

RESEARCH ARTICLE

3-Methyl-2-phenyl-1-substituted-indole derivatives as indomethacin analogs: design, synthesis and biological evaluation as potential anti-inflammatory and analgesic agents

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Abstract

In a new group of 3-methyl-2-phenyl-1-substituted-indole derivatives (**10a–f**), the indomethacin analogs were prepared via the Fisher indole synthesis reaction of propiophenone with appropriately substituted phenylhydrazine hydrochloride. This is followed by the insertion of the appropriate benzyl or benzoyl fragment. All the synthesized compounds were evaluated for their anti-inflammatory (*in vitro* and *in vivo*) and analgesic activities. The methanesulphonyl derivatives **10d**, **e** and **f** showed the highest anti-inflammatory (*in vitro* and *in vivo*) and analgesic activities. In addition, molecular docking studies were performed on compounds **10a–f** and the results were in agreement with that obtained from the *in vitro* COX inhibition assays. The significant anti-inflammatory and analgesic activities exhibited by **10d** and **10e** warrant continued preclinical development as potential anti-inflammatory and analgesic agents.

Keywords

Anti-inflammatory activity, cyclooxygenase, indomethacin, molecular modeling

History

Received 30 December 2014
Revised 12 February 2015
Accepted 12 February 2015
Published online 23 March 2015

Introduction

Non-steroidal anti-inflammatory drugs (NSAIDs) produce their activity via inhibition of cyclooxygenase (COX) catalyzed biotransformation of arachidonic acid to pro-inflammatory prostaglandins (PGs) and thromboxanes (TXs)^{1–3}. Cyclooxygenase enzyme exists in two distinct isoforms, a constitutive form (COX-1) and an inducible form (COX-2); the constitutive COX-1 is responsible for the maintenance of physiological functions, such as protection of gastric mucosa, vascular homeostasis and platelet aggregation^{4–6}. Unlike COX-1, the COX-2 isoform is induced in response to mitogenic and pro-inflammatory stimuli and it is responsible for the progression of inflammation^{7,8}. Thus, selective COX-2 inhibitor drugs (coxibs) as celecoxib (**1**), rofecoxib (**2**) and valdecoxib (**3**) were more useful for the treatment of inflammation and inflammation-associated disorders than the non-selective traditional NSAIDs as aspirin (**4**), ibuprofen (**5**) and indomethacin (**6**) (Figure 1). Unfortunately, some cardiovascular side effects including the increased incidences of high blood pressure and myocardial infarction accompanied the highly selective COX-2 inhibitors were attributed to the biochemical changes in the COX pathway^{9,10}. These adverse effects

resulted in the withdrawal of rofecoxib and valdecoxib^{9,10} (Figure 1).

Indomethacin (**6**) is one of the most potent NSAIDs, which are effective against rheumatoid arthritis, ankylosing spondylitis, osteoarthritis of large joints and other types of inflammatory diseases¹¹. At the same time, it is also one of the most ulcerogenic NSAIDs because of its high COX-1 selectivity and the acidic nature of the drug¹². From our point of view, there are two main approaches to prevent and/or treat gastroenteropathy associated with indomethacin as an example of traditional NSAIDs; the first comprises synthesis of prodrugs, where a nitric oxide (NO)-donating moiety is covalently attached to NSAID (indomethacin). This moiety should reduce the gastrointestinal toxicity through the protective effect of locally released NO on the gastric mucosa. In this regard, we previously reported some prodrugs of indomethacin and other NSAIDs, which showed higher safety profiles than of the parent NSAIDs^{13,14}. The second strategy is based on maintaining the potency of the indomethacin by keeping the main scaffold of the drug with trials to increase COX-2 selectivity via the modifications of the side groups. Accordingly, the current work described the synthesis, molecular modeling studies, *in vitro* evaluation as COX-1/COX-2 inhibitors, and the *in vivo* anti-inflammatory (AI) activity of a new series of *N*-substituted indole derivatives as indomethacin analogs **10a–f** in which; (i) the chlorobenzoyl moiety of indomethacin in position 1, which is important for anti-inflammatory activity¹⁵, is maintained in **10b**, **10e**, replaced with benzoyl in **10a**, **10d** or replaced with benzyl in **10c**, **10f**, (ii) the methyl group in position 2 was replaced with the phenyl group and this replacement is expected

