd, *J*=4.40 Hz, *J*=4.20 Hz, 4H), 5.81 (brs, 2H), 2.93 (s, 6H); HPLC 98.5 %; HRMS (ESI) m/z calcd for
C₂₄H₂₁N₅O₃S₂ [M+H]⁺. 492.1158, found 492.1172.

- 14 **1-(4-(4-Amino-3-(4-phenoxyphenyl)thieno[3,2-c]pyridin-7-yl)-1H-pyrazol-1-yl)eth** 15 **anone (14i). 14i** was synthesized by the general method described above compound **14b** 16 (25 mg, 62.5 %). Mp: 169-171 °C; MS m/z 427.2 $[M+H]^+$; ¹H NMR (400 MHz, 17 DMSO-*d*₆) δ ppm 8.70 (s, 1H), 8.48 (s, 1H), 8.28 (s, 1H), 7.61 (s, 1H), 7.52-7.44 (dd, 18 *J*=8.40 Hz, *J*=8.40 Hz, 4H), 7.24-7.20 (t, *J*=12.4 Hz, 1H), 7.16-7.13 (dd, *J*=4.40 Hz, 19 *J*=4.20 Hz, 4H), 5.66 (brs, 2H), 2.71 (s, 3H); HPLC 98.5 %; HRMS (ESI) m/z calcd for 20 C₂₄H₁₈N₄O₂S [M+H]⁺. 427.1223, found 427.1225.
- 21 (3-(Phenylcarbamoyl)phenyl)boronic acid (16a). A suspension of 3-boronobenzoic acid (1.0 g, 5.9 mmol), aniline (0.65g, 7 mmol), Et₃N (1.29 g, 12 mmol) and PyBop (3.6 22 23 g, 7mmol) in DMF(10 mL) was stirred for 12 h at r t. After completion of reaction, the 24 solution was diluted with H_2O (15 mL), and then the product was extracted three times with EtOAc (50 mL). The combined organic layer was dried over Na₂SO₄, and the 25 solvent was removed in vacuo, the crude product was purified on a silica gel column 26 using (1-5 %) CH₃OH/ DCM as eluent to afford 16a (1.47 g, 75 %) as a white solid. MS 27 m/z 241.2 [M+H]⁺. 28
- (4-(Phenylcarbamoyl)phenyl)boronic acid (16b). 16b was synthesized by the
 general method described above compound 16a (1.53 g, 40.8 %). MS m/z 241.2 [M+H]⁺.

3-(4-Aminothieno[3,2-c]pyridin-3-yl)-N-phenylbenzamide (17a). 17a was
 synthesized by the general method described above compound 12a (0.89 g, 78.3 %). MS
 m/z 345.2 [M+H]⁺.

4 4-(4-Aminothieno[3,2-c]pyridin-3-yl)-N-phenylbenzamide (17b). 17b was
5 synthesized by the general method described above compound 12a (1.21 g, 83.1 %). MS
6 m/z 345.2 [M+H]⁺.

