Tetrahedron: Asymmetry 25 (2014) 1424-1429

Contents lists available at ScienceDirect

Tetrahedron: Asymmetry

journal homepage: www.elsevier.com/locate/tetasy

Synthesis of 2,3-O-benzyl-ribose and xylose and their equilibration

Reignier Jeffrey^a, Gurdial Singh^{a,*}, Patrice G. J. Plaza-Alexander^a, Nadia Singh^a, Jonathan M. Goodman^b, Alessia Bacchi^c, Francesco Punzo^d

^a Department of Chemistry, University of the West Indies, St. Augustine, Trinidad and Tobago

^b Department of Chemistry, University of Cambridge, Lensfield Road, Cambridge CB2 1EW, UK

^c Dipartimento di Chimica, Università degli Studi di Parma, Parco Area delle Scienze 17/A, Parma 43124, Italy

^d Dipartimento di Scienze del Farmaco, Sezione Chimica, Università di Catania, Viale Andrea Doria, 6, 95125 Catania, Italy

ARTICLE INFO

Article history: Received 5 September 2014 Accepted 12 September 2014

ABSTRACT

The preparation and NMR analysis of 2,3-di-O-benzyl D-ribose and 2,3-di-O-D-xylose are described. In DMSO- d_6 the sugars adopt a conformation in which the hydroxyl groups are in an equatorial position. In CDCl₃ and CD₂Cl₂ the sugars adopts a conformation in which intramolecular hydrogen bonding plays an important role in determining the equilibrium composition. These findings were also confirmed by DFT studies and, in the solid phase, by X-ray single crystal diffraction.

© 2014 Elsevier Ltd. All rights reserved.

1. Introduction

Carbohydrates are essential biochemical building blocks for complex life forms to develop. Major strides have been made in the synthesis of complex structures encompassing carbohydrates over the last decade. However a universal strategy for glycosylation that readily results in the synthesis of oligosaccharides has remained elusive.¹ Understanding the biological activity of saccharides and of their oligomers by its very nature necessitates that we know the structure and stability of the different conformations that these molecules can adopt. A major obstacle in obtaining a basic structure is due to that fact that these can form strong hydrogen bonds in water and other polar solvents, which in turn can result in conformational changes taking place that may lead to a different biological effect being observed. As a result of these observations, a number of investigations into the conformations of 2-deoxy-p-ribose have been reported² using Fourier-transform microwave spectroscopy in conjunction with UV spectroscopy using a laser source. In aqueous solution *D*-ribose exists primarily as α - and β -pyranose³ forms whilst in RNA it is present as a furanose. In aqueous solution D-ribose exists as a mixture of α - and β -pyranoses that are present in their low energy chair conformations, ${}^{4}C_{1}$ and ${}^{1}C_{4}$. These configurations proceed through ring inversion leading to an axial or equatorial OH group for each conformer. Additionally, extra stabilisation provided by the anomeric effect results in the preference for an axial OH group at the C₁ carbon over the empirically expected equatorial orientation⁴ (Scheme 1).



Scheme 1.

We have recently reported NMR equilibration studies leading to the synthesis of arabinofuranosides using an iterative protocol.⁵ Herein we report on our findings relating to 2,3-*O*-dibenzylriboand xylo-furanosides and the former use for the synthesis of oligosaccharides.

Following on from these studies, we reasoned that ribosides and their xylo analogues should be capable of providing access to oligosaccharides using similar procedures. In addition these studies should provide insight into the conformation of these pentoses and the stereochemical effects as a result of the change in the substitution patterns at C-2 and C-3. Furthermore, we wished to control the mutarotation of these partially protected sugars with the specific goal of being able to selectively prepare furanosides from a mixture of isomers. The success of the proposed strategy depends upon the selectivity of the attack of the primary hydroxyl group on the glycosyl intermediate species that results.⁵ At the outset we wished to study the equilibrium composition of these two sugars and compare the experimental findings with theoretical predictions arising from the use of the density functional theory with respect to the possible conformations arising in solution and the effect the solvents have on these.





Tetrahedron

^{*} Corresponding author. E-mail address: Gurdial.Singh@sta.uwi.edu (G. Singh).



Scheme 2. Preparation of 2,3-di-O-benzyl ribose 5a and 2,3-di-O-benzyl xylose 5b.

2. Results and discussion

2,3-Di-O-benzylated D-ribose was synthesized from the commercially available D-ribose (Scheme 2). The methyl furanoside⁶ was protected selectively at the primary hydroxyl group using trityl chloride⁷ followed by benzylation to give the fully protected sugars. Removal of the trityl group and the anomeric methyl group was accomplished in one step to yield the desired compounds. 2,3-Di-O-benzyl-D-xylose was also synthesized from commercially available D-xylose using the same procedures.

The pure 2,3-di-O-benzyl derivatives **5a** and **5b** obtained after column chromatography were analysed using one dimensional and two-dimensional NMR in three solvents: $CDCl_3$, CD_2Cl_2 and DMSO- d_6 . The equilibrium proportions of the various isomers in solution were determined by integration of the anomeric proton NMR signals.

