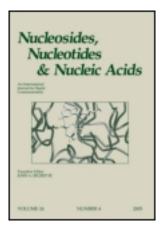
This article was downloaded by: [Texas A&M University Libraries] On: 24 June 2013, At: 20:44 Publisher: Taylor & Francis Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



# Nucleosides, Nucleotides and Nucleic Acids

Publication details, including instructions for authors and subscription information: http://www.tandfonline.com/loi/lncn20

### Design and Synthesis of LNA-Based Mercaptoacetamido-Linked Nucleoside Dimers

Vivek K. Sharma<sup>a</sup>, Sunil K. Singh<sup>a</sup>, Kapil Bohra<sup>a</sup>, Chandra Shekhar Reddy L.<sup>a</sup>, Vinod Khatri<sup>a</sup>, Carl E. Olsen<sup>b</sup> & Ashok K. Prasad<sup>a</sup> <sup>a</sup> Bioorganic Laboratory, Department of Chemistry, University of Delhi, Delhi, India

<sup>b</sup> Faculty of Life Sciences, Department of Natural Sciences, University of Copenhagen, Frederiksberg, Denmark Published online: 14 Apr 2013.

To cite this article: Vivek K. Sharma , Sunil K. Singh , Kapil Bohra , Chandra Shekhar Reddy L. , Vinod Khatri , Carl E. Olsen & Ashok K. Prasad (2013): Design and Synthesis of LNA-Based Mercaptoacetamido-Linked Nucleoside Dimers, Nucleosides, Nucleotides and Nucleic Acids, 32:5, 256-272

To link to this article: <u>http://dx.doi.org/10.1080/15257770.2013.783218</u>

### PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: <u>http://www.tandfonline.com/page/terms-and-conditions</u>

This article may be used for research, teaching, and private study purposes. Any substantial or systematic reproduction, redistribution, reselling, loan, sub-licensing, systematic supply, or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae, and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand, or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.



### DESIGN AND SYNTHESIS OF LNA-BASED MERCAPTOACETAMIDO-LINKED NUCLEOSIDE DIMERS

## Vivek K. Sharma,<sup>1</sup> Sunil K. Singh,<sup>1</sup> Kapil Bohra,<sup>1</sup> Chandra Shekhar Reddy L.,<sup>1</sup> Vinod Khatri,<sup>1</sup> Carl E. Olsen,<sup>2</sup> and Ashok K. Prasad<sup>1</sup>

<sup>1</sup>Bioorganic Laboratory, Department of Chemistry, University of Delhi, Delhi, India <sup>2</sup>Faculty of Life Sciences, Department of Natural Sciences, University of Copenhagen, Frederiksberg, Denmark

□ Three LNA-based mercaptoacetamido-linked nonionic nucleoside dimers  $T^{L}$ -S-T, T-S- $T^{L}$ , and  $T^{L}$ -S- $T^{L}$  have been synthesized by HOBT and HBTU catalyzed condensation of silyl-protected 2-S-(thymidin-5'-yl)mercaptoacetic acid or 2-S-(2'-O, 4'-C-methylenethymidin-5'-yl)mercaptoacetic acid with 3'-amino-3'-deoxy-5'-O-DMT-2'-O, 4'-C-methylenethymidine or with 3'-amino-3'-deoxy-5'-O-DMT-β-thymidine followed by desilylation of the protected dimers. The 3'-O-phosphoramidite derivative of one of the nucleoside dimers was successfully prepared by condensation with [P(-Cl)(-OCH\_2CH\_2CN){-N(1Pr)\_2}] in DCM in the presence of N,N-diisopropylethylamine (DIPEA), which is a building block for the preparation of mercaptoacetamido-linked oligonucleotides of therapeutic applications.

[Supplementary materials are available for this article. Go to the publisher's online edition of *Nucleosides, Nucleotides & Nucleic Acids* for the following free supplemental resource: supplementary information.doc.]

**Keywords** Locked nucleic acid; phosphate backbone modification; mercaptoacetamido-linkage; phosphoramidite-derivative

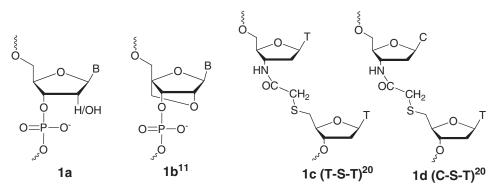
### INTRODUCTION

The recent development in antisense, antigene, and RNA interference technologies by using chemically modified oligonucleotides (ONs) has attracted a great deal of both chemists and biologists.<sup>[1–5]</sup> The modified

Address correspondence to Ashok K. Prasad, Bioorganic Laboratory, Department of Chemistry, University of Delhi, Delhi-110 007, India. E-mail: ashokenzyme@yahoo.com

Received 29 January 2013; accepted 4 March 2013.

We are thankful to the University of Delhi for providing financial support under DU-DST Purse and R&D Grants. We express our sincere gratitude to Dr. Vaijayanti A. Kumar and Mrs. Anita D. Gunjal, National Chemical Laboratory, Pune, for their help in the synthesis of phosphoramidite derivative of one of the mercaptoacetamido-linked nonionic nucleoside dimers. We are also thankful to CIF-USIC University of Delhi, Delhi, for NMR spectral data. V.K.S., S.K.S., K.B., and V.K. thank CSIR, New Delhi and C.R.L. thanks DBT, New Delhi for the award of JRF/SRF.



**FIGURE 1** Structure of DNA/RNA **1a**; LNA **1b**<sup>[11]</sup>; and Mercaptoacetamido-linked dimers **1c** and **1d**<sup>[20]</sup>. B = Nucleobases; T = Thymine; C = Cytosine.

oligonucleotides are also being increasingly utilized in nucleic acid nanotechnology, oligonucleotide-based diagnostics, gene-function determination, and drug target validation.<sup>[6–8]</sup> The novel utility of these agents resides in their ability to selectively prevent the expression of a particular diseaseassociated gene in a sequence specific manner. The ongoing synthetic studies have been focused on chemical modifications of backbone,<sup>[9]</sup> base,<sup>[10]</sup> and sugar<sup>[11]</sup> functionalities of the natural DNA/RNA (**1a**, Figure 1) and have resulted in significant progress toward establishing oligonucleotides as viable therapeutic agents. One such modification in the sugar moiety has resulted in locked nucleic acid (LNA **1b**, Figure 1), where the furanose conformation is locked in an *N*-type (C3'-endo) form by the introduction of a 2'-0,4'-C methylene-linkage.<sup>[11-13]</sup> LNA has been found to be very useful for antisense applications, since incorporation of one or more LNA monomer unit(s) into an ON shows extraordinary thermal stability when hybridized with either DNA, RNA, or LNA itself.<sup>[11,12,14]</sup>

Further, oligonucleotide analogues with differently modified backbone such as thioformacetal,<sup>[15]</sup> 5'-*N*-carbamate,<sup>[16]</sup> methylene(methy limino),<sup>[17]</sup> amide-,<sup>[18]</sup> triazole,<sup>[19]</sup> etc. have been designed and synthesized to circumvent the physical and biological limitations of natural phosphodiester linkage. Recently, Kumar et al.<sup>[20]</sup> have synthesized two 5-atom mercaptoacetamido-linked dimers, T-S-T 1c and C-S-T 1d (Figure 1), as compared to the 4-atom-linked natural DNA. The UV- $T_m$  experiments for binding affinity of these mixed backbone modified ONs with complementary DNA and RNA sequences revealed important results such as significantly higher RNA-binding selectivity ( $\Delta T_m$ /mod upto + 13°C) as compared to complementary DNA. Replacement of phosphate backbone with mercaptoacetamido linkage in an ONs have several distinct advantages in terms of its antisense properties, i.e., (i) it has strong binding affinity toward both natural DNA and RNA, which may be due to the neutral backbone that does not have any repulsive interactions with the anionic phosphate backbone;

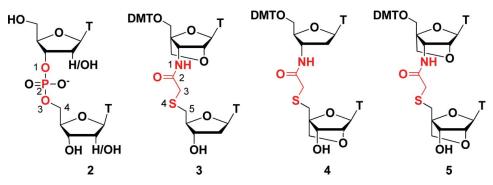
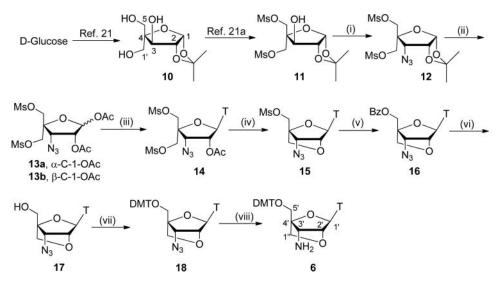


FIGURE 2 Structures of natural dinucleotide 2; designed LNA-based mercaptoacetamido-linked nonionic nucleoside dimers 3, 4, and 5. The natural phosphate linkage O-P-O in 2 has been replaced by mercaptoacetamido-linkage S-CH<sub>2</sub>-CO-NH in 3, 4, and 5, as shown in red (Color figure available online).