- 3-(4-Amino-7-iodothieno[3,2-c]pyridin-3-yl)-N-phenylbenzamide (18a). 18a was
 synthesized by the general method described above compound 11 (0.41 g, 47.1 %). MS
 m/z 470.2 [M+H]⁺.
- 10 **4-(4-Amino-7-iodothieno[3,2-c]pyridin-3-yl)-N-phenylbenzamide** (18b). 18b was 11 synthesized by the general method described above compound 11 (0.56 g, 50.3 %). MS 12 m/z 470.2 $[M+H]^+$. ¹H NMR (400 MHz, DMSO-*d*₆) δ ppm 10.41 (s, 1H), 8.10-8.08 (d, *J* 13 = 4.80 Hz, 2H), 8.06 (s, 1H), 7.82-7.80 (d, *J* = 8.00 Hz, 2H), 7.69 (s, 1H), 7.65-7.63 (d, *J* 14 = 8.00 Hz, 2H), 7.39-7.35 (t, *J* = 16.00 Hz, 2H), 7.14-7.12 (d, *J* = 8.00 Hz, 1H), 5.62 (brs, 15 2H).
- 3-(4-Amino-7-(1-(2-(methylamino)-2-oxoethyl)-1H-pyrazol-4-yl)thieno[3,2-c]pyrid 16 in-3-yl)-N-phenylbenzamide (19a). 19a was synthesized by the general method 17 described above compound 13 (21 mg, 45.1 %). Mp: 171-173 °C; MS m/z 483.2 [M+H]⁺; 18 19 ¹H NMR (400 MHz, DMSO- d_6) δ ppm 10.35 (s, 1H), 8.21 (s, 1H), 8.13-8.08 (m, 4H), 7.97 (s, 1H), 7.79-7.77 (d, J = 8.00 Hz, 2H), 7.75-7.68 (q, 3H), 7.38-7.34 (d, J = 16.00 Hz, 20 2H), 7.13-7.11 (d, J = 8.00 Hz, 1H), 5.41 (s, 2H), 4.88 (s, 2H), 2.65 (s, 3H). HPLC 21 22 96.1 %; HRMS (ESI) m/z calcd for $C_{26}H_{22}N_6O_2S [M+H]^+$. 483.1597, found 483.1601. 23 4-(4-Amino-7-(1-(2-(methylamino)-2-oxoethyl)-1H-pyrazol-4-yl)thieno[3,2-c]pyrid 24 in-3-yl)-N-phenylbenzamide (19b). 19b was synthesized by the general method described above compound **13** (27 mg, 49.8 %). Mp: 178-180 °C; MS m/z 483.2 [M+H]⁺; 25 26 ¹H NMR (400 MHz, DMSO- d_6) δ ppm 10.42 (s, 1H), 8.21 (s, 1H), 8.13-8.10 (m, 4H), 7.97 (s, 1H), 7.83-7.81 (d, J = 8.00 Hz, 2H), 7.68-7.66 (d, 3H), 7.40-7.36 (d, J = 16.00 Hz, 27 2H), 7.14-7.12 (d, J = 8.00 Hz, 1H), 5.47 (s, 2H), 4.88 (s, 2H), 2.65 (s, 3H). HPLC 98.2%; 28 HRMS (ESI) m/z calcd for $C_{26}H_{22}N_6O_2S [M+H]^+$. 483.1598, found 483.1603. 29 30 3-Chloro-N-(4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)phenyl)benzamide

1 (21). 3-chlorobenzoyl chloride (175 mg, 1 mmol) in anhydrous THF (2 mL) was added dropwise to a solution of 4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)aniline (219 mg, 2 3 1 mmol) and Et₃N (152 g, 1.5 mmol) in 10 mL anhydrous THF of cooled to 0°C, and the 4 reaction mixture was stirred at 0 °C for 1 h and at room temperature for 2 h. The solution 5 was diluted with H_2O (5 mL), and then the product was extracted three times with EtOAc (20 mL). The combined organic layer was dried over Na_2SO_4 , and the solvent was 6 removed in vacuo, the residue was used in the next reaction without further purification. 7 8 MS m/z 357.2 [M+H]⁺.

N-(4-(4-Aminothieno[3,2-c]pyridin-3-yl)phenyl)-3-chlorobenzamide (22). 22 was
synthesized by the general method described above compound 10 (151 mg, 53.7 %). MS
m/z 379.1 [M+H]⁺.

N-(4-(4-Amino-7-iodothieno[3,2-c]pyridin-3-yl)phenyl)-3-chlorobenzamide (23).
13 was synthesized by the general method described above compound 3 (102 mg, 43.2 %). MS m/z 505.1 [M+H]⁺.