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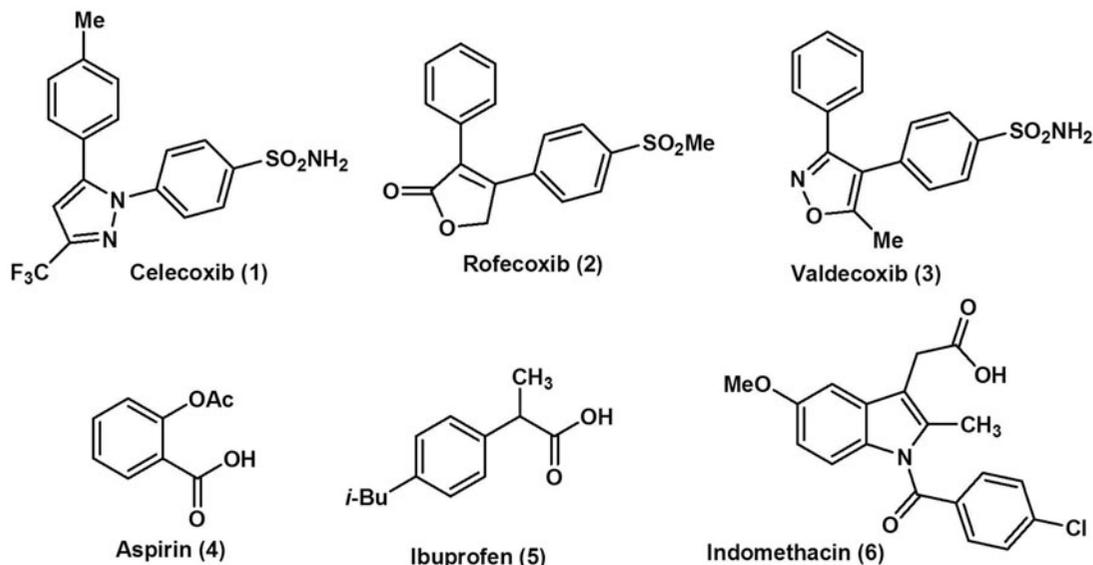
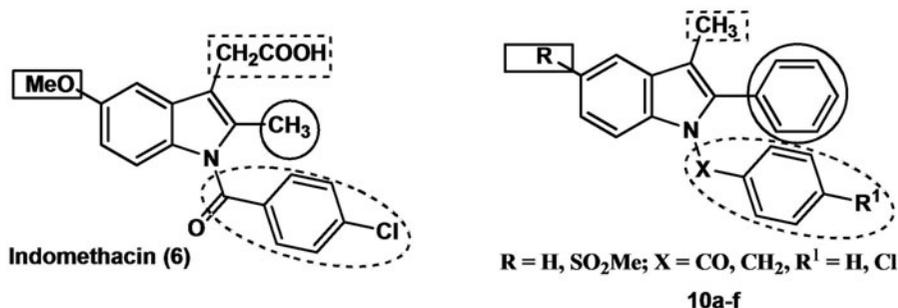


Figure 1. Chemical structures of some selective cyclooxygenase-2 (COX-2) inhibitor drugs (1–3) and some traditional non-selective NSAIDs (4–6).

Figure 2. Chemical structures of the traditional NSAID indomethacin (6) and the designed indomethacin analogs 10a–f.



to maximize the interaction with the hydrophobic residues within COX-2 active site and to enhance COX-2 selectivity, since the proposed compounds will be too large to fit into the smaller COX-1 active site, (iii) the $-\text{CH}_2\text{COOH}$ moiety in position 3 was replaced with a methyl group and this replacement is expected to decrease the acidity of the new compounds, which will consequently decrease the local action exerted by the direct contact of the new compounds with gastric mucosa, and (iv) the methoxy group in position 5 is replaced with H in **10a–c** or with COX-2 pharmacophore methanesulfonyl moiety (SO_2Me) in **10d–f** to study the effect of SO_2Me moiety on COX selectivity and anti-inflammatory activity (Figure 2).

Results and discussion

Chemistry

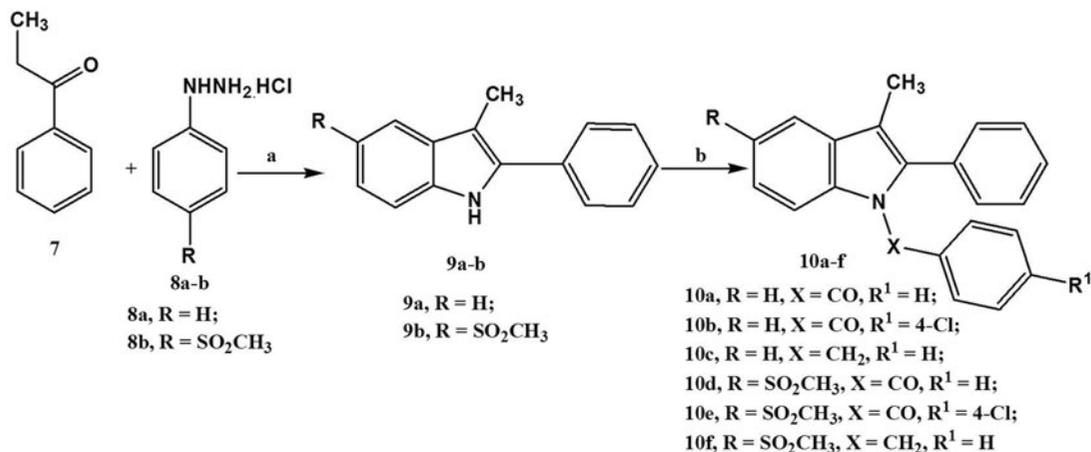
The synthesis of the target 1-substituted-2-phenyl-3-methyl indole derivatives (**10a–f**) was accomplished in accordance with the sequence of reactions depicted in Scheme 1. Reaction of propiophenone (**7**) with either phenylhydrazine hydrochloride (**8a**) or 4-methylsulfonylphenylhydrazine hydrochloride (**8b**) in ethanol under the Fischer indole synthesis reaction conditions afforded the respective 2-phenyl-3-methylindole derivative **9a** (79%) or **9b** (82%). Replacement of the chlorine atom present in benzoyl chloride, 4-chlorobenzoyl chloride or benzyl chloride by the indole derivatives (**9a–b**) in DMF under basic conditions (NaH) afforded the target products (**10a–f**) in moderate

yield (42–58%). All the prepared compounds have been characterized by IR, ^1H NMR, ^{13}C NMR, mass spectra and elemental analyses.

