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1-F-LPA: X = H, Y = OH1-F-LPE: $X = CH_2CH_2NH_2, Y = OH$ 1-F-LPC: $X = CH_2CH_2N^{\oplus}Me_3, Y = O^{\ominus}$

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Abstract An efficient method of synthesizing fluorine-containing analogues of 1-lysoglycerophospholipids (1-LPLs) by introducing a palmitoyl moiety starting from bis(2,2,2-trifluoroethyl)phosphonoacetate (Still–Gennari reagent) is described. The method effectively employs Horner–Wadsworth–Emmons reagents as masked 1-LPL derivatives to prepare a series of analogues of 1-lysophosphatidic acid (1-LPA), 1-lysophosphatidylethanolamine (1-LPE), and 1-lysophosphatidylcholine (1-LPC).

Key words lysoglycerophospholipid, Horner–Wadsworth–Emmons reaction, fluorine, lysophosphatidic acid, lysophosphatidylethanolamine, lysophosphatidylcholine

Lysoglycerophospholipids (LPLs), in which only one acyl chain is attached to the glycerol moiety at the sn-2 position (1-LPL) or *sn*-1 position (2-LPL), are of considerable current interest as important signaling molecules in living biological systems.¹ Intramolecular acyl chain migration is known to give an equilibrium mixture of 1-LPL and 2-LPL under physiological conditions; the equilibrium generally favors 2-LPL, as shown in Scheme 1.2 On the other hand, fluorine is the most electronegative element of the periodic table and can be considered a reasonable surrogate of a hydroxy group.3 Thus, replacement of a hydroxy group of 1-LPL by a fluorine atom is an important strategy for blocking the acyl migration of 1-LPL to 2-LPL.4 In this context, we have been intrigued with sn-2 palmitoyl 1-F-LPA (1), 1-F-LPE (2), and 1-F-LPC (3), which are fluorine-containing analogues of 1lysophosphatidic acid (1-LPA), 1-lysophosphatidylethanolamine (1-LPE), and 1-lysophosphatidylcholine (1-LPC), respectively. However, the literature contains only one report on the synthesis of 1, by Prestwich et al., 4b and there are no reports on the synthesis of 2 or 3.

RCO₂
$$\xrightarrow{sn-1}$$
 $\xrightarrow{sn-2}$ $\xrightarrow{sn-2}$ $\xrightarrow{1,2\text{-acyl migration}}$ $\xrightarrow{RCO_2 \cdots (R)}$ $\xrightarrow{RCO_2$

Recently, we reported a novel approach to synthesize glycerophospholipids (PLs) based on the Horner-Wadsworth–Emmons (HWE) reaction of easily handled mixed phosphonoacetate, which serves as a masked precursor of PLs.⁵ Herein we describe the facile synthesis of fluorine-containing analogues **1–3** using HWE reagent **6** as a key intermediate, which was derived from methyl bis(2,2,2-

trifluoroethyl)phosphonoacetate (Still–Gennari reagent, $\mathbf{4}$)^{6,7} and fluorine-containing chiral alcohol $\mathbf{5}$ as shown in Scheme 1.

For the synthesis of 5, (S)-2,2-dimethyl-1,3-dioxolane-4-methanol [(S)-solketal, 7]8 was chosen as the starting material (Scheme 2). The p-methoxybenzylation of 7 with pmethoxybenzyl chloride (PMBCl) in the presence of sodium hydride in DMF provided 8 in 96% yield. Deprotection of the acetonide of 8 under acidic conditions afforded diol 9 in 99% yield. Selective protection of the primary hydroxy group of diol 9 with triphenylchloromethane (TrCl) in the presence of triethylamine and N.N-dimethylaminopyridine (DMAP) gave secondary alcohol 10 in 93% yield. Benzylation of 10 with benzyl bromide in the presence of sodium hydride in DMF resulted in the formation of **11** in 94% yield. Selective removal of the triphenylmethyl group of 11 was easily performed with p-toluenesulfonic acid in methanol to afford primary alcohol 12 in 96% yield. Deoxyfluorination of alcohol 12 by a combination of perfluoro-1-butanesulfonyl fluoride (PBSF), triethylamine, and triethylamine trihvdrofluoride provided the corresponding fluoride 13 in 96% yield. The desired chiral alcohol **5** was obtained in 93% yield by oxidative cleavage of the PMB ether of 13 using 1,2dichloro-4,5-dicyanobenzoquinone (DDQ) as an oxidant in a dichloromethane/water mixed solvent system.¹⁰

Nucleophilic substitution of the chiral alcohol **5** at the phosphorus center of Still–Gennari reagent (**4**) in the presence of 1.37 equivalents of 1,8-diazabicyclo[5.4.0]undec-7-

ene (DBU) and molecular sieves (type 3A) furnished the key intermediate **6** as an inseparable diastereomeric mixture (ca. 1:1) in 87% yield as shown in Scheme 3.^{5,11}

1-F-LPA (1), the fluorine-containing analogue of 1-lysophosphatidic acid (1-LPA), was prepared using an HWE reaction of mixed phosphonoacetate 17 with benzaldehyde as the key reaction as shown in Scheme 4. The nucleophilic substitution of 2-(trimethylsilyl)ethanol (14)12 at the phosphorus center of the key intermediate 6 in the presence of excess amounts of 14 and DBU gave phosphonoacetate 15 in 76% yield. After removal of the benzyl group of 15 by hydrogenolysis using a palladium on carbon (Pd/C) catalyst, palmitic acid was incorporated into the resultant secondary alcohol 16 with 2-methyl-6-nitrobenzoic anhydride (MNBA)^{13,14} and DMAP to afford **17** in 71% yield (two steps). The HWE reaction of 17 with benzaldehyde in the presence of lithium hexamethyldisilazide (LHMDS) provided the expected phosphodiester 18, then deprotection of the 2-(trimethylsilyl)ethyl group of the resultant 18 furnished 1-

F-LPA (1) in 79% yield (two steps). In the synthetic strategy, HWE reagent 17 should be regarded as the masked precursor of *sn*-2 palmitoyl 1-F-LPA (1). Compounds 15–17 were all obtained as inseparable diastereomeric mixtures (ca. 1:1), similar to the key intermediate 6.

Subsequently, we explored the preparation of 1-F-LPE (2) and 1-F-LPC (3) based on the synthetic route of 1-F-LPA (1) through 6 as the common key intermediate. The divergent synthesis of 1-F-LPE (2), fluorine-containing analogues of 1-lysophosphatidylethanolamine (1-LPE), is shown in Scheme 5. The substitution of tert-butyl (2-hydroxyethyl)carbamate (N-Boc-ethanolamine, 19) instead of 14 at the phosphorus center of 6 afforded phosphonoacetate 20 in 67% yield. Hydrogenolysis of 20 using a Pd/C catalyst, followed by condensation of 21 with palmitic acid in the presence of MNBA and DMAP, gave 22 in 56% yield (two steps). The HWE reaction of **22** with benzaldehyde in the presence of LHMDS, followed by deprotection of the Boc group of the resultant phosphodiester 23 under acidic conditions using hydrogen chloride in 1,4-dioxane, afforded 1-F-LPE (2) as hydrochloride salt in 48% yield (two steps).