(ii) it provides a very high degree of nuclease resistance; and (iii) it enhances membrane permeability due to elimination of negative charge from the backbone.

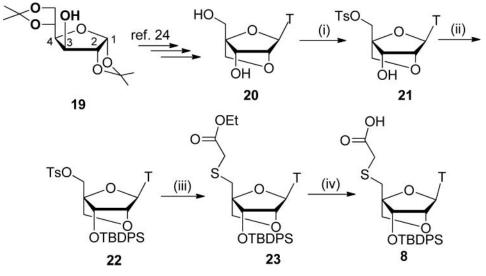
In this article we have designed and synthesized three LNA-based mercaptoacetamido-linked nonionic nucleoside dimers i.e., 1-(3'-deoxy-5'-O-DMT-2'-O,4'-C-methylenethymidin-3'-yl)-4-(5''-deoxy- $\beta$ -thymidin-3'-yl)-4-(5''-deoxy-2''-O,4''-C-methylenethymidin-5''-yl) mercaptoacetamide (T-S-T<sup>L</sup>, **4**); and 1-(3'-deoxy-5'-O-DMT-2'-O,4'-C-methylenethymidin-3'-yl)-4-(5''-deoxy-2''-O,4''-C-methylenethymidin-5''-yl) mercaptoacetamide (T<sup>L</sup>-S-T<sup>L</sup>, **5**); containing the features of both, LNA- and mercaptoacetamido-linked nucleoside dimers (Figure 2) and 3'-O-phosphoramidite derivative **27** of one nucleoside dimer, which is a building block for the preparation of mercaptoacetamido-linked oligonucleotides.

It was envisaged to synthesize the nucleoside dimer T<sup>L</sup>-S-T 3 by the condensation of 3'-amino-3'-deoxy-5'-O-DMT-2'-O.4'-C-methylenethymidine (6) with 2-S-( $\beta$ -thymidin-5'-yl)mercaptoacetic acid (9); nucleoside dimers T-S-T<sup>L</sup> 4 and T<sup>L</sup>-S-T<sup>L</sup> 5 by condensing 3'-amino-3'-deoxy-5'-O-DMT- $\beta$ thymidine (7); and nucleoside 6 with 2-S-(2'-O,4'-C-methylenethymidin-5'yl)mercaptoacetic acid (8). The aminonucleoside **6** has been synthesized 1,2-O-isopropylidene-5-O-methanesulfonyl-4-C-methanesulfonyloxyfrom methyl- $\beta$ -L-threofuranose 11, which was obtained from D-glucose through trihydroxyfuranoside 10 following the procedure of Matin<sup>[21]</sup> in an overall yields of 39%. The dimesylated compound 11 was converted into 3-azido-3-deoxy-1,2-O-isopropylidene-5-O-methanesulfonyl-4-C-methanesulfonyloxy methyl- $\alpha$ -D-*ribo*furanose 12 in two steps, i.e., triflation of the lone hydroxyl group using triflic anhydride in DCM-pyridine followed by nucleophilic displacement of the triflate with sodium azide in DMF in combined yields of 50%. Acetolysis of compound 12 with acetic acid-acetic anhydride-conc. H<sub>2</sub>SO<sub>4</sub> yielded mixture of anomers **13a–13b** in 84% yield, which on Vorbrüggen coupling<sup>[22]</sup> with thymine resulted into the formation of monoacetylated nucleoside 14 in 75% yield. The deacetylation followed by cyclization was affected on monoacetylated nucleoside 14 using 2M NaOH solution in water:dioxane to afford bicyclic nucleoside 15 in 90% yield. The nucleoside 15 was demesylated to obtain 3'-azido-3'-deoxy-2'-O, 4'-Cmethylenethymidine (17) in an overall yield of 72% through benzoylation with sodium benzoate in dry DMF followed by basic hydrolysis of the resulted benzoate 16 using 2M NaOH in THF:water (1:1) (Scheme 1). The azidobicyclic nucleoside 17 was converted into DMT-protected aminonucleoside 6 by reaction with DMT-Cl in pyridine followed by reduction of the resulted compound 18 with PPh3-pyridine and aq. ammonia solution in overall yields of 68%. The synthesis of 3'-amino-3'-deoxy-5'-O-DMT- $\beta$ -thymidine (7) was accomplished from  $\beta$ -thymidine through DMT-protection, C2-O-C3'-anhydro formation, azidation followed by reduction of the azide group with  $PPh_3$ -pyridine/aq. ammonia solution in overall yields of 49% following literature procedure.<sup>[23]</sup>



**SCHEME 1** Synthesis of 3'-amino-3'-deoxy-5'-O-DMT-2'-O,4'-C-methylenethymidine (**6**). *Reagents & conditions:* (i) (a) Trifluoromethanesulfonic anhydride, DCM:pyridine (10:1),  $-10^{\circ}$ C, (b) NaN<sub>3</sub>, DMF, 100°C; (ii) AcOH, Ac<sub>2</sub>O, H<sub>2</sub>SO<sub>4</sub> (100:10:0.1),  $0^{\circ}$ C; (iii) T = Thymine, *N*, *O-bis*-trimethylsilyl acetamide, trimethylsilyltrifluoromethane sulfonate, acetonitrile, 80°C; (iv) 2M NaOH, dioxane:water (1:1),  $0^{\circ}$ C; (v) Sodium benzoate, DMF, 100°C; (vi) 2M NaOH, water:THF (1:1),  $0^{\circ}$ C; (vii) 4,4'-Dimethoxytrityl chloride, pyridine, 25°C; (vii) PPh<sub>3</sub>, pyridine, 28% aqueous ammonia solution, 25°C.

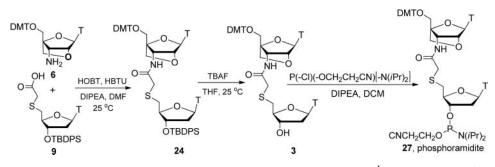
The synthesis of 2-S-(2'-O,4'-C-methylenethymidin-5'-yl)mercaptoacetic acid (8) was accomplished from 2'-O,4'-C-methylenethymidine (LNA T, 20), which was synthesized from diacetone-D-glucose 19 following multistep synthetic procedure as reported in the literature (Scheme 2).<sup>[24]</sup> The primary OH group in bicyclic nucleoside 20 was tosylated using *p*-toluenesulfonyl



**SCHEME 2** Synthesis of 2-*S*-(5'-deoxy-3'-*O*-tert-butyldiphenylsilyl-2'-*O*,4'-*C*-methylenethymidine-5'yl)mercaptoacetic acid (8). *Reagents & conditions:* (i) *p*-Toluenesulfonyl chloride, pyridine, 0°C; (ii) *tert*-butyl diphenylsilyl chloride, imidazole, DMF, 25°C; (iii) NaH, DMF, ethyl mercaptoacetate, 0°C; and (iv) 2M NaOH, methanol.

chloride in pyridine to afford nucleoside **21** in 65% yield, which on silylation with *tert*-butyldiphenylsilyl chloride in DMF in the presence of imidazole afforded 5'-*O*-*p*-toluenesulfonyl-3'-*O*-*tert*-butyldiphenylsilyl-2'-*O*,4'-Cmethylenethymidine (**22**) in 90% yield. The targeted compound **8** was synthesized in overall yields of 77% in two steps from compound **22**, i.e., substitution of tosyl group with ethyl mercaptoacetate using sodium hydride in DMF followed by de-esterification of the resulted compound **23** using 2M NaOH in methanol (Scheme 2). The synthesis of 2-*S*-( $\beta$ -thymidin-5'-yl)mercaptoacetic acid (**9**) was accomplished from  $\beta$ -thymidine through tosylation, silylation, ethoxycarbonylmethylthionation followed by de-esterification in an overall yields of 51% using similar method applied for the synthesis of nucleoside **8**. The structure of compounds **6–23** was established on the basis of its spectral data (IR, <sup>1</sup>H-, <sup>13</sup>C NMR, and HRMS) analysis. Further, the structure of known compounds **6**, **7**, **10–13**, and **17–20** was confirmed by comparing its physical and spectral data with those reported in the literature.<sup>[23–28]</sup>

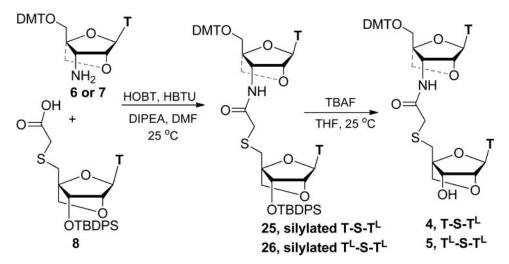
Finally, the synthesis of LNA-based mercaptoacetamido-linked nonionic nucleoside dimer T<sup>L</sup>-S-T **3** was accomplished by HOBT/HBTU catalyzed coupling of 3'-amino-nucleoside **6** and mercaptoacetic acid **9** in DMF to afford 1-(3'-deoxy-5'-O-DMT-2'-O,4'-C-methylenethymidin-3'-yl)-4-(5''-deoxy-3''-O-TBDPS- $\beta$ -thymidin-5''-yl)mercaptoacetamide **24** in 65% yield. The mercaptoacetamide **24** was desilylated using tetrabutylammoniun fluoride to obtain the desired mercaptoacatamido-linked nonionic nucleoside dimer **3** in 75% yield (Scheme 3).