The ¹H NMR spectrum of **5a** in deuterated chloroform showed the presence of four major anomers; the α - and β -pyranoses and the α - and β -furanoses. Neither the acyclic aldehyde nor hydrate was detected by natural abundance of NMR. The anomeric protons of the furanoses resonated at lower fields than those of the pyranoses, with chemical shifts of 5.30 and 5.31 ppm for the α - and β -anomers, respectively, (Table 1).

Table 1

Che	mical	shift	data	for	compound	s 5a	and	5b	in	CDC	l_3^a
-----	-------	-------	------	-----	----------	-------------	-----	----	----	-----	---------

		5a		5b	
α-Furanose	H-1, C-1	5.30	96.3	5.47	95.5
OH ON	H-2, C-2 H-3, C-3	3.61 3.69	76.9 62.4	4.00 4.18	82.4 82.9
] >""OH	H-4, C-4	3.82	80.6	4.35	81.2
BnO OBn	H-5a, 5b, C-5	3.83, 3.86	62.2	3.73,3.89	62.2
β-Furanose	H-1, C-1	5.31	99.8	5.30	101.2
он	H-2, C-2	4.22	82.0	4.02	87.4
	H-3, C-3	4.25	82.0	4.16	82.0
	H-4, C-4	3.69	72.8	4.29	77.7
BnO OBn	H-5a, 5b, C-5*	3.92, 3.88	64.6	3.72, 3.83	61.9
α-Pyranose	H-1, C-1	5.21	93.5	5.01	91.8
HO _{////} O	H-2, C-2 H-3, C-3	3.75 3.88	66.0 77.8	3.52 3.83	77.2 76.8
Provent	H-4, C-4	3.77	65.9	3.63	67.6
OBn OBn	H-5a, 5b, C-5*	3.89, 3.93	67.6	3.83, 3.85	64.9
β-Pyranose	H-1, C-1	4.91	92.4	4.96	94.1
HO	H-2, C-2	3.96	77.3	3.48	76.0
	H-3, C-3	3.87	78.0	3.72	76.6
BnorthoH	H-4, C-4	3.54	61.7	3.69	67.0
 OBn	H-5a, 5b, C-5	4.18, 4.20	76.8	3.83, 3.85	61.7

^a All values are given as chemical shifts in ppm.

H-5a and H-5b unassigned.

The ratios of the four anomers were obtained by integration of the anomeric resonance in its 1 H spectrum (Table 2).

Table 2

Percent pyranose and furanose forms of **5a** in CDCl₃, DMSO-d₆ and CD₂Cl₂*

	CDCl ₃	DMSO-d ₆	CD_2Cl_2
α-Furanose	18	_	15
β-Furanose	13	-	17
α-Pyranose	24	-	21
β-Pyranose	45	100	47

* Ratios calculated using spectra obtained immediately after dissolution in the respective solvents.

Benzylation of the 2,3-hydroxy groups of ribose results in a thermodynamic equilibrium which results in the β -pyranose being the sole isomer that is observed in DMSO and the solid state, in contrast to the unprotected sugar which gives a mixture of all four isomers in water (Table 3).

Table 3

% pyranose and furanose forms of D-ribose 1a and D-xylose 1b in D_2O^{3b}

	1a	1b
α-Furanose	6.5	<1
β-Furanose	13.5	<1
α-Pyranose	21.5	36.5
β-Pyranose	58.5	63.0

For **5a** in CDCl₃ and CD₂Cl₂, the composition of the four isomers were similar with the two pyranose forms occurring in 69% and 68%, respectively, indicative of similar solvent effects. In the more polar DMSO- d_6 only the β -pyranose isomer was detected upon immediate dissolution and subsequent acquisition of the NMR data. The resonances observed, excluding the benzyl signals are detailed in Table 4 for the sake of clarity. This indicated that in solid **5a**, the β -pyranose isomer was the sole observed component upon dissolution and immediate accumulation of the spectroscopic data. The anomeric resonance occurred at δ 4.88 and in its ¹³C spectrum the resonance was observed at δ 93.34.

 Table 4

 Chemical shift data for 5a in DMSO-d₆

	β -Pyranose in DMSO- d_6	
OH-1	6.62 <i>J</i> = 6.0	0 Hz
H-1, C-1	4.88	93.34
H-2, C-2	3.32	79.02
H-3, C-3	3.93	76.5
OH-4, H-4, C-4	4.62, 3.70	67.4
H-5, H-5′, C-5	3.53, 3.60	63.7

We were able to crystallise the pure β -anomer of **5a** from ethyl acetate by allowing the slow evaporation of the solvent to obtain thin white needles with a mp of 82–83 °C, $[\alpha]_D = -19.6$ (c 1.0, CHCl₃). With this isomer in hand we undertook single X-ray crystallographic analysis. This confirmed that crystalline 5a existed in a pyranose structure, which was in agreement with the solution structure observed in DMSO. The crystal structure belongs to the monoclinic P21 space group with a = 10.558(3), b = 6.878(2), c = 12.275(4) and $b = 97.543(4)^{\circ}$ with Z = 2 thus showing two independent molecules in the unit cell. The ring puckering analysis performed on the crystalline structure⁸ evidenced the presence of a chair conformation as reported in Table 5. More interestingly, the structure presents no solvent accessible voids, being available only two small volumes of less than 15 Å³ which are not suitable for any small solvent molecule to fit, and a resulting high packing index⁹ (64.2%). The solid state hydrogen bond network is reported in Table 6. There are both inter- and intra-molecular hydrogen bonds as well as weak π stacking interactions between the aromatic rings of the structure, particularly along the 1 - x, -1/2 + y, 1 - z direction (Fig. 1).