Biological evaluation

Anti-inflammatory activity

In vitro cyclooxygenase inhibition assay. *In vitro* structure–activity relationships acquired by this group of indomethacin derivatives (**10a–f**) showed that they are relatively weak inhibitors of COX-1 ($\text{IC}_{50} = 7.78\text{--}42.36\ \mu\text{M}$) when compared with the reference drug indomethacin ($\text{IC}_{50} = 0.63\ \mu\text{M}$). However, they are moderately-to-highly potent inhibitors of COX-2 ($\text{IC}_{50} = 1.65$ to $8.58\ \mu\text{M}$ range) in comparison with indomethacin ($\text{IC}_{50} = 11.63\ \mu\text{M}$) (Table 1). Compounds having a SO_2Me moiety (COX-2 pharmacophore) were generally more potent inhibitors of COX-2 than the corresponding analogs free of SO_2Me (**10e** > **10b**; **10f** > **10c**; but **10a** and **10d** were approximately of the same potency). In general, all compounds were more selective to COX-2 enzyme with selectivity index (S.I.) = 0.90 to 25.67 compared to indomethacin (COX-2 S.I. = 0.055). The selectivity could be attributed to the size of compounds (**10a–f**), which is too large to fit into the smaller COX-1 active site. Moreover, the presence of COX-2 pharmacophore in some analogs (**10d–f**) increased the selectivity of COX-2 isozyme. The most potent COX-2 inhibitor (**10e**) with $\text{IC}_{50} = 1.65\ \mu\text{M}$ and S.I. = 25.65 had indomethacin as a chlorobenzoyl moiety in addition to SO_2Me moiety



Scheme 1. Reagents and conditions: (a) glacial acetic acid, reflux, 10h; (b) benzoyl chloride, 4-chlorobenzoyl chloride or benzyl chloride, NaH, DMF, RT, overnight.

Table 1. *In vitro* COX-1 and COX-2 inhibition of compounds **10a–f**, and reference drug (indomethacin).

Compounds	COX Inhibition (IC ₅₀ μM)*		Selectivity index†
	COX-1	COX-2	
10a	10.39 ± 1.36	6.54 ± 1.61	1.58
10b	18.72 ± 1.64	8.21 ± 2.10	2.28
10c	7.78 ± 2.01	8.58 ± 2.14	0.90
10d	32.23 ± 2.36	7.98 ± 1.91	13.65
10e	42.36 ± 2.45	1.65 ± 1.02	25.67
10f	17.65 ± 2.79	2.36 ± 1.58	7.47
Indomethacin	0.63 ± 0.02	11.36 ± 1.6	0.055

*Values are expressed as mean ± SEM (*n* = 3).

†Selectivity index (COX-1 IC₅₀/COX-2 IC₅₀).

(COX-2 pharmacophore) which was 467 times more COX-2 selective than indomethacin (COX-2 IC₅₀ = 11.36 μM, S.I. = 0.055).

In vivo anti-inflammatory activity. The anti-inflammatory activity of the test compounds compared to indomethacin was tested using the carrageen-induced rat paw edema test, which is widely used as a primary test for the screening of new anti-inflammatory agents. Results indicated that after 1 h, indomethacin derivatives **10a, b** and **c** showed percentage anti-inflammatory activity of about 5.8–26.9%, while indomethacin derivatives with SO₂Me **10d, e** and **f** showed higher percentage of anti-inflammatory activity (67.9–73.5%). After 3 h, compounds **10a, b** and **c** showed percentage anti-inflammatory activity of 8.8–17.4% and compounds **10d, e** and **f** showed percentage anti-inflammatory activity of 66.8–80.1%. Similarly after 6 h, compounds **10a, b** and **c** showed percentage activity of 6.1–21.2%, while compounds **10d, e** and **f** showed percentage anti-inflammatory activity of 57.9–78.8%. 1-Benzyl-3-methyl-2-phenyl-1*H*-indole (**10c**) displayed the lowest anti-inflammatory activity at all-time intervals (5.8–8.8%), while (4-chlorophenyl)-(5-methanesulfonyl-3-methyl-2-phenyl-indol-1-yl)-methanone (**10e**) showed the highest anti-inflammatory activity (73.5%) at one-hour interval (more than indomethacin, 71.2%). The benzoyl derivative (**10d**) showed good anti-inflammatory activity at three- and six-hour intervals (80.1 and 78.8%, respectively) comparable to that of indomethacin (86.3 and 88.4%, respectively) and it was the most potent among the six compounds **10a–f** at these time intervals. These data indicate that; (i) presence of

Table 2. Anti-inflammatory activities of compounds **10a–f**, and reference drug (indomethacin) in carrageenan-induced rat paw edema assay.