Furthermore, Scheme 6 shows the synthesis of 1-F-LPC (**3**), a fluorine-containing analogue of 1-lysophosphatidylcholine (1-LPC). 2-Bromoethanol (**24**) was used in the reac-

tion with the key intermediate **6** to afford the corresponding phosphonoacetate **25** in 72% yield in a manner similar to that described for the reaction of **6** with **14** and **19**. After hydrogenolysis of **25** using a Pd/C catalyst, condensation of the resultant **26** with palmitic acid using MNBA and DMAP provided **27** in 72% yield (two steps). The HWE reaction of **27** with benzaldehyde in the presence of LHMDS gave phosphodiester **28**, then amination of the resultant **28** in the presence of excess amounts of trimethylamine in ethanol furnished 1-F-LPC (**3**) in 65% yield (two steps). ¹⁵

In conclusion, we have described a novel and efficient method of synthesizing *sn*-2 palmitoyl 1-F-LPA (1), 1-F-LPE (2), and 1-F-LPC (3) as 1,2-acyl migration-blocked analogues of 1-LPLs. Considering the operational ease based on the use of HWE reagents as fluorine-containing masked analogues of 1-LPLs via the common key intermediate **6**, we believe this synthetic strategy will be valuable for the chemistry and biochemistry of phospholipids classified as glycerophospholipids (PLs) and sphingophospholipids (SPLs).

All melting points were determined on a Yanagimoto micro melting point apparatus and are uncorrected. IR spectra were obtained using a JASCO FT/IR-6200 IR Fourier transform spectrophotometer. $^1\!H$ NMR (500 MHz) and $^{13}\!C$ NMR (125 MHz) spectra were recorded on a Bruker AV500 spectrometer. Chemical shifts are given in δ values (parts per million) using TMS as an internal standard. Mass spectra

chased.

(ESI) were recorded on a Waters LCT Premier spectrometer. Elemental

combustion analyses were performed using a J-SCIENCE LAB JM10

analyzer. Optical rotations were recorded using a P-2200 JASCO digi-

tal polarimeter. All reactions were monitored by TLC employing 0.25

mm silica gel plates (Merck 5715; 60 F₂₅₄). Column chromatography

was carried out on silica gel [Silica Gel 60N (Kanto Chemical) or COS-

MOSIL 75 SL-II-PREP (Nacalai Tesque)]. Anhydrous THF, CH₂Cl₂, DMF,

and toluene were used as purchased from Kanto Chemical. DBU and

Et₃N were distilled prior to use. All other reagents were used as pur-

ic layers were washed with H₂O (30 mL), dried (anhyd MgSO₄), filtered, and concentrated in vacuo. The oily residue was purified by column chromatography [Silica Gel 60N: n-hexane-EtOAc (3:1 to 4:1)] to afford **10** (2.67 g, 93%) as a yellow oil; $[\alpha]_D^{28}$ -0.6 (c 1.01, CHCl₂).

¹H NMR (500 MHz, CDCl₃): δ = 7.43–7.41 (m, 6 H), 7.30–7.20 (m, 11 H), 6.86 (d, J = 8.7 Hz, 2 H), 4.46 (s, 2 H), 3.96 (sept, J = 5.6 Hz, 1 H), 3.80 (s, 3 H), 3.57 (dd, J = 9.7, 4.3 Hz, 1 H), 3.52 (dd, J = 9.6, 6.2 Hz, 1 H)H), 3.22 (dd, I = 9.4, 5.7 Hz, 1 H), 3.19 (dd, I = 9.4, 5.3 Hz, 1 H), 2.39 (d, I = 4.8 Hz, 1 H).

 13 C NMR (125 MHz, CDCl₃): δ = 159.2, 143.8, 130.0, 129.3, 128.6, 127.8, 127.0, 113.7, 86.6, 73.0, 71.2, 69.9, 64.5, 55.2.

HRMS (ESI): m/z [M + Na]⁺ calcd for $C_{30}H_{30}O_4Na$: 477.2042; found: 477.2047.

(S)-4-{[(4-Methoxybenzyl)oxy]methyl}-2,2-dimethyl-1,3-dioxolane (8)16

To a suspension of NaH (50-72% in mineral oil, 187 mg, 3.90-5.61 mmol) in anhyd DMF (10 mL) was added 7 (0.5 mL, 4.05 mmol), and the reaction mixture was stirred at 0 °C for 30 min under argon. After the addition of PMBCI (0.58 mL, 4.28 mmol), the mixture was allowed to warm to r.t. and then stirred for 2 h under argon. H₂O (10 mL) was added to the mixture, and then extracted with EtOAc-n-hexane (1:1) $(3 \times 50 \text{ mL})$. The combined organic layers were dried (anhyd MgSO₄), filtered, and concentrated in vacuo. The oily residue was purified by column chromatography [Silica Gel 60N: n-hexane-EtOAc (4:1)] to afford **8** (986 mg, 96%) as a colorless oil; $[\alpha]_D^{24} + 22.5$ (c 1.01, CHCl₃).

IR (neat): 2986, 2935, 2865, 1613, 1514, 1457, 1371, 1249 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): δ = 7.28–7.24 (m, 2 H), 6.90–6.85 (m, 2 H), 4.52 (d, J = 11.7 Hz, 1 H), 4.48 (d, J = 11.7 Hz, 1 H), 4.28 (quint, J = 6.2)Hz, 1 H), 4.04 (dd, J = 8.3, 6.4 Hz, 1 H), 3.80 (s, 3 H), 3.72 (dd, J = 8.3, 6.3 Hz, 1 H), 3.53 (dd, J = 9.8, 5.6 Hz, 1 H), 3.42 (dd, J = 9.8, 5.7 Hz, 1 H), 1.41 (s, 3 H), 1.36 (s, 3 H).

¹³C NMR (125 MHz, CDCl₃): δ = 159.3, 130.1, 129.3, 113.8, 109.4, 74.8, 73.2, 70.8, 67.0, 55.3, 26.8, 25.4.

HRMS (ESI): m/z [M + Na]⁺ calcd for $C_{14}H_{20}O_4Na$: 275.1259; found: 275.1238.

Anal. Calcd for C₁₄H₂₀O₄: C, 66.65; H, 7.99. Found: C, 66.35; H, 8.08.

(R)-3-[(4-Methoxybenzyl)oxy]propane-1,2-diol (9)^{16,17}

A mixture of 8 (912 mg, 3.61 mmol), aq 1 M HCl (10 mL, 10 mmol), and THF (10 mL) was stirred at r.t. for 2 h. Then, sat. aq NaHCO₃ (10 mL) was added to the reaction mixture and extracted with EtOAc (3 \times 70 mL). The combined organic layers were dried (anhyd MgSO₄), filtered, and concentrated in vacuo. The oily residue was purified by column chromatography [Silica Gel 60N: CHCl3-MeOH (15:1)] to afford **9** (760 mg, 99%) as a white solid; mp 37.5–39 °C (white powder, CHCl₃-n-hexane); $[\alpha]_{D}^{25}$ -2.2 (c 1.23, CHCl₃).