Prior to the commencement of these studies we had undertaken density functional theory (DFT) calculations of 2,3-di-O-benzyl and

Table 5

Ring puckering coordinates and their e.s.d. for the Cremer and Pople analysis performed on the anomeric ring and its resulting conformation

Q (Å)	q_2 (Å)	ϕ_2 (°)	q_{3} (Å)	θ (°)	Conformation
0.570 (5)	0.016 (5)	353 (20)	-0.570 (5)	177.9 (5)	Chair

2,3-di-O-methyl ribosides along with the xylose analogues, using B3LYP/6-31G^{**} and Schrodinger's Jaguar software, version 7.6, on geometries generated by conformation searches using MacroModel (BatchMin v9.7) and the OPLS_2005 force field. These calculations indicated that the major isomer in DMSO, (Table 7), would have a β -pyranose structure. This is in agreement with our experimental findings in both the solution and solid states. When dissolved in CD₂Cl₂ and CDCl₃ the crystalline β -ribopyranoside afforded a mixture of furanosides and pyranosides.

In DMSO- d_6 the anomer with the least steric interactions was the most abundant.¹⁰ The hydrogen bond interaction between the solvent and the hydroxyl groups increases the effective size of the substituents. In CDCl₃, which does not interact with the solute via hydrogen bonding, other effects come into play including the anomeric effect and ring strain or torsional strain. Intramolecular hydrogen bonding also influences the equilibrium because conformations containing these interactions are preferentially stabilized.¹¹

In order to account for possible stereoelectronic effects we undertook DFT calculations with *D*-arabinose. These suggested that in a polar solvent the pyranose structure should predominate. This was at variance with our experimental findings⁵ wherein it was established that the furanose and pyranose ring systems occurred in equal percentages. These findings suggest that the stereochemistry of the C-2 hydroxy group plays a predominant role in determining the ring size of pentoses. This is not too surprising since the C-2 hydroxy function in both ribose and xylose is in the preferred equatorial orientation whilst in the case of *D*-arabinose it has a higher energy axial geometry.

Table 6

Potential hydrogen bonds analysis. Bond lengths, together with their standard uncertainties (s.u.) are given in Å; bond angles (D-H···A) in (°)

Bond type	Donor	-H	Acceptor	D-H	$H{\cdots}A$	$D{\cdots}A$	$D{-}H{\cdots}A$
Inter- ⁱ	03	-H30	02	0.79 (7)	1.96 (7)	2.738 (5)	166 (7)
Inter- ⁱⁱ	C19	-H19	03	0.93	2.59	3.503 (7)	166
Intra-	02	-H20	05	0.83 (7)	2.00 (8)	2.696 (6)	141 (7)
Intra-	C19	-H19	03	0.93	2.59	3.503 (7)	166

Symmetry codes: (i) *x*, -1 + *y*, *z*; (ii) *x*, 1 + *y*, *z*.



Figure 1. Packing diagram viewed down the *b* axis. Hydrogens are shown as fixed-size spheres of an arbitrary radius of 0.15 Å and the ellipsoids probability level set to 30% for the sake of clarity.

	2,3,Di-O-Bn ribose			2,3-Di-O-Me ribose		2,3-Di-O-Bn xylose		
	Model %	Exp DMSO %	Exp CDCl ₃ %	ab initio	B3LYP	Model DMSO	Exp DMSO	Exp CDCl ₃
α-Furanose	0.0	0.0	18.0	0.0	0.6	0.0	0.0	6.9
β-Furanose	0.0	00	13.0	0.0	0.0	0.0	0.0	5.6
α-Pyranose	4.8	0.0	24.0	2.0	44.2	13.0	44.4	31.2
β-Pyranose	95.2	100.0	45.0	98.0	55.2	87.0	55.6	56.3
	Arabinose			2,3-Di-O-Me arabinose		2,3-Di-O-Bn arabinose		
	Model dry	Model H ₂ O	Literature ^{3b}	Model DMS	0	Model DMSO	Exp DMSO	Exp CDCl ₃
α-Furanose	0.0	0.0	2.5	6.7		8.7	35.0	26.0
β-Furanose	0.0	0.0	2.0	48.1		11.9	16.0	21.0
α-Pyranose	2.0	10.0	60.0	0.0		0.0	24.0	26.0
β-Pyranose	98.0	90.0	35.5	45.2		79.4	25.0	27.0

Differences in the chemical shifts and the coupling constants of the anomeric protons when the solvent was changed from CDCl₃ to DMSO- d_6 were related to a change in conformation, particularly in the pyranose forms. Both 2,3-di-O-benzyl-β-D-ribopyranose **5a** and 2,3-di-O-benzyl-β-D-xylopyranose 5b experienced conformational changes due to the change in solvent as indicated by the values of the $I_{1,2}$ coupling constants (Table 8).

