

Accepted Manuscript

Selective killing of cancer cells with triterpenoic acid amides - The substantial role of an aromatic moiety alignmentx

Sven Sommerwerk, Lucie Heller, Julia Kufs, René Csuk



PII: S0223-5234(16)30543-8

DOI: [10.1016/j.ejmech.2016.06.053](https://doi.org/10.1016/j.ejmech.2016.06.053)

Reference: EJMECH 8714

To appear in: *European Journal of Medicinal Chemistry*

Received Date: 18 May 2016

Revised Date: 26 June 2016

Accepted Date: 28 June 2016

Please cite this article as: S. Sommerwerk, L. Heller, J. Kufs, R. Csuk, Selective killing of cancer cells with triterpenoic acid amides - The substantial role of an aromatic moiety alignmentx, *European Journal of Medicinal Chemistry* (2016), doi: 10.1016/j.ejmech.2016.06.053.

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Selective killing of cancer cells with triterpenoic acid amides – the substantial role of an aromatic moiety alignment

Sven Sommerwerk, Lucie Heller, Julia Kufs, René Csuk*

Martin-Luther-University Halle-Wittenberg, Organic Chemistry, Kurt-Mothes-Str.2, D-06120 Halle (Saale) Germany

Corresponding author:

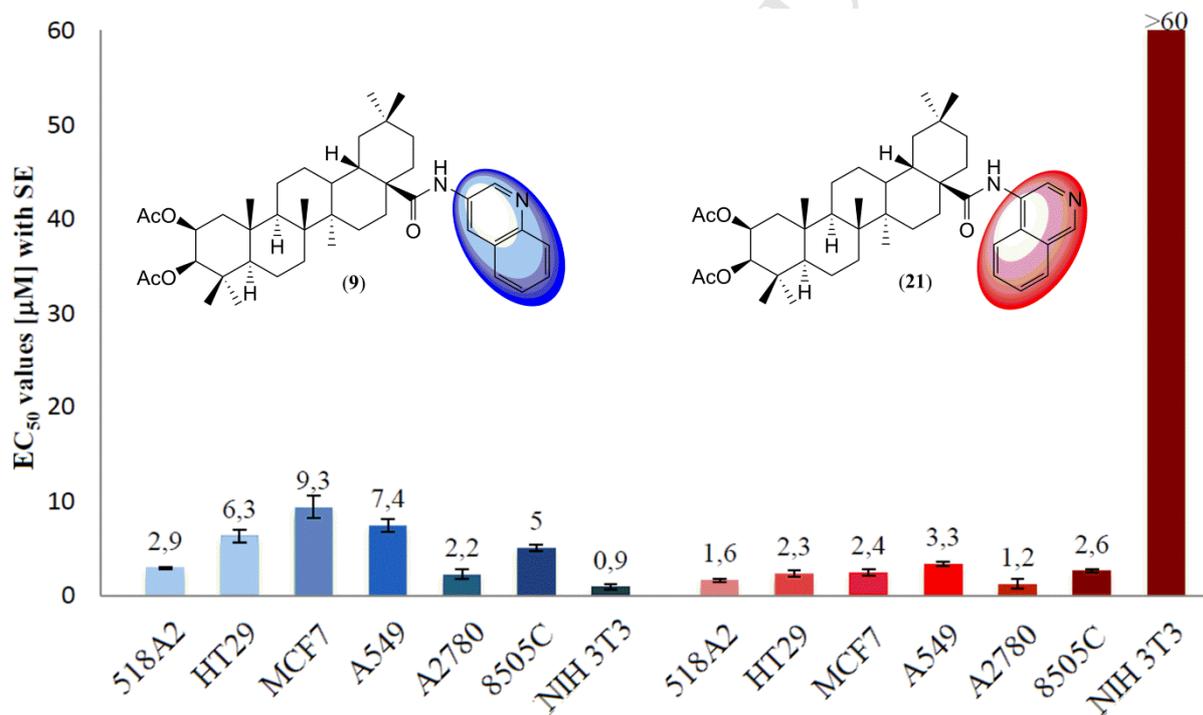
Prof. Dr. René Csuk
Bereich Organische Chemie
Martin-Luther Universität Halle-Wittenberg
Kurt-Mothes-Str. 2
D-06120 Halle (Saale)
Germany
Tel. +49 (0) 345 5525660
Fax: +49 (0) 345 5527030
Email: rene.csuk@chemie.uni-halle.de

Graphical Abstract:

Selective killing of cancer cells with triterpenoic acid amides – the substantial role of an aromatic moiety alignment

Sven Sommerwerk, Lucie Heller, Julia Kufs, René Csuk*

Martin-Luther-University Halle-Wittenberg, Organic Chemistry, Kurt-Mothes-Str.2, D-06120 Halle (Saale) Germany



A small structural difference (3-quinolinyl / 4-isoquinolinyl) has a strong impact on cytotoxicity and malignant/non-malignant cell selectivity.

Abstract

2,3-Di-*O*-acetyl-triterpenoic acid derived amides possessing a (2 β , 3 β) configuration in ring A and two acetyl groups were previously shown to possess high cytotoxicity for human tumor cell lines but to exhibit low cytotoxicity for non-malignant mouse fibroblasts. In this study, augustic acid (**1**) and 2-*epi*-corosolic acid (**2**) were chosen as starting points for the synthesis of analogs. While augustic acid derived 3-quinolinyl amide **9** gave low EC₅₀ values in SRB assays but was cytotoxic for all lines, the isomeric 4-isoquinolinyl amide **21** was very cytotoxic for the tumor cell lines but significantly less cytotoxic for the mouse fibroblasts NIH 3T3. In addition, a triacetylated 4-isoquinolinyl derivative of asiatic acid (**28**) gave EC₅₀ = 80 nM (for A2780 ovarian cancer cells). As shown by additional experiments (acridine orange/propidium iodide staining, fluorescence spectroscopy and cell cycle investigations) these compounds act mainly by apoptosis.

Keywords: maslinic acid, oleanolic acid, asiatic acid, tumor cells, SRB assay, apoptosis

1. Introduction

Despite of improved technology and medical progress for the last decades, cancer remains one of the world's leading causes of death finally resulting in approximately 8 million deaths globally per year.[1] Options to treat this family of diseases include surgery, radiotherapy, chemotherapy or a combination thereof. Many of the chemotherapeutics commonly applied for the treatment of cancer are characterized altogether by a high degree of non-selective toxicity for the cells. For example, *cis*-platinum (one of the most common used chemotherapeutics accounting to annually sales of approximately 3 billion Euro) is highly active for many types of human tumors [2] (e.g. ovarian cancer as well as cancer of the head, neck, testes but also for treating small cell lung cancer), but this drug also exhibits strong nephro- and neurotoxic effects.[3] Thus, a main goal of an effective treatment of cancer pursues to kill cancer cells selectively without any (or negligible) damage to non-malignant cells. Modern therapeutic schemes to achieve these goals include, for example, prodrug targeting,[4] advanced drug delivery systems [5, 6] or by applying light [7, 8] or heat [9, 10] or by a selective drug targeting using magnetic triggering.[11, 12]

In the course of investigating a small library of maslinic acid derivative compounds, we discovered an interesting molecule named **EM2** (Fig. 1), and this di-*O*-acetylated benzylamide of maslinic acid exhibited rather low EC_{50} values (sulforhodamine B assays [13] (SRB), $EC_{50} = 0.5 \mu\text{M}$ for human ovarian cancer cells) while being significantly less toxic for non-malignant mouse fibroblasts (NIH 3T3, $EC_{50} = 33.8 \mu\text{M}$).[14] We considered **EM2** as an ideal basis to start additional investigations concerning the biological activity of pentacyclic triterpenoid analogs carrying an additional nitrogen containing heterocycle attached to ring E by a suitable spacer. In addition, we became also interested in possible effects of the presence/absence of acetyl groups in ring A onto cytotoxicity and/or selectivity of the compounds. Hence, a panel of analogs was synthesized, and their cytotoxicity measured using SRB assays. For selected compounds (showing low EC_{50} values for the cancer cells and high EC_{50} values for the mouse fibroblasts), cell cycle investigations, annexin V/propidium iodide staining experiments as well as visual inspections of the cells by fluorescence microscopy were carried out.

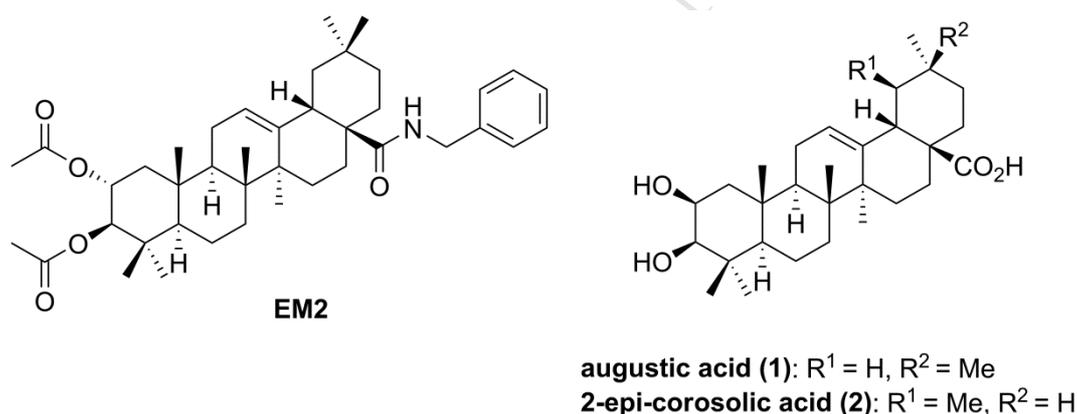
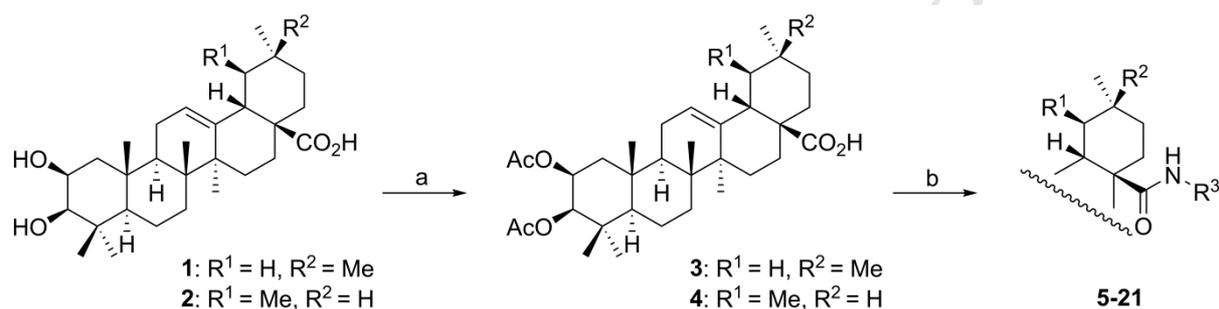


Fig. 1. Structures of **EM2**, augustic acid (**1**) and 2-epi-corosolic acid (**2**).

2. Results and discussion

Augustic acid (**1**, Fig. 1) and 2-epi-corosolic acid (**2**) were chosen as starting points for the synthesis of analogs. Both of these compounds carry two hydroxyl functions in ring A showing a (2 β , 3 β)-configuration, and they are easily accessed by synthesis.[15-18] Acetylation of **1** or **2** gave diacetates **3** and **4** (Scheme 1) whose reactions under Schotten-Baumann conditions with oxalyl chloride followed by the addition of an amine furnished amides **5–21** in moderate to excellent isolated yields between 38 and 90%. Amides **5–21** were

characterized by NMR, IR, UV-vis spectroscopy and ESI-MS. Thus, in the IR spectra of these amides a strong absorption band at $\nu = 1506\text{--}1534\text{ cm}^{-1}$ was detected being characteristic for the presence of a NHCO moiety. In their UV-vis spectra the aromatic chromophore gave characteristic absorption maxima between $\lambda = 244\text{--}279\text{ nm}$ for the pyridinyl substituted compounds while for the quinolinyl amides absorption maxima $\lambda = 241\text{--}335\text{ nm}$ were detected. As far as their ^1H NMR spectra are concerned, the signal for the NH moiety was detected between $\delta = 10.30\text{--}10.37\text{ ppm}$ for the 8-quinolinyl amides while all of the other compounds this signal was found between $\delta = 7.77\text{--}8.10\text{ ppm}$, respectively.[19, 20]



Scheme 1. Synthesis of compounds **3–21**: (a) Ac_2O , TEA, DMAP, DCM, 25°C , 20h, 65–79%; (b) oxalyl chloride, TEA, DMF, DCM, 25°C , 2h; then: $\text{R}^3\text{-NH}_2$, TEA, DMAP, DCM, 25°C , 2d, 38–90%.

The compounds were subjected to SRB assays,[13] the results of which are summarized in Table 1.