15 N-(4-(4-Amino-7-(1-methyl-1H-pyrazol-4-yl)thieno[3,2-c]pyridin-3-yl)phenyl)-3-c 16 hlorobenzamide (24a). 24a was synthesized by the general method described above 17 compound 12a (30 mg, 38.5 %). Mp: 221-223 °C; MS m/z 460.1 [M+H]⁺; ¹H NMR (400 18 MHz, CDCl₃) δ ppm 8.01 (s, 1H), 7.96 (s, 1H), 7.89-7.88 (m, 2H), 7.78-7.75 (t, J = 12.4019 Hz, 4H), 7.57-7.44 (m, 4H), 4.73(s, 2H), 4.01 (s, 3H). HPLC 99.0 %; HRMS (ESI) m/z 20 calcd for C₂₄H₁₈ClN₅OS [M+H]⁺. 460.0993, found 460.1002.

21 **N-(4-(4-Amino-7-(1-(2-(methylamino)-2-oxoethyl)-1H-pyrazol-4-yl)thieno[3,2-c]p** 22 **yridin-3-yl)phenyl)-3-chlorobenzamide (24b)**. **24b** was synthesized by the general 23 method described above compound **12a** (38 mg, 35.8 %). Mp: 220-222 °C; MS m/z 517.1 24 $[M+H]^+$; ¹H NMR (400 MHz, CDCl₃) δ ppm 8.01-8.00 (m, 3H), 7.96-7.86 (m, 2H), 25 7.80-7.88 (d, *J* = 12.40 Hz, 3H), 7.55-7.47 (m, 4H), 7.21 (s, 1H), 6.45 (s, 1H), 5.01 (s, 26 2H), 4.90 (s, 2H), 2.85 (s, 3H). HPLC 99.0 %; HRMS (ESI) m/z calcd for 27 $C_{26}H_{21}CIN_6O_2S [M+H]^+$. 517.1208, found 517.1225.

28

29 **5.2 BTK enzymatic assay**

30 The HTRF kinase assay (components supplied as kit by Cisbio) was chosen for BTK

enzyme assays. It uses time resolved fluorescence resonance energy transfer (TR-FRET) 1 to detect production of a phosphorylated substrate. A peptide substrate is labeled with a 2 biotin that can bind to XL665 labeled streptavidin, and the anti-phosphoresidue antibody 3 is labeled with Eu⁺. Upon phosphorylation of the substrate, the antibody binds to 4 5 phosphorylated substrate that enables TR-FRET detection in homogenous assay format. All the reagents used for the BTK kinase assays including their resources are BTK kinase 6 7 (Invitrogen), HTRF kinEASE-TK kit (Cisbio Bioassays), ATP(Sigma), DTT(Sunshine), MgCl₂ and MnCl₂ (Sigma). The assay buffer was composed of 50 mM HEPES (pH 7.0), 8 5 mM MgCl₂, 5mM DTT, 0.1 % NaN3, 0.1 % BSA and 0.1 mM orthovanadate. The 9 10 HTRF assays were preformed according to the manual in the kit. All reagents were dispensed into each well plate according to the orders as follow: (1) BTK enzyme: 11 0.5ng/µl, 4 µl; (2) Each compound as well as control: 0.008-50 mM; (3) Reagent: 22.4 12 μ M ATP and 0.15 μ M substrate, 2 μ l; (4) Inculation: Ambient, at 25°C, 5 min; (5) 13 Reagent: antibody and XL-665, 8 µl. Then following 1 hour incubation at room 14 15 temperature fluorescence was measured on the PHERAStar FS microplate reader (BMG Lab Technologies). Signal was expressed in terms of HTRF ratio (fluorescence intensity 16 at 665 nm/fluorescence intensity at 620 nm). 17