IR (KBr): 3330, 2934, 2870, 1612, 1514, 1250 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): δ = 7.26–7.22 (m, 2 H), 6.90–6.86 (m, 2 H), 4.47 (s, 2 H), 3.90-3.84 (m, 1 H), 3.80 (s, 3 H), 3.71-3.65 (m, 1 H), 3.63-3.57 (m, 1 H), 3.55-3.48 (m, 2 H), 2.85 (br s, 1 H), 2.41 (br s, 1 H). ¹³C NMR (125 MHz, CDCl₃): δ = 159.4, 129.8, 129.5, 113.9, 73.3, 71.5, 70.7, 64.1, 55.3.

HRMS (ESI): m/z [M + Na]⁺ calcd for $C_{11}H_{16}O_4Na$: 235.0946; found: 235.0924.

Anal. Calcd for C₁₁H₁₆O₄: C, 62.25; H, 7.60. Found: C, 61.95; H, 7.61.

(S)-1-[(4-Methoxybenzyl)oxy]-3-(trityloxy)propan-2-ol (10)¹⁷

To a solution of 9 (1.34 g, 6.31 mmol) in anhyd DMF (10 mL) were added Et₃N (2.63 mL, 18.9 mmol), DMAP (38 mg, 0.311 mmol), and TrCl (3.06 g, 11.0 mmol) at r.t. under argon. The reaction mixture was

(S)-({2-(Benzyloxy)-3-[(4-methoxybenzyl)oxy]propoxy}methanetriyl)tribenzene (11)

To a solution of 10 (1.70 g, 3.74 mmol) and benzyl bromide (533 μL , 4.48 mmol) in anhyd DMF (8 mL) was added NaH (50-72% in mineral oil; washed with several portions of anhyd n-pentane, 179 mg, 7.46 mmol) at 0 °C under argon. The reaction mixture was stirred at r.t. for 18 h under argon. Then, H₂O (10 mL) was added to the mixture and extracted with EtOAc-n-hexane (1:1) (3 × 50 mL). The combined organic layers were washed with H₂O (2 × 30 mL), dried (anhyd MgSO₄), filtered, and concentrated in vacuo. The oily residue was purified by column chromatography [Silica Gel 60N: n-hexane-EtOAc (8:1 to 10:1)] to afford **11** (1.91 g, 94%) as a yellow oil; $[\alpha]_D^{23}$ -6.8 (*c* 0.88, CHCl₃).

IR (neat): 3060, 3031, 2931, 2868, 1612, 1513, 1449, 1248 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): δ = 7.45–7.42 (m, 6 H), 7.36–7.15 (m, 16 H), 6.85-6.81 (m, 2 H), 4.66 (d, J = 12.0 Hz, 1 H), 4.63 (d, J = 12.0 Hz, 1 H), 4.44 (d, I = 11.7 Hz, 1 H), 4.42 (d, I = 11.7 Hz, 1 H), 3.77 (s, 3 H), 3.79-3.72 (m, 1 H), 3.63 (dd, J = 10.2, 4.6 Hz, 1 H), 3.59 (dd, J = 10.2, 5.8 Hz, 1 H), 3.29-3.22 (m, 2 H).

¹³C NMR (125 MHz, CDCl₃): δ = 159.1, 144.1, 138.7, 130.5, 129.1, 128.7, 128.3, 127.7, 127.4, 126.9, 113.7, 86.6, 77.7, 72.9, 72.2, 70.3, 63.6, 55.2.

HRMS (ESI): m/z [M + Na]⁺ calcd for $C_{37}H_{36}O_4Na$: 567.2511; found: 567.2511.

(R)-2-(Benzyloxy)-3-[(4-methoxybenzyl)oxy]propan-1-ol (12)¹⁸

To a solution of 11 (100 mg, 0.184 mmol) in MeOH (1.8 mL) was added p-TsOH·H₂O (39 mg, 0.205 mmol) at r.t. The reaction mixture was stirred for 3 h. Then, aq 5 N NaOH (1 mL) was added to the mixture and extracted with CHCl₃ (3 × 10 mL). The combined organic layers were dried (anhyd MgSO₄), filtered, and concentrated in vacuo. The oily residue was purified by column chromatography [Silica Gel 60N: *n*-hexane–EtOAc (3:1)] to afford **12** (53.2 mg, 96%) as a colorless oil; $[\alpha]_{D}^{23}$ +17.8 (c 0.98, CHCl₃).

IR (neat): 3440, 3031, 2868, 1612, 1514, 1455, 1248 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): δ = 7.36–7.26 (m, 5 H), 7.26–7.22 (m, 2 H), 6.89-6.85 (m, 2 H), 4.69 (d, J = 11.8 Hz, 1 H), 4.60 (d, J = 11.8 Hz, 1 H), 4.47 (d, J = 11.6 Hz, 1 H), 4.46 (d, J = 11.6 Hz, 1 H), 3.80 (s, 3 H), 3.77-3.71 (m, 1 H), 3.70-3.63 (m, 2 H), 3.62-3.54 (m, 2 H), 2.16 (br t, 1 H).

¹³C NMR (125 MHz. CDCl₂): δ = 159.2. 138.2. 130.0. 129.3. 128.4. 127.8, 113.8, 78.0, 73.2, 72.1, 69.9, 62.9, 55.3.

stereomer), 33.8 (d, ${}^{1}J_{C,P}$ = 141.5 Hz for one diastereomer). HRMS (ESI): m/z [M + Na]⁺ calcd for $C_{15}H_{19}F_{4}O_{6}PNa$: 425.0753; found: 425.0747.

HRMS (ESI): m/z [M + Na]⁺ calcd for $C_{18}H_{22}O_4Na$: 325.1416; found: 325.1416.

(S)-1-{[2-(Benzyloxy)-3-fluoropropoxy]methyl}-4-methoxybenzene (13)

To a solution of **12** (1.14 g, 3.77 mmol) in anhyd THF (10 mL) were added Et₃N (6.24 mL, 45.0 mmol), Et₃N·(HF)₃ (3.66 mL, 22.5 mmol), and PBSF (3.96 mL, 22.5 mmol) at r.t. under argon. The reaction mixture was stirred for 3 h. Then, H₂O (10 mL) was added to the mixture and extracted with CHCl₃ (3 × 50 mL). The combined organic layers were dried (anhyd MgSO₄), filtered, and concentrated in vacuo. The oily residue was purified by column chromatography [Silica Gel 60N: n-hexane–EtOAc (7:1)], then again by column chromatography [n-hexane–EtOAc (10:1)] to afford **13** (1.10 g, 96%) as a colorless oil; $\alpha \ln^{24} + 13.4$ (c 1.02, CHCl₃).

IR (neat): 2865, 1613, 1514, 1455, 1249 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): δ = 7.36–7.25 (m, 5 H), 7.25–7.21 (m, 2 H), 6.89–6.85 (m, 2 H), 4.68 (d, J = 12.0 Hz, 1 H), 4.67 (d, J = 12.0 Hz, 1 H), 4.55 (ddd, ${}^2J_{H,F}$ = 47.3 Hz, ${}^2J_{H,H}$ = 9.8 Hz, ${}^3J_{H,H}$ = 3.7 Hz, 1 H), 4.47–4.45 (m, 2 H), 4.51 (ddd, ${}^2J_{H,F}$ = 47.5 Hz, ${}^2J_{H,H}$ = 9.9 Hz, ${}^3J_{H,H}$ = 5.5 Hz, 1 H), 3.80 (s, 3 H), 3.86–3.78 (m, 1 H), 3.60–3.53 (m, 2 H).