Compounds	Anti-inflammatory activity (%)		
	1 h	3 h	6 h
10a	26.9	16.2	21.2
10b	17.3	17.4	15.7
10c	5.8	8.8	6.1
10d	73.1	80.1	78.8
10e	73.5	71.8	71.8
10f	67.9	66.8	57.9
Indomethacin	71.2	86.3	88.4

All test compounds (**10a–f**, indomethacin) were administered (10 mg/kg) 30 min prior to testing. The anti-inflammatory activity % is expressed according to the following equation:

AI (%) = $(V_c - V_t/V_c) \times 100$, where V_c represents the paw volume of control group of animals and V_t represents the paw volume in drug treated animals.

methanesulfonyl (SO₂Me) moiety (in compounds **10d, e** and **f**) increases the anti-inflammatory activity for this class of compounds, and (ii) maintaining chlorobenzoyl moiety of indomethacin as in compound **10e** or its replacement with benzoyl moiety (compound **10d**) maintains or increases the anti-inflammatory activity, while its replacement with benzyl moiety (compound **10f**) sharply decreased the anti-inflammatory activity (Table 2).

Analgesic activity

The analgesic activity of the target compounds (**10a–f**) and indomethacin as a reference drug was also investigated using acetic acid-induced writhing test. Acetic acid-induced writhing was significantly reduced in mice receiving SO₂Me containing compounds **10d, e** and **f**, and the degree of inhibition of the writhing response by these compounds was 49.1, 65.1 and 59.3%, respectively, compared to the vehicle-treated control group. The analgesic activity of SO₂Me free compounds (**10a, b**, and **c**) was relatively low (33.0, 30.2 and 18.9%, respectively) (Figure 3). From these results, the analgesic activities of these compounds are clearly parallel to their anti-inflammatory activities.

Molecular modeling

To know the plausible mode of interactions of the prepared compounds **10a–f** within the COX-1 and COX-2 isozymes,

Figure 3. Analgesic effect of test compounds **10a–f** compared to indomethacin using acetic acid-induced writhing in mice. Data represent the mean value \pm SD of four mice per group. Statistical comparisons between basal and post-drug values were analyzed for statistical significance using the one-way ANOVA followed by Dunnett's test and denoted by * $p < 0.05$, # $p < 0.01$.

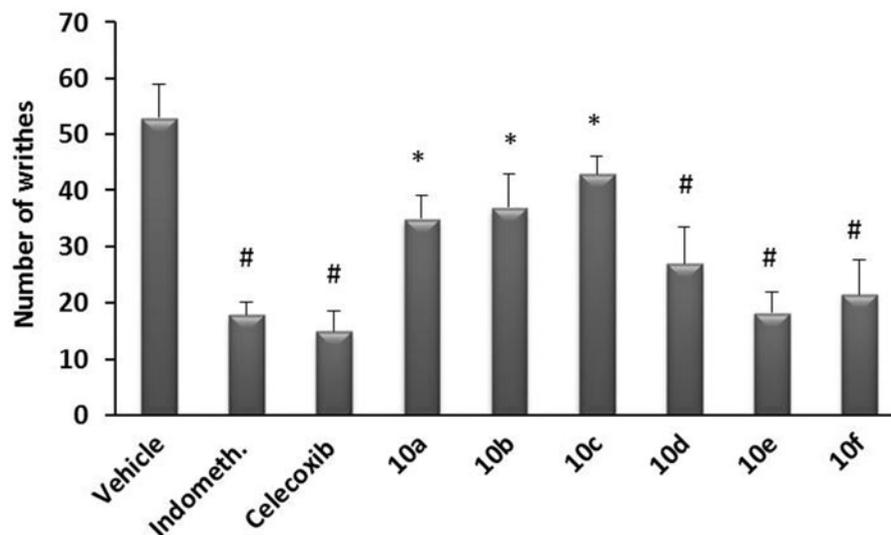


Table 3. Molecular modeling data for compounds **10a–f**, indomethacin and valdecoxib during docking in the COX-1 (PDB:ID 4COX) and COX-2 (PDB:ID 2AW1) active sites.

Compound	COX-1			COX-2		
	Affinity (kcal/mol)	Distance (in Å) from main residue	Functional group	Affinity (kcal/mol)	Distance (in Å) from main residue	Functional group
10a	-5.5167	2.66	Arg120	-14.5783	2.70	Asn67
10b	-9.0055	-	-	-11.4647	2.76	Asn67
10c	-12.1998	-	-	-15.6805	3.03	Asn62
10d	-5.6238	2.70	Arg120	-20.8529	2.90	His94
10e	-5.0689	-	-	-20.7351	3.05	Thr199
10f	-4.7538	-	-	-21.7493	2.53	Thr199
Indomethacin	-17.0967	2.96	Tyr385	-	2.93	His94
Valdecoxib	-	2.76	Arg120	-17.0929	2.96	Thr199
					2.83	Thr199
					3.06	His94
					2.90	Thr199
					3.28	His94
					2.73	Thr199
					2.80	Thr199

molecular docking experiments were performed using X-ray, crystal structure data for COX-1 and COX-2 were obtained from the protein data bank^{16,17}. Indomethacin (**6**) – as a selective COX-1 inhibitor – was used as a ligand for COX-1 isoform while, valdecoxib (**3**) was used as a ligand for COX-2 isoform. The main interactions of indomethacin with COX-1 resulted in two main residues Tyr385 and Arg120 amino acids, sequentially forming two hydrogen bonding interactions with –N and –CO of indomethacin at a distance equal to 2.96 and 2.76 Å, respectively. On the other hand, docking valdecoxib into COX-2 isozyme afforded three hydrogen bonding interactions; (i) NH₂ with His94 (3.28 Å), (ii) NH₂ with Thr199 (2.73 Å), and (iii) SO₂ with Thr199 (2.80 Å).