18

19 **5.3 ELISA-based kinase selectivity assay.**

In vitro kinase inhibition assays were carried out as described elsewhere. Briefly, 20 96-well plates were pre-coated with 0.2 mg/mL poly (Glu-Tyr, 4:1) (Sigma) overnight at 21 37°C. 0.05 mL aliquot of 0.01 mmol/L ATP diluted in kinases and reaction medium were 22 23 added. Test compounds at various concentrations diluted in 0.01mL of 1 % DMSO (V/V) were added. The reaction mixtures were incubated for 60 min at 37°C. The wells were 24 25 washed with PBS containing 0.1 % T-PBS for three times. The 0.1 mL Phosphorylated 26 tyrosine substrate was added. The kinase reaction was incubated for for 60 min at 37°C, and then the wells were washed three times and then anti-mouse IgG (ZSGB-BIO; 27 ZB-2305; 0.1 mL /well) coupled with horseradish peroxidase (HRP) was added and 28 incubated for another 30 min. The TMB reaction was quenched by addition of 0.05mL of 29 30 2 M H₂SO₄. The optical density was measured at 450 nm by an ELISA reader. The IC₅₀

values were calculated for test compounds by using a regression analysis of the
 concentration/inhibition data.

3

4

5.4 Pharmacokinetic Profiles of compound 14b in SD rats

5 Compound 14g were administered to 3 male SD rats (weight ranging from 180 g to 240 g) at doses of 2.5 mg/kg for iv administration as well as doses of 10 mg/kg for po 6 7 administration for another 3 male SD rats. The dosing volume was 5 mL/kg. After administration, blood samples were collected at the point including 2 min, 5 min, 15 min, 8 9 30 min, 1 h, 2 h, 4 h, 6 h, 8 h, 10 h, and 24 h for analyses, the collected blood samples were centrifuged at 4000 rpm for 5 min at 4 °C, and then analyzed after protein 10 precipitation. LC/MS/MS analysis of compound 14g was performed under optimized 11 12 conditions to obtain the best sensitivity and selectivity of the analyte in selected reaction 13 monitoring mode (SRM) containing an internal standard. Plasma concentration-time data 14 were measured by a noncompartmental approach using the software WinNonlin Enterprise, version 5.2 (Pharsight Co., Mountain View, CA). 15

16

17 **5.5 Molecular docking**

The molecular docking procedure was referred to CDOCKER protocol within 18 19 Discovery Studio 2.5. The protein-ligand complex crystal structure of **B43** bound to Btk 20 (PDB code 3GEN) was chosen as the template. The initial 3D conformation of compound 21 14g was optimized in the ChemBio3D Ultra using MM2 energy minimization method. 22 The whole Btk enzyme was typed with CHARMm force field and the hydrogen atoms 23 were added. The water molecules were deleted except those occupying at the ligand 24 pocket. Binding site was defined as a sphere. Compound 14g was docked in the defined 25 active site. A set of 10 starting random conformations was generated. The top scoring pose of compound 14g was retained and selected for analyzing. 26

27

28 References and Notes

- [1] Lou, Y.; Owens, T.D.; Kuglstatter, A.; Kondru, R.K.; Goldstein, D.M. J. Med. Chem. 2012, 55,
 4539.
- 31 [2] Xu, D.; Kim, Y.; Postelnek, J.; Vu, M.D.; Hu, D.Q.; Liao, C.; Bradshaw, M.; Hsu, J.; Zhang,