SCHEME 3 Synthesis of mercaptoacetamido-linked nucleoside dimer  $T^{L}$ -S-T 3 and its 3'-O-phosphoramidite.

Further, nucleoside dimers T-S-T<sup>L</sup> **4** and T<sup>L</sup>-S-T<sup>L</sup> **5** were synthesized in two steps, i.e. first by HOBT/HBTU catalyzed coupling of nucleoside **8** with 3'-aminonucleosides **7** and **6** to afford the silylated nucleoside dimers **25** and **26** in 62% yield in each cases. In the second step, desilylation of protected mercaptoacetamides **25** and **26** using tetrabutylammoniun fluoride afforded the desired mercaptoacatamido-linked nucleoside dimers **4** and **5** in 80% and 70% yields, respectively (Scheme 4).

One of the LNA-based mercaptoacetamido-linked nonionic nucleoside dimer T<sup>L</sup>-S-T **3** was condensed with [P(-Cl) (-OCH<sub>2</sub>CH<sub>2</sub>CN){-N(iPr)<sub>2</sub>}] in DCM in the presence of *N*,*N*-diisopropylethylamine (DIPEA) and converted into its corresponding 3'-*O*-phosphoramidite **27**, a building block for the synthesis of mercaptoacetamido-linked oligonucleotide of therapeutic application (Scheme 3). The structure of protected nucleoside dimers **24**, **25**, and **26**; hydroxyl dimers T<sup>L</sup>-S-T **3**, T-S-T<sup>L</sup> **4**, and T<sup>L</sup>-S-T<sup>L</sup> **5**; and phosphoramidite



SCHEME 4 Synthesis of mercaptoacetamido-linked nucleoside dimers T-S-T<sup>L</sup> 4 and T<sup>L</sup>-S-T<sup>L</sup> 5.

**27** was unambiguously established on the basis of their spectral (IR, <sup>1</sup>H-, <sup>13</sup>C-, <sup>31</sup>P NMR spectra, and HRMS) data analysis.

Synthesis of three LNA-based mercaptoacetamido-linked nonionic nucleoside dimers  $T^{L}$ -S-T **3**, T-S-T<sup>L</sup> **4**, and  $T^{L}$ -S-T<sup>L</sup> **5** has been achieved using commonly available starting compound, i.e., D-glucose or thymidine. Phosphoramidite derivative of one of the nucleoside dimers,  $T^{L}$ -S-T **3** has been prepared, which can be used for the synthesis of LNA-based mercaptoacetamido-linked nonionic oligonucleotides of potential antisense application.

### **EXPERIMENTAL SECTION**

Melting points (m.p.) were determined on Buchi M-560 instrument and are uncorrected. The IR spectra were recorded on a Perkin–Elmer model 2000 FTIR spectrometer by making KBr disc for solid samples and thin film for oils. The <sup>1</sup>H- and <sup>13</sup>C NMR spectra were recorded on a Bruker Avance spectrometer at 300 MHz and 75.5 MHz or at Jeol alpha-400 spectrometer at 400 MHz and 100.5 MHz, using TMS as internal standard. HR-ESI-TOF-MS analyses were carried out on a microTOF-Q instrument from Bruker Daltonics, Bremen. The optical rotations were measured with Rudolph autopol II automatic polarimeter using light of 546 nm wavelength. Analytical TLCs were performed on precoated Merck silica-gel  $60F_{254}$  plates; the spots were detected either under UV light or by charring with 4% alcoholic H<sub>2</sub>SO<sub>4</sub>. Silica gel (100–200 mesh) was used for column chromatography.

**3**'-**Amino-3**'-**deoxy-5**'-*O***-<b>DMT**-*β*-**thymidine** (**7**).<sup>23</sup> It was synthesized from *β*-thymidine according to the literature procedure as light yellow foam solid in 49% yield. m.p.: 110–112°C;  $[\alpha]_D^{24} = +1.9$  (c = 0.1, CHCl<sub>3</sub>); HR-ESI-TOF-MS: m/z 566.2241 ([M+Na]<sup>+</sup>), calcd. for  $[C_{31}H_{33}N_3O_6+Na]^+$  566.2262.

**4-C-Hydroxymethyl-1,2-O-isopropylidene**-β-L-threo-pento-furanose (**10**).<sup>21</sup> It was synthesized according to the literature procedure as white solid in 65% yield. m.p.: 99–100°C (lit<sup>21a</sup> mp: 98–99°C);  $[\alpha]_D^{24} = -5.69$  (*c* = 0.34, MeOH); {lit<sup>21a</sup> [ $\alpha$ ]<sub>D</sub> = -5.73 (*c* 0.34, MeOH)}; HR-ESI-TOF-MS *m/z* 243.0840 ([M+Na]<sup>+</sup>), calcd. for [C<sub>9</sub>H<sub>16</sub>O<sub>6</sub>+Na]<sup>+</sup> 243.0839.

**1,2-O-Isopropylidene-5-O-methanesulfonyl-4-C-methanesulfonyloxymet hyl-β-L-threo-pentofuranose** (11).<sup>21a</sup> It was synthesized from 10 according to the literature procedure as white solid in 60% yield. m.p.: 131–132°C (Lit.<sup>21a</sup> m.p.: 132–133°C),  $[\alpha]_D^{25} = -8.4$  (*c* 0.25, CHCl<sub>3</sub>) {Lit.<sup>21a</sup>  $[\alpha]_D^{25} =$ -8.0 (*c* 0.25, CHCl<sub>3</sub>)}; HR-ESI-TOF-MS *m/z* 399.0387 ([M+Na]<sup>+</sup>), calcd. for  $[C_{11}H_{20}O_{10}S_2+Na]^+$  399.0390.

3-Azido-3-deoxy-1,2-*O*-isopropylidene-5-*O*-methanesulfonyl-4-*C*-methan esulfonyloxymethyl- $\beta$ -D-*ribo*furanose (12).<sup>25b</sup> To a stirred solution of compound 11 (5.0 g, 13.30 mmol) in dichloromethane:anhydrous pyridine (10:1, 45 mL) was added dropwise the solution of trifluoromethanesulfonyl

chloride (3.35 mL, 19.95 mmol). After stirring the mixture for 1.5 hours at  $-10^{\circ}$ C, 50 mL ice-cold water was added and the mixture was extracted with chloroform (3 × 100 mL). The chloroform layer was washed with saturated bicarbonate solution (3 × 50 mL), and dried over anhydrous sodium sulfate. Excess of solvent was evaporated at reduced pressure and the residue thus obtained was heated with NaN<sub>3</sub> (0.95 g, 14.62 mmol) in DMF (70 mL) at 100°C to afford compound **12** (2.70 g) in an overall yield of 50% as sticky colorless solid.  $R_f = 0.5$  (5% methanol in chloroform).  $[\alpha]_D^{35} = +78.7$  (*c* 0.1, CHCl<sub>3</sub>). HR-ESI-TOF-MS *m/z* 424.0449 ([M+Na]<sup>+</sup>), calcd. for  $[C_{11}H_{19}N_3O_9S_2+Na]^+$  424.0455.

1,2-Di-*O*-acetyl-3-azido-3-deoxy-5-*O*-methanesulfonyl-4-*C*-methanesul fonyloxymethyl- $\alpha$ , $\beta$ -D-*ribo*furanose (13a–13b).<sup>27</sup> It was synthesized from 12 according to the literature procedure as colorless viscous oil 13a–13b ( $\alpha$ : $\beta$ = *ca*. 2:8, 4.22 g, 84% yield). HR-ESI-TOF-MS *m*/*z* 468.0352 ([M+Na]<sup>+</sup>), calcd. for [C<sub>12</sub>H<sub>19</sub>N<sub>3</sub>O<sub>11</sub>S<sub>2</sub>+Na]<sup>+</sup> 468.0353.