The docking results including the energy associated with intermolecular interactions (affinity in kcal/mol) obtained upon computational docking for all compounds (**10a–f**, indomethacin and valdecoxib) within COX-1 and COX-2 active sites and the

hydrogen bonding interactions between the amino acid residues and functional groups of the compounds are summarized in Table 3. For COX-1, all six compounds **10a–f** indicated lower binding interactions (affinity in kcal/mol ranges from –5.5167 to –12.1998 with no or one hydrogen bonding interaction) than indomethacin (–17.0967 with two hydrogen bonding interactions). In case of COX-2, while compounds **10a–c** showed appreciable binding interactions (affinity in kcal/mol ranges from –11.4647 to –15.6805 with one or two hydrogen bonding interactions), compounds **10d–f** (have SO₂Me as COX-2 pharmacophore) showed excellent binding interactions (affinity in kcal/mol ranges from –20.7351 to –21.7493 with two or three hydrogen bonding interactions) in comparison with valdecoxib (–17.0929 with three hydrogen bonding interactions). All the docked compounds were in response to COX-2 rather than COX-1 and that is consistent with their experimentally observed potent COX-2 inhibition (Figure 4).

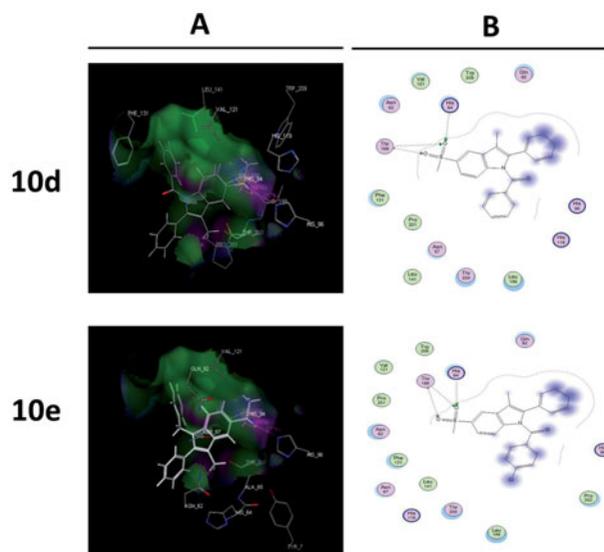


Figure 4. Binding of the most active compounds **10d** and **e** inside COX-2 active site. (A) The proposed binding mode inside the active site of COX-2 resulting from docking, the most important amino acids are shown together with their respective numbers. (B) 2D interaction.

Conclusion

Six new compounds **10a–f**, as indomethacin derivatives, were synthesized for biological evaluation as anti-inflammatory and analgesic activities. The *in vitro* anti-inflammatory activity studies showed that all compounds were more COX-2 selective (COX-2 S.I. = 0.90 to 25.67) than indomethacin (COX-2 S.I. = 0.055), especially compound **10e** (467 times more COX-2 selective than the reference drug indomethacin). While, the *in vivo* anti-inflammatory activity studies indicated that compounds **10d**, **e** and **f** (have SO₂Me as COX-2 pharmacophore) displayed good anti-inflammatory activity (57.9–80.1%) comparable to that of indomethacin (71.2–88.4%), especially **10d** (73.1–80.1%) and **10e** (71.8–73.5%). In addition, the docking experiments showed that all the docked compounds were fitted to COX-2 rather than COX-1 and compounds **10d–f** (having SO₂Me as COX-2 pharmacophore) showed excellent binding interactions (–20.7351 to –21.7493 kcal/mol) with two or three hydrogen bonding interactions in comparison with the COX-2 selective drug (valdecoxib, –17.0929 kcal/mol and three hydrogen bonding interactions), which was in parallel correlation with the *in vitro* COX inhibition evaluation. Also, the analgesic activities of these compounds **10a–f** are clearly parallel to their anti-inflammatory activities. Because of their relatively potent anti-inflammatory and analgesic activities, the compounds **10d** and **10e** warrant continued preclinical development as potential anti-inflammatory and analgesic agents.

Experimental

Chemistry

Melting points were determined on a Thomas-Hoover capillary apparatus and are uncorrected. Infrared (IR) spectra were recorded as films on NaCl plates using a Nicolet 550 Series II Magna FT-IR spectrometer (Middleton, WI). ¹H NMR and ¹³C NMR spectra were measured on a Bruker Avance III 400 MHz (Bruker BioSpin AG, Fällanden, Switzerland) for ¹H and 100 MHz for ¹³C with BBFO Smart Probe and Bruker 400 AEON Nitrogen-Free Magnet, Faculty of Pharmacy, Beni-Suef University, Egypt, in DMSO-*d*₆ with TMS as the internal standard, where *J* (coupling constant) values are estimated in

Hertz (Hz) and chemical shifts were recorded in ppm on δ scale. Mass spectra (MS) were recorded on a Hewlett Packard 5988 spectrometer (Palo Alto, CA). Microanalyses for C, H and N were carried out on Perkin-Elmer 2400 analyzer (Perkin-Elmer, Norwalk, CT) at the Micro analytical unit of Cairo University, Egypt, and all compounds were within $\pm 0.4\%$ of the theoretical values. All other reagents, purchased from the Aldrich Chemical Company (Milwaukee, WI), were used without further purification. 4-Methylsulfonylphenylhydrazine hydrochloride (**8b**) was prepared according to a previous procedure¹⁸.

