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Conformationally constrained N_1 -arylsulfonyltryptamine derivatives as 5-HT₆ receptor antagonists

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Abstract—Several series of conformationally constrained N_1 -arylsulfonyltryptamine derivatives were prepared and tested for 5-HT₆ receptor binding affinity and ability to modulate cAMP production in a cyclase assay. The 3-piperidin-3-yl-, 3-(1-methylpyrrolidin-2-ylmethyl)-, and 3-pyrrolidin-3-yl-1*H*-indole arrays (**8**–13) appear to be able to adopt a conformation that allows high affinity 5-HT₆ receptor binding, while the β -carboline array 14 binds with a significantly weaker (10- to 100-fold) affinity. N_1 -Benzenesulfo-nyl-3-piperidin-3-yl-1*H*-indole **9a** is a high affinity full agonist with EC₅₀ = 24 nM. Several of the N_1 -arylsulfonyl-3-(1-methylpyrrolidin-2-ylmethyl)-1*H*-indole derivatives behave as very potent antagonists ((*S*)-11r, (*S*)-11t; IC₅₀ = 0.8, 1.0 nM). © 2005 Elsevier Ltd. All rights reserved.

The human 5-HT₆ receptor was cloned in 1996 and consists of a 440 amino acid residue, seven-transmembrane protein, with <40% protein sequence homology with the other 5-HT receptors.¹ The 5-HT₆ receptor is positively coupled to adenylyl cyclase,² and located almost exclusively in the central nervous system, with highest density in the cerebral cortex, nucleus accumbens, caudate-putamen, and hippocampus, and moderate densities in the thalamus and substantia nigra.³ A wide range of antipsychotic agents and antidepressants have high affinity for the 5-HT₆ receptor,⁴ stimulating considerable effort to understand its role in treatment of CNS disorders, including schizophrenia, depression, and impaired learning and memory.^{5–10}

Among the first 5-HT₆ receptor antagonists was a series of bismethylaminopyrimidinyl- and bismethylaminopyridinyl-sulfonamides, Ro-04-6790 (1) and Ro-63-0563 (2) (Fig. 1).¹¹ Subsequently, a series of piperazinyl-benzenesulfonamide antagonists SB-271046 (3)¹² and SB-

357134 (4),¹³ with improved affinity and physical properties, were reported. N_1 -arylsulfonyltryptamines, such as MS-245 (5; $K_i = 2.3$ nM), have also been shown to



Figure 1. Structures of 5-HT₆ receptor ligands.

Keywords: N_1 -Arylsulfonyltryptamine; 5-HT6; Serotonin; Receptor; Antagonist.

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bind to the 5-HT₆ receptor with high affinity,^{14,15} as have the conformationally constrained N_1 -arylsulfonyl-3-(1,2,3,6-tetrahydropyridin-4-yl)-1*H*-indole (**6**; $K_i = 2 \text{ nM})^{16,17}$ and the (9-arylsulfonyl-2,3,4,9-tetrahydro-1*H*-carbazol-4-yl)-methylamine (**7**; $K_i = 1.5 \text{ nM})$.¹⁸

In this report, we present the results of a study into the structural-activity relationships of a series of conformationally constrained N_1 -arylsulfonyltryptamine derivatives. The effect of indole substitution on 5-HT₆ receptor binding affinity of N_1 -arylsulfonyltryptamine derivatives has been studied and, in general, substitution on the 4-, 5-, 6-, or 7-positions does not increase the affinity, relative to the unsubstituted indole^{14,16}. Thus, the focus of this work is on the conformationally constrained aminoethyl portion and the N_1 -arylsulfonyl substituents.

Seven arrays, represented by the generic structures 8–14 (Fig. 2), were prepared. Array 8 was prepared, as shown in Scheme 1, by coupling indole with 1-benzyl-3-piperdinone hydrate in refluxing isopropylalcohol (IPA) with excess 1 N KOH to give 3-(1-benzyl-1,2,5,6-tetrahydropyridin-3-yl)-1*H*-indole 15.¹⁹ Removal of the benzyl group was carried out by a modified version of a proce-dure by Olofson et al.²⁰ in which the benzylamine was stirred at room temperature with α -chloroethyl chloroformate (ACE-Cl) in dichloroethane (DCE) to give the α -chloroethyl carbamate, which was cleaved to the free amine by refluxing in MeOH. Subsequent protection gives the N-Boc core 16, which was coupled with a set of sulfonyl chlorides in THF with potassium tert-butoxide. Finally, the Boc group was removed by stirring with 2 N HCl in dioxane. The compounds were purified by reverse-phase semi-preparative HPLC.²¹