¹³C NMR (125 MHz, CDCl₃): δ = 159.3, 138.2, 130.0, 129.3, 128.4, 127.8, 127.7, 113.8, 83.5 (d, $^{1}J_{CF}$ = 170.8 Hz), 76.6 (d, $^{2}J_{CF}$ = 19.0 Hz), 73.2, 72.4, 68.4 (d, $^{3}J_{CF}$ = 8.0 Hz), 55.3.

HRMS (ESI): m/z [M + Na]⁺ calcd for $C_{18}H_{21}FO_3Na$: 327.1372; found: 327.1374.

(S)-2-(Benzyloxy)-3-fluoropropan-1-ol (5)¹⁹

To a solution of **13** (1.40 g, 4.60 mmol) in CH₂Cl₂ (47 mL)/H₂O (3 mL) was added DDQ (1.15 g, 5.07 mmol) at 0 °C. The reaction mixture was stirred at r.t. for 8 h. Then, sat. aq Na₂CO₃ (10 mL) was added to the mixture and extracted with CHCl₃ (3 × 50 mL). The combined organic layers were dried (anhyd MgSO₄), filtered, and concentrated in vacuo. The oily residue was purified by column chromatography [Silica Gel 60N: n-hexane–EtOAc (3:1)] to afford **5** (791 mg, 93%) as a colorless oil; $[\alpha]_D^{24}$ +14.8 (c 1.01, CHCl₃).

 $IR\ (neat):\ 3416,\ 2953,\ 2885,\ 1455,\ 1348,\ 1209,\ 1119,\ 1060\ cm^{-1}.$

¹H NMR (500 MHz, CDCl₃): δ = 7.38–7.27 (m, 5 H), 4.72 (d, J = 11.7 Hz, 1 H), 4.62 (d, J = 11.7 Hz, 1 H), 4.53 (ddd, ${}^2J_{\rm H,F}$ = 47.2 Hz, ${}^2J_{\rm H,H}$ = 9.9 Hz, ${}^3J_{\rm H,H}$ = 4.4 Hz, 1 H), 4.51 (ddd, ${}^2J_{\rm H,F}$ = 47.3 Hz, ${}^2J_{\rm H,H}$ = 9.9 Hz, ${}^3J_{\rm H,H}$ = 5.2 Hz, 1 H), 3.78–3.70 (m, 2 H), 3.67–3.60 (m, 1 H), 2.12 (br s, 1 H).

 13 C NMR (125 MHz, CDCl₃): δ = 137.9, 128.6, 128.0, 127.9, 82.8 (d, 1 J_{C,F} = 170.8 Hz), 77.9 (d, 2 J_{C,F} = 19.1 Hz), 72.5, 61.4 (d, 3 J_{C,F} = 7.4 Hz).

HRMS (ESI): m/z [M + Na]⁺ calcd for $C_{10}H_{13}FO_2Na$: 207.0797; found: 207.0784.

Methyl 2-{[(S)-2-(Benzyloxy)-3-fluoropropoxy](2,2,2-trifluoroethoxy)phosphoryl}acetate (6)

A solution of alcohol **5** (607 mg, 3.30 mmol) in anhyd toluene (120 mL) was added to a solution of methyl bis(2,2,2-trifluoroethyl)phosphonoacetate (**4**; 773 μ L, 3.62 mmol), DBU (673 μ L, 4.51 mmol), and molecular sieves 3A (1.28 g) in anhyd toluene (44 mL) at r.t. under argon. After stirring the reaction mixture at r.t. for 1.5 h, aq 1 M HCl (10 mL) was added and then extracted with CHCl₃ (3 × 50 mL). The combined organic layers were dried (anhyd MgSO₄), filtered, and concentrated in vacuo. The oily residue was purified by column chromatography [Silica Gel 60N: *n*-hexane–EtOAc (3:2)] to afford **6** (diastereomeric mixture, 1.16 g, 87%) as a colorless oil.

Methyl 2-{[(S)-2-(Benzyloxy)-3-fluoropropoxy][(2-trimethylsilyl)ethoxy|phosphoryl}acetate (15)

DBU (283 μ L, 1.90 mmol) was added to a solution of **6** (300 mg, 0.746 mmol), 2-(trimethylsilyl)ethanol (**14**; 319 μ L, 2.24 mmol), and molecular sieves 3A (300 mg) in anhyd toluene (10 mL) at r.t. under argon. After stirring the reaction mixture at r.t. for 17 h, aq 1 M HCl (10 mL) was added and then extracted with CHCl₃ (3 × 50 mL). The combined organic layers were dried (anhyd MgSO₄), filtered, and concentrated in vacuo. The oily residue was purified by column chromatography [Silica Gel 60N: *n*-hexane–EtOAc (3:2)] to afford **15** (diastereomeric mixture, 240 mg, 76%) as a colorless oil.

¹H NMR (500 MHz, CDCl₃): δ = 7.38–7.27 (m, 5 H), 4.72 (d, J = 11.8 Hz, 1 H), 4.68 (d, J = 11.8 Hz, 1 H), 4.56 (ddd, ${}^2J_{\rm H,F}$ = 47.0 Hz, ${}^2J_{\rm H,H}$ = 9.9 Hz, ${}^3J_{\rm H,H}$ = 4.6 Hz, 1 H), 4.53 (ddd, ${}^2J_{\rm H,F}$ = 47.0 Hz, ${}^2J_{\rm H,H}$ = 9.9 Hz, ${}^3J_{\rm H,H}$ = 5.3 Hz, 1 H), 4.36–4.29 (m, 1 H), 4.26–4.13 (m, 3 H), 3.90–3.81 (m, 1 H), 3.723/3.715 (2 s, 3 H), 3.05–2.93 (m, 2 H), 1.12–1.06 (m, 2 H), 0.03/0.02 (2 s, 9 H).

 13 C NMR (125 MHz, CDCl₃): δ = 166.1 (d, $^2J_{C,P}$ = 4.7 Hz), 137.62, 137.59, 128.5, 127.96, 127.95, 127.9, 127.8, 82.0 (d, $^1J_{C,F}$ = 171.7 Hz), 76.0, 75.9, 75.81, 75.76, 72.34, 72.32, 65.62, 65.56, 65.5, 65.4, 64.13, 64.08, 64.03, 63.97, 63.91, 63.85, 52.58, 52.55, 34.2 (d, $^1J_{C,P}$ = 136.1 Hz for one diastereomer), 34.1 (d, $^1J_{C,P}$ = 135.3 Hz for one diastereomer), 19.80, 19.76, –1.52, –1.53.

HRMS (ESI): m/z [M + Na]* calcd for $C_{18}H_{30}FO_6PSiNa$: 443.1431; found: 443.1431.