2'-O-Acetyl-3'-azido-3'-deoxy-5'-O-methanesulfonyl-4'-C-methanesulfony loxymethyl-thymidine (14). N,O-bis(Trimethysilyl)acetamide (BSA) (12.5 mL, 50.29 mmol) was added to an anomeric mixture of compound 13a-13b (5.60 g, 12.57 mmol) and thymine (2.38 g, 18.86 mmol) in dry acetonitrile (120 mL), under nitrogen atmosphere. The reaction mixture was refluxed for 1 hour to get a clear solution. Reaction mixture was brought to room temperature (r.t.) and then trimethylsilyl triflate (TMSOTf) (4.0 mL, 21.37 mmol) was added, and refluxing of the reaction was continued further for 14 hours. The solution was cooled to r.t., extracted with chloroform  $(3 \times 100 \text{ ml})$ , and washed with saturated aqueous bicarbonate solution. The organic layer was further washed with water and brine, dried over sodium sulfate, concentrated under reduced pressure and the residue thus obtained was purified on silica gel column chromatography using methanol in chloroform as a gradient solvent system to give 14 (4.82 g, 75%) as light yellow foam solid.  $R_f = 0.5$  (10% methanol in chloroform). m.p.: 70–72°C.  $[\alpha]_D^{35} = -9.9$  (c 0.1, CHCl<sub>3</sub>). IR (thin film) vmax: 3029, 2942, 2123, 1753, 1695, 1358, 1216, 1176, 1085, 1003, 966, 823, and 756  $\text{cm}^{-1}$ . <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz): δ 9.52 (1H, brs), 7.08 (1H, s), 5.73–5.65 (2H, m), 4.94 (1H, I = 6.6 Hz, d), 4.49-4.33 (4H, m), 3.14 (3H, s), 3.11 (3H, s),2.20 (3H, s), 1.92 (3H, s); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75.5 MHz): δ 169.97, 163.64, 150.31, 137.99, 112.05, 92.55, 83.52, 74.35, 68.13, 67.15, 62.86, 37.86, 37.76, 20.49, 12.23; HR-ESI-TOF-MS m/z 534.0563 ([M+Na]<sup>+</sup>), calcd. for  $[C_{15}H_{21}N_5O_{11}S_2+Na]^+$  534.0571.

3'-Azido-3'-deoxy-5'-O-methanesulfonyl-2'-O,4'-C-methylenethymidine (15). To a stirred solution of compound 14 (4.70 g, 9.19 mmol) in dioxane:water (22 mL, 1:1) was added 2M NaOH (21.4 mL) and reaction mixture was stirred at r.t. for 1 hour. The reaction mixture was extracted with ethyl acetate ( $3 \times 100$  ml), washed with bicarbonate solution and brine, dried over sodium sulfate, and concentrated under reduced pressure. The residue thus obtained was purified by silica gel column chromatography using methanol in chloroform as gradient solvent system to give **15** (2.88 g, 90%) as off white solid.  $R_f = 0.5$  (10% methanol in chloroform). m.p.: 77–79°C.  $[\alpha]_D^{35} = + 64.4$  (*c* 0.1, CHCl<sub>3</sub>); IR (thin film) *v*max: 3179, 3026, 2121, 1691, 1464, 1355, 1276, 1175, 1111, 1059, 1024, 998, 966, 812, and 757 cm<sup>-1</sup>; <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>, 300 MHz):  $\delta$  11.39 (1H, brs), 7.45 (1H, s), 5.55 (1H, s), 4.79 (1H, *J* = 12.0 Hz, d), 4.64 (1H, *J* = 12.0 Hz, d), 4.60 (1H, s), 4.20 (1H, s), 3.88–3.80 (2H, m), 3.32 (3H, s), 1.79 (3H, s); <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>, 75.5 MHz):  $\delta$  163.85, 149.93, 134.35, 108.67, 86.59, 85.84, 78.52, 70.89, 65.09, 60.60, 36.96, 12.27; HR-ESI-TOF-MS *m*/*z* 396.0579 ([M+Na]<sup>+</sup>), calcd. for [C<sub>12</sub>H<sub>15</sub>N<sub>5</sub>O<sub>7</sub>S+Na]<sup>+</sup> 396.0584.

3'-Azido-5'-O-benzoyl-3'-deoxy-2'-O-4'-C-methylenethymidine (16). A mix ture of compound 15 (3.5 g, 10.02 mmol) and sodium benzoate (2.89 g, 20.05 mmol) was suspended in anhydrous DMF (170 mL) and was stirred for 4 hours at 100°C. The mixture was cooled to r.t., filtered, and concentrated under reduced pressure. The residue thus obtained was redissolved in ethyl acetate (200 mL) and washed with saturated NaHCO<sub>3</sub> ( $2 \times 200$  mL) and  $H_2O$  (4 × 200 mL). The organic phase was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and concentrated under reduced pressure. The residue thus obtained was purified by silica gel column chromatography using methanol in chloroform as gradient solvent system to afford the nucleoside 16 (3.19 g, 80%) as white solid.  $R_f = 0.5$  (8% methanol in chloroform). m.p.: 168–170°C;  $[\alpha]_D^{25} = +$ 89.2 (c 0.1, CHCl<sub>3</sub>); IR (thin film) vmax: 3178, 3041, 2959, 2827, 2130, 1709, 1686, 1467, 1270, 1111, 1060, 1026, 940, 907, 871, and 719 cm<sup>-1</sup>; <sup>1</sup>H NMR  $(DMSO-d_6, 300 \text{ MHz}): \delta 11.40 (1\text{H}, \text{brs}), 8.05 (2\text{H}, I = 8.1 \text{ Hz}, \text{d}), 7.74-7.57$ (3H, m), 7.34 (1H, s), 5.56 (1H, s), 4.79 (2H, s), 4.61 (1H, s), 4.37 (1H, s), 3.94 (2H, J = 16.8 and 8.7 Hz, dd), 1.59 (3H, s); <sup>13</sup>C NMR (DMSO- $d_{6.8}$ 75.5 MHz):  $\delta$  165.16, 163.80, 149.90, 134.11, 133.81, 129.33, 128.92, 108.53, 86.59, 86.45, 78.51, 71.14, 66.32, 60.65, 12.04; HR-ESI-TOF-MS m/z 422.1062  $([M+Na]^+)$ , calcd. for  $[C_{18}H_{17}N_5O_6+Na]^+$  422.1071.

**3'-Azido-3'-deoxy-2'-O,4'-C-methylenethymidine** (17).<sup>28</sup> To a stirred solution of compound **16** (3.11 g, 7.78 mmol) in THF:water (1:1, 50 mL) was added 2M NaOH (11.8 mL), and the mixture was stirred at 0°C for 1 hour. After completion of reaction acetic acid (3.0 mL) was added, and the solution was concentrated under reduced pressure. Purification was performed on silica gel column using methanol in chloroform as gradient solvent system to yield azido-LNA **17** (2.07 g, 90%) as a white powder. m.p.: 94–95°C (Lit.<sup>28</sup> m.p.: 94–96°C);  $[\alpha]_D^{35} = + 87.1$  (*c* 0.1, MeOH); HR-ESI-TOF-MS *m/z* 318.0816 ([M+Na]<sup>+</sup>), calcd. for [C<sub>11</sub>H<sub>13</sub>N<sub>5</sub>O<sub>5</sub>+Na]<sup>+</sup> 318.0809.

3'-Azido-3'-deoxy-5'-O-DMT-2'-O-4'-C-methylenethymidine (18).<sup>25a</sup> It was synthesized from 17 according to the literature procedure to afford 18 (0.45 g, 75%) as white foam solid. m.p.: 125–126°C, (Lit.<sup>25a</sup> m.p.:

125–128°C);  $[\alpha]_D^{22} = +19.1$  (*c* 0.75, acetone); HR-ESI-TOF-MS: *m/z* 620.2107 ([M+Na]<sup>+</sup>), calcd. for  $[C_{32}H_{31}N_5O_7+Na]^+$  620.2116.

**3'-Amino-3'-deoxy-5'-O-DMT-2'-O-4'-C-methylenethymidine (6).**<sup>25a</sup> It was synthesized from **18** according to the literature procedure to afford **6** (0.10 g, 90% yield) as light yellow foam solid. m.p.: 130–132°C, (Lit.<sup>25a</sup> m.p.: 131–134°C).  $[\alpha]_D^{22} = +$  22.9 (*c* 1, acetone). HR-ESI-TOF-MS: *m/z* 594.2214 ([M+Na]<sup>+</sup>), calcd. for  $[C_{32}H_{33}N_3O_7+Na]^+$  594.2211.

**2'-0,4'-C-methylenethymidine** (20).<sup>24</sup> Compound **20** was synthesized from D-glucose according to the literature procedure as a white solid material. m.p.: 196–197°C (Lit.<sup>24</sup> m.p.: 196–198°C);  $[\alpha]_D^{23} = +57.9$  (c = 1, EtOH); HR-ESI-TOF-MS: m/z 293.0752 ([M+Na]<sup>+</sup>), calcd. for  $[C_{11}H_{14}N_2O_6+Na]^+$  293.0744.