Coupling of indole with 1-Boc-3-piperdinone leads to the formation of the enamine isomer 17, due to the electron with-drawing nature of the *N*-substituent.²² Hydrogenation gives the reduced *N*-Boc-3-piperidin-3-yl-1*H*-indole 18. Array 9 was then prepared, as described above for array 8.



Figure 2. Conformationally constrained N_1 -arylsulfonyltryptamine arrays.



Scheme 1. Reagents and conditions: (a) 1-benzyl-3-piperidone, KOH, IPA, Δ ; (b) (i) ACE-Cl, DCE, rt; (ii) MeOH, Δ ; (c) (Boc)₂O, K₂CO₃, acetone, water, rt; (d) ArSO₂Cl, *t*-BuOK, THF, rt; (e) 2 N HCl, dioxane, rt; (f) 1-Boc-3-piperidone, KOH, IPA, Δ ; (g) H₂, Pd/C MeOH, rt.

The reaction of the magnesium salt of indole with either (S)- or (R)-N-Cbz-proline acid chloride in toluene afforded the 3-ketoindole **19** (Scheme 2: (S)-enantiomer shown).²³ Removal of the Cbz group was effected by hydrogenation over palladium on carbon with formic acid in methanol. The keto group was reduced with LiAlH₄ in refluxing THF and the pyrrolidine protected as the N-Boc derivative **20**. Array **10** was prepared, as described above for array **8**. Reduction of **19** with LiAlH₄ in refluxing THF afforded the 3-(1-methyl-pyrrolidin-2-ylmethyl)-1*H*-indole **21**, which was converted to array **11** by sulfonylation.

The synthesis of the N_1 -arylsulfonyl-3-(1-alkyl-pyrrolidin-3-yl)-1*H*-indole arrays **12–13** was carried out by coupling indole with *N*-methyl- and *N*-benzylmaleimide in refluxing acetic acid, followed by reduction with LiAlH₄ to give the 3-(1-methyl- and 3-(1-benzylpyrrolidin-3-yl)-1*H*-indole cores **22–23** (Scheme 3)²³ that were sulfonylated.



Scheme 2. Reagents and conditions: (a) (*S*)-*N*-Cbz-Pro-Cl, EtMgBr, DCM, 0 °C; (b) HCO₂H, Pd/C, MeOH, rt; (c) LiAlH₄, THF, Δ ; (d) (Boc)₂O, K₂CO₃, acetone, water, rt; (e) ArSO₂Cl, *t*-BuOK, THF, rt.



Scheme 3. Reagents and conditions: (a) *N*-methyl- or *N*-benzylmaleimide, AcOH, Δ ; (b) LiAlH₄, THF, Δ ; (c) ArSO₂Cl, *t*-BuOK, THF, rt.

The 9-arylsulfonyl-2,3,4,9-tetrahydro-1H- β -carboline array 14 was prepared by Boc protection of the β -carboline, followed by sulfonylation and deprotection, as described for array 8.

All the conformationally constrained N_1 -arylsulfonyltryptamine derivatives were assayed for their ability to displace [³H]-LSD from cloned human 5-HT₆ receptors stably expressed in HeLa cells.¹⁶ N_1 -Benzenesulfonyl-3-(1,2,5,6-tetrahydropyridin-3-yl)- 1*H*-indole **8a** had excellent affinity ($K_i = 4.6$ nM) for the 5-HT₆ receptor. Mono- and dihalo, alkyl and alkoxy substituted arylsulfonamides (**8b–8q**; $K_i = 8.5-48$ nM) all had slightly reduced affinity, as did the heteroaryl sulfonamides (**8t–8w**; $K_i = 8-58$ nM) (Table 1).