Table 1. Cytotoxicity of selected compounds (EC_{50} values in μM from SRB assays after 96 hours of treatment; the values are averaged from three independent experiments performed each in triplicate; confidence interval $\text{CI} = 95\%$; n.d. not detected). Human cancer cell lines: 518A2 (melanoma), HT29 (colorectal adenocarcinoma), MCF7 (breast adenocarcinoma), A549 (lung adenocarcinoma), A2780 (ovarian carcinoma), 8505C (thyroid carcinoma); NIH 3T3: nonmalignant mouse fibroblasts.

EC_{50}	R^1	R^2	R^3	518A2	HT29	MCF7	A549	A2780	8505C	NIH 3T3
1	H	Me	-	25.7 ± 0.7	>30	25.1 ± 2.2	n.d.	n.d.	n.d.	>30
2	Me	H	-	>30	>30	29.1 ± 0.7	>30	24.9 ± 2.7	>30	>30
3	H	Me	-	13.8 ± 1.0	23.1 ± 0.6	16.1 ± 3.8	n.d.	n.d.	n.d.	>30

4	Me	H	-	17.9±0.6	16.2±1.2	10.4±2.9	15.6±0.4	17.9±0.6	15.6±0.5	12.7±0.7
5	H	Me	3-pyridinyl	1.7±0.5	2.2±0.2	2.0±0.2	2.8±0.4	0.9±0.1	2.3±0.4	2.4±0.2
6	Me	H	3-pyridinyl	2.0±0.1	3.2±0.1	3.5±0.3	3.5±0.2	2.0±0.1	2.7±0.3	4.2±0.3
7	H	Me	4-pyridinyl	4.2±0.5	2.7±0.2	3.1±0.7	4.0±0.3	1.6±0.7	4.1±0.3	1.8±0.2
8	Me	H	4-pyridinyl	8.3±0.2	9.4±0.6	9.0±1.3	10.1±0.9	8.3±0.2	8.5±0.1	5.7±0.9
9	H	Me	3-quinolinyl	2.9±0.1	6.3±0.7	9.3±1.3	7.4±0.7	2.2±0.6	5.0±0.4	0.9±0.3
10	Me	H	3-quinolinyl	2.5±0.2	6.5±1.4	3.2±0.4	3.4±0.4	2.8±0.4	3.1±0.2	2.2±0.2
11	H	Me	4-quinolinyl	2.4±0.2	4.1±0.3	4.3±0.4	4.2±0.3	2.0±0.9	3.4±0.2	2.9±0.8
12	Me	H	4-quinolinyl	6.4±0.3	5.5±0.6	4.0±1.0	8.2±0.7	3.5±0.1	6.6±0.8	5.2±0.9
13	H	Me	5-quinolinyl	1.2±0.2	2.4±0.2	2.5±0.1	3.0±0.4	0.7±0.1	2.3±0.2	8.6±1.8
14	Me	H	5-quinolinyl	2.0±0.8	3.6±0.5	5.5±1.0	3.7±0.5	2.7±0.5	3.2±0.3	>60
15	H	Me	6-quinolinyl	2.9±0.1	4.4±0.4	5.8±0.8	4.9±0.5	2.0±0.5	3.8±0.2	0.6±0.1
16	Me	H	6-quinolinyl	4.2±0.2	9.2±1.0	6.5±1.3	5.3±0.4	4.2±0.1	4.3±0.1	0.9±0.1
17	H	Me	7-quinolinyl	1.6±0.2	1.3±0.1	1.4±0.2	2.6±0.1	1.0±0.1	1.6±0.1	0.7±0.1
18	Me	H	7-quinolinyl	2.4±0.2	2.0±0.2	1.6±0.4	3.1±0.0	1.8±0.0	2.5±0.5	1.8±0.1
19	H	Me	8-quinolinyl	6.2±1.0	>30	24.6±9.7	>30	5.1±0.9	16.1±3.5	>30
20	Me	H	8-quinolinyl	13.3±1.4	>30	>30	>30	6.7±0.9	>30	>30
21	H	Me	4-isoquinolinyl	1.6±0.2	2.3±0.4	2.4±0.4	3.3±0.3	1.2±0.6	2.6±0.2	>60

Compounds **5–8** carrying a pyridine substituent showed an improved cytotoxicity as compared to acetylated acids **3** and **4**. Thus, compound **5** gave an $EC_{50} = 0.9 \mu\text{M}$ for the human carcinoma cell line A2780 but selectivity between the human tumor cell lines and non-malignant mouse fibroblasts NIH 3T3 remained low for these compounds. A different performance, however, was observed during the biological evaluation of quinolinyl substituted compounds – depending on their substitution pattern. While the cytotoxicity of compounds **9–12** and **15–18** was similar to that of the pyridinyl substituted amides, 8-quinolinyl derivatives (**19** and **20**) showed selective cytotoxicity towards different human tumor cell lines. However, their overall cytotoxicity was low, and their solubility in water was quite poor. As a consequence, we discarded these compounds from additional investigations. 5-Quinolinyl substituted compounds **13** and **14**, however, performed better because they gave rather low EC_{50} values, they were cytotoxic to the human tumor cell lines but of significantly lowered cytotoxicity for the non-malignant mouse fibroblasts. From these findings we presumed a potentially existing connection between cytotoxicity and the orientation of the

aromatic system, and – as a consequence – we synthesized 4-isoquinolinyl compound **21**. Compounds **9** (a 3-quinolinyl derivative) and **21** are formally distinct from each other by a re-orientation of the aromatic substituent. (Fig. 2)

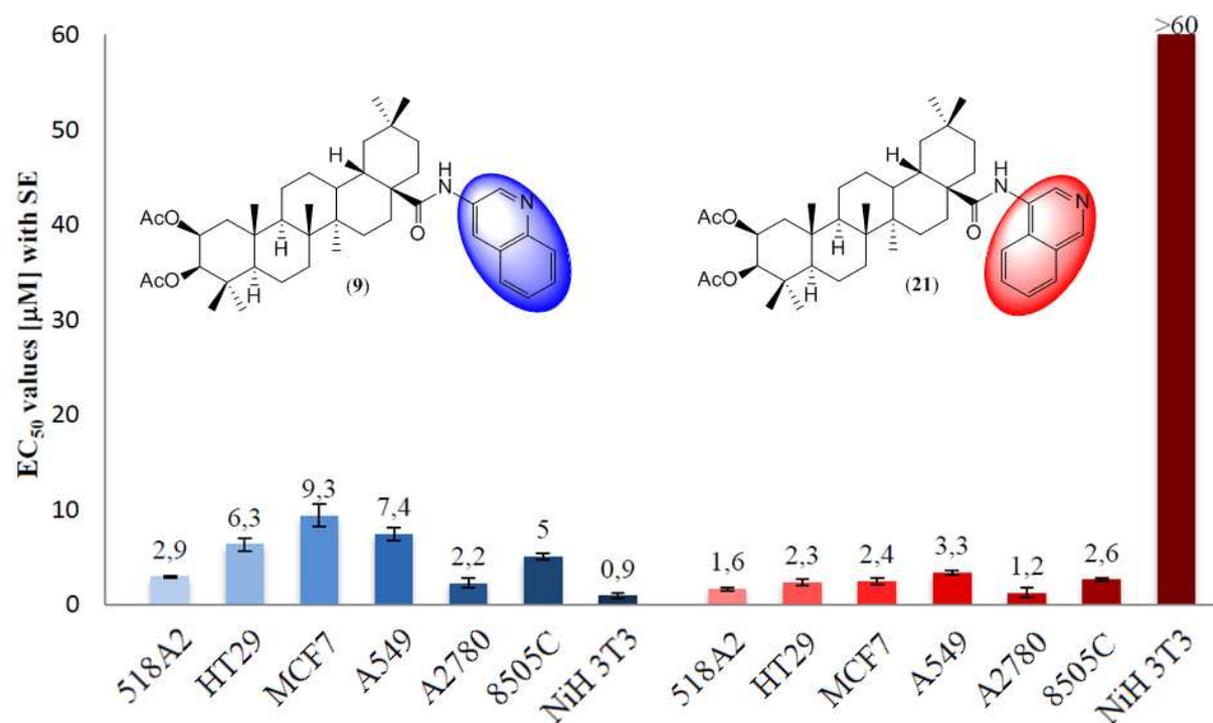
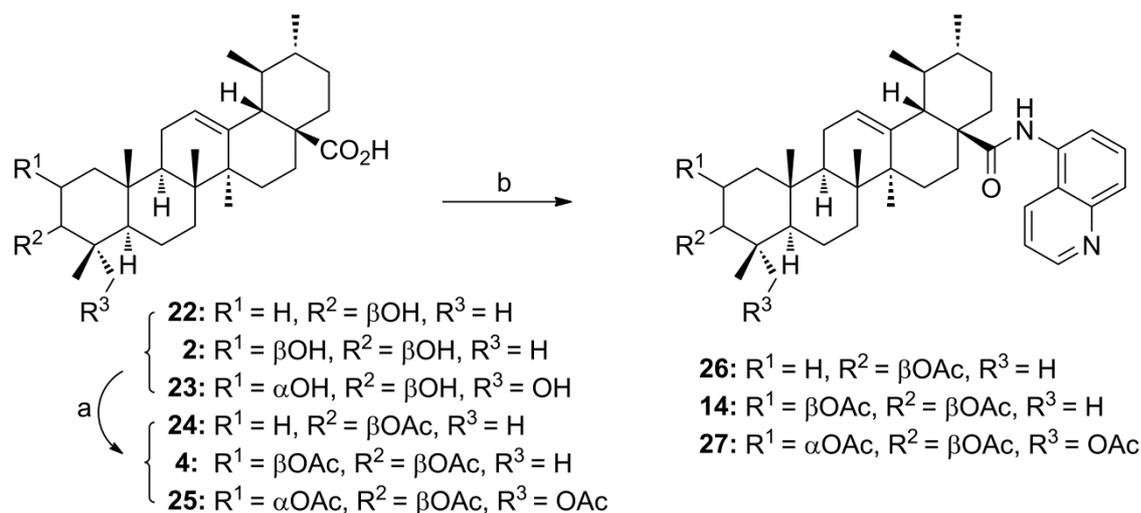


Fig. 2. Comparison of cytotoxicity of 3-quinolinyl amide **9** and 4-isoquinolinyl amide **21** (EC₅₀ values from SRB assays in μM) for different human tumor cell lines and nonmalignant mouse fibroblasts (NIH 3T3).

While this re-orientation had only a slight but significant effect in gaining high cytotoxicity, its impact on the selectivity for tumor cells was dramatically enhanced, and selectivity factors greater/equal 50 were observed.

To get a first impression of the impact of the presence of the hydroxyl groups of ring A, some derivatives of ursolic and asiatic acid were synthesized (Scheme 2).



Scheme 2. Synthesis of compounds **4**, **14**, **24–27**: (a) Ac_2O , TEA, DMAP, DCM, 25 °C, 20 h, 65–89%; (b) oxalyl chloride, TEA, DMF, DCM, 25°C, 2h; then 5-aminoquinoline, TEA, DMAP, DCM, 25 °C, 2 d, 52–87%.

The cytotoxic activities of acetylated amides of ursolic acid [21–26] has already been investigated by several groups in detail while for asiatic acid mainly amides without additional acetyl groups in ring A have been investigated so far.^{27,28} Screening of our analogs for their cytotoxic activity showed (cf. Table 2) gave for compound **26** (an ursolic acid derived 5-quinolinyl derivative) an almost complete loss of selectivity between cancer cells and nonmalignant mouse fibroblasts. For analog **27** (derived from asiatic acid), however, selectivity was maintained, and an increased cytotoxicity was gained.

Table 2. Cytotoxicity of selected compounds (EC_{50} values in μM from SRB assays after 96 hours of treatment; the values are averaged from three independent experiments performed each in triplicate; confidence interval CI = 95 %). Human cancer cell lines: 518A2 (melanoma), HT29 (colorectal adenocarcinoma), MCF7 (breast adenocarcinoma), A549 (lung adenocarcinoma), A2780 (ovarian carcinoma), 8505C (thyroid carcinoma); NIH 3T3: nonmalignant mouse fibroblasts.

EC_{50}	518A2	HT29	MCF7	A549	A2780	8505C	NIH 3T3
14	2.0±0.8	3.6±0.5	5.5±1.0	3.7±0.5	2.7±0.5	3.2±0.3	> 60
26	3.5±0.9	5.1±0.4	3.9±0.1	5.1±0.6	4.4±0.6	3.5±0.0	3.5±0.2

27	0.5±0.1	0.8±0.1	1.4±0.1	1.1±0.1	0.4±0.2	0.9±0.0	10.2±4.0
-----------	---------	---------	---------	---------	---------	---------	----------

Within the panel of compounds investigated so far, two factors appear particularly critical for obtaining cytotoxic as well as tumor-selective compounds, viz. the presence of an isoquinolinyl moiety at position 28 as well as a minimum of three *O*-acetyl groups attached to ring A. Combining these factors led to the synthesis of compound **28**, an 4-isoquinolinyl derivative of asiatic acid whose EC₅₀ values are depicted in Figure 3.