(2S)-1-Fluoro-3-({(2-methoxy-2-oxoethyl)[2-(trimethylsi-lyl)ethoxy]phosphoryl}oxy)propan-2-yl Palmitate (17)

A mixture of **15** (43 mg, 0.102 mmol) and 10% Pd/C (21 mg, 0.0197 mmol) in MeOH (1 mL) was stirred at r.t. for 30 min under $\rm H_2$ atmosphere. The reaction mixture was filtered and concentrated in vacuo to afford **16**. A solution of **16** in anhyd $\rm CH_2CI_2$ (3 mL) was added to a solution of palmitic acid (52 mg, 0.203 mmol), 2-methyl-6-nitrobenzoic anhydride (70 mg, 0.203 mmol), and DMAP (37 mg, 0.303 mmol) in anhyd $\rm CH_2CI_2$ (10 mL) at r.t. under argon. The mixture was stirred for 40 min. Aq 1 M HCl (2 mL) was added to the mixture and then extracted with $\rm CHCI_3$ (3 × 10 mL). The combined organic layers were dried (anhyd MgSO₄), filtered, and concentrated in vacuo. The residue was purified by column chromatography [Silica Gel 60N: n-hexane–EtOAc (3:2)] to afford **17** (diastereomeric mixture, 41 mg, 71%, two steps) as a colorless oil.

¹H NMR (500 MHz, CDCl₃): δ = 5.26–5.16 (m, 1 H), 4.64–4.60 (m, 1 H), 4.55–4.51 (m, 1 H), 4.38–4.16 (m, 4 H), 3.74 (s, 3 H), 3.00 (d, ${}^2J_{\rm H,P}$ = 21.6 Hz, 1 H for one diastereomer), 2.98 (d, ${}^2J_{\rm H,P}$ = 21.5 Hz, 1 H for one

diastereomer), 2.37 (br td, 2 H), 1.67–1.59 (m, 2 H), 1.34–1.22 (m, 24 H), 1.11 (t, J = 8.6 Hz, 2 H), 0.88 (t, J = 7.1 Hz, 3 H), 0.047/0.045 (2 s, 9 H).

¹³C NMR (125 MHz, CDCl₃): δ = 172.83, 172.82, 165.9 (d, ${}^{2}J_{C,P}$ = 6.2 Hz), 80.8 (d, ${}^{1}J_{C,F}$ = 172.8 Hz for one diastereomer), 80.7 (d, ${}^{1}J_{C,F}$ = 173.4 Hz for one diastereomer), 70.3, 70.23, 70.18, 70.13, 70.07, 70.0, 65.7, 65.6, 63.33, 63.28, 63.22, 63.17, 52.6, 34.14, 34.11 (d, ${}^{1}J_{C,P}$ = 136.1 Hz for one diastereomer), 34.07 (d, ${}^{1}J_{C,P}$ = 135.4 Hz for one diastereomer), 31.9, 29.7, 29.69, 29.66, 29.6, 29.5, 29.4, 29.3, 29.1, 24.8, 22.7, 19.83, 19.82, 19.79, 19.77, 14.1, –1.51.

HRMS (ESI): m/z [M + Na]⁺ calcd for $C_{27}H_{54}FO_7PSiNa$: 591.3258; found: 591.3248.

(S)-1-Fluoro-3-(phosphonooxy)propan-2-yl Palmitate [1-F-LPA (1)]^{4b}

To a solution of **17** (70 mg, 0.123 mmol) in anhyd THF (1.26 mL) was added LHMDS (1.1 mol/L in n-hexane, 137 μ L, 0.151 mmol) and the solution was stirred at 0 °C for 5 min under argon. After the addition of benzaldehyde (15 μ L, 0.147 mmol), the reaction mixture was allowed to warm to r.t. and then stirred for 50 min under argon. Aq 1 M HCl (2 mL) was added to the mixture and then extracted with CHCl₃ (3 × 10 mL). The combined organic layers were dried (anhyd MgSO₄), filtered, and concentrated in vacuo. The crude **18** was dissolved in anhyd CH₂Cl₂ (1.25 mL), and TFA (385 μ L, 5.03 mmol) was added. After stirring at r.t. for 1 h, the mixture was concentrated in vacuo. The residue was purified by column chromatography [COSMOSIL 75 SL-II-PREP: CHCl₃-MeOH (100:1 to 6:1)] to afford 1-F-LPA (1) (39.9 mg, 79%, two steps) as a white solid; mp 80–93 °C; $[\alpha]_D^{27}$ +4.8 (c 0.79, CHCl₃).

IR (KBr): 2918, 2849, 1729, 1468, 1179 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): δ = 5.23 (br d, 1 H), 4.93 (br s, 2 H), 4.58 (br d, 2 H), 4.23–4.10 (m, 2 H), 2.44–2.31 (m, 2 H), 1.68–1.55 (m, 2 H), 1.40–1.10 (m, 24 H), 0.88 (t, *J* = 7.1 Hz, 3 H).

¹³C NMR (125 MHz, CDCl₃): δ = 174.2, 80.8 (d, ¹J_{C,F} = 173.5 Hz), 70.6 (d, ²J_{C,F} = 17.3 Hz), 64.0, 34.2, 31.9, 29.74, 29.72, 29.69, 29.68, 29.5, 29.4, 29.3, 29.1, 24.8, 22.7, 14.1.

HRMS (ESI): m/z [M - H]⁻ calcd for $C_{19}H_{37}FO_6P$: 411.2312; found: 411.2310.

Methyl 2-([(S)-2-(Benzyloxy)-3-fluoropropoxy]{2-[(tert-butoxy-carbonyl)amino]ethoxy}phosphoryl)acetate (20)

To a solution of **6** (53.4 mg, 0.133 mmol), *tert*-butyl (2-hydroxyethyl)carbamate (**19**; 64.2 mg, 0.398 mmol), and molecular sieves 3A (107 mg) in anhyd toluene (2 mL) was added DBU (59.4 μ L, 0.398 mmol) at r.t. under argon. After stirring at r.t. for 20 h, MeOH (537 μ L, 13.3 mmol) was added and the reaction mixture was stirred at r.t. for another 15 min. The mixture was filtered, and a 1/15 M phosphate buffer (pH 7.4, 10 mL) was added to the filtrate, and then extracted with CHCl₃ (3 × 10 mL). The combined organic layers were dried (anhyd MgSO₄), filtered, and concentrated in vacuo. The oily residue was purified by column chromatography [Silica Gel 60N: n-hexane–EtOAc (1:2 to 1:3)] to afford **20** (diastereomeric mixture, 41.3 mg, 67%) as a colorless oil.

 1 H NMR (500 MHz, CDCl₃): δ = 7.38–7.28 (m, 5 H), 5.15 (br s, 1 H), 4.70 (dd, J = 11.7, 2.9 Hz, 1 H), 4.68 (d, J = 11.7 Hz, 1 H), 4.61–4.54 (m, 1 H), 4.51–4.45 (m, 1 H), 4.38–4.28 (m, 1 H), 4.23–4.05 (m, 3 H), 3.89–3.81 (m, 1 H), 3.74/3.73 (2 s, 3 H), 3.43–3.28 (m, 2 H), 3.08–2.96 (m, 2 H), 1.44 (s, 9 H).