1		J.; Pashine, A.; Srinivasan, D.; Woods, J.; Levin, A.; O'Mahony, A.; Owens, T.D.; Lou, Y.;
2		Hill, R.J.; Narula, S.; DeMartino, J.; Fine, J.S.; J. Pharmacol. Exp. Ther. 2012, 341, 90.
3	[3]	Ghia, P.; Lancet Oncol. 2014, 15, 1043.
4	[4]	https://www.thomson-pharma.com/.
5	[5]	Akinleye, A.; Chen, Y.; Mukhi, N.; Song, Y.; Liu, D.; J. Hematol. Oncol. 2013, 6, 59.
6	[6]	Robak, T.; Robak, E. Expert, Opin. Investig. Drugs. 2012, 21, 921.
7	[7]	Hendriks, R.W.; Nat. Chem. Biol. 2011, 7, 4.
8	[8]	Pan, Z.; Scheerens, H.; Li, S.; Schultz, B.E.; Sprengeler, P.A.; Burrill, L.C.; Mendonca, R.V.;
9		Sweeney, M.D.; Scott, K.C.K.; Grothaus, P.G.; Jeffery, D.A.; Spoerke, J.M.; Honigberg, L.A.;
10		Young, P.R.; Dalrymple, S.A.; Palmer, J.T.; ChemMedChem. 2007, 2, 58.
11	[9]	Kim, K.H.; Maderna, A.; Schnute, M.E.; Hegen, M.; Mohan, S.; Miyashiro, J.; Lin, L.; Li, E.;
12		Keegan, S.; Lussier, J.; Wrocklage, C.; Nicherson-Nutter, C.L.; Wittwer, A.J.; Soutter, H.;
13		Caspers, N.; Han, S.; Kurumbail, R.; Dunussi-Joannopoulos, K.; Douhan, J.; Wissner, A.;
14		Bioorg. Med. Chem. Lett. 2011, 21, 6258.
15	[10]	Di Paolo, J.A.; Huang, T.; Balazs, M.; Barbosa, J.; Barck, K.H.; Bravo, B.J.; Carano, R.A.;
16		Darrow, J.; Davies, D.R.; DeForge, L.E.; Diehl, L.; Ferrando, R.; Gallion, S.L.; Giannetti, A.M.;
17		Gribling, P.; Hurez, V.; Hymowitz, S.G.; Jones, R.; Kropf, J.E.; Lee, W.P.; Maciejewski, P.M.;
18		Mitchell, S.A.; Rong, H.; Staker, B.L.; Whitney, J.A.; Yeh, S.; Young, W.B.; Yu, C.; Zhang, J.;
19		Reif, K.; Currie, K.S. Nat. Chem. Biol. 2011, 7, 41.
20	[11]	Young, W.B.; Barbosa, J.; Blomgren, P.; Bremer, M.C.; Crawford, J.J.; Dambach, D.; Gallion,
21		S.; Hymowitz, S.G.; Kropf, J.E.; Lee, S.H.; Liu, L.; Lubach, J.W.; Macaluso, J.; Maciejewski,
22		P.; Maurer, B.; Mitchell, S.A.; Ortwine, D.F.; Paolo, J.D.; Reif, K.; Scheerens, H.; Schmitt, A.;
23		Sowell, C.G.; Wang, X.; Wong, H.; Xiong, J.M.; Xu, J.; Zhao, Z.; Currie, K.S.; Europ. J. Med.
24		Chem. 2014 , 86, 664.
25	[12]	Puig de la Bellacasa, R.; Roue, G.; Balsas, P.; Perez-Galan, P.; Teixido, J.; Colomer, D.; Borrel,
26		J.I.; Bioorg. Med. Chem. Lett. 2014, 24, 2206.
27	[13]	Shi, Q.; Tebben, A.; Dyckman, A.J.; Li, H.; Liu, C.; Lin, J.; Spergel, S.; Burke, J.R.;
28		McIntyre, K.W.; Olini, G.C.; Strnad, J.; Surti, N.; Muckelbauer, J.K.; Chang, C.; An, Y.;
29		Cheng, L.; Ruan, Q.; Leftheris, K.; Carter, P.H.; Tino, J.; De Lucca, G.V. Bioorg. Med. Chem.
30		Lett. 2014 , 24, 2206.
31	[14]	Lou, Y.; Han, X.; Kuglstatter, A.; Kondru, R.K.; Sweeney, Z.K.; Soth, M.; McIntosh, J.;
32		Litman, R.; Suh, J.; Kocer, B.; Davis, D.; Park, J.; Frauchiger, S.; Dewdney, N.; Zecic, H.;
33		Taygerly, J.P.; Sarma, K.; Hong, J.; Hill, R.J.; Gabriel, T.; Goldstein, D.M.; Owens, T.D. J.
34		Med. Chem. 2015 , 58, 512.
35	[15]	Lou, Y.; Sweeney, Z.K.; Kuglstatter, A.; Davis, D.; Goldstein, D.M.; Han, X.; Hong, J.; Kocer,
36		B.; Kondru, R.K.; Litman, R.; McIntosh, J.; Sarma, K.; Suh, J.; Taygerly, J.; Owens, T.D.;
37		Bioorg. Med. Chem. Lett. 2015, 25, 367.
38	[16]	Zhao, X.; Xin, M.; Huang, W.; Ren, Y.; Jin, Q.; Tang, F.; Jiang, H.; Wang, Y.; Yang, J.; Mo, S.;
39		Xiang, H.; Bioorg. Med. Chem. 2015, 23, 348.
40	[17]	Zhao, X.; Huang, W.; Wang, Y.; Xin, M.; Jin, Q.; Cai, J.; Tang, F.; Zhao, Y.; Xiang, H.; Bioorg.
41		Med. Chem. 2015 , 23, 891.
42	[18]	M. S. Schnute, A. Huangr, E. Saiah. Bruton's Tyrosine Kinase (BTK), in: J.I. Levin, S. Laufer
43		(Eds.), Anti-Inflammatory Drug Discovery, RSC Publishing Inc., Southend-on-Sea, United
44		Kingdom, 2012 , pp. 297–326.
45	[19]	Honigberg, L. A.; Smith, A. M.; Sirisawad, M.; Verner, E.; Loury, D.; Chang, B.; Li, S.; Pan,