In CDCl₃, 2,3-di-O-benzyl-β-D-ribopyranose 5a exists primarily as the ${}^{1}C_{4}$ conformer as indicated by the $J_{1,2}$ value of 1.99 Hz (Table 8). In DMSO- d_6 , however, a $J_{1,2}$ value of 5.6 Hz suggests that this anomer exists as a mixture of the ${}^{1}C_4$ and ${}^{4}C_1$ conformers, with the equilibrium favouring the latter.^{3e} Similarly, 2,3-di-O-benzyl- β -D-xylopyranose **5b** exists primarily in the ${}^{4}C_{1}$ conformation, in which all substituents are in an equatorial position, in DMSO- d_6 but as a mixture of the two chair conformations in the less polar, non-hydrogen bonding CDCl₃ with the equilibrium in favour of the ${}^{1}C_{4}$ conformer. In 2,3-di-O-benzyl- α ,D-xylopyranose **5b** the conformation was assigned based on the well-known fact that an equatorial proton resonates at a lower field than a chemically similar but axial proton. Thus 2,3-di-O-benzyl-α,D-xylopyranose **5b**, exists primarily as the ${}^{1}C_{4}$ conformer in CDCl₃.¹² while 2,3-di-Obenzyl-α, p-ribopyranose **5a** exists as a mixture of both conformers in either solvent. These observations suggest that intramolecular hydrogen bonding is present in the absence of a hydrogen bonding solvent.¹³ This intramolecular hydrogen bonding occurs even in the presence of bulky benzyl substituents and stabilizes the conformer in which these groups are axial, in spite of unfavourable 1,3-diaxial interactions¹⁴ (Fig. 2).

Having this information in hand, we will begin to investigate the application of these findings to the synthesis of furano-oligosaccharides. In particular we wish to undertake an iterative synthesis of 1-O-linked ribosides. We reason that we should be able to selectively react the equilibrium mixture in order to obtain the desired furano linked ribosides.

Table 8

Comparison of chemical shifts in ppm and coupling constants (in Hz) of anomeric protons of compounds 5a and 5b

	5a			5b			
	CDCl ₃	CD_2Cl_2	DMSO-d ₆	CDCl ₃	CD_2Cl_2	DMSO-d ₆	
α-Furanose	5.30 (6.1)	5.27 (4.0)		5.47 (4.2)	5.44 (3.85)		
β-Furanose	5.31 (3.8)	5.35 (4.7)		5.30	5.23		
α-Pyranose	4.91 (10.0)	4.88 (10.0)		5.01 (2.2)	5.01 (2.28)	5.20 (5.4)	
β-Pyranose	5.21 (1.99)	5.23	4.88 (5.6)	4.96 (2.6)	4.92 (3.5)	4.47 (6.6)	





2,3-di-O-benzyl-α-D-ribopyranose



Figure 2. Major chair conformations of 5a and 5b.

3. Conclusion

In conclusion, we have investigated the equilibria present in 2,3-di-O-benzylated pentose sugars and established that in polar solvents, the β -pyranose form is the major isomer, whilst in less polar solvents a mixture of both furanose and pyranose isomers can be found.

4. Experimental

4.1. General

All chemicals used were reagent or HPLC grade and used as supplied without prior purification unless otherwise stated. Solvents: DCM and diethyl ether were dried over calcium hydride for 24 h, distilled and stored over 4 Å molecular sieves. Flash column chromatography was carried out using forced flow of the indicated solvent on Merck (230–400 mesh) silica gel. TLC was performed using pre-coated silica gel 60 F_{254} plates; compounds were visualized using acidic ammonium molybdate solution [ammonium molybdate(VI) tetrahydrate (25 g) in 1 M H_2SO_4 (500 mL)]. Anhydrous reactions were performed under argon in oven dried apparatus; anhydrous transfers were carried out with standard syringe techniques.

¹H, ¹³C and ³¹P NMR spectra were recorded on Bruker 600, 400 MHz spectrometers in the deuterated solvent stated. For ease of interpretation, protons are expressed as multiples of 1. Refer to the main text for equilibrium ratios. IR spectra were recorded on a Perkin Elmer Spectrum RXI FT-IR spectrometer. High resolution mass data was obtained using a Bruker Daltonics micrOTof-Q instrument in the electron spray ionization mode. Melting points were measured in open capillaries and are uncorrected.

4.2. Crystallographic structure determination

The single crystals used for structure determination were mounted on a glass capillary; the small dimensions and some twinning did not allow collection of high quality data and resolution, but were nevertheless perfectly suitable for satisfactory structural determination. Single crystal X-ray diffraction data were collected at *T* = 293 K using the MoK α radiation (λ = 0.71073 Å) on a SMART APEX2 diffractometer. Lorentz, polarization and absorption corrections were applied.¹⁵ The structure was solved by direct methods using SIR2004¹⁶ and refined by full-matrix least-squares on all F2 using SHELXL-2013¹⁷ implemented in the Olex2 package.¹⁸ Hydrogen atoms were introduced in calculated positions riding on their carrier atoms with the exception of hydroxylic hydrogens that were located on the difference map and refined isotropically. Anisotropic displacement parameters were refined for all non-hydrogen atoms. Due to slight twinning the Flack parameter does not converge to zero, although the absolute structure is assessed by chemical argumentation. Hydrogen bonds have been analysed with SHELXL2013¹⁷ and PARST97¹⁹ and extensive use was made of the Cambridge Crystallographic Data Centre packages²⁰ for the analysis of crystal packing. CCDC 1022046 contains the Supplementary crystallographic data for this paper. Copies of the data can be obtained free of charge on application to CCDC, 12 Union Road, Cambridge CB2 1EZ, UK (Fax: (+44) 1223 336 033; e-mail: deposit@ccdc.cam.ac.uk).