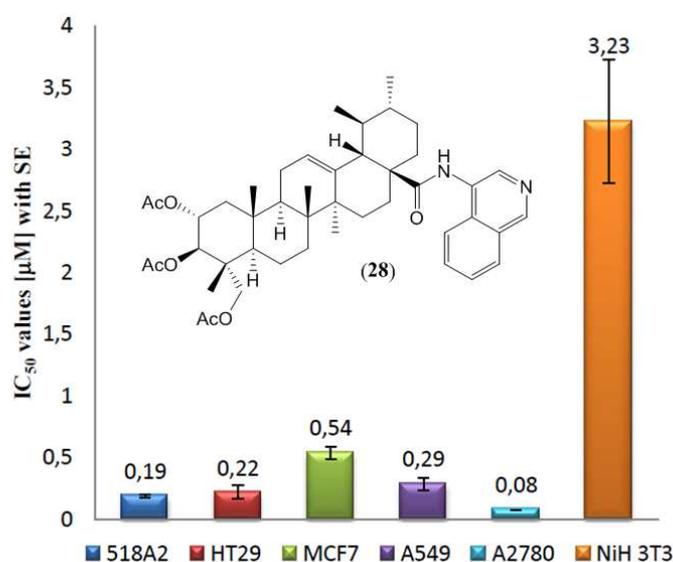


Fig. 3. Cytotoxicity of compound **28** [EC₅₀ values (in μM from SRB assays)] for different human tumor cell lines and non-malignant mouse fibroblasts (NIH 3T3).

Hence, we managed the synthesis of a triterpenoid derivative showing both low EC₅₀ values (for example, EC₅₀ = 80 nM for A2780 human ovarian cancer) but also high tumor/nonmalignant cell selectivity (EC₅₀ = 3.23 μM for NIH 3T3, thus a selectivity index of 40).

To have a better understanding of the cytotoxicity of these compounds (and their mode of action) some additional experiments were carried out (Fig. 4). A2780 Ovarian cancer cells were incubated with compounds **9**, **11**, **13**, **15**, **21**, **27** and **28**, and annexin V/propidium iodide staining experiments as well as quantitative FACS experiments were performed.

Incubation of the cells with each of the compounds for two days showed a significant increase in the number of cells having died by apoptosis; even more cells had died by secondary necrosis. (Fig. 4 B)

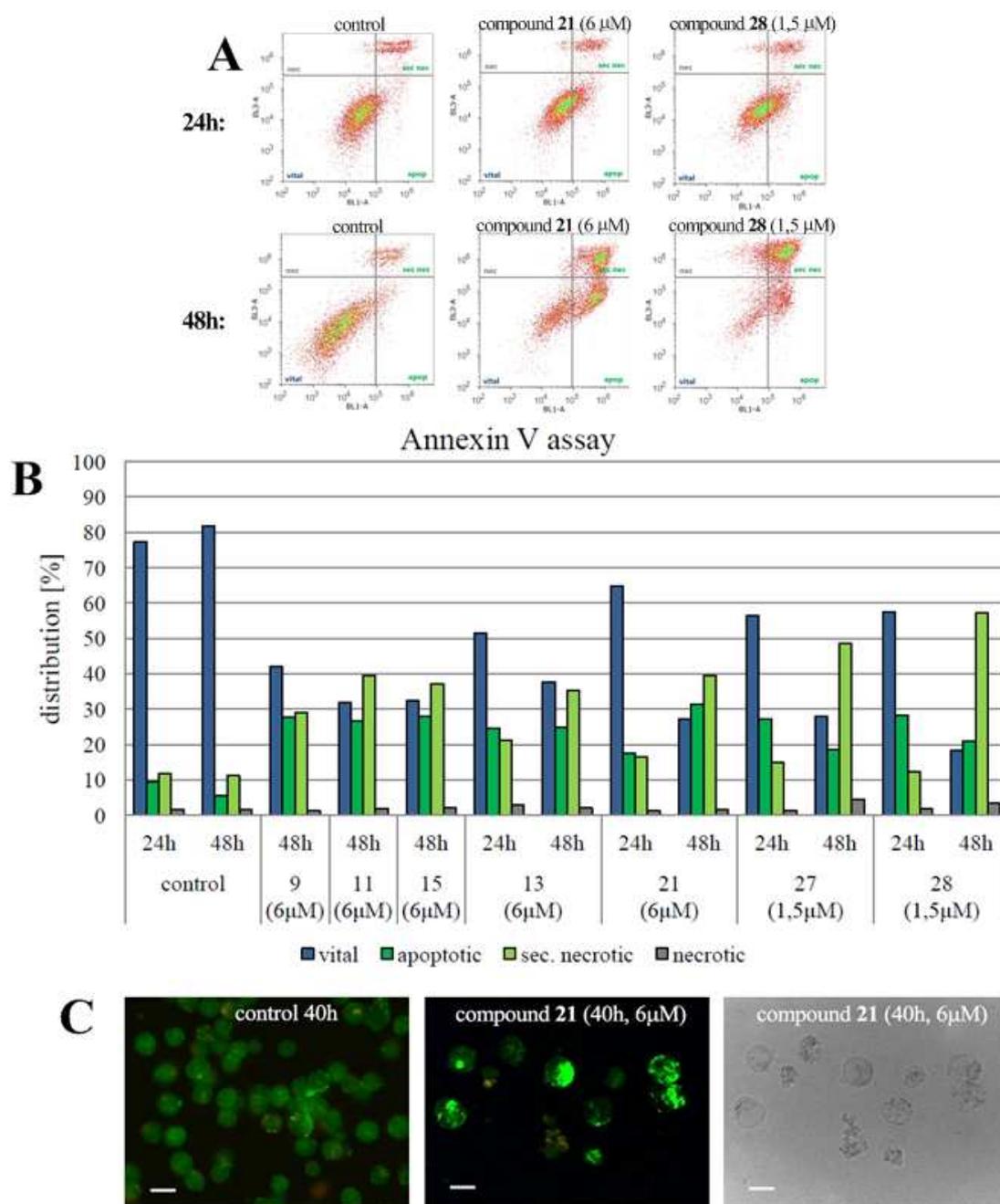


Fig. 4. (A) Representative samples of FACS based annexin V/PI assay of A2780 cells after 24h and 48h treatment with **21** and **28**. (B) Evaluation of annexin V/PI assay: treatment of A2780 cells with derivatives **9**, **11**, **13**, **15**, **21**, **27** and **28** after 24h and 48h. Diagram with arithmetic population distribution [%], determined by Attune[®] Cytometric Software v1 1.2.5. (C) Fluorescence microscopic and bright-field microscopic images (scale bar = 20 μm): control (A2780, 40h); treatment of A2780 cells with **21**.

Treatment of the cells with compound **28** for 24 h, however, showed the number of apoptotic cells significantly higher than that of cells having died of secondary necrosis. For example, treatment of A2780 cells with compound **28** (concentration as low as 1.5 μM , treatment for 24 h) gave an increase in the number of apoptotic cells (as compared to control) of 20% while the number of cells having died of necrosis or secondary necrosis remained unchanged. Prolongation of the time of incubation (up to 48 hours) led to a decrease of apoptotic cells by 10% while the number of secondary necrotic cells increased by 45% (compared to control).

Furthermore, inspection of dead A2780 cells (after having been incubated with **21**, 6 μM , 40 h) by fluorescence microscopy and application of an acridine orange/propidium iodide staining (AO/PI, Fig. 4C) allowed the identification of apoptosis induced modifications of the cell membrane. Comparison of cells (treated either with **21** or untreated) showed for the untreated cells the complete absence of apoptosis while the cells that were treated with **21** showed an unsymmetrical distribution of AO (typically crescent and granular) [27] and membrane blebbing – both findings are characteristic hallmarks for the presence of apoptosis. Some of the cells were shining in orange – these cells were at a secondary necrotic stage.

For the investigation of the cell cycle, A2780 cells were incubated with compounds **9**, **11**, **13**, **15**, **21**, **27** and **28** for 24h and 48h, respectively. Inspection of the results showed after an incubation for 24 h a decrease of the G1 peak together with a simultaneous increase of cells in the S phase and a complete loss of the G2 peak (Fig. 5). After incubation of the cells for 48 h, a decrease of the S phase was observed, and a subG1 peak appeared. This hypodiploidic peak can be explained by a cleavage of DNA; the presence of this peak is usually regarded as one of the typical hallmarks of apoptotic cells. The results from some additional investigations of the cell cycle as well as quantitative FACS measurements have been compiled in the supplementary material.

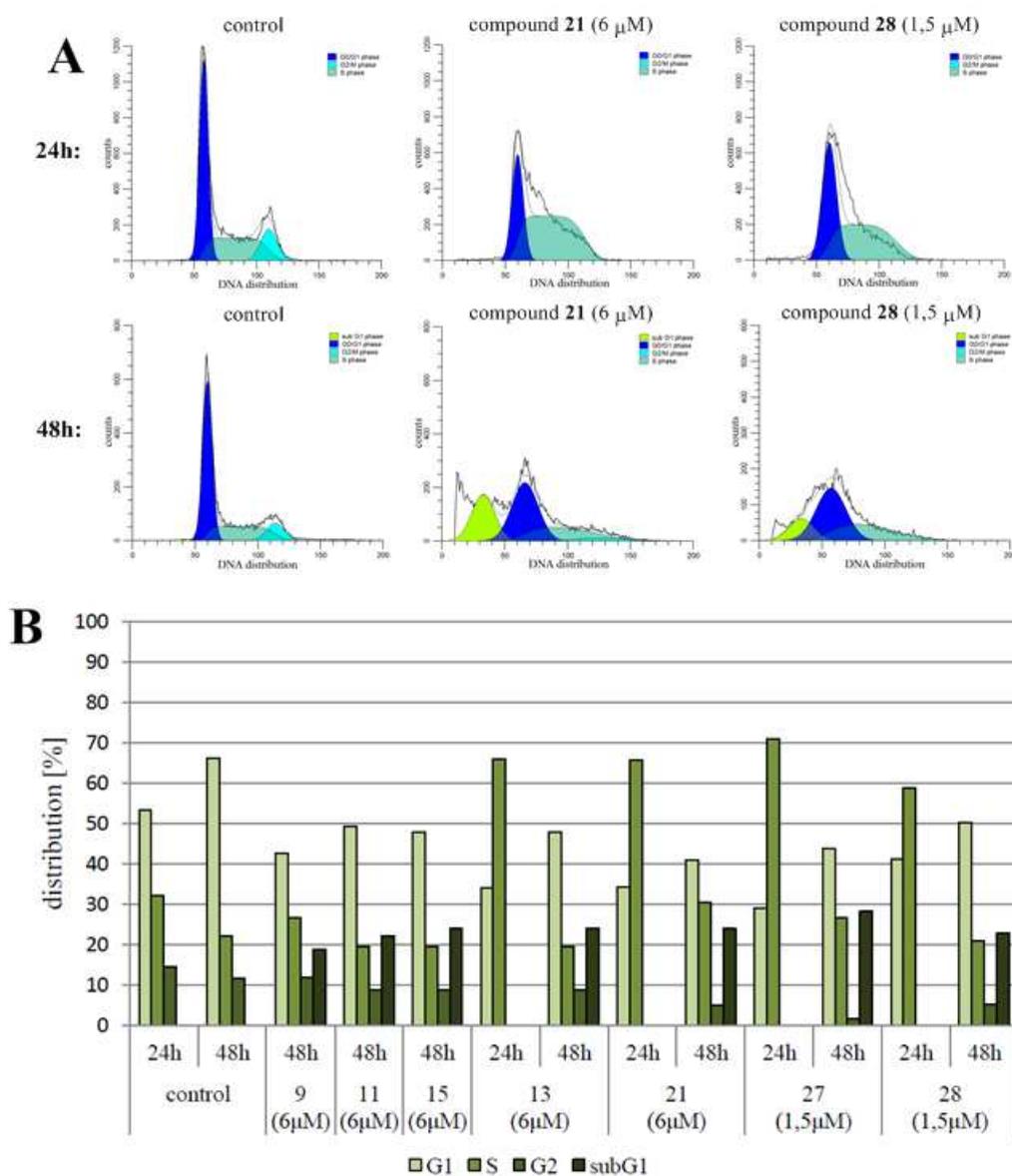


Fig. 5. (A): Representative examples for cell cycle evaluation of A2780 cells after 24h and 48h treatment with derivatives **21** and **28** via ModFit V 4.0.5. (B): Diagram with arithmetic means of the population distribution [%] of A2780 cells treated with **9**, **11**, **13**, **15**, **21**, **27** and **28** for 24h and 48h.

3. Conclusion

In this study amides of 2,3-O-acetyl-augustic acid and -2-epi-corosolic acid were synthesized, and their cytotoxicity was determined photometric SRB assays. While augustic acid derived 3-quinolinylyl amide **9** gave low EC_{50} values in SRB assays ($EC_{50} = 2.2$ to $9.3 \mu\text{M}$) but was cytotoxic for all lines including non-malignant mouse fibroblasts ($EC_{50} = 0.9 \mu\text{M}$), the

isomeric 4-isoquinolinyll amide **21** was even higher cytotoxic for the tumor cell lines (EC_{50} = 1.2 to 2.6 μ M) but significantly less cytotoxic for the mouse fibroblasts NIH 3T3 (EC_{50} > 60 μ M). In addition, a triacetylated 4-isoquinolinyll derivative of asiatic acid (**28**) gave rather low EC_{50} = 80 nM (for A2780 ovarian cancer cells) values. As shown by additional experiments (acridine orange/propidium iodide staining, fluorescence spectroscopy and cell cycle investigations) these compounds act mainly by apoptosis. These derivatives can function as ideal starting points for further screening and testing.