(2S)-1-[({2-[(tert-Butoxycarbonyl)amino]ethoxy}{2-methoxy-2-oxoethyl)phosphoryl)oxy|-3-fluoropropan-2-y| Palmitate (22)

A mixture of **20** (54.3 mg, 0.117 mmol) and 10% Pd/C (24.9 mg, 0.0234 mmol) in MeOH (1 mL) was stirred at r.t. for 30 min under $\rm H_2$ atmosphere. The reaction mixture was filtered and concentrated in vacuo to afford **21**, which was used in the next reaction without further purification. To a solution of palmitic acid (60 mg, 0.234 mmol), 2-methyl-6-nitrobenzoic anhydride (80.6 mg, 0.234 mmol), and DMAP (42.9 mg, 0.351 mmol) in anhyd $\rm CH_2Cl_2$ (3 mL) was added a solution of **21** in anhyd $\rm CH_2Cl_2$ (2 mL) at r.t. under argon. The mixture was stirred for 30 min. A 1/15 M phosphate buffer (pH 7.4, 3 mL) was added to the mixture, and then extracted with CHCl₃ (3 × 10 mL). The combined organic layers were dried (anhyd MgSO₄), filtered, and concentrated in vacuo. The residue was purified by column chromatography [Silica Gel 60N: n-hexane–EtOAc (1:1)] to afford **22** (diastereomeric mixture, 40.2 mg, 56%, two steps) as a colorless oil.

¹H NMR (500 MHz, CDCl₃): δ = 5.26–5.14 (m, 2 H), 4.65–4.59 (m, 1 H), 4.55–4.49 (m, 1 H), 4.38–4.10 (m, 4 H), 3.76 (s, 3 H), 3.48–3.35 (m, 2 H), 3.04 (d, $^2J_{\rm H,P}$ = 21.6 Hz, 0.97 H for one diastereomer), 3.02 (br dd, 1.03 H for one diastereomer), 2.37 (t, J = 7.6 Hz, 2 H), 1.67–1.60 (m, 2 H), 1.45 (s, 9 H), 1.34–1.23 (m, 24 H), 0.88 (t, J = 7.1 Hz, 3 H).

 ^{13}C NMR (125 MHz, CDCl₃): δ = 172.89, 172.88, 165.9 (d, $^2J_{C,P}$ = 5.8 Hz), 155.8, 80.7 (d, $^1J_{C,F}$ = 173.3 Hz for one diastereomer), 80.6 (d, $^1J_{C,F}$ = 173.4 Hz for one diastereomer), 79.6, 70.16, 70.11, 70.06, 70.00, 69.95, 69.90, 66.41, 66.36, 63.6, 63.53, 63.48, 63.43, 63.38, 52.8, 41.04, 41.01, 34.1, 33.8 (d, $^1J_{C,P}$ = 137.3 Hz for one diastereomer), 33.7 (d, $^1J_{C,P}$ = 137.1 Hz for one diastereomer), 31.9, 29.71, 29.69, 29.66, 29.62, 29.5, 29.4, 29.3, 29.1, 28.4, 24.8, 22.7, 14.1.

HRMS (ESI): m/z [M + Na]⁺ calcd for $C_{29}H_{55}FNO_{9}PNa$: 634.3496; found: 634.3470.

(2S)-1-{[(2-Aminoethoxy)(hydroxy)phosphoryl]oxy}-3-fluoropropan-2-yl Palmitate Hydrochloride [1-F-LPE (2)·HCl]

To a solution of **22** (69.2 mg, 0.113 mmol) in anhyd THF (1 mL) was added LHMDS (1.02 mol/L in n-hexane, 133 μ L, 0.136 mmol) and the solution was stirred at 0 °C for 5 min under argon. After adding benzaldehyde (13.9 μ L, 0.136 mmol), the reaction mixture was allowed to warm to r.t. and stirred for 15 min under argon. A 1/15 M phosphate buffer (pH 7.4, 3 mL) was added to the mixture, and then concentrated in vacuo. The residue was purified by column chromatography [COSMOSIL 75 SL-II-PREP: CHCl₃–MeOH (100:1 to 2:1)] to afford **23**, which was used in the next reaction without further purification. A mixture of **23** and 4 M HCl in 1,4-dioxane (0.57 mL, 2.26 mmol) in CH₂Cl₂ (1 mL) was stirred at r.t. for 1.5 h and concentrated in vacuo. The residue was purified by column chromatography [Silica Gel 60N: CHCl₃–MeOH (2:1)] to afford 1-F-LPE (**2**)·HCl (26.6 mg, 48%, two steps) as a white solid; mp 188–191 °C; $[\alpha]_D^{27}$ +9.7 [c 0.55, CHCl₃–MeOH (2:1)].

IR (KBr): 2920, 2851, 1741, 1467, 1252, 1225, 1082 cm⁻¹.

¹³C NMR (125 MHz, CDCl₃): δ = 172.9, 165.7 (d, ${}^{2}J_{C,P}$ = 5.4 Hz), 80.7 (d, ${}^{1}J_{C,F}$ = 173.3 Hz for one diastereomer), 80.6 (d, ${}^{1}J_{C,F}$ = 172.9 Hz for one diastereomer), 70.12, 70.08, 70.06, 70.02, 69.95, 69.91, 69.86, 66.0, 65.97, 65.95, 65.92, 63.7, 63.63, 63.58, 63.53, 63.5, 63.4, 52.8, 34.14, 34.12, 33.94 (d, ${}^{1}J_{C,P}$ = 138.7 Hz for one diastereomer), 33.89 (d, ${}^{1}J_{C,P}$ = 138.2 Hz for one diastereomer), 31.9, 29.7, 29.69, 29.66, 29.65, 29.62, 29.5, 29.4, 29.3, 29.1, 24.8, 22.7, 14.1.

HRMS (ESI): m/z [M + Na]⁺ calcd for $C_{24}H_{45}BrFO_7PNa$: 597.1968; found: 597.1911.

¹H NMR [500 MHz, CDCl₃–CD₃OD (2:1)]: δ = 5.23–5.13 (m, 1 H), 4.61 (dd, $^2J_{\rm H,F}$ = 47.1 Hz, $^3J_{\rm H,H}$ = 4.1 Hz, 2 H), 4.07–3.99 (m, 4 H), 3.13–3.09 (m, 2 H), 2.39–2.34 (m, 2 H), 1.67–1.58 (m, 2 H), 1.36–1.22 (m, 24 H), 0.89 (t, J = 7.1 Hz, 3 H).

¹³C NMR [125 MHz, CDCl₃–CD₃OD (2:1)]: δ = 173.9, 81.5 (d, ${}^{1}J_{CF}$ = 171.8 Hz), 71.3 (dd, ${}^{2}J_{CF}$ = 19.4 Hz, ${}^{3}J_{CP}$ = 8.4 Hz), 62.8 (dd, ${}^{3}J_{CF}$ = 7.4 Hz, ${}^{2}J_{CP}$ = 5.2 Hz), 61.8 (d, ${}^{2}J_{CP}$ = 5.2 Hz), 40.8 (d, ${}^{3}J_{CP}$ = 5.4 Hz), 34.4, 32.2, 29.92, 29.89, 29.7, 29.6, 29.5, 29.3, 25.1, 22.9, 14.2.