4.3. General procedure for preparation of 2,3-di-O-benzyl sugars

The starting sugar (5.0 g, 33.3 mmol) was suspended in methanol,⁶ after which *p*-TSA (290 mg, 1.5 mmol) was added and the

mixture was stirred at 40 °C for 20 h. The solvent was removed in vacuo and the crude product dissolved in pyridine (50 mL). Trityl chloride⁷ (9.3 g. 33.3 mmol) was added and the mixture was stirred at rt overnight. The reaction mixture was then poured into an ice-water mixture and allowed to warm up to room temperature. The mixture was then extracted with chloroform $(3 \times 50 \text{ mL})$. The organic layers were then combined and washed using saturated aqueous ammonium chloride $(2 \times 50 \text{ mL})$ and water $(2 \times 50 \text{ mL})$. The organic layer was then dried (Na_2SO_4) and the solvents removed in vacuo. The solvent was removed under reduced pressure and the residue obtained purified via column chromatography (CHCl₃ followed by CHCl₃/MeOH; 9:1). The tritylated methyl glycoside was then benzylated under standard benzylation conditions. Deprotection of the trityl and methyl groups was carried out by refluxing in a solution of CH₃CN/H₂O/ TFA (4:3:1: 1:20 w/v) at 95 °C for 12 h. The solvent was then removed in vacuo and the residue neutralized using saturated aqueous NaHCO₃ (20 mL) and solid NaHCO₃. The mixture was then extracted using EtOAc (2×20 mL). The organic layers were combined and dried (Na₂SO₄) and the solvent removed under reduced pressure. The residue obtained was purified via column chromatography using CHCl₃/MeOH (95:5).

4.4. 2,3-Di-O-benzyl-D-ribose 5a

The resultant light yellow oil solidified on standing, 75% yield overall. A portion of this was recrystallized from EtOAc mp of 82–83 °C, $[\alpha]_D^{23} = -19.6$ (c 1.0, CHCl₃). NMR (600 MHz) CDCl₃ δ_H (ppm): shared resonances: 7.2-7.4 (40H, m, aromatic protons), 4.5-4.8 (4H, benzylic protons + 1 hydroxyl proton), 4.25 (1H, 1 hydroxyl proton), 3.5-4.0 (3H, 3 hydroxyl protons) 1.55-2.85 (3H, 3 hydroxyl protons); α -furanose: 5.30 (1H, d, J = 6.1 Hz, H-1), 3.61 (1H, H-2), 3.69 (1H, H-3), 3.82 (1H, H-4), 3.83 (1H, H-5_a), 3.86 (1H, H-5_b); β -furanose: 5.31 (1H, d, J = 3.8 Hz, H-1), 4.22 (1H, H-2), 4.25 (1H, H-3), 3.69 (1H, H-4), 3.92 (1H, H-5_b), 3.88 (1H, H-5_a); β-pyranose: 4.91 (1H, d, *J* = 1.99 Hz, H-1), 3.96 (1H, H-2), 3.87 (1H, H-3), 3.54 (1H, H-4), 4.18 (1H, H-5_a), 4.20 (1H, H-5_b); α-pyranose: 5.21 (1H, d, *J* = 10 Hz, H-1), 3.75 (1H, H-2), 3.88 (1H, H-3), 3.77 (1H, H-4), 3.89 (1H, H-5_b), 3.93 (1H, H-5_a); ¹³C (150 MHz) CDCl₃ δ_C (ppm): shared resonances: 138.1, 137.7, 137.7, 137.6, 137.4, 137.3, 137.3, 128.7, 128.7, 128.7, 128.7, 128.7128.6, 128.5, 128.5, 128.4, 128.4, 128.4, 128.3, 128.3, 128.2, 128.1, 128.0, 128.0, 128.0, 128.0, 127.9, 127.9, 127.9, 127.9, 127.7, 127.7, 127.7, 127.7 (aromatic-C), 76.1, 73.8, 73.6, 73.0, 73.0, 72.8, 72.3, 70.6 (benzylic-C); α-furanose: 96.3 (C-1), 76.9 (C-2), 62.4 (C-3), 80.6 (C-4), 62.2 (C-5); β-furanose: 99.8 (C-1), 82.0 (C-2), 82.0 (C-3),72.8 (C-4), 64.6 (C-5); α-pyranose: 92.4 (C-1), 77.3 (C-2), 78.0 (C-3), 61.7 (C-4), 76.8 (C-5); β-pyranose: 93.5 (C-1), 66.0 (C-2), 77.8 (C-3), 65.9 (C-4), 67.6 (C-5).