4. Experimental Part

4.1. General

Information about equipment and general techniques has been compiled in the supplementary materials section.

4.2. Syntheses

4.2.1. *2 β , 3 β -Diacetyloxy-olean-12-en-28-oic acid (3)*

To a solution of **1** (950 mg, 2.01 mmol) in dry DCM (100 mL), acetic anhydride (820 mg, 8.04 mmol), triethylamine (915 mg, 9.05 mmol) and DMAP (25 mg, 0.20 mmol) were added, and the mixture was stirred at room temperature for 20 hours. Et₂O (500 mL) was added, and the organic layer was washed with diluted HCl (0.1 M, 1 \times 500 mL), water (2 \times 500 mL) and brine (1 \times 250 mL), dried (MgSO₄), filtrated and concentrated in vacuum. The residue was subjected to column chromatography (silica gel, hexane/ethyl acetate, 8:2) to provide **3** (884 mg, 79%) as a white solid; m.p. 322 $^{\circ}$ C (decomp.) (lit.[28]: 148–150 $^{\circ}$ C); R_F = 0.24 (silica gel, hexane/ethyl acetate, 8:2); $[\alpha]_D^{25}$ = +83.81 $^{\circ}$ (c = 0.32, CHCl₃); IR (KBr): ν = 2944 s , 1744 v_s , 1698 s , 1464 w , 1436 w , 1366 m , 1252 s , 1196 m , 1160 w , 1058 m , 1030 m cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ = 5.33–5.30 (m , 1 H, H-2), 5.27 (dd , J = 3.6, 3.6 Hz, 1 H, H-12), 4.62 (d , J = 3.9 Hz, 1 H, H-3), 2.82 (dd , J = 13.9, 4.5 Hz, 1 H, H-18), 2.08 (s , 3 H, H-32), 2.02 (s , 3 H, H-34), 2.01–1.91 (m , 3 H, H-1a + H-16a + H-11a), 1.89–1.69 (m , 3 H, H-11b + H-22a + H-15a), 1.64–1.41 (m , 7 H, H-6a + H-6b + H-16b + H-19a + H-22b + H-9 + H-7a), 1.38–1.28 (m , 3 H, H-7b + H-21a + H-1b), 1.24–1.11 (m , 2 H, H-21b + H-19b), 1.20 (s , 3 H, H-25), 1.12 (s , 3 H, H-27), 1.09–1.03 (m , 1 H, H-15b), 1.04 (s , 3 H, H-23), 1.00–0.96 (m , 1 H, H-5), 0.92 (s , 3 H, H-29), 0.90 (s , 3 H, H-30), 0.90 (s , 3 H, H-24), 0.76 (s , 3 H, H-26) ppm; ¹³C NMR (125 MHz, CDCl₃): δ = 184.3 (C-28), 170.9 (C-33), 170.4 (C-31), 143.9 (C-13), 122.6 (C-12), 78.1 (C-3), 69.8 (C-2), 55.4 (C-5), 48.2 (C-9), 46.7 (C-17), 45.9 (C-19),

42.0 (C-1), 41.8 (C-14), 41.1 (C-18), 39.5 (C-8), 37.5 (C-4), 37.0 (C-10), 33.9 (C-21), 33.2 (C-30), 32.6 (C-22), 32.6 (C-7), 30.8 (C-20), 29.3 (C-24), 27.7 (C-15), 26.2 (C-27), 23.7 (C-29), 23.6 (C-11), 22.9 (C-16), 21.4 (C-32), 21.0 (C-34), 18.1 (C-6), 17.8 (C-23), 17.5 (C-26), 16.1 (C-25) ppm; MS (ESI): m/z (%) = 557.2 ($[M+H]^+$, 13), 574.3 ($[M+NH_4]^+$, 100), 579.4 ($[M+Na]^+$, 21), 1249.1 ($[2M+Na]^+$, 87); analysis calculated for $C_{34}H_{52}O_6$ (556.77): C 73.34, H 9.41; found: C 73.18, H 9.57.

4.2.2. 2β , 3β -Diacetyloxy-urs-12-en-28-oic acid (4)

Following the procedure given for **1**, from **2** compound **4** (728 mg, 65%) was obtained as a white solid; m.p. 247–252 °C; R_F = 0.48 (silica gel, hexane/ethyl acetate, 7:3); $[\alpha]_D = +75.00^\circ$ (c = 0.39, $CHCl_3$); IR (KBr): ν = 3410 m , 2950 s , 2872 m , 1746 vs , 1698 s , 1458 m , 1398 m , 1370 s , 1254 vs , 1234 s , 1194 m , 1160 m , 1056 m , 1030 s , 972 m cm^{-1} ; 1H NMR (500 MHz, $CDCl_3$): δ = 5.32 (ddd , J = 3.8, 3.8, 3.5 Hz, 1 H, H-2), 5.24 (dd , J = 3.6, 3.6 Hz, 1 H, H-12), 4.63 (d , J = 3.9 Hz, 1 H, H-3), 2.18 (d , J = 11.2 Hz, 1 H, H-18), 2.08–1.84 (m , 4 H, H-11a + H-11b + H-15a + H-16a), 2.04 (s , 3 H, H-32), 2.02 (s , 3 H, H-34), 1.97 (dd , J = 6.8, 3.9 Hz, 1 H, H-1a), 1.75–1.63 (m , 3 H, H-16b + H-22a + H-22b), 1.61–1.43 (m , 5 H, H-6a + H-6b + H-7a + H-9 + H-21a), 1.39–1.24 (m , 4 H, H-1b + H-7b + H-19 + H-21b), 1.22 (s , 3 H, H-25), 1.14–1.03 (m , 1 H, H-15b), 1.07 (s , 3 H, H-27), 1.04 (s , 3 H, H-23), 1.03–0.96 (m , 2 H, H-5 + H-20), 0.95 (d , J = 6.3 Hz, 3 H, H-30), 0.90 (s , 3 H, H-24), 0.86 (d , J = 6.4 Hz, 3 H, H-29), 0.79 (s , 3 H, H-26) ppm; ^{13}C NMR (125 MHz, $CDCl_3$): δ = 183.7 (C-28), 170.9 (C-33), 170.4 (C-31), 138.3 (C-13), 125.7 (C-12), 78.1 (C-3), 69.8 (C-2), 55.3 (C-5), 52.6 (C-18), 48.1 (C-17), 48.1 (C-9), 42.2 (C-14), 42.1 (C-1), 39.8 (C-8), 39.1 (C-19), 39.0 (C-20), 37.5 (C-4), 36.9 (C-22), 36.9 (C-10), 32.9 (C-7), 30.7 (C-21), 29.3 (C-24), 28.0 (C-15), 24.1 (C-16), 23.8 (C-27), 23.5 (C-11), 21.4 (C-32), 21.3 (C-30), 21.0 (C-34), 18.1 (C-6), 17.8 (C-23), 17.3 (C-26), 17.2 (C-29), 16.3 (C-25) ppm; MS (ESI): m/z (%) = 555.5 ($[M-H]^-$, 63), 1111.2 ($[2M-H]^-$, 100), 1133.5 ($[2M-2H+Na]^-$, 28); analysis calculated for $C_{34}H_{52}O_6$ (556.77): C 73.34, H 9.41; found: C 73.09, H 9.61.

4.2.3. 2β , 3β -Diacetyloxy-olean-12-en-28-oic acid 3-pyridinyl amide (5)

To a solution of **3** (100 mg, 0.18 mmol) in dry DCM (10 mL), oxalyl chloride (70 mg, 0.55 mmol), triethylamine (3 mg, 0.03 mmol) and DMF (2 mg, 0.03 mmol) were added, and the mixture was stirred at room temperature for 2 hours. The solvent was removed under reduced pressure, the residue was dissolved in dry THF (1 × 10 mL), the solvent was removed, and the residue was immediately dissolved in dry DCM (10 mL). The solution was

cooled to 0 °C and triethylamine (24 mg, 0.24 mmol), DMAP (2 mg, 0.02 mmol) as well as 3-aminopyridine (52 mg, 0.55 mmol) were added. After 2 days of stirring, Et₂O (100 mL) was added, the organic was washed with diluted HCl (0.1 M, 1 × 100 mL), water (2 × 100 mL) and brine (1 × 50 mL), dried (MgSO₄), filtrated and evaporated to dryness. Column chromatography (silica gel, hexane/ethyl acetate, 7:3) afforded **5** (85 mg, 75%) as a white solid; m.p. 156–160 °C; R_F = 0.60 (silica gel, chloroform/ethyl acetate, 1:1); $[\alpha]_D^{25} = +70.84^\circ$ ($c = 0.34$, CHCl₃); UV-vis (CHCl₃): λ_{\max} (log ϵ) = 244 nm (3.81), 279 nm (3.30); IR (KBr): $\nu = 3388m, 2950s, 2878m, 1746vs, 1684m, 1586m, 1524s, 1480s, 1418m, 1366s, 1252vs, 1234s, 1192m, 1158m, 1056m, 1030m, 708m$ cm⁻¹; ¹H NMR (500 MHz, CDCl₃): $\delta = 8.40$ (*d*, $J = 2.5$ Hz, 1 H, H-36), 8.31 (*dd*, $J = 4.7, 1.4$ Hz, 1 H, H-37), 8.22 (*ddd*, $J = 8.4, 2.5, 1.5$ Hz, 1 H, H-39), 7.77 (*s*, 1 H, NH), 7.24 (*dd*, $J = 8.3, 4.7$ Hz, 1 H, H-38), 5.57 (*dd*, $J = 3.4, 3.4$ Hz, 1 H, H-12), 5.31 (*ddd*, $J = 3.7, 3.7, 3.7$ Hz, 1 H, H-2), 4.60 (*d*, $J = 3.9$ Hz, 1 H, H-3), 2.67 (*dd*, $J = 12.7, 3.2$ Hz, 1 H, H-18), 2.12–1.90 (*m*, 4 H, H-16a + H-11a + H-11b + H-1a), 2.04 (*s*, 3 H, H-32), 2.02 (*s*, 3 H, H-34), 1.87–1.65 (*m*, 4 H, H-19a + H-22a + H-22b + H-16b), 1.64–1.52 (*m*, 3 H, H-15a + H-6a + H-9), 1.52–1.22 (*m*, 7 H, H-7a + H-7b + H-6b + H-21a + H-21b + H-1b + H-19b), 1.19 (*s*, 3 H, H-27), 1.14 (*s*, 3 H, H-25), 1.09 (*ddd*, $J = 14.1, 2.8, 2.8$ Hz, 1 H, H-15b), 1.02 (*s*, 3 H, H-23), 0.98–0.92 (*m*, 1 H, H-5), 0.93 (*s*, 6 H, H-29 + H-30), 0.88 (*s*, 3 H, H-24), 0.71 (*s*, 3 H, H-26) ppm; ¹³C NMR (125 MHz, CDCl₃): $\delta = 177.3$ (C-28), 170.8 (C-33), 170.4 (C-31), 145.3 (C-37), 145.2 (C-13), 141.1 (C-36), 134.9 (C-35), 127.2 (C-39), 123.8 (C-38), 123.5 (C-12), 78.0 (C-3), 69.6 (C-2), 55.2 (C-5), 48.1 (C-9), 47.5 (C-17), 46.7 (C-19), 42.7 (C-18), 42.5 (C-14), 42.1 (C-1), 39.7 (C-8), 37.4 (C-4), 36.8 (C-10), 34.2 (C-21), 33.0 (C-30), 32.5 (C-22), 32.4 (C-7), 30.9 (C-20), 29.2 (C-24), 27.4 (C-15), 25.9 (C-27), 24.2 (C-16), 23.9 (C-11), 23.7 (C-29), 21.4 (C-32), 21.0 (C-34), 18.0 (C-6), 17.8 (C-23), 17.2 (C-26), 16.1 (C-25) ppm; MS (ESI): m/z (%) = 633.5 ([M+H]⁺, 100), 1265.1 ([2M+H]⁺, 16); analysis calculated for C₃₉H₅₆N₂O₅ (632.87): C 74.01, H 8.92, N 4.43; found: C 73.82, H 9.03, N 4.19.