The racemic N_1 -arylsulfonyl-3-piperidin-3-yl-1*H*-indole series **9** showed similar SAR trends with the unsubstituted benzenesulfonamide having high affinity (**9a**; $K_i = 2 \text{ nM}$) and the substituted benzenesulfonamides and heteroarylsulfamides having equal or slightly lower binding affinity (**9f–9x**; $K_i = 2-44 \text{ nM}$). The enantiomers of **9a** were separated²⁴ and showed a 3-fold difference in binding affinity ($K_i = 1 \text{ nM vs. 3 nM}$).

In general, the (*S*)-*N*₁-arylsulfonyl-3-pyrrolidin-2-ylmethyl-1*H*-indole series **10** had lower 5-HT₆ receptor affinity (**10a–10x**; $K_i = 20–283$ nM), with the exception of 4-aminophenylsulfonamide (**10m**; $K_i = 7$ nM). Increased binding affinity for 4-aminophenylsulfonamide tryptamines has been previously reported.²⁵ The corresponding (*R*)-*N*₁-arylsulfonyl-3-pyrrolidin-2-ylmethyl-1*H*-indoles were not prepared. Both (*R*)- and (*S*)-enantiomers of *N*₁-arylsulfonyl-3-(1-methyl-pyrrolidin-2-ylmethyl)-1*H*-indole series **11** had high affinity (**11a–11x**; $K_i = 3–19$ nM). The unsubstituted phenyl-, 3-, and 4-halophenyl-, 4-aminophenyl-, and 5-halothiophenesulfonyl derivatives had the highest affinity. Interestingly, even the large 5-chloro-3-methylbenzothiophene analog retains high affinity (**11x**; $K_i = 11$ nM).

All members of the racemic N_1 -arylsulfonyl-3-(1-methyl-pyrrolidin-3-yl)-1*H*-indole array **12** had excellent 5-HT₆ receptor affinity (**12c–12r**; $K_i = 1-5$ nM). Surprisingly, members of the N_1 -arylsulfonyl-3-(1-benzyl-pyrrolidin-3-yl)-1*H*-indole array **13** also had high affinity (**13k–13y**; $K_i = 1-9$ nM), in spite of a large *N*benzyl substituent. This was in contrast to the N_1 -arylsulfonyl-3-(1,2,3,6-tetrahydropyridin-4-yl)-1*H*-indole series where the larger *N*-Bn derivatives had significantly weaker affinity than the corresponding N-Me analogs.¹⁶

The 9-arylsulfonyl-2,3,4,9-tetrahydro-1*H*- β -carbolines (**14a–14m**; *K*_i = 41–252 nM) had weaker 5-HT₆ receptor affinity.

Each of the compounds 5–9, 11–12, and 14 was expanded to a set of roughly 200 molecule conformations (OMEGA software, OpenEye Scientific), generating 2400 total molecule conformations that were aligned (ROCS software, OpenEye Scientific) with each other. An overlap score was assigned to each alignment based on the molecule shape and pharmacophore orientation. These alignments were analyzed to find out the conformation that had the largest average overlap with the remaining conformations. This conformation was defined as the reference conformation. A consensus of those conformations, which match that of the reference conformation, was obtained and is shown in Figure 3.

With the N_1 -phenylsulfonylindole groups aligned, the basic amine of compounds 5–9 and 11–12 can adopt a similar position. This is in contrast to 14 that cannot adopt a conformation where the basic amine overlays with the basic amine of the other molecules.

Glennon has shown that in the N_1 -arylsulfonylskatole series absence of a basic amine results only in a 3-fold loss in affinity, relative to the tryptamine with the 4-aminophenylsulfonyl group.²⁵ However, we show here that compound **14m** ($K_i = 116$ nM), which has this 4-aminophenylsulfonyl group and where the basic amine is restricted to a position that does not overlap with the basic amines of the other cores, is about 100-fold less potent than the corresponding N_1 -(4-aminophenylsulfonyl)-3-(1-methyl-pyrrolidin-3-yl)-1*H*-indole **12m** ($K_i = 1$ nM).