¹H NMR (600 MHz) CD₂Cl₂ δ_H (ppm): shared resonances: 7.2– 7.4 (40H, m, aromatic protons), 4.4-4.8 (4H, benzylic protons + 1 hydroxyl proton), 3.5-4.0 (3H, 3 hydroxyl protons) 3.09 (1H, br s; hydroxyl proton), 1.24–2.73 (3H; hydroxyl protons); α-furanose: 5.29 (1H, d, J = 4.0 Hz, H-1), 3.72 (1H, H-2), 3.90 (1H, H-3), 3.60 (1H, H-4), 3.77 (1H, H-5_a), 3.73 (1H, H-5_b); β -furanose: 5.32 (1H, d, J = 4.7 Hz, H-1), 3.92 (1H, H-2), 4.21 (1H, H-3), 3.63 (1H, H-4), 4.83 (1H, H-5_b), 4.84 (1H, H-5_a); β-pyranose: 5.35 (1H, H-1), 3.98 (1H, H-2), 4.21 (1H, H-3), 3.88 (1H, H-4), 4.19 (1H, H-5_a), 4.22 (1H, H-5_b); α -pyranose: 4.88 (1H, d, J = 10 Hz, H-1), 3.64 (1H, H-2), 3.52 (1H, H-3), 3.77 (1H, H-4), 3.44 (1H, H-5_b), 3.46 (1H, H-5_a); ¹³C (150 MHz) CD₂Cl₂ δ_C (ppm): shared resonances: 138.8, 138.4, 138.3, 138.2, 138.1, 138.1, 129.0, 128.9, 128.8, 128.8, 128.8, 128.5, 128.4, 128.4, 128.4, 128.3, 128.3, 128.3, 128.3, 128.2, 128.2, 128.1, 128.0 (aromatic-C), 73.5, 73.4, 73.1, 73.0, 72.8, 71.0 (benzylic-C, overlapping resonances); α-furanose: 97.8

(C-1), 77.9 (C-2), 67.8 (C-3), 62.2 (C-4), 65.2 (C-5); β-furanose: 100.3 (C-1), 78.6 (C-2), 77.4 (C-3),78.6 (C-4), 74.2 (C-5); α-pyranose: 92.8 (C-1), 76.9 (C-2), 63.0 (C-3), 66.5 (C-4), 67.8 (C-5); β-pyranose: 94.0 (C-1), 78.0 (C-2), 82.4 (C-3), 81.6 (C-4), 85.6 (C-5).

¹H NMR (600 MHz) DMSO-*d*₆ δ_H (ppm): 7.2–7.4 (10H, m, aromatic protons), 4.71–4.77 (4H, benzylic protons) 4.62 (1H, br s, hydroxyl proton, OH-4); β-pyranose: 6.62 (1H, d, *J* = 6.0 Hz; OH-1), 4.88 (1H, overlapping dd, *J* = 6.8 Hz, H-1), 3.32 (1H, H-2), 3.93 (1H, H-3), 3.70 (1H, H-4), 3.60 (1H, dd, *J* = 10.8 Hz, 3.6 Hz, H-5_a), 3.53 (1H, dd, *J* = 7.8 Hz, 3.6 Hz, H-5_b); ¹³C (150 MHz) DMSO-*d*₆ δ_C (ppm): 139.3, 138.8, 128.1, 128.0, 127.3, 127.3, 127.2, 127.0 (aromatic-C), 72.3, 71.7 (benzylic-C); β pyranose: 93.3 (C-1), 79.0 (C-2), 76.5 (C-3), 67.4 (C-4), 63.7 (C-5); *m*/*z* found (M⁺) 330.1453; C₁₉H₂₂O₅ requires 330.1467.

4.5. 2,3-Di-O-benzyl-D-xylose 5b

Colourless oil which solidified after storage under vacuum to give an off-white solid. 69% yield overall; mp 80-82 °C, $[\alpha]_{D}^{26} = -4.4$ (c 1.1 CHCl₃); $v_{max}(film)/cm^{-1}$ 3406, 2928, 1721, 1453, 1357, 1274, 1215 and 1094. ¹H NMR (600 MHz) CDCl₃ δ_H (ppm): shared resonances: 7.2-7.4 40H/m (aromatic protons), 4.4–4.7 (16H; benzylic protons), 3.40–4.20 (4H; hydroxyl protons); 2.55–3.30 (4H; hydroxyl protons); α -furanose: 5.47 (1H, d, J = 4.2 Hz, H-1), 4.00 (1H, H-2), 4.18 (1H, H-3), 4.35 (1H, H-4), 3.73 (H-5_a) 3.89 (H-5_b); β -furanose: 5.30 (1H, s, H-1), 4.02 (1H, H-2), 4.16 (1H, H-3), 4.29 (1H, H-4), 3.72 (1H, H-5_a), 3.83 (1H, H-5_b); α-pyranose: 5.01 (1H, d, J = 2.2 Hz, H-1), 3.52 (1H, H-2), 3.83 (1H, H-3), 3.63 (1H, H-4), 3.83 (1H, H-5_a), 3.85 (1H, H-5_b); β-pyranose: 4.96 (1H, d, J = 2.6 Hz, H-1), 3.48 (1H, H-2), 3.72 (1H, H-3), 3.69 (1H, H-4), 3.55 (1H, H-5_a), 4.13 (1H, H-5_b) $^{13}\mathrm{C}$ (150 MHz) CDCl₃ δ_{C} (ppm): shared resonances: 137.9, 137.3, 137.2, 137.1, 137.0, 136.9, 128.7, 128.7, 128.7, 128.6, 128.6, 128.6, 128.6, 128.4, 128.4, 128.4, 128.3, 128.3, 128.2, 128.1, 128.1, 128.1, 128.00, 128.0, 127.9, 127.9, 127.8, 127.7 (aromatic-C) 73.7, 73.7.6, 73.3, 73.1, 73.1, 72.8, 72.5, 72.0 (benzylic-C); α-furanose: 95.5 (C-1), 82.4 (C-2), 82.9 (C-3), 81.2 (C-4), 62.2 (C-5); β-furanose: 101.2 (C-1), 87.4 (C-2), 82.0 (C-3), 77.7 (C-4), 61.9 (C-5); α-pyranose: 91.8 (C-1), 77.2 (C-2), 76.8 (C-3), 67.6 (C-4), 64.9 (C-5); β-pyranose: 94.1 (C-1), 76.0 (C-2), 76.6 (C-3), 67.0 (C-4), 61.7 (C-5);