4.2.4. 2 β , 3 β -Diacetyloxy-ursan-12-en-28-oic acid 3-pyridinyl amide (**6**)

As described for **5**, compound **6** (120 mg, 56%) was obtained from **4** and 3-aminopyridine as a white solid; m.p. 146–149 °C; R_F = 0.42 (silica gel, hexane/ethyl acetate, 4:6); $[\alpha]_D^{25} = +51.20^\circ$ ($c = 0.35$, CHCl₃); UV-vis (CHCl₃): λ_{\max} (log ϵ) = 244 nm (4.02), 279 nm (3.57); IR (KBr): $\nu = 3394m, 2928s, 2872m, 1746vs, 1684m, 1618w, 1586m, 1526s, 1480s, 1485m, 1418s, 1398m, 1370s, 1326m, 1252vs, 1234vs, 1192m, 1156m, 1106w, 1078m, 1056m, 1030s, 972m, 944m, 754m, 706m$ cm⁻¹; ¹H NMR (500 MHz, CDCl₃): $\delta = 8.39$ (*d*, $J = 2.2$ Hz, 1 H,

H-36), 8.31 (*dd*, $J = 4.7, 1.1$ Hz, 1 H, H-37), 8.19 (*ddd*, $J = 8.4, 2.4, 1.4$ Hz, 1 H, H-39), 7.76 (*s*, 1 H, NH), 7.24 (*dd*, $J = 8.3, 4.7$ Hz, 1 H, H-38), 5.51 (*dd*, $J = 3.5, 3.5$ Hz, 1 H, H-12), 5.32 (*ddd*, $J = 3.8, 3.8, 3.5$ Hz, 1 H, H-2), 4.61 (*d*, $J = 3.9$ Hz, 1 H, H-3), 2.12–1.93 (*m*, 6 H, H-1a + H-11a + H-11b + H-16a + H-18 + H-22a), 2.04 (*s*, 3 H, H-32), 2.02 (*s*, 3 H, H-34), 1.87–1.81 (*m*, 1 H, H-16b), 1.75–1.67 (*m*, 1 H, H-15a), 1.62–1.45 (*m*, 6 H, H-6a + H-7a + H-9 + H-19 + H-21a + H-22b), 1.42 (*d*, $J = 11.8, 2.9$ Hz, 1 H, H-6b), 1.39–1.26 (*m*, 3 H, H-1b + H-7b + H-21b), 1.15 (*s*, 3 H, H-25), 1.13 (*s*, 3 H, H-27), 1.11–1.08 (*m*, 1 H, H-15b), 1.04–0.90 (*m*, 2 H, H-5 + H-20), 1.02 (*s*, 3 H, H-23), 0.98 (*d*, $J = 6.0$ Hz, 3 H, H-30), 0.92 (*d*, $J = 6.5$ Hz, 3 H, H-29), 0.89 (*s*, 3 H, H-24), 0.70 (*s*, 3 H, H-26) ppm; ^{13}C NMR (125 MHz, CDCl_3): $\delta = 177.1$ (C-28), 170.8 (C-33), 170.4 (C-31), 145.2 (C-37), 141.0 (C-36), 140.5 (C-13), 135.0 (C-35), 127.2 (C-39), 126.3 (C-12), 123.8 (C-38), 78.0 (C-3), 69.6 (C-2), 55.2 (C-5), 54.4 (C-18), 49.0 (C-17), 48.1 (C-9), 43.0 (C-14), 42.2 (C-1), 40.0 (C-19), 39.8 (C-8), 39.2 (C-20), 37.4 (C-4), 37.2 (C-22), 36.8 (C-10), 32.7 (C-7), 31.0 (C-21), 29.3 (C-24), 27.9 (C-15), 25.2 (C-16), 23.8 (C-11), 23.5 (C-27), 21.4 (C-32), 21.3 (C-30), 21.0 (C-34), 18.0 (C-6), 17.8 (C-23), 17.4 (C-29), 17.1 (C-26), 16.3 (C-25) ppm; MS (ESI): m/z (%) = 633.7 ($[\text{M}+\text{H}]^+$, 100), 1265.5 ($[\text{2M}+\text{H}]^+$, 46); analysis calculated for $\text{C}_{39}\text{H}_{56}\text{N}_2\text{O}_5$ (632.87): C 74.01, H 8.92, N 4.43; found: C 73.84, H 9.09, N 4.35.

4.2.5. $2\beta, 3\beta$ -Diacetyloxy-olean-12-en-28-oic acid 4-pyridinyl amide (7)

As described for **5**, compound **7** (62 mg, 55%) was obtained from **3** and 4-aminopyridine as a white solid; m.p. 171–174 °C; $R_F = 0.31$ (silica gel, chloroform/ethyl acetate, 1:1); $[\alpha]_D = +32.92^\circ$ ($c = 0.31$, MeOH); UV-vis (MeOH): λ_{max} (log ϵ) = 248 nm (4.21); IR (KBr): $\nu = 3386m, 2950s, 2878m, 1746vs, 1696m, 1588s, 1506s, 1474m, 1412m, 1366s, 1326m, 1252vs, 1236s, 1200m, 1158m, 1056m, 1030m, 1012m, 992m, 822m$ cm^{-1} ; ^1H NMR (500 MHz, CD_3OD): $\delta = 8.39$ – 8.36 (*m*, 2 H, H-37), 7.70–7.67 (*m*, 2 H, H-36), 5.42 (*dd*, $J = 3.5, 3.5$ Hz, 1 H, H-12), 5.32 (*ddd*, $J = 3.7, 3.7, 3.7$ Hz, 1 H, H-2), 4.58 (*d*, $J = 3.9$ Hz, 1 H, H-3), 2.99 (*dd*, $J = 13.2, 3.4$ Hz, 1 H, H-18), 2.20 (*ddd*, $J = 14.0, 14.0, 3.8$ Hz, 1 H, H-16a), 2.07–1.90 (*m*, 3 H, H-11a + H-11b + H-1a), 2.02 (*s*, 3 H, H-32), 1.99 (*s*, 3 H, H-34), 1.86–1.75 (*m*, 2 H, H-19a + H-16b), 1.73–1.61 (*m*, 4 H, H-22a + H-22b + H-15a + H-9), 1.61–1.42 (*m*, 4 H, H-6a + H-6b + H-7a + H-21a), 1.38 (*dd*, $J = 15.0, 3.7$ Hz, 1 H, H-1b), 1.32–1.18 (*m*, 3 H, H-7b + H-21b + H-19b), 1.21 (*s*, 3 H, H-27), 1.21 (*s*, 3 H, H-25), 1.12 (*ddd*, $J = 13.9, 3.0, 3.0$ Hz, 1 H, H-15b), 1.06 (*s*, 3 H, H-23), 1.05–1.01 (*m*, 1 H, H-5), 0.98 (*s*, 3 H, H-29), 0.94 (*s*, 3 H, H-30), 0.90 (*s*, 3 H, H-24), 0.72 (*s*, 3 H, H-26) ppm; ^{13}C NMR (125 MHz, CD_3OD): $\delta = 179.4$ (C-28), 172.5 (C-33), 172.1 (C-31), 150.5 (C-37), 148.4 (C-35), 145.3 (C-13),

124.0 (C-12), 115.8 (C-36), 79.6 (C-3), 70.8 (C-2), 56.4 (C-5), 49.3 (C-9), 48.9 (C-17), 47.4 (C-19), 43.1 (C-14), 42.8 (C-1), 42.3 (C-18), 40.7 (C-8), 38.3 (C-4), 37.9 (C-10), 35.0 (C-21), 33.6 (C-7), 33.5 (C-30), 33.4 (C-22), 31.6 (C-20), 29.5 (C-24), 28.5 (C-15), 26.6 (C-27), 24.6 (C-11), 24.1 (C-29), 23.8 (C-16), 21.2 (C-32), 20.8 (C-34), 19.0 (C-6), 18.2 (C-23), 17.7 (C-26), 16.6 (C-25) ppm; MS (ESI): m/z (%) = 633.4 ($[M+H]^+$, 100); analysis calculated for $C_{39}H_{56}N_2O_5$ (632.87): C 74.01, H 8.92, N 4.43; found: C 73.79, H 9.13, N 4.22.

4.2.6. $2\beta, 3\beta$ -Diacetyloxy-ursan-12-en-28-oic acid 4-pyridinyl amide (**8**)

As described for **5**, compound **8** (41 mg, 56%) was obtained from **4** and 4-aminopyridine as a white solid; m.p. 160–163 °C; R_F = 0.23 (silica gel, hexane/ethyl acetate, 3:7); $[\alpha]_D = +41.60^\circ$ ($c = 0.86$, $CHCl_3$); UV-vis ($CHCl_3$): λ_{max} (log ϵ) = 248 nm (3.90); IR (KBr): $\nu = 3422s$, 2950 m , 2872 w , 1744 m , 1690 m , 1636 m , 1588 m , 1506 m , 1432 s , 1414 s , 1384 vs , 1328 s , 1252 m , 1234 m , 1196 w , 1156 w , 1052 w , 1032 w , 1010 w cm^{-1} ; 1H NMR (500 MHz, CD_3OD): $\delta = 8.36$ (dd , $J = 5.0, 1.5$ Hz, 2 H, H-37), 7.67 (dd , $J = 4.9, 1.6$ Hz, 2 H, H-36), 5.42 (dd , $J = 3.6, 3.6$ Hz, 1 H, H-12), 5.32 (ddd , $J = 3.9, 3.9, 3.2$ Hz, 1 H, H-2), 4.59 (d , $J = 4.0$ Hz, 1 H, H-3), 2.37 (d , $J = 10.4$ Hz, 1 H, H-18), 2.23–2.15 (m , 1 H, H-16a), 2.03–1.95 (m , 3 H, H-1a + H-11a + H-11b), 2.01 (s , 3 H, H-32), 1.99 (s , 3 H, H-34), 1.91–1.82 (m , 2 H, H-15a + H-16b), 1.82–1.76 (m , 1 H, H-22a), 1.70–1.46 (m , 7 H, H-6a + H-6b + H-7a + H-9 + H-19 + H-21a + H-22b), 1.44–1.36 (m , 2 H, H-1b + H-21b), 1.35–1.30 (m , 1 H, H-7b), 1.19 (s , 3 H, H-25), 1.17 (s , 3 H, H-27), 1.14–1.08 (m , 2 H, H-15b + H-20), 1.07–1.01 (m , 1 H, H-5), 1.06 (s , 3 H, H-23), 1.00 (d , $J = 6.4$ Hz, 3 H, H-30), 0.95 (d , $J = 6.5$ Hz, 3 H, H-29), 0.90 (s , 3 H, H-24), 0.70 (s , 3 H, H-26) ppm; ^{13}C NMR (125 MHz, CD_3OD): $\delta = 179.4$ (C-28), 172.5 (C-33), 172.1 (C-31), 150.5 (C-37), 148.4 (C-35), 140.1 (C-13), 127.1 (C-12), 115.7 (C-36), 79.6 (C-3), 70.7 (C-2), 56.3 (C-5), 53.8 (C-18), 50.2 (C-17), 49.1 (C-9), 43.4 (C-14), 43.0 (C-1), 40.9 (C-8), 40.7 (C-19), 40.1 (C-20), 38.3 (C-4), 37.8 (C-10), 37.8 (C-22), 33.8 (C-7), 31.7 (C-21), 29.5 (C-24), 28.9 (C-15), 25.0 (C-16), 24.5 (C-11), 24.3 (C-27), 21.5 (C-30), 21.1 (C-32), 20.8 (C-34), 19.0 (C-6), 18.2 (C-23), 17.7 (C-29), 17.6 (C-26), 16.6 (C-25) ppm; MS (ESI): m/z (%) = 633.5 ($[M+H]^+$, 100), 1287.5 ($[2M+Na]^+$, 1); analysis calculated for $C_{39}H_{56}N_2O_5$ (632.87): C 74.01, H 8.92, N 4.43; found: C 73.75, H 8.83, N 4.31.