1	Z.; Thamm, D. H.; Miller, R. A.; Buggy, J. J. Proc. Natl. Acad. Sci. U. S. A. 2010, 107,
2	13075-13080.
3	[20] Wan, H.L.; Wang, Z.R.; Li, L.L.; Cheng, C.; Ji, P.; Liu, J.J.; Zhang, H.; Zou, J.; Yang, S.Y.;
4	Chem. Biol. Drug Des. 2012, 80, 366.
5	[21] Marcotte, D.J.; Liu, Y.T.; Arduini, R.M.; Hession, C.A.; Miatkowski, K.; Wildes, C.P.; Cullen,
6	P.F.; Hong, V.; Hopkins, B.T.; Mertsching, E.; Jenkins, T.J.; Romanowski, M.J.; Baker, D.P.;
7	Silvian, L.F.; Protein Sci. 2010 , 19, 429.
8	[22] (a) Xin, M.; Wen, J.; Tang, F.; Tu, C.; Shen, H.; Zhao, X. Bioorg. Med. Chem. Lett. 2013 , 23,
9	6777; (b) Xin, M.; Wen, J.; Tang, F.; Tu, C.; Huang, W.; Shen, H.; Zhao, X.; Cheng, L.; Wang,
10	M.; Zhang, L. Bioorg. Med. Chem. Lett. 2014 , 24, 983; (c) Xin, M.; Zhang, L.; Tang, F.; Tu, C.;
11	Wen, J.; Zhao, X.; Liu, Z.; Cheng, L.; Shen, H. Bioorg. Med. Chem. 2014, 22, 1429; (d) Xin,
12	M.; Zhang, L.; Shen, H; Wen, J.; Tu, C.; Liu, Z.; Cheng, L.; Zhao, X. Med. Chem. Res. 2014,
13	23, 3/84. (e) Zhang, L.; Xin, M.; Wen, J.; Tang, F.; Tu, C.; Shen, H.; Wei, P.; Chin. J. Org.
14	Chem. 2014, 34, 1407.
13	[25] MIyazaki, T.; Nakano, M.; Salo, H.; Truesdale, A. I.; Stuart, J.D.; Nartev, E.N.; Hightower,
10	K.E.; Kane-Carson, L.; Bloorg. Med. Chem. Lett. 2007, 17, 250.
17	





1



- 2 3
- Figure 3. Docking mode of compound 14g with BTK enzyme (PDB code: 3GEN). Selected residues
- 4 Thr-474, Glu-475, Met-477, H₂O-36 and Cys-481 are shown. Hydrogen bonds are shown as green
- 5 dashed lines.