5'-O-p-Toluenesulfonyl-2'-O,4'-C-methylenethymidine (21). p-Toluene sulfonyl chloride (12 mmol) dissolved in 10 mL of dry pyridine was added drop wise during 4 hours to the solution of **20** (10 mmol) in 30 mL of dry pyridine at 0°C. The reaction mixture was stirred for additional 4 hours at r.t.. Pyridine was removed under reduced pressure and residue thus obtained was dissolved in 100 mL ethyl acetate and the organic layer was washed with  $(2 \times 50 \text{ mL})$  10% aq. NaHCO<sub>3</sub> solution followed by cold water  $(2 \times 50 \text{ mL})$ . The organic layer was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and was evaporated at reduced pressure. The crude product so obtained was purified over silica gel column using methanol in chloroform as gradient solvent system to afford **21** as white solid (2.75 g, 65% yield).  $R_f = 0.4$  (10% methanol in chloroform); m.p.: 100–102°C;  $[\alpha]_D^{22} = +71.7$  (c = 0.1, MeOH); IR (KBr)  $\nu_{max}$ : 3401, 3198, 3070, 2952, 1690, 1597, 1466, 1363, 1275, 1176, 1054, 983, and  $810 \text{ cm}^{-1}$ ; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz):  $\delta$  9.72 (1H, brs), 7.82 (2H, I = 8.2 Hz, d), 7.46 (1H, s), 7.39 (2H, J = 8.2 Hz, d), 5.52 (1H, s), 4.60 (1H, s), 4.41 (2H, s), 4.23 (1H, s), 4.00 (1H, brs), 3.99 (1H, I = 8.0 Hz, d), 3.79 (1H, I = 8.0 Hz, d), 3.77.9 Hz, d), 2.46 (3H, s), 1.85 (3H, s); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75.5 MHz): δ 164.35, 150.20, 149.55, 145.64, 134.36, 132.30, 130.21, 110.56, 87.18, 86.22, 79.15, 77.01, 69.94, 64.27, 21.70, 12.45; HR-ESI-TOF-MS: m/z 447.0838 ([M+Na]<sup>+</sup>), calcd. for  $[C_{18}H_{20}N_2O_8S+Na]^+$  447.0833.

5'-O-p-Toluenesulfonyl-3'-O-tert-butyldiphenylsilyl-2'-O,4'-C-methylene thymidine (22). A solution of compound 21 (6.0 mmol), tert-butyldiphe nylsilyl chloride (7.2 mmol), imidazole (14.8 mmol) in 20 mL of anhydrous DMF was stirred for 6 hours under nitrogen atmosphere at r.t.. Excess of DMF was removed in vacuo and the residue was dissolved in 200 mL of ethyl acetate, washed with water (3 × 50 mL) followed by brine (2 × 30 mL). The organic layer was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and solvent was then removed in vacuo. The residue thus obtained was purified by silica gel column chromatography using ethyl acetate in petroleum ether as gradient solvent system to afford 22 as light yellow crystalline solid (3.51 g, 90% yield). R<sub>f</sub> = 0.5 (50% ethyl acetate in petroleum ether); m.p.: 95–96°C;  $[α]_D^{23} = + 6.2$  (*c* 0.1, MeOH); IR (KBr)  $ν_{max}$ : 3181, 3048, 2955, 2859, 1689, 1597, 1466, 1370, 1274, 1178, 1104, 1054, 823, 744, and 705 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz): δ 8.71 (1H, brs), 7.81–7.34 (15H, m), 5.47 (1H, s), 4.84–4.35 (2H, m), 4.14 (1H, J = 7.2 Hz, d), 4.02–3.79 (3H, m), 2.48 (3H, s), 1.74 (3H, s), 1.03 (9H, s); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75.5 MHz): δ 163.48, 149.13, 145.60, 135.61, 135.33, 133.71, 132.35, 131.98, 131.39, 130.45, 130.37, 130.15, 128.06, 127.95, 110.20, 86.86, 85.81, 78.47, 71.50, 63.70, 26.90, 21.72, 19.04, 12.37; HR-ESI-TOF-MS: m/z 663.2162 ([M+H]<sup>+</sup>), calcd. for [C<sub>34</sub>H<sub>38</sub>N<sub>2</sub>O<sub>8</sub>SSi+H]<sup>+</sup> 663.2191.

Ethyl 2-S-(3'-O-tert-butyldiphenylsilyl-5'-deoxy-2'-O,4'-C-methylenethymi din-5'-yl)mercaptoacetate (23). A solution of NaH (0.29 g, 60% in hexane, 7.25 mmol.) and ethyl mercaptoacetate (5.5 mmol) in anhydrous DMF (5 mL) was stirred for 30 minutes. Compound 22 (4.50 mmol) dissolved in DMF (5 mL) was then added slowly at 0°C. The reaction mixture was stirred for 1 hour at r.t.. Excess of DMF was removed under reduced pressure, and the residue was dissolved in 200 mL ethyl acetate. The organic layer was washed with water  $(3 \times 50 \text{ mL})$  and with brine  $(2 \times 30 \text{ mL})$ . The organic layer was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and solvent was removed in vacuo. The crude thus obtained was purified on silica gel column using methanol in chloroform as gradient solvent system to afford 23 as viscous oil (2.26 g, 82% yield).  $R_f = 0.5$  (50% ethyl acetate in petroleum ether);  $[\alpha]_D^{23} = +41.2$  (*c* 0.1, MeOH); IR (KBr) v<sub>max</sub>: 3183, 3069, 2931, 2858, 1690, 1459, 1275, 1155, 1113, 1051, 943, 856 and 744 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz): δ 9.20 (1H, brs), 7.63–7.31 (11H, m), 5.51 (1H, s), 4.22–4.14 (3H, m), 3.93 (2H, s), 3.91 (1H, s), 3.70 (1H, s), 3.33-3.11 (3H, m), 1.78 (3H, I = 1.0 Hz, d), 1.28 (3H, I = 1.0 Hz, d),I = 7.2 Hz, t), 1.09 (9H, s); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75.5 MHz):  $\delta$  169.80, 163.62, 149.25, 145.90, 135.66, 135.57, 135.43, 134.24, 132.16, 131.80, 130.39, 130.34, 130.130.14, 127.98, 127.93, 109.85, 88.39, 86.96, 78.75, 73.61, 72.87, 61.60, 34.79,28.88, 26.75, 21.07, 19.10, 12.18; HR-ESI-TOF-MS: m/z 633.2028 ([M+Na]<sup>+</sup>), calcd. for  $[C_{31}H_{38}N_2O_7SSi+Na]^+$  633.2061.

2-S-(5'-Deoxy-3'-O-tert-butyldiphenylsilyl-2'-O,4'-C-methylenethymidin-5'-yl)mercaptoacetic acid (8). Sodium hydroxide (2M, 5 mL) solution was added to a solution of 23 (5.30 mmol) in 10 mL methanol. The reaction mixture was stirred for 30 minutes. After completion of reaction on analytical TLC examination, the sodium salt of the acid and excess NaOH present in the solution was neutralized by DOWEX-50H<sup>+</sup> resin. The resin was filtered and washed with 2:1 mixture of methanol:water. The filtrate was concentrated and dried under vacuum to afford 8 as white solid (2.32 g, 94% yield).  $R_f = 0.5$  (50% methanol in chloroform); m.p.: 208–210°C;  $[\alpha]_D^{23} = +34.5$  (*c* 0.1, MeOH); IR (KBr)  $\nu_{max}$ : 3449, 3064, 2931, 2858, 1689, 1580, 1469, 1427, 1389, 1273, 1156, 1110, 1052, 855 and 704 cm<sup>-1</sup>; <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>, 300 MHz):  $\delta$  11.26 (1H, brs), 7.58–7.32 (11H, m), 5.30 (1H, s), 4.04 (1H, I = 8.1 Hz, d), 3.82 (1H, I = 8.1 Hz, d), 3.74 (1H, s), 3.61 (1H, s), 3.28 (2H, s), 3.12 (2H, s), 1.64 (3H, s), 1.02 (9H, s); <sup>13</sup>C NMR (DMSO- $d_6$ , 75.5 MHz):  $\delta$  172.60, 163.58, 149.39, 135.13, 134.95, 133.72, 132.17, 131.44, 130.17, 127.87, 127.77, 108.69, 88.34, 86.03, 78.10, 73.14, 72.46, 38.58, 27.69, 26.47, 18.63, 12.22; HR-ESI-TOF-MS: m/z 605.1731 ([M+Na]<sup>+</sup>), calcd. for [C<sub>29</sub>H<sub>34</sub>N<sub>2</sub>O<sub>7</sub>SSi+Na]<sup>+</sup> 605.1748.