4.2.7. $2\beta, 3\beta$ -Diacetyloxy-olean-12-en-28-oic acid 3-quinolinyl amide (**9**)

As described for **5**, compound **9** (88 mg, 72%) was obtained from **3** and 3-aminoquinoline as a white solid; m.p. 174–177 °C; R_F = 0.69 (silica gel, chloroform/ethyl acetate, 1:1); $[\alpha]_D = +110.93^\circ$ ($c = 0.32$, $CHCl_3$); UV-vis ($CHCl_3$): λ_{max} (log ϵ) = 254 nm (4.60), 322 nm (3.69),

335 nm (3.70); IR (KBr): $\nu = 3388w, 2948s, 2876m, 1746vs, 1684m, 1608w, 1534s, 1486s, 1468m, 1420m, 1364s, 1252vs, 1196m, 1158m, 1056m, 1030m \text{ cm}^{-1}$; $^1\text{H NMR}$ (500 MHz, CDCl_3): $\delta = 8.91$ (*d*, $J = 2.4$ Hz, 1 H, H-43), 8.55 (*d*, $J = 2.6$ Hz, 1 H, H-36), 8.03–8.00 (*m*, 2 H, H-38 + NH), 7.78 (*d*, $J = 8.0$ Hz, 1 H, H-39), 7.61 (*ddd*, $J = 8.4, 6.9, 1.4$ Hz, 1 H, H-40), 7.52 (*ddd*, $J = 8.1, 6.9, 1.2$ Hz, 1 H, H-41), 5.65 (*dd*, $J = 3.5, 3.5$ Hz, 1 H, H-12), 5.32 (*ddd*, $J = 3.8, 3.8, 3.8$ Hz, 1 H, H-2), 4.61 (*d*, $J = 3.9$ Hz, 1 H, H-3), 2.75 (*dd*, $J = 12.9, 3.6$ Hz, 1 H, H-18), 2.18–1.95 (*m*, 4 H, H-16a + H-11a + H-11b + H-1a), 2.04 (*s*, 3 H, H-32), 2.02 (*s*, 3 H, H-34), 1.90–1.71 (*m*, 4 H, H-19a + H-22a + H-22b + H-16b), 1.68–1.52 (*m*, 3 H, H-15a + H-6a + H-9), 1.52–1.23 (*m*, 7 H, H-7a + H-7b + H-6b + H-21a + H-21b + H-1b + H-19b), 1.22 (*s*, 3 H, H-27), 1.15–1.09 (*m*, 1 H, H-15b), 1.12 (*s*, 3 H, H-25), 1.01 (*s*, 3 H, H-23), 0.98–0.94 (*m*, 1 H, H-5), 0.97 (*s*, 3 H, H-29), 0.96 (*s*, 3 H, H-30), 0.88 (*s*, 3 H, H-24), 0.70 (*s*, 3 H, H-26) ppm; $^{13}\text{C NMR}$ (125 MHz, CDCl_3): $\delta = 177.6$ (C-28), 170.9 (C-33), 170.4 (C-31), 145.4 (C-37), 145.3 (C-13), 143.8 (C-36), 131.6 (C-35), 129.1 (C-38), 128.5 (C-42), 128.3 (C-40), 128.0 (C-39), 127.4 (C-41), 123.8 (C-43), 123.7 (C-12), 78.0 (C-3), 69.6 (C-2), 55.2 (C-5), 48.1 (C-9), 47.7 (C-17), 46.8 (C-19), 42.8 (C-18), 42.6 (C-14), 42.1 (C-1), 39.7 (C-8), 37.4 (C-4), 36.8 (C-10), 34.3 (C-21), 33.1 (C-30), 32.5 (C-22), 32.4 (C-7), 30.9 (C-20), 29.2 (C-24), 27.4 (C-15), 26.0 (C-27), 24.4 (C-16), 23.9 (C-11), 23.7 (C-29), 21.4 (C-32), 21.0 (C-34), 18.0 (C-6), 17.8 (C-23), 17.2 (C-26), 16.1 (C-25) ppm; MS (ESI): m/z (%) = 683.5 ($[\text{M}+\text{H}]^+$, 100); analysis calculated for $\text{C}_{43}\text{H}_{58}\text{N}_2\text{O}_5$ (682.93): C 75.62, H 8.56, N 4.10; found: C 75.47, H 8.77, N 3.98.

4.2.8. $2\beta, 3\beta$ -Diacetyloxy-ursan-12-en-28-oic acid 3-quinolinyl amide (**10**)

As described for **5**, compound **10** (60 mg, 65%) was obtained from **4** and 3-aminoquinoline as a white solid; m.p. 141–143 °C; $R_F = 0.38$ (silica gel, hexane/ethyl acetate, 6:4); $[\alpha]_D = +84.40^\circ$ ($c = 0.31$, CHCl_3); UV-vis (CHCl_3): λ_{max} (log ϵ) = 254 nm (4.60), 322 nm (3.66), 336 nm (3.66); IR (KBr): $\nu = 3396m, 2924s, 2870m, 2852m, 1744vs, 1684m, 1608w, 1576w, 1534s, 1486m, 1468m, 1458m, 1420m, 1396m, 1364s, 1252vs, 1234s, 1196m, 1158w, 1144w, 1116w, 1102w, 1078w, 1056m, 1030m \text{ cm}^{-1}$; $^1\text{H NMR}$ (500 MHz, CDCl_3): $\delta = 8.86$ (*d*, $J = 2.5$ Hz, 1 H, H-43), 8.55 (*d*, $J = 2.6$ Hz, 1 H, H-36), 8.02 (*d*, $J = 9.1$ Hz, 1 H, H-38), 7.79–7.77 (*m*, 1 H, H-41), 7.63–7.59 (*m*, 1 H, H-39), 7.54–7.50 (*m*, 1 H, H-40), 5.59 (*dd*, $J = 3.6, 3.6$ Hz, 1 H, H-12), 5.32 (*ddd*, $J = 3.8, 3.8, 3.5$ Hz, 1 H, H-2), 4.61 (*d*, $J = 3.9$ Hz, 1 H, H-3), 2.17–1.97 (*m*, 6 H, H-1a + H-11a + H-11b + H-16a + H-18 + H-22a), 2.04 (*s*, 3 H, H-32), 2.02 (*s*, 3 H, H-34), 1.92–1.86 (*m*, 1 H, H-16b), 1.79–1.70 (*m*, 1 H, H-15a), 1.68–1.46 (*m*, 6 H, H-6a + H-7a + H-9 + H-19 + H-21a + H-22b), 1.44–1.34 (*m*, 3 H, H-1b + H-

6b + H-21b), 1.32–1.26 (*m*, 1 H, H-7b), 1.17–1.10 (*m*, 1 H, H-15b), 1.15 (*s*, 3 H, H-27), 1.12 (*s*, 3 H, H-25), 1.08–0.98 (*m*, 4 H, H-20 + H-30), 1.00 (*s*, 3 H, H-23), 0.97–0.93 (*m*, 1 H, H-5), 0.95 (*d*, $J = 6.5$ Hz, 3 H, H-29), 0.88 (*s*, 3 H, H-24), 0.68 (*s*, 3 H, H-26) ppm; ^{13}C NMR (125 MHz, CDCl_3): $\delta = 177.4$ (C-28), 170.9 (C-33), 170.4 (C-31); 145.3 (C-42), 143.8 (C-43), 140.7 (C-13), 131.7 (C-35), 129.1 (C-41), 128.5 (C-37), 128.2 (C-39), 128.0 (C-40), 127.3 (C-38), 126.4 (C-12), 123.7 (C-36), 78.0 (C-3), 69.6 (C-2), 55.2 (C-5), 54.4 (C-18), 49.1 (C-17), 48.1 (C-9), 43.0 (C-14), 42.2 (C-1), 40.0 (C-19), 39.8 (C-8), 39.2 (C-20), 37.4 (C-22), 37.2 (C-4), 36.8 (C-10), 32.7 (C-7), 31.0 (C-21), 29.3 (C-24), 28.0 (C-15), 25.4 (C-16), 23.8 (C-11), 23.5 (C-27), 21.4 (C-32), 21.3 (C-30), 21.0 (C-34), 18.0 (C-6), 17.8 (C-23), 17.4 (C-29), 17.2 (C-26), 16.3 (C-25) ppm; MS (ESI): m/z (%) = 683.5 ($[\text{M}+\text{H}]^+$, 100), 1365.3 ($[\text{2M}+\text{H}]^+$, 38); analysis calculated for $\text{C}_{43}\text{H}_{58}\text{N}_2\text{O}_5$ (682.93): C 75.62, H 8.56, N 4.10; found: C 75.51, H 8.89, N 3.99.

4.2.9. 2β , 3β -Diacetyloxy-olean-12-en-28-oic acid 4-quinolinyl amide (**11**)

As described for **5**, compound **11** (49 mg, 40%) was obtained from **3** and 4-aminoquinoline as a white solid; m.p. 226–227 °C; $R_F = 0.72$ (silica gel, chloroform/ethyl acetate, 1:1); $[\alpha]_D = +17.60^\circ$ ($c = 0.29$, CHCl_3); UV-vis (CHCl_3): λ_{max} (log ϵ) = 299 nm (4.03), 315 nm (3.88); IR (KBr): $\nu = 2946s$, 2870 m , 1744 vs , 1692 m , 1624 m , 1570 m , 1522 s , 1494 s , 1458 s , 1432 s , 1396 m , 1366 m , 1314 m , 1252 vs , 1236 s , 1162 m , 1056 m , 1030 m , 1012 m cm^{-1} ; ^1H NMR (500 MHz, CDCl_3): $\delta = 8.83$ (*d*, $J = 5.1$ Hz, 1 H, H-37), 8.58 (*s*, 1 H, NH), 8.49 (*d*, $J = 5.1$ Hz, 1 H, H-36), 8.13 (*d*, $J = 8.3$ Hz, 1 H, H-39), 7.76–7.71 (*m*, 2 H, H-41 + H-42), 7.59 (*ddd*, $J = 8.3, 7.0, 1.2$ Hz, 1 H, H-40), 5.76 (*dd*, $J = 3.5, 3.5$ Hz, 1 H, H-12), 5.30 (*ddd*, $J = 3.8, 3.8, 3.8$ Hz, 1 H, H-2), 4.60 (*d*, $J = 3.9$ Hz, 1 H, H-3), 2.85 (*dd*, $J = 12.9, 3.8$ Hz, 1 H, H-18), 2.16 (*ddd*, $J = 14.0, 14.0, 3.8$ Hz, 1 H, H-16a), 2.03–1.82 (*m*, 6 H, H-11a + H-11b + H-1a + H-19a + H-22a + H-16b), 2.02 (*s*, 3 H, H-32), 2.02 (*s*, 3 H, H-34), 1.78 (*ddd*, $J = 14.0, 14.0, 4.3$ Hz, 1 H, H-22b), 1.66 (*ddd*, $J = 13.9, 13.9, 4.3$ Hz, 1 H, H-15a), 1.61–1.41 (*m*, 4 H, H-9 + H-6a + H-21a + H-7a), 1.41–1.21 (*m*, 5 H, H-6b + H-19b + H-1b + H-21b + H-7b), 1.23 (*s*, 3 H, H-27), 1.14 (*ddd*, $J = 13.7, 3.1, 3.1$ Hz, 1 H, H-15b), 0.99 (*s*, 3 H, H-25), 0.98 (*s*, 3 H, H-29), 0.98 (*s*, 6 H, H-30 + H-23), 0.96–0.91 (*m*, 1 H, H-5), 0.87 (*s*, 3 H, H-24), 0.51 (*s*, 3 H, H-26) ppm; ^{13}C NMR (125 MHz, CDCl_3): $\delta = 177.5$ (C-28), 170.9 (C-33), 170.4 (C-31), 151.6 (C-37), 149.0 (C-35), 145.3 (C-13), 140.7 (C-38), 130.9 (C-39), 129.4 (C-41), 126.3 (C-40), 124.1 (C-12), 120.2 (C-43), 118.9 (C-42), 110.3 (C-36), 78.0 (C-3), 69.6 (C-2), 55.3 (C-5), 48.8 (C-17), 48.1 (C-9), 46.9 (C-19), 43.1 (C-18), 42.5 (C-14), 42.0 (C-1), 39.6 (C-8), 37.4 (C-4), 36.7 (C-10), 34.2 (C-21), 33.1 (C-30), 32.8 (C-22), 32.4 (C-7), 30.9

(C-20), 29.2 (C-24), 27.4 (C-15), 26.2 (C-27), 24.5 (C-16), 23.8 (C-29), 23.7 (C-11), 21.4 (C-32), 21.0 (C-34), 17.9 (C-6), 17.7 (C-23), 16.8 (C-26), 16.0 (C-25) ppm; MS (ESI): m/z (%) = 683.4 ($[M+H]^+$, 100); analysis calculated for $C_{43}H_{58}N_2O_5$ (682.93): C 75.62, H 8.56, N 4.10; found: C 75.41, H 8.81, N 4.00.