High affinity compounds were assayed for their ability to modulate cAMP production in a cyclase assay (Table 1).¹⁶ None of the N_1 -arylsulfonyl-3-(1,2,5,6-tetrahydropyridin-3-yl)-1*H*-indole derivatives 8 showed any antagonist activity; however, several compounds of the series did show a modest ability to stimulate cAMP production indication agonist activity. Several racemic N_1 -arylsulfonyl-3-piperidin-3-yl-1H-indole analogs had agonist activity. Of particular interest is 9a that stimulated cAMP production with an EC₅₀ value of 24 nM and behaved as a full agonist, and although the enantiomers had only 3-fold difference in affinity ($K_i = 1 \text{ nM}$ vs. 3 nM) all the agonist activity could be attributed to one enantiomer, while the diastomer failed to show any functional activity. The N_1 -(4-methoxybenzene)sulfonyl-3-piperidin-3-yl-1*H*-indole 9l ($IC_{50} = 7 \text{ nM}$) behaved as a partial antagonist.

Several of the N_1 -arylsulfonyl-3-(1-methylpyrrolidin-2ylmethyl)-1*H*-indole derivatives were very potent antagonists in the cAMP stimulation assay, in particular, (*S*)-11*r* and (*S*)-11*t* were full antagonists with IC₅₀ values of approximately 1 nM. Members of the N_1 -arylsulfonyl-3-(1-methyl- and N_1 -arylsulfonyl-3-(1-benzyl-