General procedure for synthesis of 3-methyl-2-phenylindole derivatives **9a–b**

A mixture of propiophenone (**7**, 1.3 g, 0.01 mol) and the appropriate phenyl hydrazine hydrochloride (**8a** or **8b**, 0.01 mol) in glacial acetic acid (30 mL) was heated under reflux for 10 h. After cooling, the reaction mixture was poured into ice-cold water. The separated solid was filtered, dried and crystallized from acetone to give pure compounds **9a** and **9b**. Physical and spectral data are listed below.

3-Methyl-2-phenyl-1H-indole (9a). Buff solid¹⁹; Yield 79%; m.p. 136–138 °C; IR (KBr) 3331 (NH), 3023 (CH aromatic), 2924 (CH aliphatic) cm⁻¹; ¹H NMR (DMSO-*d*₆) δ 2.44 (s, 3H, CH₃), 7.04 (t, 1H, *J* = 7.2 Hz, phenyl H-4), 7.12–7.16 (m, 1H, indole H-5), 7.35–7.42 (m, 2H, indole H-6, H-7), 7.50–7.56 (m, 3H, phenyl H-3, H-5 and indole H-4), 7.70 (d, 2H, *J* = 7.6 Hz, phenyl H-2, H-6), 11.19 (s, 1H, NH, D₂O exchangeable); ¹³C NMR (DMSO-*d*₆) δ 10.30, 107.22, 111.48, 118.88, 119.03, 122.01, 127.43, 127.95, 129.16, 129.85, 133.55, 134.19, 136.38; EIMS (*m/z*) 208 (M+1, 17.88%), 207 (M⁺, 100%), 206 (M-1, 89.24%). Anal. Calcd for C₁₅H₁₃N: C, 86.92; H, 6.32; N, 6.76. Found: C, 86.79; H, 6.22; N, 7.01.

5-Methanesulfonyl-3-methyl-2-phenyl-1H-indole (9b). Brown solid; Yield 82%; m.p. 197–199 °C; IR (KBr) 3413 (NH), 3048 (CH aromatic), 2912, 2862 (CH aliphatic), 1307, 1158 (SO₂) cm⁻¹; ¹H NMR (DMSO-*d*₆) δ 2.47 (s, 3H, CH₃), 3.19 (s, 3H, SO₂CH₃), 7.42 (t, 1H, *J* = 7.6 Hz, phenyl H-4), 7.53–7.59 (m, 3H, phenyl H-3, H-5 and indole H-7), 7.65–7.72 (m, 3H, phenyl H-2, H-6 and indole H-6), 8.16 (s, 1H, indole H-4), 11.81 (s, 1H, NH, D₂O exchangeable); ¹³C NMR (DMSO-*d*₆) δ 10.08 (CH₃), 45.02 (SO₂CH₃), 108.84, 112.02, 119.23, 120.19, 128.21, 128.23, 129.18, 129.32, 131.51, 132.57, 136.95, 138.44; EIMS (*m/z*) 286 (M+1, 24.21%), 285 (M⁺, 100%). Anal. Calcd for C₁₆H₁₅NO₂S: C, 67.34; H, 5.30; N, 4.91. Found: C, 67.55; H, 5.58; N, 4.69.

General procedure for synthesis of 3-methyl-2-phenyl-1-substituted-indole derivatives (**10a–f**)

To a solution of the respective 2-phenyl indole derivative **9a** or **9b** (2.5 mmol) in dry DMF (5 mL), NaH (0.11 g, 4.5 mmol) was slowly added with stirring at room temperature for 30 min. Then, the reaction flask was cooled down on a bath of ice followed by a slow addition for a solution of the appropriate alkyl or acyl halide (2.5 mmol) in DMF (5 mL) and the mixture was stirred at room temperature overnight. The reaction mixture was poured into ice-cold water and extracted with hexane (4 \times 10 mL). The organic layer was dried and crystallized from methanol/chloroform mixture (4:1) to give pure compounds **10a–f**. Physical and spectral data are listed below.

(3-Methyl-2-phenyl-indol-1-yl)-phenyl-methanone (10a). Buff solid; Yield 54%; m.p. 300–302 °C; IR (KBr) 3273 (CH aromatic), 2924 (CH aliphatic), 1675 (C=O) cm⁻¹; ¹H NMR

(DMSO- d_6) δ 2.50 (s, 3H, CH_3), 7.04 (t, 1H, $J = 7.2$ Hz, phenyl H-4), 7.11–7.15 (m, 1H, indolyl H-5), 7.34–7.42 (m, 2H, indolyl H-6, H-7), 7.45 (m, 7H, phenyl H-2, H-3, H-5, H-6, indolyl H-4 and benzoyl H-3, H-5), 7.68–7.95 (m, 3H, benzoyl H-2, H-4, H-6); ^{13}C NMR (DMSO- d_6) δ 9.63 (CH_3), 113.82, 116.35, 119.78, 123.29, 124.89, 127.93, 128.27, 128.46, 129.61, 129.77, 130.10, 131.19, 132.43, 133.22, 135.37, 136.95, 169.47 (C=O); EIMS (m/z) 312 (M+1, 3.43%), 311 (M^+ , 13.64%), 105 ($C_7H_5O^+$, 100%). Anal. Calcd for $C_{22}H_{17}NO$: C, 84.86; H, 5.50; N, 4.50. Found: C, 85.13; H, 5.56; N, 4.38.