4.2.10. 2β , 3β -Diacetyloxy-ursan-12-en-28-oic acid 4-quinolinyl amide (**12**)

As described for **5**, compound **12** (47 mg, 38%) was obtained from **4** and 4-aminoquinoline as a white solid; m.p. 162–164 °C; R_F = 0.56 (silica gel, chloroform/ethyl acetate, 1:1); $[\alpha]_D = -9.12^\circ$ ($c = 0.27$, $CHCl_3$); UV-vis ($CHCl_3$): λ_{max} (log ϵ) = 300 nm (4.03), 315 nm (3.90); IR (KBr): $\nu = 2928s, 2872m, 1744vs, 1692m, 1570m, 1518s, 1494s, 1458s, 1394m, 1370m, 1310m, 1252vs, 1234s, 1196m, 1158w, 1056m, 1030m\text{ cm}^{-1}$; 1H NMR (500 MHz, $CDCl_3$): $\delta = 8.82$ ($d, J = 5.2$ Hz, 1 H, H-37), 8.56 (s , 1 H, NH), 8.46 ($d, J = 5.2$ Hz, 1 H, H-36), 8.18 ($d, J = 8.5$ Hz, 1 H, H-39), 7.77–7.73 (m , 2 H, H-41 + H-42), 7.63–7.58 (m , 1 H, H-40), 5.69–5.66 (m , 1 H, H-12), 5.32–5.28 (m , 1 H, H-2), 4.59 ($d, J = 3.6$ Hz, 1 H, H-3), 2.22–1.91 (m , 7 H, H-18 + H-16a + H-16b + H-11a + H-11b + H-22a + H-1a), 2.01 (s , 6 H, H-32 + H-34), 1.81–1.31 (m , 10 H, H-15a + H-22b + H-21a + H-21b + H-19 + H-9 + H-7a + H-6a + H-6b + H-1b), 1.31–1.24 (m , 1 H, H-7b), 1.18–1.12 (m , 1 H, H-15b), 1.16 (s , 3 H, H-27), 1.11–0.91 (m , 2 H, H-20 + H-5), 1.03–0.98 (m , 12 H, H-23 + H-25 + H-29 + H-30), 0.87 (s , 3 H, H-24), 0.54 (s , 3 H, H-26) ppm; ^{13}C NMR (125 MHz, $CDCl_3$): $\delta = 177.4$ (C-28), 170.9 (C-33), 170.4 (C-31), 151.1 (C-37), 148.4 (C-35), 141.3 (C-38), 140.5 (C-13), 130.4 (C-39), 129.6 (C-41), 126.7 (C-12), 126.5 (C-40), 120.2 (C-43), 118.9 (C-42), 110.3 (C-36), 78.0 (C-3), 69.6 (C-2), 55.3 (C-5), 54.6 (C-18), 50.3 (C-17), 48.1 (C-9), 42.9 (C-14), 42.1 (C-1), 40.1 (C-19), 39.8 (C-8), 39.2 (C-20), 37.4 (C-22), 37.4 (C-4), 36.7 (C-10), 32.6 (C-7), 31.0 (C-21), 29.2 (C-24), 27.9 (C-15), 25.5 (C-16), 23.8 (C-11), 23.5 (C-27), 21.4 (C-32), 21.3 (C-30), 21.0 (C-34), 18.0 (C-6), 17.7 (C-23), 17.5 (C-29), 16.9 (C-26), 16.1 (C-25) ppm; MS (ESI): m/z (%) = 683.5 ($[M+H]^+$, 100); analysis calculated for $C_{43}H_{58}N_2O_5$ (682.93): C 75.62, H 8.56, N 4.10; found: C 75.33, H 8.69, N 3.85.

4.2.11. 2β , 3β -Diacetyloxy-olean-12-en-28-oic acid 5-quinolinyl amide (**13**)

As described for **5**, compound **13** (82 mg, 67%) was obtained from **3** and 5-aminoquinoline as a white solid; m.p. 167–168 °C; R_F = 0.60 (silica gel, chloroform/ethyl acetate, 1:1); $[\alpha]_D = +38.91^\circ$ ($c = 0.34$, $CHCl_3$); UV-vis ($CHCl_3$): λ_{max} (log ϵ) = 242 nm (4.49), 307 nm (3.77), 317 nm (3.77); IR (KBr): $\nu = 3386w, 2948s, 2866m, 1746vs, 1684m, 1594w, 1572w, 1522m, 1488s, 1396m, 1364s, 1252vs, 1196m, 1156m, 1056m, 1030m, 798m\text{ cm}^{-1}$; 1H NMR (500

MHz, CDCl₃): δ = 8.93 (*dd*, J = 4.2, 1.5 Hz, 1 H, H-40), 8.14–8.12 (*m*, 2 H, H-42 + H-36), 8.10 (*s*, 1 H, NH), 7.92 (*ddd*, J = 8.5, 0.9, 0.9 Hz, 1 H, H-38), 7.69 (*dd*, J = 8.4, 7.7 Hz, 1 H, H-37), 7.43 (*dd*, J = 8.6, 4.2 Hz, 1 H, H-41), 5.63 (*dd*, J = 3.5, 3.5 Hz, 1 H, H-12), 5.30 (*ddd*, J = 3.9, 3.9, 3.9 Hz, 1 H, H-2), 4.60 (*d*, J = 3.9 Hz, 1 H, H-3), 2.86 (*dd*, J = 12.8, 4.1 Hz, 1 H, H-18), 2.14 (*ddd*, J = 14.0, 14.0, 3.9 Hz, 1H, H-16a), 2.02 (*s*, 6 H, H-32 + H-34), 1.99–1.94 (*m*, 3 H, H-11a + H-11b + H-1a), 1.92–1.79 (*m*, 4 H, H-19a + H-22a + H-22b + H-16b), 1.74 (*ddd*, J = 14.1, 14.1, 4.4 Hz, 1H, H-15a), 1.61–1.36 (*m*, 5 H, H-9 + H-6a + H-6b + H-7a + H-21a), 1.36–1.24 (*m*, 4 H, H-19b + H-1b + H-21b + H-7b), 1.23 (*s*, 3 H, H-27), 1.15 (*ddd*, J = 13.8, 3.1, 3.1 Hz, 1H, H-15b), 1.06 (*s*, 3 H, H-25), 1.00 (*s*, 3 H, H-23), 1.00 (*s*, 3 H, H-29), 0.98–0.94 (*m*, 1 H, H-5), 0.97 (*s*, 3 H, H-30), 0.88 (*s*, 3 H, H-24), 0.67 (*s*, 3 H, H-26) ppm; ¹³C NMR (125 MHz, CDCl₃): δ = 177.0 (C-28), 170.8 (C-33), 170.4 (C-31), 150.3 (C-40), 148.8 (C-39), 145.4 (C-13), 132.9 (C-35), 129.7 (C-37), 129.3 (C-42), 126.7 (C-38), 123.5 (C-12), 122.2 (C-43), 120.8 (C-41), 120.2 (C-36), 78.0 (C-3), 69.6 (C-2), 55.3 (C-5), 48.1 (C-9), 48.1 (C-17), 46.9 (C-19), 43.0 (C-18), 42.6 (C-14), 42.0 (C-1), 39.7 (C-8), 37.4 (C-4), 36.8 (C-10), 34.3 (C-21), 33.1 (C-30), 33.1 (C-22), 32.5 (C-7), 30.9 (C-20), 29.2 (C-24), 27.4 (C-15), 26.1 (C-27), 24.2 (C-16), 23.8 (C-29), 23.7 (C-11), 21.4 (C-32), 21.0 (C-34), 18.0 (C-6), 17.7 (C-23), 17.4 (C-26), 16.1 (C-25) ppm; MS (ESI): m/z (%) = 683.6 ([M+H]⁺, 100); analysis calculated for C₄₃H₅₈N₂O₅ (682.93): C 75.62, H 8.56, N 4.10; found: C 75.53, H 8.71, N 3.76.

4.2.12. 2 β , 3 β -Diacetyloxy-ursan-12-en-28-oic acid 5-quinolinyl amide (**14**)

As described for **5**, compound **14** (43 mg, 58%) was obtained from **4** and 5-aminoquinoline as a white solid; m.p. 144–146 °C; R_F = 0.35 (silica gel, hexane/ethyl acetate, 4:6); $[\alpha]_D^{25}$ = +18.90° (c = 1.02, CHCl₃); UV-vis (CHCl₃): λ_{max} (log ϵ) = 241 nm (4.42), 308 nm (3.71), 317 nm (3.71); IR (KBr): ν = 3346 m , 2948 s , 2872 m , 1744 vs , 1658 s , 1620 m , 1596 m , 1572 m , 1518 s , 1488 s , 1458 s , 1398 s , 1366 s , 1320 m , 1252 vs , 1234 vs , 1194 s , 1156 m , 1114 m , 1078 m , 1056 s , 1030 s , 996 m , 972 m , 944 m , 798 s , 754 s cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ = 8.92 (*dd*, J = 4.2, 1.5 Hz, 1 H, H-40), 8.13 (*d*, J = 8.6 Hz, 1 H, H-42), 8.03 (*s*, 1 H, NH), 8.02 (*d*, J = 6.8 Hz, 1 H, H-36), 7.92 (*d*, J = 8.5 Hz, 1 H, H-38), 7.68 (*dd*, J = 8.1, 8.1 Hz, 1 H, H-37), 7.41 (*dd*, J = 8.6, 4.2 Hz, 1 H, H-41), 5.55 (*dd*, J = 3.3, 3.3 Hz, 1 H, H-12), 5.30 (*ddd*, J = 3.7, 3.7, 3.5 Hz, 1 H, H-2), 4.60 (*d*, J = 3.9 Hz, 1 H, H-3), 2.19 (*d*, J = 10.5 Hz, 1 H, H-18), 2.17–2.10 (*m*, 1 H, H-16a), 2.06–1.95 (*m*, 4 H, H-1a + H-11a + H-11b + H-22a), 2.02 (*s*, 3 H, H-32), 2.01 (*s*, 3 H, H-34), 1.95–1.80 (*m*, 2 H, H-15a + H-16b), 1.77–1.69 (*m*, 1 H, H-22b), 1.63–1.47 (*m*, 5 H, H-6a + H-7a + H-9 + H-19 + H-21a), 1.46–1.29 (*m*, 4 H, H-1b

+ H-6b + H-7b + H-21b), 1.18–1.12 (*m*, 1 H, H-15b), 1.16 (*s*, 3 H, H-27), 1.11–1.03 (*m*, 1 H, H-20), 1.08 (*s*, 3 H, H-25), 1.01 (*d*, $J = 3.9$ Hz, 3 H, H-30), 1.01 (*s*, 3 H, H-23), 0.99–0.93 (*m*, 1 H, H-5), 0.97 (*d*, $J = 6.4$ Hz, 3 H, H-29), 0.88 (*s*, 3 H, H-24), 0.71 (*s*, 3 H, H-26) ppm; ^{13}C NMR (125 MHz, CDCl_3): $\delta = 176.8$ (C-28), 170.8 (C-33), 170.4 (C-31), 150.3 (C-40), 148.8 (C-39), 140.4 (C-13), 133.0 (C-35), 129.6 (C-37), 129.5 (C-42), 126.7 (C-38), 126.2 (C-12), 122.3 (C-43), 120.7 (C-41), 120.5 (C-36), 78.0 (C-3), 69.6 (C-2), 55.3 (C-5), 54.6 (C-18), 49.5 (C-17), 48.1 (C-9), 43.0 (C-14), 42.1 (C-1), 40.1 (C-19), 39.9 (C-8), 39.3 (C-20), 37.8 (C-22), 37.4 (C-4), 36.8 (C-10), 32.9 (C-7), 31.0 (C-21), 29.2 (C-24), 27.9 (C-15), 25.2 (C-16), 23.7 (C-27), 23.6 (C-11), 21.4 (C-32), 21.3 (C-30), 21.0 (C-34), 18.0 (C-6), 17.8 (C-23), 17.5 (C-26), 17.5 (C-29), 16.2 (C-25) ppm; MS (ESI): m/z (%) = 683.5 ($[\text{M}+\text{H}]^+$, 100), 705.3 ($[\text{M}+\text{Na}]^+$, 3), 1365.3 ($[\text{2M}+\text{H}]^+$, 15), 1387.6 ($[\text{2M}+\text{Na}]^+$, 10), 1403.1 ($[\text{2M}+\text{K}]^+$, 3); analysis calculated for $\text{C}_{43}\text{H}_{58}\text{N}_2\text{O}_5$ (682.93): C 75.62, H 8.56, N 4.10; found: C 75.39, H 8.82, N 4.03.

4.2.13. 2β , 3β -Diacetyloxy-olean-12-en-28-oic acid 6-quinolinyl amide (**15**)

As described for **5**, compound **15** (109 mg, 89%) was obtained from **3** and 6-aminoquinoline as a white solid; m.p. 167–169 °C; $R_F = 0.63$ (silica gel, chloroform/ethyl acetate, 1:1); $[\alpha]_D = +101.63^\circ$ ($c = 0.37$, CHCl_3); UV-vis (CHCl_3): λ_{max} (log ϵ) = 247 nm (4.70), 274 nm (3.86), 282 nm (3.84), 322 nm (3.70), 335 nm (3.68); IR (KBr): $\nu = 3396w$, 2948s, 2878m, 1744vs, 1684m, 1626w, 1600w, 1532s, 1496m, 1464m, 1430m, 1364s, 1252vs, 1194m, 1156w, 1118w, 1056m, 1030m, 1012m cm^{-1} ; ^1H NMR (500 MHz, CDCl_3): $\delta = 8.81$ (*dd*, $J = 4.2, 1.7$ Hz, 1 H, H-39), 8.49 (*d*, $J = 2.4$ Hz, 1 H, H-43), 8.09 (*dd*, $J = 8.1, 1.6$ Hz, 1 H, H-41), 8.01 (*d*, $J = 9.0$ Hz, 1 H, H-37), 7.94 (*s*, 1 H, NH), 7.39–7.34 (*m*, 2 H, H-40 + H-36), 5.62 (*dd*, $J = 3.6, 3.6$ Hz, 1 H, H-12), 5.31 (*ddd*, $J = 3.8, 3.8, 3.8$ Hz, 1 H, H-2), 4.61 (*d*, $J = 3.9$ Hz, 1 H, H-3), 2.73 (*dd*, $J = 12.7, 3.5$ Hz, 1 H, H-18), 2.17–1.94 (*m*, 4 H, H-16a + H-11a + H-11b + H-1a), 2.03 (*s*, 3 H, H-32), 2.02 (*s*, 3 H, H-34), 1.90–1.70 (*m*, 4 H, H-19a + H-22a + H-22b + H-16b), 1.70–1.52 (*m*, 3 H, H-15a + H-9 + H-6a), 1.52–1.23 (*m*, 7 H, H-7a + H-7b + H-21a + H-21b + H-6b + H-1b + H-19b), 1.21 (*s*, 3 H, H-27), 1.15–1.08 (*m*, 1 H, H-15b), 1.14 (*s*, 3 H, H-25), 1.01 (*s*, 3 H, H-23), 0.98–0.94 (*m*, 1 H, H-5), 0.96 (*s*, 3 H, H-29), 0.95 (*s*, 3 H, H-30), 0.88 (*s*, 3 H, H-24), 0.70 (*s*, 3 H, H-26) ppm; ^{13}C NMR (125 MHz, CDCl_3): $\delta = 177.1$ (C-28), 170.9 (C-33), 170.4 (C-31), 149.4 (C-40), 145.7 (C-41), 145.4 (C-13), 136.0 (C-35), 136.0 (C-38), 130.2 (C-42), 129.1 (C-37), 123.4 (C-12), 123.1 (C-43), 121.7 (C-39), 116.0 (C-36), 78.0 (C-3), 69.7 (C-2), 55.3 (C-5), 48.1 (C-9), 47.5 (C-17), 46.8 (C-19), 42.8 (C-18), 42.5 (C-14), 42.1 (C-1), 39.7 (C-8), 37.5 (C-4), 36.8 (C-10), 34.3 (C-21), 33.1

(C-30), 32.6 (C-22), 32.4 (C-7), 30.9 (C-20), 29.2 (C-24), 27.4 (C-15), 26.0 (C-27), 24.4 (C-16), 23.9 (C-11), 23.7 (C-29), 21.4 (C-32), 21.0 (C-34), 18.0 (C-6), 17.7 (C-23), 17.1 (C-26), 16.2 (C-25) ppm; MS (ESI): m/z (%) = 683.5 ($[M+H]^+$, 100); analysis calculated for $C_{43}H_{58}N_2O_5$ (682.93): C 75.62, H 8.56, N 4.10; found: C 75.59, H 8.80, N 3.85.