$-\mathbf{u}$	Table 1. 5-HT	's binding affinity	and functional a	ctivity of constra	ined N ₁ -arv	lsulfonvltryptamines
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Compound	Ar-SO ₂	$K_i (nM)^a$	cAMP assay IC ₅₀ (nM) ^b	I _{max} (%)	cAMP assay EC ₅₀ (nM) ^c	E_{\max} (%)
8a	Ph A F Pl	4.6 ± 0.4			159.0 ± 8.5	41.0 ± 1.4
8b	2-F-Ph	9.2 ± 0.5			51.0 ± 0.7	52.0 ± 0.0
8C 8d	2-CI-Ph 2 CF Ph	13.0 ± 1.8 21.7 ± 0.0				
8e	3-F-Ph	85 ± 13			269 5 + 15 9	515 ± 04
8f	3-Cl-Ph	14.0 ± 0.6			200.0 = 10.0	51.5 = 0.1
8g	3-CF ₃ -Ph	44.7 ± 2.1				
8h	4-F-Ph	10.5 ± 1.1			344.5 ± 11.0	62.0 ± 0.7
8i	4-Cl-Ph	22.0 ± 0.9				
81	4-MeO-Ph	16.0 ± 0.6				
8n 8a	2,4-diF-Ph 2,2 diCl Ph	13.0 ± 1.4				
80 8n	2,3-diCI-Ph 3 4-diE-Ph	20.0 ± 2.9 26.0 ± 1.5				
8a	3.4-diCl-Ph	48.0 ± 2.9				
8t	5-Cl-thienyl	22.7 ± 2.7				
8v	diMe-oxazole	58.0 ± 2.4				
8w	5-Cl-1,3-diMe-pyrazole	8.0 ± 0.2			89.5 ± 8.8	50.0 ± 0.0
9a (racemic)	Ph	2.0 ± 0.1	0.0		24.0 ± 3.5	100.0 ± 0.0
9a (enantiomer 1)	Ph Dh	3.0 ± 0.1	0.0	0.0	0.0	0.0
9a (enantiomer 2)	rn 3 Cl Ph	1.0 ± 0.1 2.0 ± 0.1			30.U ± 0.4	100.0 ± 0.0
91 9i	J-CI-FII 4-CF2-Ph	2.0 ± 0.1 10.0 ± 0.3				
9l	4-MeO-Ph	10.0 ± 0.3 10.0 ± 1.0	7.3 ± 1.0	65.5 ± 1.1		
9p	3,4-diF-Ph	7.0 ± 1.0			66.0 ± 5.0	60.5 ± 0.4
9s	thienyl	9.8 ± 2.6				
9t	5-Cl-thienyl	3.0 ± 0.3			58.5 ± 0.4	93.0 ± 1.4
9x	5-Cl-3-Me-benzothiophene	44.0 ± 3.0				
(S)-10a (S) 10a	Ph 2 Br Dh	27.7 ± 7.6 42.0 ± 7.0				
(S)-10C (S)-10f	2-DI-PII 3-Cl-Ph	43.0 ± 7.0 22.0 ± 2.0				
(S)-10i	4-Cl-Ph	49.0 ± 4.0				
(S)-10j	4-I-Ph	34.0 ± 4.0				
(<i>S</i>)-10k	4-Me-Ph	33.0 ± 5.0				
(<i>S</i>)-101	4-MeO-Ph	40.0 ± 5.0				
(S)-10m	$4-NH_2-Ph$	7.0 ± 1.0	7.6 ± 0.3	79.2 ± 0.1		
(S)-10q (S)-10r	3,4-diMeO-Ph	49.0 ± 3.0 23.0 + 3.0				
(S)-10t	5-Cl-thienvl	25.0 ± 3.0 25.0 ± 3.0				
(S)-10u	5-Br-thienyl	20.0 ± 1.0				
(<i>S</i>)-10x	5-Cl-3-Me-benzothiophene	283.7 ± 4.5				
(<i>R</i>)-11a	Ph	8.0 ± 1.0	0.6 ± 0.1	75.0 ± 0.7		
(<i>R</i>)-11u	5-Br-thienyl	4.0 ± 0.2	014104	070 1 1 4	102.8 ± 18.5	76.5 ± 0.4
(S)-11a (S) 11c	Pn 2 Br Ph	5.0 ± 1.0 7.0 ± 1.0	81.4 ± 0.4 76.5 ± 10.2	87.0 ± 1.4 87.5 ± 1.0		
(S)-11f	2-DI-I II 3-Cl-Ph	4.0 ± 0.3	962 ± 23	77.0 ± 0.0		
(S)-11i	4-Cl-Ph	7.0 ± 0.4	8.2 ± 1.7	86.0 ± 0.0		
(<i>S</i>)-11j	4-I-Ph	3.0 ± 0.2	49.5 ± 13.8	99.5 ± 0.4		
(<i>S</i>)-11k	4-Me-Ph	5.0 ± 1.0	56.3 ± 14.4	78.5 ± 0.4		
(<i>S</i>)-111	4-MeO-Ph	19.3 ± 6.3				
(S)-11m (S) 11a	4-NH ₂ -Ph 3 4 diCl Ph	3.0 ± 0.1	24.5 ± 1.1 7 3 ± 0.6	100.0 ± 0.0 86.5 ± 0.2		
(S)-11r	3 4-diMeO-Ph	12.0 ± 3.0 8.0 ± 0.2	0.8 ± 0.0	100.0 ± 0.0		
(S)-11t	5-Cl-thienyl	4.0 ± 0.1	1.0 ± 0.0	99.0 ± 0.0		
(<i>S</i>)-11u	5-Br-thienyl	3.0 ± 0.1	112.0 ± 12.7	96.0 ± 1.4		
(<i>S</i>)-11x	5-Cl-3-Me-benzothiophene	11.0 ± 2.0	11.0 ± 0.0	87.5 ± 1.1		
12c	2-Br-Ph	1.0 ± 0.1	53.3 ± 12.2	98.5 ± 1.1		
12h 12i	4-F-Ph 4-Cl-Ph	2.0 ± 0.1 1.0 ± 0.2	52.5 ± 5.3 85 5 + 11 7	83.5 ± 1.8 86.5 ± 1.1		
12i	4-U-FII 4-I-Ph	1.0 ± 0.2 1.0 ± 0.2	43.6 ± 5.3	935 ± 0.4		
12m	4-NH ₂ -Ph	1.0 ± 0.2 1.0 ± 0.2	15.5 ± 2.5	89.5 ± 1.8		
12q	3,4-diCl-Ph	3.0 ± 0.4	107.1 ± 18.3	100.0 ± 0.0		
12r	3,4-diMeO-Ph	5.0 ± 1.0	93.7 ± 0.3	100.0 ± 0.0		
13c	2-Br-Ph	14.0 ± 1.0	21.0 ± 5.6	62.0 ± 1.0		
13k 13m	4-Me-Ph	8.0 ± 1.0	123.0 ± 2.1	72.0 ± 1.4		
1300 13a	4-1NT12-1711 3 4-diCl-Ph	1.0 ± 0.2 15.0 ± 0.3	23.0 ± 0.7 61.0 ± 9.2	97.0 ± 2.1 63.0 ± 1.4		
1.54	5, - utor i ii	15.0 ± 0.5	01.0 ± 7.2	0.0 ± 1.4		