HRMS (ESI): m/z [M - H - HCl]⁻ calcd for $C_{21}H_{42}FNO_6P$: 454.2734; found: 454.2732.

Anal. Calcd for $C_{21}H_{44}$ CIFNO₆P: C, 51.26; H, 9.01; N, 2.85. Found: C, 51.23; H, 8.85; N, 2.98.

Methyl 2-{[(S)-2-(Benzyloxy)-3-fluoropropoxy](2-bromoethoxy)phosphoryl}acetate (25)

To a solution of **6** (50 mg, 0.124 mmol), 2-bromoethanol (**24**; 13 μ L, 0.186 mmol), and molecular sieves 3A (50 mg) in anhyd toluene (0.8 mL) was added DBU (28 μ L, 0.186 mmol) at 0 °C under argon. After stirring at 0 °C for 7 h, a 1/15 M phosphate buffer (pH 7.4, 3 mL) was added to the reaction mixture. The mixture was filtered, and then extracted with CHCl₃ (3 × 10 mL). The combined organic layers were dried (anhyd MgSO₄), filtered, and concentrated in vacuo. The oily residue was purified by column chromatography [Silica Gel 60N: n-hexane–EtOAc (2:3) to afford **25** (diastereomeric mixture, 38.4 mg, 72%) as a colorless oil.

¹H NMR (500 MHz, CDCl₃): δ = 7.34–7.28 (m, 5 H), 4.71 (dd, J = 11.7, 1.2 Hz, 1 H), 4.68 (dd, J = 11.7, 2.8 Hz, 1 H), 4.62–4.55 (m, 1 H), 4.53–4.45 (m, 1 H), 4.41–4.30 (m, 3 H), 4.25–4.17 (m, 1 H), 3.91–3.81 (m, 1 H), 3.74/3.73 (2 s, 3 H), 3.50/3.48 (2 t, J = 6.2, 6.1 Hz, 2 H), 3.06/3.04/3.03 (dd, ${}^2J_{\rm H,P}$ = 21.9 Hz, ${}^2J_{\rm H,H}$ = 14.8 Hz, dd, ${}^2J_{\rm H,P}$ = 21.5 Hz, 2H).

¹³C NMR (125 MHz, CDCl₃): δ = 165.8 (d, ${}^2J_{\text{C,P}}$ = 5.9 Hz), 137.5, 128.5, 128.0, 127.94, 127.91, 81.8 (d, ${}^1J_{\text{C,F}}$ = 172.1 Hz), 75.8, 75.7, 75.63, 75.59, 72.4, 72.3, 65.9, 65.8, 65.79, 65.74, 64.47, 64.42, 64.37, 64.34, 64.29, 64.23, 52.72, 52.70, 34.0 (d, ${}^1J_{\text{C,P}}$ = 138.9 Hz for one diastereomer), 33.9 (d, ${}^1J_{\text{C,P}}$ = 138.0 Hz for one diastereomer), 29.9, 29.83, 29.79, 29.7.

HRMS (ESI): m/z [M + Na]⁺ calcd for $C_{15}H_{21}BrFO_6PNa$: 449.0141; found: 449.0145.

(2S)-1-{[(2-Bromoethoxy)(2-methoxy-2-oxoethyl)phosphoryl]oxy}-3-fluoropropan-2-yl Palmitate (27)

A mixture of **25** (61.2 mg, 0.143 mmol) and 10% Pd/C (30 mg, 0.0282 mmol) in MeOH (1 mL) was stirred at r.t. for 15 min under $\rm H_2$ atmosphere. The reaction mixture was filtered and concentrated in vacuo to afford **26**, which was used in the next reaction without further purification. To a solution of palmitic acid (73 mg, 0.286 mmol), 2-methyl-6-nitrobenzoic anhydride (99 mg, 0.286 mmol), and DMAP (52 mg, 0.426 mmol) in anhyd CH₂Cl₂ (6 mL) was added a solution of **26** in anhyd CH₂Cl₂ (4 mL) at r.t. under argon. The mixture was stirred for 15 min. A 1/15 M phosphate buffer (pH 7.4, 3 mL) was added to the mixture, and then extracted with CHCl₃ (3 × 10 mL). The combined organic layers were dried (anhyd MgSO₄), filtered, and concentrated in vacuo. The residue was purified by column chromatography [Silica Gel 60N: n-hexane–EtOAc (1:1)] twice to afford **27** (diastereomeric mixture, 59.6 mg, 72%, two steps) as a colorless oil.

(S)-3-Fluoro-2-(palmitoyloxy)propyl [2-(Trimethylammonio)ethyl] Phosphate [1-F-LPC (3)]

To a solution of **27** (68.6 mg, 0.119 mmol) in anhyd THF (1 mL) were added LHMDS (1.02 mol/L in n-hexane, 140 μ L, 0.143 mmol) and benzaldehyde (14.6 μ L, 0.143 mmol) at 0 °C under argon. The reaction mixture was stirred for 15 min under argon. A 1/15 M phosphate buffer (pH 7.4, 3 mL) was added to the mixture, and then concentrated in vacuo. The residue was purified by column chromatography [COSMO-SIL 75 SL-II-PREP: CHCl₃-MeOH (100:1 to 3:1)] to afford **28**, which was used in the next reaction without further purification. A mixture of **28** and Me₃N (ca. 3 mol/L in EtOH, 2 mL, ca. 6 mmol) was stirred at r.t. for 5 d and then concentrated in vacuo. The residue was purified by column chromatography [Silica Gel 60N: CHCl₃-MeOH (3:1) to CHCl₃-MeOH-H₂O (5:5:1)] to afford 1-F-LPC (**3**) (38.7 mg, 65%, two steps) as a white solid; mp 65-67 °C; $[\alpha]_D^{27}$ +9.8 (c 1.09, CHCl₃).

IR (KBr): 2918, 2850, 1737, 1468, 1247, 1093 cm⁻¹.

 1H NMR (500 MHz, CDCl $_3$): δ = 5.15 (br d, 1 H), 4.69–4.52 (m, 2 H), 4.35–4.26 (m, 2 H), 3.98–3.91 (m, 2 H), 3.82–3.76 (m, 2 H), 3.36 (s, 9 H), 2.35–2.28 (m, 2 H), 1.64–1.54 (m, 2 H), 1.34–1.20 (m, 24 H), 0.88 (t, J = 7.1 Hz, 3 H).

¹³C NMR (125 MHz, CDCl₃): δ = 173.5, 82.0 (d, ${}^{1}J_{CF}$ = 171.8 Hz), 71.2 (dd, ${}^{2}J_{CF}$ = 19.6 Hz, ${}^{3}J_{CP}$ = 9.1 Hz), 66.2, 62.4, 59.6, 54.6, 34.3, 32.0, 29.78, 29.77, 29.71, 29.66, 29.5, 29.4, 29.2, 25.0, 22.7, 14.1.

HRMS (ESI): m/z [M + Na]⁺ calcd for $C_{24}H_{49}FNO_6PNa$: 520.3179; found: 520.3178.

Supporting Information

Supporting information for this article is available online at https://doi.org/10.1055/s-0036-1588826.