- 6
- 7



1

2 Scheme 1. Reagents and conditions: (a) Pd(PPh₃)₄, Na₂CO₃, EtOH, H₂O, toluene, 90 °C, 3 h, 96 %; (b) NIS, DMF, r t,

3 1 h, 70 %; (c) R₁B(OH)₂, Pd(PPh₃)₄, Na₂CO₃, DME, H₂O, 80 °C, 2-3 h, 35-90 %; (d) 1-Bocpyrazole-4-boronic acid

- 4 pinacol ester, Pd(dppf)Cl₂CH₂Cl₂, Na₂CO₃, 1,4-dioxane, 100 °C,1-2 h, 55 %; (e) CF₃COOH, DCM, rt, 12 h, 92 %; (f)



NF

1

Scheme 2. Reagents and conditions: (a) aniline, PyBop, Et₃N, DMF, r t, 12 h; (b) compound 9, Pd(PPh₃)₄,
 Na₂CO₃, DME, H₂O, 80 °C, 2-3 h; (c) NIS, DMF, r t, 1h, 50-55 %; (d) Pd(dppf)Cl₂·CH₂Cl₂, Na₂CO₃,

4 1,4-dioxane, 100 °C, 2 h; (e) m-Chlorobenzoyl chloride, Et₃N, DCM, 0-25 °C, 2 h, 85%; (f) Pd(PPh₃)₄, Na₂CO₃,

5 DME, H₂O, 80 °C, 2-3 h, 80.2 %; (g) NIS, DMF, rt, 2 h, 57 %; (h) 1-Methylpyrazole-4-boronic acid pinacol

6 ester for 24a, N-methyl-2-(4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)-1H-pyrazol-1-yl)acetamide for 24b,

 $7 \qquad Pd(dppf)Cl_2 \cdot CH_2Cl_2, Na_2CO_3, 1,4\text{-ioxane}, 100\ ^{o}C, 2\ h.$

- 8
- 9

1

2 Table 1. In vitro BTK enzymatic inhibition for thieno[3,2-c]pyrid-4-amine derivatives



4 ^a Values are means of three experiments 5

1

2 Table 2. Kinase selectivity profiles of compounds 14b and 14g

	Comeda	- t			IC ₅₀ (nM)				
	Compus	structures	BTK	JAK2	JAK3	KDR	c-Met		
	14b	N N N N	107.4	>10000	>10000	343	>10000	R	
	14g	$\underset{h \rightarrow N}{\overset{N}{\overset{N}{\overset{N}{\overset{N}{\overset{N}{\overset{N}{\overset{N}{\overset$	12.8	>10000	>10000	284	>10000	2	
3									
4						G	2		
5									
		\mathbf{X}							
	\mathcal{N}								

$\frac{14g}{10 \text{ (po)}^{b}} = \frac{2547.3}{855.5} = \frac{1459.1}{1365.7} = \frac{900.0}{13320.6} = \frac{1716.4}{8670.1} = \frac{0.43}{0.43} = \frac{0.67}{0.36} = \frac{0.11}{0.67} = \frac{1}{23.4}$ ^a Compound 14g was formulated using 5 % DMA + 10 % solutol. ^b Compound 14g was formulated using 0.5 % HMPC + 0.5 % Twen-80.
14g 10 (po) ^b 855.5 1365.7 13320.6 8670.1 1.48 1.01 0.67 23.4 ^a Compound 14g was formulated using 5 % DMA + 10 % solutol. ^b Compound 14g was formulated using 0.5 % HMPC + 0.5 % Twen-80.
^a Compound 14g was formulated using 5 % DMA + 10 % solutol. ^b Compound 14g was formulated using 0.5 % HMPC + 0.5 % Twen-80.

Table 3. PK properties study for compound 14g in SD rats ^a 1