Table 1	(continued)
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Compound	Ar-SO ₂	$K_i (nM)^a$	cAMP assay $IC_{50} (nM)^b$	I _{max} (%)	cAMP assay $EC_{50} (nM)^c$	E _{max} (%)
13u	5-Br-thienyl	9.0 ± 1.0	110.0 ± 5.7	69.0 ± 0.7		
13y	4-Me-2-NH2-thiazole	3.0 ± 0.1	110.0 ± 7.1	97.0 ± 1.4		
14a	Ph	252.7 ± 63.8				
14f	3-Cl-Ph	41.7 ± 10.7				
14k	4-Me-Ph	271.3 ± 37.3				
14m	4-NH ₂ -Ph	116.0 ± 5.5				

^a Displacement of [³H]-LSD binding to cloned h5-HT₆ receptors stably expressed in HeLa cells. Mean of three determinations.¹⁶

^b Inhibition of cAMP production in HeLa cells stably transfected with human 5-HT₆ receptors. Mean of three determinations.

^c Agonism of cAMP production in HeLa cells stably transfected with human 5-HT₆ receptors. Mean of three determinations.



Figure 3. Alignment of conformationally constrained N_1 -arylsulfonyltryptamine derivatives 5, 6, 7 (*R* and *S*), 8, 9 (*R* and *S*), 11 and 12 (*R* and *S*), and 14 (Ar = Ph). Compound 14 is shown in green, while others are shown in grey.

pyrrolidin-3-yl)-1*H*-indole array **12–13** functioned as antagonists with modest IC_{50} values (15–100 nM).

In summary, several arrays of conformationally constrained N_1 -arylsulfonyltryptamines were prepared and tested for 5-HT₆ receptor binding and functional assessment. Similar SAR, as has been reported for the N_1 -arylsulfonyl-3-(1,2,3,6-tetrahydropyridin-4-yl)-1*H*-indole derivatives, was observed with various substituted phenyl-, heteroaryl-, and fused arylsulfonyl groups being tolerated.¹⁶

The 3-piperidin-3-yl, 3-pyrrolidin-2-ylmethyl, and 3pyrrolidin-3-yl constrained aminoethyl groups (8–13) appear to be able to adopt a conformation that allows high affinity 5-HT₆ receptor binding, while the β -carboline 14 binds with a significantly weaker (10- to 100-fold) affinity.

 N_1 -Benzenesulfonyl-3-piperidin-3-yl-1H-indole **9a** is a high affinity full agonist with an EC₅₀ of 24 nM. Several of the N_1 -arylsulfonyl-3-(1-methylpyrrolidin-3-ylmethyl)-1H-indole derivatives behave as very potent antagonists ((*S*)-11r, (*S*)-11t; IC₅₀ = 0.8, 1.0 nM).