¹H NMR (600 MHz) CD₂Cl₂ δ_H (ppm): shared resonances: 7.2– 7.4 40H/m (aromatic protons), 4.4-4.7 (16H; benzylic protons), 3.40-4.20 (4H; hydroxyl protons); 2.55-3.30 (4H; hydroxyl protons); α -furanose: 5.44 (1H, d, J = 3.85 Hz, H-1), 4.02 (1H, H-2), 4.18 (1H, H-3), 4.33 (1H, H-4), 3.74 (H-5_a) 3.90 (H-5_b); β-furanose: 5.23 (1H, s), 4.01 (1H, H-2), 4.12 (1H, H-3), 4.24 (1H, H-4), 3.69 (1H, H-5_a), 3.78 (1H, H-5_b); α -pyranose: 5.01 (1H, d, J = 2.28 Hz, H-1), 3.52 (1H, H-2), 3.83 (1H, H-3), 3.63 (1H, H-4), 3.83 (1H, H-5_a), 3.85 (1H, H-5_b); β-pyranose: 4.92 (1H, d, J = 3.5 Hz, H-1), 3.47 (1H, H-2), 3.71 (1H, H-3), 3.69 (1H, H-4), 3.49 (1H, H-5_a), 4.10 (1H, H-5_b) ¹³C (150 MHz) CDCl₃ δ_C (ppm): shared resonances: 138.7, 137.9, 137.9, 137.7, 129.0, 129.0, 129.0, 129.0, 129.0, 128.6, 128.6, 128.6, 128.5, 128.4, 128.4, 128.4, 128.3, 128.3, 128.3, 128.2, 128.1 (aromatic-C) 74. 73.9, 73.5, 73.5, 73.4, 73.2, 72.8, 72.3 (benzylic-C); α-furanose: 96.0 (C-1), 82.4 (C-2), 82.8 (C-3), 81.7 (C-4), 62.6 (C-5); β-furanose: 101.7 (C-1), 88.0 (C-2), 82.8 (C-3), 78.2 (C-4), 62.2 (C-5); α-pyranose: 92.1 (C-1), 78.2 (C-2), 77.3 (C-3), 68.1 (C-4), 65.2 (C-5); β-pyranose: 94.7 (C-1), 76.9 (C-2), 77.3 (C-3), 67.5 (C-4), 62.2 (C-5);

¹H NMR (600 MHz) DMSO-*d*₆ δ_H (ppm): shared resonances: 7.2– 7.4 (20H, m, aromatic protons), 4.6–4.9 (8H, benzylic protons + 2H; hydroxyl protons); α-pyranose: 6.89 (1H, d, *J* = 6.5 Hz; OH-1), 5.14 (1H, m, H1) 3.31 (1H, H-2), 3.57 (1H, H-3), 3.45 (1H, H-4), 3.38 (1H, H-5_a), 3.53 (1H, H-5_b); β-pyranose: 5.20 (1H, d, *J* = 5.4 Hz; OH-1), 4.47 (1H, overlapping dd, *J* = 6.6 Hz, H-1), 3.09 (1H, H-2), 3.30 (1H, H-3), 3.51 (1H, H-4), 3.09 (1H, H-5_a), 3.66 (1H, H-5_b); ¹³C (150 MHz) DMSO-*d*₆ δ_C (ppm): shared resonances: 139.4, 139.1, 138.9, 138.7, 128.2, 128.1, 128.1, 128.0, 127.9, 127.9, 127.9, 127.5, 127.4, 127.3, 127.3, 127.2, 127.1, 127.0, 126.7 (aromatic-C), 74.0 (2C), 73.7, 71.5 (benzylic-C); α-pyranose: 89.9 (C-1), 79.6 (C-2), 81.2 (C-3), 70.0 (C-4), 61.1 (C-5); β-pyranose: 97.5 (C-1), 82.5 (C-2), 84.3 (C-3), 69.9 (C-4), 65.4 (C-5). *m*/*z* found (M⁺) 330.1530; C₁₉H₂₂O₅ requires 330.1467.