**2-S-(5'-Deoxy-3'-O-tert-butyldiphenylsilyl-**β-thymidin-5'-yl)mercaptoacetic acid (9). It was synthesized from β-thymidine following the same procedure used for the synthesis of mercaptoacetic acid **8** from **20** as white solid (2.70 g, 92% yield). m.p.: 69–70°C;  $[\alpha]_D^{27} = +29.13$  (*c* 0.1, MeOH); IR (thin film)  $\nu$ max: 3180, 3071, 2932, 2859, 1702, 1474, 1428, 1364, 1276, 1197, 1112, 998, 924, 823, 756, 703, and 669 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz): δ 9.48 (1H, brs), 7.65–7.61 (4H, m), 7.45–7.40 (6H, m), 7.21 (1H, *J* = 1.2 Hz, d), 6.27 (1H, *J* = 6.8 Hz, t), 4.24–4.20 (1H, m), 4.12–4.08 (1H, m), 3.12 (2H, *J* = 33.2 Hz and *J* = 14.8 Hz, dd), 2.63–2.62 (1H, m), 2.58–2.53 (1H, m), 2.37–2.31 (1H, m), 1.95–1.88 (1H, m), 1.83 (3H, s), 1.07 (9H, s); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz): δ 174.46, 164.60, 150.34, 136.02, 135.70, 132.86, 130.20, 127.97, 111.22, 86.15, 84.58, 74.52, 40.17, 34.29, 34.20, 26.81, 18.95, 12.29; HR-ESI-TOF-MS *m*/z 577.1788 ([M+Na]<sup>+</sup>), calcd. for [C<sub>28</sub>H<sub>34</sub>N<sub>2</sub>O<sub>6</sub>SSi+Na]<sup>+</sup> 577.1799.

General procedure for the synthesis of silylated dimers 24–26. To a solution of mercaptoacetic acid 8/9 (1.72 mmol) in dry DMF (5 mL), HBTU (2.06 mmol), DIPEA (5.20 mmol), and HOBt (0.1 mmol) were added and stirred for 15 minutes at 25°C. Amine 6/7 (1.70 mmol) was dissolved in 5 mL DMF and was then added into the reaction mixture and the reaction mixture was further stirred at 25°C for 4 hours. The reaction mixture was concentrated to dryness, dissolved in ethyl acetate (30 mL) and washed with aqueous 5% NaHCO<sub>3</sub> solution (2 × 20 mL). The organic layer was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and concentrated under reduced pressure. The crude product so obtained was purified by silica gel column chromatography using methanol in chloroform as gradient solvent system to afford mercaptoacetamido-linked dimer 24 and 25/26 in 65% and 62% yields, respectively.

1-(3'-Deoxy-5'-*O*-DMT-2'-*O*,4'-*C*-methylenethymidin-3'-yl)-4-(5"-deoxy-3"-*O*-tert-butyldiphenylsilyl-β-thymidin-5"-yl)mercaptoacetamide (24). It was obtained by the HOBT/HBTU catalyzed coupling reaction of acid **9** with amine **6** as white solid (1.24 g, 65% yield). m.p.: 104–105°C.  $[\alpha]_D^{27} =$ +26.71 (*c* 0.1, MeOH); IR (thin film) νmax: 3199, 3069, 2932, 2859, 1691, 1509, 1466, 1384, 1275, 1252, 1178, 1112, 1055, 1035, 978, 916, 830, 792, 756, 704, 665, and 613 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz): δ 8.90 (1H, brs), 8.58 (1H, brs), 8.01 (1H, brs), 7.62–7.60 (4H, m), 7.45–7.25 (18H, m), 6.82 (4H, *J* = 8.4 Hz, d), 6.17–5.82 (1H, m), 5.30 (1H, *J* = 5.2 Hz, d), 4.51 (1H, *J* = 8.0 Hz, d), 4.47 (1H, *J* = 3.6 Hz, d), 4.23–4.21 (1H, m), 3.97–3.91 (1H, m), 3.78–3.76 (6H, m), 3.43 (2H, m), 3.41 (1H, *J* = 7.2 Hz, d), 2.88–2.87 (2H, m), 2.46–2.41 (1H, m), 2.24–2.22 (3H, m), 1.84 (3H, s), 1.63 (3H, s), 1.06 (9H, s); <sup>13</sup>C NMR (DMSO- $d_6$ , 100.6 MHz):  $\delta$  169.71, 163.77, 163.62, 158.13, 150.43, 149.91, 144.51, 135.75, 135.38, 134.97, 133.87, 132.82, 132.59, 130.07, 129.76, 128.00, 127.72, 126.75, 113.24, 109.81, 108.84, 87.63, 86.30, 85.88, 85.43, 84.05, 79.18, 78.54, 75.36, 71.15, 57.97, 55.00, 51.18, 38.42, 33.65, 26.66, 18.59, 12.41, 12.00; HR-ESI-TOF-MS m/z 1130.4031 ([M+Na]<sup>+</sup>), calcd. for [C<sub>60</sub>H<sub>65</sub>N<sub>5</sub>O<sub>12</sub>SSi+Na]<sup>+</sup> 1130.4012.

1-(3'-Deoxy-5'-O-DMT-β-thymidin-3'-yl)-4-(5"-deoxy-3"-O-tertbutyldiphenylsilyl-2"-O,4"-C-methylenethymidin-5"-yl)mercaptoacetamide (25). It was obtained by the HOBT/HBTU catalyzed coupling reaction of acid 8 with amine 7 as white solid (1.17 g, 62% yield).  $R_f = 0.7$  (10% methaol in chloroform); m.p.: 138–140°C;  $[\alpha]_D^{24} = +18.3$  (*c* 0.05, CHCl<sub>3</sub>); IR (KBr) v<sub>max</sub>: 3338, 3064, 2930, 2857, 1686, 1507, 1459, 1252, 1176, 1112, 1051, 829, and 703 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz):  $\delta$  9.47 (2H, 2 x brs), 7.63-7.56 (5H, m), 7.41-7.19 (17H, m), 6.83 (4H, J = 8.4 Hz, d), 6.34 (1H, brs), 5.50 (1H, s), 4.66 (1H, s), 4.11 (1H, J = 7.8 Hz, d), 4.03 (1H, s), 3.87-3.82 (2H, m), 3.71 (7H, s), 3.46 (2H, s), 3.29 (2H, s), 3.06-2.88 (2H, m), 2.40–2.30 (2H, m), 1.74 (3H, s), 1.37 (3H, s), 1.06 (9H, s); <sup>13</sup>C NMR (DMSO-d<sub>6</sub>, 100.6 MHz):  $\delta$  168.56, 163.70, 163.56, 158.14, 150.38, 149.43, 144.71, 135.71, 135.45, 135.19, 135.03, 133.58, 132.12, 131.47, 130.26, 129.77, 127.92, 127.85, 127.72, 126.79, 113.23, 109.67, 108.73, 87.85, 86.17, 85.88, 83.66, 83.15, 78.23, 73.03, 72.69, 63.66, 55.05, 49.61, 36.74, 35.84, 28.30, 26.49, 22.51, 18.65, 12.26, 11.72; HR-ESI-TOF-MS: m/z 1130.3980  $([M+Na]^+)$ , calcd. for  $[C_{60}H_{65}N_5O_{12}SSi+Na]^+ 1130.4012$ .

1-(3'-Deoxy-5'-O-DMT-2'-O,4'-C-methylenethymidin-3'-yl)-4-(5"-deoxy-3"-O-tert-butyldiphenylsilyl-2"-O,4"-C-methylene-thymidin-5"-yl)mercaptoaceta mide (26). It was obtained by the HOBT/HBTU catalyzed coupling reaction of acid 8 with amine 6 as white solid (1.20 g, 62% yield).  $R_f = 0.7$  (10% methaol in chloroform); m.p.: 118–120°C;  $[\alpha]_D^{25} = +4.2$  (c 0.05, CHCl<sub>3</sub>); IR (KBr) v<sub>max</sub>: 3199, 3069, 2931, 1691, 1608, 1509, 1465, 1252, 1177, 1112, 1053, 980, 831, and 703 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz): δ 7.62–7.22 (22H, m), 6.84 (4H, I = 8.7 Hz, d), 5.64 (1H, s), 5.48 (1H, s), 4.50 (1H, s),4.45 (1H, J = 8.1 Hz, d), 4.09 (1H, J = 7.8 Hz, d), 3.92 (1H, s), 3.85 (1H, s),3.78 (7H, s), 3.73 (2H, m), 3.48 (2H, s), 3.26 (2H, I = 14.4 Hz, q), 2.95 (1H, I)= 15.0 Hz, d, 2.81 (1H, I = 15.0 Hz, d), 1.69 (6H, s), 1.06 (9H, s); <sup>13</sup>C NMR  $(DMSO-d_6, 100.6 \text{ MHz}): \delta 169.73, 163.75, 163.56, 158.12, 150.39, 149.89,$ 144.49, 135.79, 135.44, 135.28, 134.95, 133.11, 132.73, 110.01, 108.81,87.59, 86.27, 85.87, 83.21, 80.68, 78.51, 72.88, 71.06, 57.88, 57.23, 54.99, 51.10, 45.67, 38.12, 33.58, 33.22, 26.68, 18.88, 12.39, 11.93; HR-ESI-TOF: m/z 1136.3997 ( $[M+H]^+$ ), calcd. for  $[C_{61}H_{65}N_5O_{13}SSi+H]^+$  1136.4101.