References

(1) (a) D'Arrigo, P.; Servi, S. Molecules 2010, 15, 1354.
 (b) Nagamatsu, T.; Iwasawa-Kawai, Y.; Ichikawa, M.; Kawana, K.; Yamashita, T.; Osuga, Y.; Fujii, T.; Schust, D. J. Am. J. Reprod. Immunol. 2014, 72, 182. (c) Kihara, Y.; Mizuno, H.; Chun, J. Exp. Cell Res. 2015, 333, 171. (d) Li, J.; Wang, X.; Zhang, T.; Wang, C.; Huang, Z.; Luo, X.; Deng, Y. Asian J. Pharm. Sci. 2015, 10, 81. (e) Yung, Y. C.; Stoddard, N. C.; Mirendil, H.; Chun, J. Neuron 2015, 85, 669. (f) Takeda, A.; Umemoto, E.; Miyasaka, M. Transl. Cancer Res. 2015, 4, 537. (g) Fukushima, N.; Ishii, S.; Tsujiuchi,

- T.; Kagawa, N.; Katoh, K. *Cell. Mol. Life Sci.* **2015**, *72*, 2377. (h) Arifin, S. A.; Falasca, M. *Metabolites* **2016**, *6*, No. 6. (i) Mizejewski, G. J. *Curr. Drug Targets* **2017**, *18*, 874.
- (2) (a) Plückthun, A.; Dennis, E. A. Biochemistry 1982, 21, 1743.
 (b) Adlercreutz, D.; Budde, H.; Wehtje, E. Biotechnol. Bioeng. 2002, 78, 403. (c) Qian, L.; Xu, Y.; Arai, H.; Aoki, J.; McIntyre, T. M.; Prestwich, G. D. Org. Lett. 2003, 5, 4685. (d) Vikbjerg, A. F.; Mu, H.; Xu, X. J. Am. Oil Chem. Soc. 2006, 83, 609. (e) Okudaira, M.; Inoue, A.; Shuto, A.; Nakanaga, K.; Kano, K.; Makide, K.; Saigusa, D.; Tomioka, Y.; Aoki, J. J. Lipid Res. 2014, 55, 2178.
- (3) (a) O'Hagan, D.; Rzepa, H. S. Chem. Commun. 1997, 645.
 (b) Pongdee, R.; Liu, H. Bioorg. Chem. 2004, 32, 393. (c) Jeschke, P. ChemBioChem 2004, 5, 570. (d) Edmonds, M.; Peddie, V. Chem. New Zealand 2006, 70, 85. (e) Müller, K.; Faeh, C.; Diederich, F. Science 2007, 317, 1881. (f) Liu, P.; Sharon, A.; Chu, C. K. J. Fluorine Chem. 2008, 129, 743. (g) Hunter, L. Beilstein J. Org. Chem. 2010, 6, No. 38. (h) Menaa, F.; Menaa, B.; Sharts, O. N. J. Mol. Pharm. Org. Process Res. 2013, 1, 104.
- (4) (a) Xu, Y.; Prestwitch, G. D. J. Org. Chem. 2002, 67, 7158. (b) Xu, Y.; Qian, L.; Prestwitch, G. D. J. Org. Chem. 2003, 68, 5320. (c) Xu, Y.; Aoki, J.; Shimizu, K.; Umezu-Goto, M.; Hama, K.; Takanezawa, Y.; Yu, S.; Mills, G. B.; Arai, H.; Qian, L.; Prestwich, G. D. J. Med. Chem. 2005, 48, 3319.
- (5) Sano, S.; Sumiyoshi, H.; Handa, A.; Tokizane, R.; Nakao, M. Tetrahedron Lett. 2015, 56, 4686.
- (6) (a) Still, W. C.; Gennari, C. Tetrahedron Lett. 1983, 24, 4405.(b) Messik, F.; Oberthür, M. Synthesis 2013, 45, 167.
- (7) DeHoff, B.; Roy, M.-N. Methyl Bis(2,2,2-trifluoroethoxy)phosphinylacetate, In e-EROS Encyclopedia of Reagents for Organic Synthesis; Wiley: New York, 2012.

- (8) (a) Jiang, G.; Xu, Y.; Falguières, T.; Gruenberg, J.; Prestwich, G. D. Org. Lett. 2005, 7, 3837. (b) Kristinsson, B.; Haraldsson, G. G. Synlett 2008, 2178. (c) Machado, A. C. O.; da Silva, A. A. T.; Borges, C. P.; Simas, A. B. C.; Freire, D. M. G. J. Mol. Catal. B: Enzym. 2011, 69, 42.
- (9) Yin, J.; Zarkowsky, D. S.; Thomas, D. W.; Zhao, M. M.; Huffman, M. A. Org. Lett. 2004, 6, 1465.
- (10) Horita, K.; Yoshioka, T.; Yanaka, T.; Oikawa, Y.; Yonemitsu, O. *Tetrahedron* **1986**, *42*, 3021.
- (11) (a) Sano, S.; Takemoto, Y.; Nagao, Y. ARKIVOC 2003, (viii), 93.
 (b) Sano, S.; Kujime, E.; Takemoto, Y.; Shiro, M.; Nagao, Y. Chem. Pharm. Bull. 2005. 53, 131.
- (12) Reddy, J. P.; Yoakim, C. 2-(Trimethylsilyl)ethanol, In e-EROS Encyclopedia of Reagents for Organic Synthesis; Wiley: New York, 2007.
- (13) (a) Shiina, I.; Ibuka, R.; Kubota, M. Chem. Lett. 2002, 31, 286.
 (b) Shiina, I.; Kubota, M.; Oshiumi, H.; Hashizume, M. J. Org. Chem. 2004, 69, 1822. (c) Shiina, I. Tetrahedron 2004, 60, 1587.
- (14) Shiina, I. 2-Methyl-6-nitrobenzoic Anhydride (MNBA), In e-EROS Encyclopedia of Reagents for Organic Synthesis; Wiley: New York, 2009.
- (15) (a) Eibl, H.; Nicksch, A. *Chem. Phys. Lipids* **1978**, 22, 1. (b) Diembeck, W.; Eibl, H. *Chem. Phys. Lipids* **1979**, 24, 237.
- (16) (a) Lim, Z.-Y.; Thuring, J. W.; Holmes, A. B.; Manifava, M.; Ktistakis, N. T. J. Chem. Soc., Perkin Trans. 1 2002, 1067.
 (b) Pilkington, L. I.; Barker, D. Eur. J. Org. Chem. 2014, 1037.
- (17) (a) Qin, D.; Byun, H.-S.; Bittman, R. J. Am. Chem. Soc. 1999, 121, 662. (b) Gil-Mesón, A.; Roncero, A. M.; Tobal, I. E.; Basabe, P.; Díez, D.; Mollinedo, F.; Marcos, I. S. Molecules 2016, 21, 47.
- (18) Fukase, K.; Matsumoto, T.; Ito, N.; Yoshimura, T.; Kotani, S.; Kusumoto, S. *Bull. Chem. Soc. Jpn.* **1992**, *65*, 2643.
- (19) Bravo, P.; Piovosi, E.; Resnati, G. J. Chem. Soc., Perkin Trans. 1 1989, 1201.