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References and notes

- Kohen, R.; Metcalf, M. A.; Khan, N.; Druck, T.; Huebner, K.; Lachowicz, J. E.; Meltzer, H. Y.; Sibley, D. R.; Roth, B. L.; Hamblin, M. W. *J. Neurochem.* 1996, 66, 47.
- Sebben, M.; Ansanay, H.; Bockaert, J.; Dumuis, A. Neuroreport 1994, 5, 2553.
- Woolley, M. L.; Marsden, C. A.; Fone, K. C. Curr. Drug Target CNS Neurol. Disord. 2004, 3, 59.
- Roth, B. L.; Craigo, S. C.; Choudhary, M. S.; Uluer, A.; Monsma, F. J., Jr.; Shen, Y.; Meltzer, H. Y.; Sibley, D. R. *J. Pharmacol. Exp. Ther.* **1994**, *268*, 1403.
- Pullagurla, M.; Bondareva, T.; Young, R.; Glennon, R. A. Pharmacol. Biochem. Behav. 2004, 78, 263.
- Minabe, Y.; Shirayama, Y.; Hashimoto, K.; Routledge, C.; Hagan, J. J.; Ashby, C. R., Jr. Synapse 2004, 52, 20.
- Lacroix, L. P.; Dawson, L. A.; Hagan, J. J.; Heidbreder, C. A. Synapse 2004, 51, 158.
- King, M. V.; Sleight, A. J.; Woolley, M. L.; Topham, I. A.; Marsden, C. A.; Fone, K. C. *Neuropharmacology* 2004, 47, 195.
- Woolley, M. L.; Marsden, C. A.; Sleight, A. J.; Fone, K. C. Psychopharmacology (Berl) 2003, 170, 358.
- Lindner, M. D.; Hodges, D. B., Jr.; Hogan, J. B.; Orie, A. F.; Corsa, J. A.; Barten, D. M.; Polson, C.; Robertson, B. J.; Guss, V. L.; Gillman, K. W.; Starrett, J. E., Jr.; Gribkoff, V. K. J. Pharmacol. Exp. Ther. 2003, 307, 682.
- Sleight, A. J.; Boess, F. G.; Bos, M.; Levet-Trafit, B.; Riemer, C.; Bourson, A. Br. J. Pharmacol. 1998, 124, 556.
- Bromidge, S. M.; Brown, A. M.; Clarke, S. E.; Dodgson, K.; Gager, T.; Grassam, H. L.; Jeffrey, P. M.; Joiner, G. F.; King, F. D.; Middlemiss, D. N.; Moss, S. F.; Newman, H.; Riley, G.; Routledge, C.; Wyman, P. J. Med. Chem. 1999, 42, 202.
- Bromidge, S. M.; Clarke, S. E.; Gager, T.; Griffith, K.; Jeffrey, P.; Jennings, A. J.; Joiner, G. F.; King, F. D.; Lovell, P. J.; Moss, S. F.; Newman, H.; Riley, G.; Rogers, D.; Routledge, C.; Serafinowska, H.; Smith, D. R. *Bioorg. Med. Chem. Lett.* 2001, 11, 55.
- Tsai, Y.; Dukat, M.; Slassi, A.; MacLean, N.; Demchyshyn, L.; Savage, J. E.; Roth, B. L.; Hufesein, S.; Lee, M.; Glennon, R. A. *Bioorg. Med. Chem. Lett.* 2000, 10, 2295.
- Russell, M. G.; Baker, R. J.; Barden, L.; Beer, M. S.; Bristow, L.; Broughton, H. B.; Knowles, M.; McAllister, G.; Patel, S.; Castro, J. L. J. Med. Chem. 2001, 44, 3881.
- Cole, D. C.; Ellingboe, J. W.; Lennox, W. J.; Mazandarani, H.; Smith, D. L.; Stock, J. R.; Zhang, G.; Zhou, P.; Schechter, L. E. *Bioorg. Med. Chem. Lett.* 2005, 15, 379.
- 17. Slassi, A.; Edwards, L.; O'Brien, A.; Xin, T.; Tehim, A. WO 2000 063203.
- Chang-Fong, J.; Rangisetty, J. B.; Dukat, M.; Setola, V.; Raffay, T.; Roth, B.; Glennon, R. A. *Bioorg. Med. Chem. Lett.* 2004, 14, 1961.

- Gharagozloo, P.; Miyauchi, M.; Birdsall, B.; Birdsall, N. J. J. Org. Chem. 1998, 63, 1974.
- Olofson, R. A.; Martz, J. T.; Senet, J. P.; Piteau, M.; Malfroot, T. J. Org. Chem. 1984, 49, 2081.
- 21. Cole, D. C.; Pagano, N.; Kelly, M. F.; Ellingboe, J. *J. Comb. Chem.* **2004**, *6*, 78.
- 22. Gharagozloo, P.; Miyauchi, M.; Birdsall, N. J. Tetrahedron 1996, 52, 10185.
- 23. Macor, J. E.; Blank, D. H.; Fox, C. B.; Lebel, L. A.; Newman, M. E.; Post, R. J.; Ryan, K.; Schmidt, A. W.;

Schulz, D. W.; Koe, B. K. J. Med. Chem. 1994, 37, 2509.

- 24. Chiral HPLC separation. Column: Chiralcel-OJ 20×250 mm; mobile phase—87% heptane (containing 0.1% TFA) and 13% IPA; sample dissolved in EtOH 10 mg/mL; flow rate—8 mL/min. Enantiomer 1: retention time 23.4 min, enantiomer 2: retention time 27.4 min.
- 25. Pullagurla, M. R.; Dukat, M.; Setola, V.; Roth, B.; Glennon, R. A. Bioorg. Med. Chem. Lett. 2003, 13, 3355.