General procedure for the synthesis of mercaptoacetamido-linked dimers 3, 4, and 5. A solution of 24/25 or 26 (0.90 mmol) and TBAF (1.20 mL, 1M in THF) in anhydrous tetrahydrofuran (15 mL) was stirred at

r.t. for 1 hour. Excess of tetrahydrofuran was evaporated in vacuo and residue was redissolved in dichloromethane (50 mL). The solution was washed with water (2 × 20 mL) followed by brine (2 × 20 mL). The organic layer was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and evaporated to dryness. The crude thus obtained was purified by silica gel column chromatography using methanol in chloroform as gradient solvent system to afford mercaptoacetamido-linked nucleosiode dimers 3/4/5 in 75%, 80%, and 70% yields, respectively.

 $1-(3'-\text{Deoxy-}5'-O-\text{DMT-}2'-O,4'-C-\text{methylenethymidin-}3'-yl)-4-(5''-\text{deoxy-}\beta$ thymidin-5"-yl)mercaptoacetamide (3). It was obtained as white solid powder (0.59 g, 75% yield). m.p.: 173–175°C (decompose);  $[\alpha]_{\text{D}}^{30} = +29.1$  (c 0.05, MeOH); IR (KBr) vmax: 3448, 3067, 2927, 1690, 1508, 1466, 1252, 1178, 1054, 977, 895, 823, 760, 727, and 705 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$  10.36 (1H, brs), 9.87 (1H, brs), 7.61 (1H, brs), 7.47 (2H, I = 7.2 Hz, d), 7.36–7.21(10H, m), 7.14 (1H, brs), 6.84 (4H, I = 9.2 Hz and I = 2.8 Hz, dd), 5.98 (1H, s), 5.59 (1H, s), 5.04 (1H, J = 16.0 Hz, d), 4.59 (1H, J = 7.6 Hz, d), 4.25 (2H, I = 20.0 Hz, d), 3.96 (2H, I = 8.4 Hz, d), 3.78 (6H, s), 3.48 (2H, I = 23.2 Hz and I = 11.2 Hz, dd), 3.23 (2H, s), 2.79–2.71 (2H, m), 2.28–2.25 (1H, J = 12.0 Hz, d), 1.84 (3H, s), 1.68 (3H, s); <sup>13</sup>C NMR (DMSO-d<sub>6</sub>, 100 MHz): δ 169.88, 163.80, 158.15, 150.45, 149.91, 144.60, 135.95, 135.25, 134.98, 133.92, 129.86, 127.96, 127.72, 126.79, 113.27,109.77, 108.82, 87.65, 86.30, 85.89, 85.18, 83.81, 79.19, 78.53, 72.41, 71.16, 57.99, 55.05, 51.23, 39.18, 34.01, 12.40, 12.05; HR-ESI-TOF-MS *m/z* 892.2771  $([M+Na]^+)$ , calcd. for  $[C_{44}H_{47}N_5O_{12}S+Na]^+$  892.2834.

 $1-(3'-\text{Deoxy}-5'-O-\text{DMT}-\beta-\text{thymidin}-3'-\text{yl})-4-(5''-\text{deoxy}-2''-O,4''-C-\text{methylene})$ thymidin-5"-yl)mercaptoacetamide (4). It was obtained as white solid powder (0.61 g, 80% yield).  $R_f = 0.5$  (10% methaol in chloroform); m.p.: 158–160°C;  $[\alpha]_{D}^{24} = +$  102.2 (c 0.1, CHCl<sub>3</sub>); IR (KBr)  $\nu_{max}$ : 3353, 3064, 2930, 1694, 1509, 1466, 1252, 1178, 1105, 1042, 830, and 584 cm<sup>-1</sup>; <sup>1</sup>H NMR (DMSO- $d_6$ , 400 MHz):  $\delta$  11.35 (2H, brs), 8.54 (1H, J = 7.2 Hz, d), 7.53 (1H, s), 7.66 (1H, s), 7.37 (2H, I = 7.2 Hz, d), 7.30–7.20 (8H, m), 6.88 (4H, J = 8.8 Hz and J = 2.0 Hz, dd), 6.20 (1H, J = 6.4 Hz, t), 5.85 (1H, J)= 4.4 Hz, d, 5.43 (1H, s), 4.50–4.43 (1H, m), 4.13 (1H, s), 3.87 (2H, I =4.4 Hz, d), 3.81 (1 H, I = 8.0 Hz, d), 3.70-3.66 (7 H, m), 3.32-3.25 (2 H, m), 3.18-3.11 (2H, m), 3.00-2.90 (2H, m), 2.37-2.32 (1H, m), 2.18-2.12 (1H, m), 1.75 (3H, s), 1.42 (3H, s); <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>, 100 MHz): δ 168.87, 163.35, 158.14, 150.40, 149.95, 144.60, 135.80, 135.45, 135.27, 134.64, 129.80, 127.94, 127.74, 126.82, 113.25, 109.77, 108.65, 88.00, 86.51, 85.72, 83.67, 83.09, 79.09, 72.57, 70.68, 63.51, 55.07, 49.56, 45.74, 36.76, 35.98, 28.34, 12.40, 11.76; HR-ESI-TOF-MS: m/z 892.2851 ([M+Na]<sup>+</sup>), calcd. for  $[C_{44}H_{47}N_5O_{19}S+Na]^+$  892.2834.

1-(3'-Deoxy-5'-O-DMT-2'-O,4'-C-methylenethymidin-3'-yl)-4-(5"-deoxy-2"-O,4"-C-methylenethymidin-5"-yl)mercaptoacetamide (5). It was obtained as white solid powder (0.55 g, 70% yield).  $R_f = 0.5$  (10% methaol in chloroform); m.p.: 158–160°C;  $[\alpha]_D^{24} = +47.1$  (c 0.05, CHCl<sub>3</sub>); IR (KBr)  $ν_{\text{max}}$ : 3195, 2924, 2853, 1694, 1609, 1509, 1463, 1251, 1178, 1053, 978, and 832 cm<sup>-1</sup>; <sup>1</sup>H NMR (DMSO- $d_6$ , 400 MHz): δ 11.45 (2H, brs), 8.16 (1H, J =8.4 Hz, d), 7.70 (1H, s), 7.58 (1H, s), 7.44 (2H, J = 7.6 Hz, d), 7.33–7.23 (8H, m), 6.89 (4H, J = 8.8 Hz and J = 2.0 Hz, dd), 4.85 (1H, s), 5.52 (2H, J =16.0 Hz, d), 4.44 (2H, J = 7.2 Hz, d), 4.32 (1H, s), 3.85–3.84 (2H, m), 3.73 (8H, s), 3.34–3.31 (2H, m), 3.26–3.19 (2H, m), 3.04 (2H, J = 14.8 Hz, d), 1.78 (3H, s), 1.54 (3H, s); <sup>13</sup>C NMR (DMSO- $d_6$ , 100 MHz): δ 169.68, 163.69, 158.14, 149.90, 144.60, 134.97, 134.31, 133.86, 129.83, 129.78, 127.92, 127.66, 126.77, 113.25, 108.77, 87.56, 87.10, 86.47, 86.28, 85.84, 78.53, 77.48, 72.65, 72.00, 71.13, 57.93, 55.02, 50.88, 45.64, 34.59, 28.14, 12.21; HR-ESI-TOF-MS: m/z 898.2925 ([M+H]<sup>+</sup>), calcd. for [C<sub>45</sub>H<sub>47</sub>N<sub>5</sub>O<sub>13</sub>S+H]<sup>+</sup> 898.2911.