(4-Chlorophenyl)-(3-methyl-2-phenyl-indol-1-yl)-methanone

(10b). Buff solid; Yield 57%; m.p. 99–101 °C; IR (KBr) 3061 (CH aromatic), 2992 (CH aliphatic), 1685 (C=O) cm^{-1} ; 1H NMR (DMSO- d_6) δ 2.40 (s, 3H, CH_3), 7.10–7.34 (m, 8H, 5 phenyl protons and indolyl H-5, H-6, H-7), 7.52–7.85 (m, 5H, indolyl H-4 and 4 benzoyl protons); ^{13}C NMR (DMSO- d_6) δ 9.64 (CH_3), 114.05, 116.66, 119.81, 123.55, 125.15, 127.84, 128.51, 128.86, 130.21, 130.52, 131.95, 132.27, 134.19, 136.17, 136.89, 137.76, 168.63 (C=O); EIMS (m/z) 347 (M+2, 9.70%), 345 (M^+ , 25.82%), 139 ($C_7H_4ClO^+$, 100%). Anal. Calcd for $C_{22}H_{17}NO$: C, 76.41; H, 4.66; N, 4.05. Found: C, 76.35; H, 4.49; N, 4.23.

1-Benzyl-3-methyl-2-phenyl-1H-indole **(10c)**. Yellow solid²⁰; Yield 49%; m.p. 110–112 °C; IR (KBr) 3059, 3027 (CH aromatic), 2923, 2859 (CH aliphatic) cm^{-1} ; 1H NMR (DMSO- d_6) δ 2.23 (s, 3H, CH_3), 5.31 (s, 2H, CH_2), 6.83 (d, 2H, $J = 6.8$ Hz, benzyl H-2, H-6), 7.08–7.21 (m, 5H, benzyl H-3, H-4, H-5 and indole H-5, H-6), 7.33 (d, 1H, $J = 7.6$ Hz, indole H-7), 7.39–7.51 (m, 5H, phenyl protons), 7.57 (d, 1H, $J = 7.6$ Hz, indole H-4); ^{13}C NMR (DMSO- d_6) δ 9.76 (CH_3), 47.06 (CH_2), 108.76, 110.89, 119.13, 119.65, 122.23, 126.43, 127.40, 128.52, 128.76, 128.87, 129.05, 130.71, 131.84, 136.81, 137.70, 138.87; EIMS (m/z) 298 (M+1, 12.57%), 297 (M^+ , 53.35%), 91 ($C_7H_7^+$, 100%). Anal. Calcd for $C_{22}H_{19}N$: C, 88.85; H, 6.44; N, 4.71. Found: C, 88.93; H, 6.56; N, 4.63.

(5-Methanesulfonyl-3-methyl-2-phenyl-indol-1-yl)-phenyl-methanone **(10d)**. Buff solid; Yield 55%; m.p. 151–153 °C; IR (KBr) 3061 (CH aromatic), 2925, 2859 (CH aliphatic), 1677 (C=O), 1321, 1178 (SO_2) cm^{-1} ; 1H NMR (DMSO- d_6) δ 2.41 (s, 3H, CH_3), 3.19 (s, 3H, SO_2CH_3), 7.21–7.72 (m, 11H, phenyl protons, benzoyl protons and indolyl H-7), 7.76 (d, 1H, $J = 7.2$ Hz, indolyl H-6), 8.29 (s, 1H, indolyl H-4); ^{13}C NMR (DMSO- d_6) δ 10.08 (CH_3), 45.02 (SO_2CH_3), 108.84, 112.03, 116.31, 119.60, 120.20, 127.33, 128.23, 128.64, 129.18, 129.31, 129.49, 130.35, 130.82, 131.60, 132.57, 138.79, 169.56 (C=O); EIMS (m/z) 390 (M+1, 0.27%), 389 (M^+ , 0.93%), 285 ($C_{16}H_{15}NSO_2^+$, 100%). Anal. Calcd for $C_{23}H_{19}NO_3S$: C, 70.93; H, 4.92; N, 3.60. Found: C, 80.13; H, 5.04; N, 3.81.

(4-Chlorophenyl)-(5-methanesulfonyl-3-methyl-2-phenyl-indol-1-yl)-methanone **(10e)**. Buff solid; Yield 58%; m.p. 204–206 °C; IR (KBr) 3047 (CH aromatic), 2943, 2914 (CH aliphatic), 1672 (C=O), 1317, 1175 (SO_2) cm^{-1} ; 1H NMR (DMSO- d_6) δ 2.33 (s, 3H, CH_3), 3.27 (s, 3H, SO_2CH_3), 7.23–7.29 (m, 5H, phenyl protons), 7.37 (d, 2H, $J = 8.0$ Hz, benzoyl H-3, H-5), 7.56 (d, 2H, $J = 8.0$ Hz, benzoyl H-2, H-6), 7.81 (d, 2H, $J = 8.8$ Hz, indolyl H-7), 7.86 (d, 2H, $J = 8.8$ Hz, indolyl H-6), 8.29 (s, 1H, indolyl H-4); ^{13}C NMR (DMSO- d_6) δ 9.51 (CH_3), 44.58 (SO_2CH_3), 114.61, 116.61, 119.58, 123.30, 128.40, 128.69, 129.02, 130.24, 130.31, 131.48, 132.19, 133.35, 135.82, 138.42, 138.51, 139.00, 168.56 (C=O); EIMS (m/z) 425 (M+2, 4.52%), 423 (M^+ , 11.15%), 139 ($C_7H_4ClO^+$, 100%). Anal. Calcd for $C_{23}H_{18}ClNO_3S$: C, 65.17; H, 4.28; N, 3.30. Found: C, 65.32; H, 4.41; N, 3.21.