Acknowledgments

We thank the University of the West Indies for scholarships (PGJP-A, R.J.).

References

- (a) Davis, B. G. J. Chem. Soc., Perkin Trans. 1 2000, 2137–2160; (b) Boons, G. J. Tetrahedron 1996, 52, 1095–1121; (c) Demchenko, A. V. Lett. Org. Chem. 2005, 2, 580–589.
- (a) Peña, I.; Cocinero, E. J.; Cabezas, C.; Lesarri, A.; Mata, S.; Écija, P.; Daly, A. M.; Cimas, Á.; Bermffldez, C.; Basterretxea, F. J.; Blanco, S.; Fernández, J. A.; López, J. C.; Castaño, F.; Alonso, J. L. *Angew. Chem., Int. Ed.* **2013**, *52*, 11840–11845; (b) Nikolaenko, T. Yu.; Bulavin, L. A.; Govoruna, D. N. J. Appl. Spectrosc. **2011**, *78*, 751–754. For the quantum mechanical interpretation of the IR of 2-dexoy ribose see:.
- (a) Angyal, S. J. Adv. Carbohydr. Chem. Biochem. 1984, 42, 15–68; (b) Angyal, S. J.; Pickles, V. A. Aust. J. Chem 1972, 1695–1710; (c) Angyal, S. J.; Pickles, V. A. Carbohydr. Res. 1967, 4, 269–270; (d) Bishop, C. T.; Cooper, F. P. Can. J. Chem. 1963, 41, 2743–2746; (e) Mackie, W.; Perlin, A. S. Can. J. Chem. 1966, 44, 2039–2049; (f) Lemieux, R. U.; Stevens, J. D. Can. J. Chem. 1966, 44, 249–262.
- (a) Juaristi, E.; Cuevas, G. Tetrahedron 1992, 48, 5019–5087; (b) Perrin, C. L.; Armstrong, K. B.; Fabian, M. A. J. Am. Chem. Soc. 1994, 116, 715–722.
- 5. Plaza, P. G. J.; Singh, G. Tetrahedron: Asymmetry 2010, 21, 2167–2171.
- Jiang, S.; McCullough, K. J.; Mekki, B.; Singh, G.; Wightman, R. H. J. Am. Chem. Soc., Perkin Trans. 1: Org. Bio-org. Chem. 1997, 12, 1805–1814.
- 7. Saeki, H.; Iwashige, T. Chem. Pharm. Bull. 1968, 16, 1129-1132.
- (a) Cremer, D.; Pople, J. A. J. Am. Chem. Soc. 1975, 97, 1354–1358; (b) García Álvarez, J. L.; Carriedo, G. A.; Amato, M. E.; Lombardo, G. M.; Punzo, F. Eur J. Inorg. Chem. 2010, 28, 4483–4491.
- Kitajgorodskij, A. I. Molecular Crystals and Molecules; Academic Press: New-York, 1973.
- 10. Angyal, S. J.; Christofides, J. C. J. Chem. Soc., Perkin Trans. 2 1996, 1485-1491.
- 11. Reuben, J. J. Am. Chem. Soc. 1985, 107, 5867-5870.
- 12. Snider, B. B.; Vo, H. V.; O'Neil, S. V. J. Org. Chem. 1998, 63, 4732–4740.
- 13. Muddasani, P. R.; Bernet, B. B.; Vasella, A. *Helv. Chim. Acta* **1994**, *77*, 257–290.
- 14. Liu, Q.; Brady, J. W. J. Am. Chem. Soc. 1996, 118, 12276–12286.
- (a) SAINT: SAX, Area Detector Integration, Siemens Analytical Instruments, Madison, Wisconsin, USA.; (b) SADABS: Siemens Area Detector Absorption Correction Software, Sheldrick, G., University of Goettingen: Germany, 1996.
- 16. Sir2004 Burla, M. C.; Caliandro, R.; Camalli, M.; Carrozzini, B.; Cascarano, G. L.; De Caro, G. L.; Giacovazzo, C.; Polidori, G.; Spagna, R. A Program for automatic solution and refinement of Crystal Structures; Istituto di Ricerca per lo Sviluppo di Metodologie Cristallografiche: CNR, Bari, 2004.
- Sheldrick, G. Shelxl2013. Program for structure refinement; University of Goettingen: Germany, 2013.
- Dolomanov, O. V.; Bourhis, L. J.; Gildea, R. J.; Howard, J. A. K.; Puschmann, H. J. Appl. Crystallogr. 2009, 42, 339–341.
- 19. Nardelli, M. J. Appl. Crystallogr. 1995, 28, 659.
- (a) Allen, F. H.; Kennard, O.; Taylor, R. Acc. Chem. Res. **1983**, *16*, 146–153; (b) Bruno, I. J.; Cole, J. C.; Edgington, P. R.; Kessler, M.; Macrae, C. F.; McCabe, P.; Pearson, J.; Taylor, R. Acta Crystallogr. **2002**, *B58*, 389–397.