1-(3'-Deoxy-5'-O-DMT-2'-O,4'-C-methylenethymidin-3'-yl)-4-(3"-O-βcyanoethyl-N,N-diisopropylphosphoramidite-5"-deoxy- $\beta$ -thymidin-5"yl)mercaptoacetamide (27). To a stirred solution of compound 3 (0.1g, 0.115 mmol) in dry DCM (2 mL), N,N-Diisopropylethylamine(25  $\mu$ L, 0.283 mmol) and 2-cyanoethyl N,N-diisopropylamino chlorophosphoramidite (64  $\mu$ L, 0.283 mmol) were added under argon atmosphere at 0°C. The reaction mixture was stirred at r.t. for 30 minutes. After completion of reaction on analytical TLC examination, the reaction mixture was diluted with dry DCM (20 mL) and washed with 5% NaHCO<sub>3</sub> solution ( $2 \times 10$  mL). The organic phase was dried and concentrated under reduced pressure. Purification was done by flash column chromatography using hexane: ethyl acetate (1:8) solvent gradient. The phosphoramidite 27 was dried overnight over P<sub>2</sub>O<sub>5</sub> and KOH in a desiccator before characterization. It was obtained as white solid,  $R_f = 0.5$  (ethyl acetate); <sup>31</sup>P NMR (CDCl<sub>3</sub>):  $\delta$ 148.96, 148.70; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$  7.59 (1H, s), 7.49 (1H, J = 4.0 Hz, d), 7.38–7.10 (11H, m), 6.86 (4H, I = 8.0 Hz, d), 6.01–5.99 (1H, m), 5.84-5.83 (1H, m), 5.68 (1H, s), 5.04-4.93 (3H, m), 4.56-4.46 (3H, m), 3.95-3.60 (11H, m), 3.50-3.48 (2H, m), 3.24-3.21 (2H, m), 2.88-2.87 (3H, m), 2.66-2.64 (2H, m), 1.72 (3H, s), 1.49 (3H, s), 1.30 (6H, J = 8.0 Hz, d), 1.20 (6H, J = 8.0 Hz, d); HR-ESI-TOF-MS:  $m/z \ 1092 \ ([M+Na]^+)$ , calcd. for  $[C_{53}H_{64}N_7O_{13}PS+Na]^+$  1092.

#### REFERENCES

- Snead, N.M.; Rossi, J.J. RNA interference trigger variants: getting the most out of RNA for RNA interference-based therapeutics. *Nucleic Acid Ther.* 2012, 22, 139–146.
- Campbell, M.A.; Wengel, J. Locked vs. unlocked nucleic acids (LNA vs. UNA): contrasting structures work towards common therapeutic goals. *Chem. Soc. Rev.* 2011, 40, 5680–5689.
- Herdewijn, P. Nucleic acids with a six-membered 'carbohydrate' mimic in the backbone. *Chem. Biodivers.* 2010, 7, 1–59.
- 4. Crooke, S.T. Progress in antisense technology. Annu. Rev. Med. 2004, 55, 61-95.
- 5. Novina, C.D.; Sharp, P.A. The RNAi revolution. Nature 2004, 430, 161-164.
- 6. Yan, H. Nucleic acid nanotechnology. Science 2004, 306, 2048-2049.

- Wengel, J. Nucleic acid nanotechnology-towards Ångström-scale engineering. Org. Biomol. Chem. 2004, 2, 277–280.
- Chen, X.; Roloff, A.; Seitz, O. Consecutive signal amplification for DNA detection based on de novo fluorophore synthesis and host–guest chemistry. *Angew. Chem. Int. Ed. Engl.* 2012, 51, 4479–4483.
- 9. Corey, D.R. Chemical modification: the key to clinical application of RNA interference? J. Clin. Invest. **2007**, 117, 3615–3622.
- Mickelfield, J. Backbone modification of nucleic acids: synthesis, structure and therapeutic applications. *Current Med. Chem.* 2001, 8, 1157–1179.
- Wengel, J. Synthesis of 3'-C- and 4'-C-branched oligodeoxynucleotides and the development of locked nucleic acid (LNA). Acc. Chem. Res. 1999, 32, 301–310.
- Hildebrandt-Eriksen, E.S.; Aarup, V.; Persson, R.; Hansen, H.F.; Munk, M.E.; Ørum, H. A locked nucleic acid oligonucleotide targeting microRNA 122 is well-tolerated in cynomolgus monkeys. *Nucleic Acid Ther.* 2012, 22, 152–161.
- Veedu, R.N.; Wengel, J. Locked nucleic acids: promising nucleic acid analogs for therapeutic applications. *Chem Biodivers.* 2010, 7, 536–542.
- Baraasch, D.A.; Corey, D.R. Locked nucleic acid (LNA): fine-tuning the recognition of DNA and RNA. *Chem. Biol.* 2001, 8, 1–7.
- Zhang, J.; Shaw, J.T.; Matteucci, M.D. Synthesis and hybridization property of an oligonucleotide containing a 3'-thioformcetal linked pentathymidylate. *Bioorg. Med. Chem. Lett.* 1999, 9, 319– 322.
- Waldner, A.; Mesmaeker, A. De. Synthesis of oligodeoxyribonucleotides containing dimers with carbamate moieties as replacement of the natural phosphodiester linkage. *Bioorg. Med. Chem. Lett.* 1994, 4, 405–408.
- Bhat, B.; Swayze, E.E.; Wheeler, P.; Dimock, S.; Perbost, M.; Sanghvi, Y.S. Synthesis of novel nucleic acid mimics *via* the stereoselective intermolecular radical coupling of 3'-Iodo nucleosides and formaldoximes. *J. Org. Chem.* 1996, 61, 8186–8199.
- Lauristen, A.; Wengel, J. Oligodeoxynucleotides containing amide-linked LNA-type dinucleotides: synthesis and high-affinity nucleic acid hybridization. *Chem. Commun.* 2002, 530–531.
- Isobe, H.; Fujino, T.; Yamazaki, N.; Niekowski, M.G.; Nakamura, E. Triazole-linked analogue of deoxyribonucleic acid (TLDNA): design, synthesis, and double-strand formation with natural DNA. *Org. Lett.* 2008, 10, 3729–3732.
- Gogoi, K.; Gunjal, A.D.; Phalgune, U.D.; Kumar, V.A. Synthesis and RNA binding selectivity of oligonucleotides modified with five-atom thioacetamido nucleic acid backbone structures. *Org. Lett.* 2007, 9, 2697–2700.
- (a) Martin, M.M. Synthesis of D-glucose derived oxetane: 1,2-O-isopropylidene-4-(S)-3-O,4-C-methylene-5-O-methanesulfonyl-β-L-three-pento-1,4-furanose. J. Appl. Sci. Res. 2008, 4, 1478–1482
  b) Youssefyeh, R.D.; Verheyden, J.P.H.; Moffatt, J.G. 4'-Substituted nucleosides: synthesis of some 4'-hydroxymethyl nucleosides. J. Org. Chem. 1979, 44, 1301–1309.
- Vorbrüggen, H.; Lagoja, I.M.; Herdewijn, P. Synthesis of ribonucleosides by condensation using trimethylsilyl triflate. *Curr. Protoc. Nucleic Acid Chem.* 2007, 1, 1.13.1–1.13.16.
- Teng, B.; Bai, Y.; Chang, Y.; Chen, S.; Li, Z. Technetium-99m-labeling and synthesis of thymidine analogs: potential candidates for tumor imaging. *Bioorg. Med. Chem. Lett.*, 2007, 17, 3440– 3444.
- (a) Koshkin, A.A.; Singh, S.K.; Nielson, P.; Rajwanshi, V.K.; Kumar, R.; Meldgaard, M.; Wengel, J. LNA (Locked Nucleic Acids): synthesis of the adenine, cytosine, guanine, 5-methylcytosine, thymine and uracil bicyclonucleoside monomers, oligomerisation, and unprecedented nucleic acid recognition. *Tetrahedron* 1998, 54, 3607–3630 b) Koshkin, A.A.; Fensholdt, J.; Pfundheller, H.M.; Lomholt, C. A simplified and efficient route to 2'-0, 4'-C-methylene-linked bicyclic ribonucleosides (Locked Nucleic Acid). J. Org. Chem. 2001, 66, 8504–8512.
- 25. (a) Obika, S.; Abdur Rahman, S.M.; Song, B.; Onoda, M.; Koizumi, M.; Morita, K.; Imanishi, T. Synthesis and properties of 3'-amino-2',4'-BNA, a bridged nucleic acid with a N3'-P5' phosphoramidate linkage *Bioorg*. Med. Chem. **2008**, 16, 9230–9232. b) Olsen, A. G.; Nielsen, C.; Wengel, J. Synthesis and evaluation of anti-HIV activity of 3-azido-4-(hydroxymethyl)tetrahydrofuran derivatives containing 2-(thymin-1-yl)methyl, 2-(cytosin-1-yl)methyl or 2-(adenin-9-yl)methyl substituents—a new series of AZT analogues. *J. Chem. Soc. Perkin Trans.* 1 **2001**, 900–904.

### V. K. Sharma et al.

- Bryld, T.; Sørensen, M.H.; Nielsen, P.; Koch, T.; Nielsen, C.; Wengel, J. Synthesis and antiviral evaluation of novel conformationally locked nucleosides and masked 5-phosphate derivatives thereof. *J. Chem. Soc. Perkin Trans.* 1 2002, 1655–1662.
- Obika, S.; Andoh, J.; Sugimoto, T.; Miyashita, K.; Imanishi, T. Synthesis of a conformationally locked AZT analogue, 3'-azido-3'-deoxy-2'-O,4'-C-methylene-5-methyluridine. *Tetrahedron Lett.* 1999, 40, 6465–6468.

<sup>26.</sup> Czernecki, S.; Valery, J.M. U.S. Patent 5,124,442.