1-Benzyl-5-methanesulfonyl-3-methyl-2-phenyl-1H-indole

(10f). Buff solid; Yield 42%; m.p. 235–237 °C; IR (KBr) 3031 (CH aromatic) 2921, 2861 (CH aliphatic), 1343, 1182 (SO_2) cm^{-1} ; 1H NMR (DMSO- d_6) δ 2.47 (s, 3H, CH_3), 3.19 (s, 3H, SO_2CH_3), 5.40 (s, 2H, CH_2), 6.83 (d, 2H, $J = 7.6$ Hz, benzyl H-2, H-6), 7.19–7.21 (m, 3H, benzyl H-3, H-4, H-5), 7.41–7.65 (m, 7H, phenyl protons and indole H-6, H-7), 8.18 (s, 1H, indole H-4); ^{13}C NMR (DMSO- d_6) δ 10.06 (CH_3), 45.03 (SO_2CH_3), 47.35 (CH_2), 110.55, 111.59, 112.04, 119.38, 120.19, 127.66, 128.08, 128.23, 129.00, 129.12, 129.32, 130.73, 130.93, 132.16, 138.12, 138.72; EIMS (m/z) 376 (M+1, 13.95%), 375 (M^+ , 47.96%), 91 ($C_7H_7^+$, 100%). Anal. Calcd for $C_{23}H_{21}NO_2S$: C, 73.57; H, 5.64; N, 3.73. Found: C, 73.42; H, 5.53; N, 3.81.

Biological evaluation

Animals

Swiss albino mice (20–25 g) and Wistar rats (150–175 g) were obtained from the National Research Center, Cairo, Egypt. They were used throughout the study and were kept at controlled conditions (temperature 23 ± 2 °C, humidity $60 \pm 10\%$) at a 12/12 h light/dark cycle. All procedures relating to animal care and treatments were conducted in accordance with protocols approved by the Research Ethical Committee of Faculty of Pharmacy Beni-Suef University (2014-Beni-Suef, Egypt).

COX-1/COX-2 inhibition colorimetric assay

The *in vitro* inhibition of COX-1/COX-2 was measured using colorimetric COX (ovine) Inhibitor Screening Assay Kit (Cayman Chemical, Ann Arbor, MI) according to manufacturer's instructions. This assay directly measures $PGF_{2\alpha}$ that was produced by stannous chloride reduction of COX-derived PGH_2 by enzyme immunoassay. All assays were conducted in triplicates and IC_{50} values are the average of three determinations for each compound.

Carrageenan-induced rat paw edema assay

Rats were administered with vehicle, test compounds, indomethacin or celecoxib at a dose of 10 mg/kg by oral gavages. Immediately thereafter, the rats received 100 μ L of vehicle or carrageenan (1% in saline) s.c. on the plantar surface of the left hind paw under mild anesthesia, essentially, as reported before²¹. The paw edema was evaluated by measuring paw-volume changes at 1, 3 and 6 h after carrageenan injection using a plethysmometer. The right hind paw served as a reference of non-inflamed paw for comparison. Results are expressed as paw-volume change (mL).

Acetic acid-induced writhing test

The writhing tests were carried as described before²². Briefly, mice were administered orally vehicle, test compounds, indomethacin or celecoxib at a dose of 10 mg/kg 30 min prior to intraperitoneal administration of 0.7% v/v acetic acid solution (10 mL/kg body weight). The number of writhes (i.e. abdominal constriction followed by dorsiflexion and extension) occurring during a 20-min period beginning 5 min after acetic acid injection was measured. The results are expressed as the number of writhes per 20-min period.

Statistical analysis

Comparisons among the groups were analyzed for statistical significance using the one-way ANOVA followed by Dunnett's test. Data were represented as mean \pm standard deviation (SD). Differences were considered significant at $*p < 0.05$, and $**p < 0.01$, respectively.

Molecular modeling and docking

The crystal structures of indomethacin bound at COX-1 (PDB: ID 4COX)¹⁷ and valdecoxib bound at the COX-2 (PDB:ID 2AW1) active sites¹⁸ [obtained from the protein data bank at Research Collaboration for Structural Bioinformatics (RSCB) Protein Database (PDB)]. Docking of the co-crystallized ligand should be carried out to study the scoring energy(s), root mean standard deviation (RMSD) and amino acid interactions. The RMSD, which is a measure of superposing was 0.50 Å for the lead compounds. Docking was performed using London dG force and refinement of the results was done using force field energy. Preparation of the synthesized compounds for docking was achieved via their 3D structure built by Molecular Operating Environment (MOE, Version 2005.06, Chemical Computing Group Inc., Montreal, Quebec, Canada). Certain procedures were taken before docking, which include: 3D protonation of the structures, running conformational analysis using systemic search, selecting the least energetic conformer and applying the same docking protocol used with ligands. Docking for the synthesized compounds was applied. Amino acid interactions and the hydrogen bond lengths are summarized in Table 3.

Declaration of interest

The authors have declared no conflict of interest.

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