4.2.14. 2β , 3β -Diacetyloxy-ursan-12-en-28-oic acid 6-quinolinyl amide (**16**)

As described for **5**, compound **16** (200 mg, 87%) was obtained from **4** and 6-aminoquinoline as a white solid; m.p. 156–159 °C; R_F = 0.39 (silica gel, hexane/ethyl acetate, 4:6); $[\alpha]_D = +76.70^\circ$ ($c = 0.31$, $CHCl_3$); UV-vis ($CHCl_3$): λ_{max} (log ϵ) = 248 nm (4.62), 284 nm (3.77), 322 nm (3.60), 335 nm (3.60); IR (KBr): $\nu = 3394m$, 2928s, 2872m, 1744vs, 1682s, 1626m, 1600m, 1576m, 1534s, 1496s, 1458s, 1430s, 1398m, 1378s, 1364s, 1252vs, 1234vs, 1194s, 1156m, 1138m, 1118m, 1078m, 1056m, 1030s, 966m, 944m, 934m, 828m, 752m cm^{-1} ; 1H NMR (500 MHz, $CDCl_3$): $\delta = 8.81$ (*dd*, $J = 4.2, 1.7$ Hz, 1 H, H-39), 8.46 (*d*, $J = 2.4$ Hz, 1 H, H-43), 8.09 (*d*, $J = 6.8$ Hz, 1 H, H-41), 8.01 (*d*, $J = 9.0$ Hz, 1 H, H-37), 7.95 (*s*, 1 H, NH), 7.37–7.34 (*m*, 2 H, H-40 + H-36), 5.57 (*dd*, $J = 3.5, 3.5$ Hz, 1 H, H-12), 5.31 (*ddd*, $J = 3.8, 3.8, 3.8$ Hz, 1 H, H-2), 4.61 (*d*, $J = 3.9$ Hz, 1 H, H-3), 2.16–2.03 (*m*, 4 H, H-11a + H-11b + H-16a + H-18), 2.03 (*s*, 3 H, H-32), 2.02 (*s*, 3 H, H-34), 2.03–1.97 (*m*, 2 H, H-1a + H-22a), 1.88 (*dd*, $J = 8.3, 6.3$ Hz, 1 H, H-16b), 1.80–1.72 (*m*, 1 H, H-15a), 1.67–1.46 (*m*, 6 H, H-6a + H-7a + H-9 + H-19 + H-21a + H-22b), 1.44–1.34 (*m*, 3 H, H-1b + H-6b + H-21b), 1.32–1.26 (*m*, 1 H, H-7b), 1.16–1.10 (*m*, 1 H, H-15b), 1.15 (*s*, 3 H, H-27), 1.13 (*s*, 3 H, H-25), 1.06–0.98 (*m*, 1 H, H-20), 1.00 (*d*, $J = 2.3$ Hz, 3 H, H-30), 0.99 (*s*, 3 H, H-23), 0.97–0.93 (*m*, 1 H, H-5), 0.95 (*d*, $J = 6.4$ Hz, 3 H, H-29), 0.88 (*s*, 3 H, H-24), 0.68 (*s*, 3 H, H-26) ppm; ^{13}C NMR (125 MHz, $CDCl_3$): $\delta = 176.9$ (C-28), 170.8 (C-33), 170.4 (C-31), 149.4 (C-39), 145.6 (C-38), 140.6 (C-13), 136.1 (C-35), 136.1 (C-41), 130.2 (C-37), 129.1 (C-42), 126.2 (C-12), 123.0 (C-36), 121.7 (C-40), 115.9 (C-43), 78.0 (C-3), 69.6 (C-2), 55.2 (C-5), 54.4 (C-18), 48.9 (C-17), 48.1 (C-9), 43.0 (C-14), 42.2 (C-1), 40.0 (C-19), 39.8 (C-8), 39.3 (C-20), 37.4 (C-4), 37.2 (C-22), 36.8 (C-10), 32.7 (C-7), 31.0 (C-21), 29.2 (C-24), 28.0 (C-15), 25.3 (C-16), 23.8 (C-11), 23.5 (C-27), 21.4 (C-32), 21.3 (C-30), 21.0 (C-34), 18.0 (C-6), 17.8 (C-23), 17.5 (C-29), 17.1 (C-26), 16.3 (C-25) ppm; MS (ESI): m/z (%) = 683.7 ($[M+H]^+$, 100), 1365.3 ($[2M+H]^+$, 34), 1388.3 ($[2M+Na]^+$, 4); analysis calculated for $C_{43}H_{58}N_2O_5$ (682.93): C 75.62, H 8.56, N 4.10; found: C 75.41, H 8.74, N 3.91.

4.2.15. 2 β , 3 β -Diacetyloxy-olean-12-en-28-oic acid 7-quinolinyl amide (**17**)

As described for **5**, compound **17** (104 mg, 85%) was obtained from **3** and 7-aminoquinoline as a white solid; m.p. 167–170 °C; R_F = 0.67 (silica gel, chloroform/ethyl acetate, 1:1); $[\alpha]_D^{25} = +48.44^\circ$ ($c = 0.27$, CHCl₃); UV-vis (CHCl₃): λ_{\max} (log ϵ) = 247 nm (4.85), 275 nm (4.14), 324 nm (3.99), 335 nm (3.95); IR (KBr): $\nu = 2948s, 2866m, 1744vs, 1684m, 1624m, 1526m, 1496s, 1458s, 1430s, 1364m, 1252vs, 1192w, 1156m, 1056m, 1030m$ cm⁻¹; ¹H NMR (500 MHz, CDCl₃): $\delta = 8.86$ (*dd*, $J = 4.3, 1.7$ Hz, 1 H, H-38), 8.10 (*dd*, $J = 8.3, 1.1$ Hz, 1 H, H-42), 8.07 (*dd*, $J = 8.9, 2.1$ Hz, 1 H, H-43), 7.98 (*s*, 1 H, NH), 7.90 (*d*, $J = 2.0$ Hz, 1 H, H-36), 7.77 (*d*, $J = 8.9$ Hz, 1 H, H-40), 7.32 (*dd*, $J = 8.2, 4.3$ Hz, 1 H, H-39), 5.61 (*dd*, $J = 3.4, 3.4$ Hz, 1 H, H-12), 5.32 (*ddd*, $J = 3.7, 3.7, 3.7$ Hz, 1 H, H-2), 4.62 (*d*, $J = 3.9$ Hz, 1 H, H-3), 2.73 (*dd*, $J = 12.3, 3.4$ Hz, 1 H, H-18), 2.16–1.92 (*m*, 4 H, H-16a + H-11a + H-11b + H-1a), 2.04 (*s*, 3 H, H-32), 2.02 (*s*, 3 H, H-34), 1.90–1.63 (*m*, 5 H, H-19a + H-22a + H-22b + H-16b + H-15a), 1.62–1.36 (*m*, 5 H, H-9 + H-21a + H-7a + H-6a + H-6b), 1.36–1.24 (*m*, 4 H, H-1b + H-19b + H-21b + H-7b), 1.21 (*s*, 3 H, H-27), 1.15–1.08 (*m*, 1 H, H-15b), 1.14 (*s*, 3 H, H-25), 1.01 (*s*, 3 H, H-23), 0.99–0.94 (*m*, 1 H, H-5), 0.96 (*s*, 3 H, H-29), 0.95 (*s*, 3 H, H-30), 0.88 (*s*, 3 H, H-24), 0.75 (*s*, 3 H, H-26) ppm; ¹³C NMR (125 MHz, CDCl₃): $\delta = 177.0$ (C-28), 170.9 (C-33), 170.4 (C-31), 150.9 (C-38), 148.8 (C-37), 145.2 (C-13), 139.6 (C-35), 135.9 (C-42), 128.7 (C-40), 125.5 (C-41), 123.6 (C-12), 121.1 (C-43), 120.1 (C-39), 117.1 (C-36), 78.0 (C-3), 69.7 (C-2), 55.3 (C-5), 48.2 (C-9), 47.6 (C-17), 46.8 (C-19), 42.8 (C-18), 42.5 (C-14), 42.1 (C-1), 39.7 (C-8), 37.5 (C-4), 36.8 (C-10), 34.3 (C-21), 33.1 (C-30), 32.6 (C-22), 32.4 (C-7), 30.9 (C-20), 29.2 (C-24), 27.4 (C-15), 26.0 (C-27), 24.3 (C-16), 23.9 (C-11), 23.8 (C-29), 21.4 (C-32), 21.0 (C-34), 18.0 (C-6), 17.8 (C-23), 17.2 (C-26), 16.2 (C-25) ppm; MS (ESI): m/z (%) = 683.6 ([M+H]⁺, 100), 1365.4 ([2M+H]⁺, 15); analysis calculated for C₄₃H₅₈N₂O₅ (682.93): C 75.62, H 8.56, N 4.10; found: C 75.43, H 8.69, N 3.86.

4.2.16. 2 β , 3 β -Diacetyloxy-ursan-12-en-28-oic acid 7-quinolinyl amide (**18**)

As described for **5**, compound **18** (83 mg, 68%) was obtained from **4** and 7-aminoquinoline as a white solid; m.p. 157–162 °C; R_F = 0.51 (silica gel, chloroform/ethyl acetate, 1:1); $[\alpha]_D^{25} = +24.50^\circ$ ($c = 0.28$, CHCl₃); UV-vis (CHCl₃): λ_{\max} (log ϵ) = 248 nm (4.90), 275 nm (4.24), 325 nm (4.06), 335 nm (4.02); IR (KBr): $\nu = 2948s, 2872m, 1744vs, 1684m, 1624m, 1582w, 1528m, 1496s, 1456s, 1430s, 1364m, 1252vs, 1236s, 1194m, 1156w, 1056m, 1032m$ cm⁻¹; ¹H NMR (500 MHz, CDCl₃): $\delta = 8.86$ (*d*, $J = 3.9$ Hz, 1 H, H-38), 8.09 (*d*, $J = 8.2$ Hz, 1 H, H-42), 8.04 (*d*, $J = 9.2$ Hz, 1 H, H-43), 7.99 (*s*, 1 H, NH), 7.87 (*s*, 1 H, H-36), 7.76 (*d*,

$J = 8.9$ Hz, 1 H, H-40), 7.31 (*dd*, $J = 8.2, 4.2$ Hz, 1 H, H-39), 5.57–5.54 (*m*, 1 H, H-12), 5.33–5.30 (*m*, 1 H, H-2), 4.61 (*d*, $J = 3.8$ Hz, 1 H, H-3), 2.16–1.96 (*m*, 6 H, H-11a + H-11b + H-16a + H-22a + H-1a + H-18), 2.03 (*s*, 3 H, H-32), 2.02 (*s*, 3 H, H-34), 1.92–1.85 (*m*, 1 H, H-16b), 1.78 (*ddd*, $J = 13.6, 13.6, 3.7$ Hz, 1 H, H-15a), 1.69–1.46 (*m*, 6 H, H-6a + H-7a + H-9 + H-19 + H-21a + H-22b), 1.44–1.25 (*m*, 4 H, H-6b + H-21b + H-7b + H-1b), 1.16–1.09 (*m*, 1 H, H-15b), 1.15 (*s*, 3 H, H-27), 1.13 (*s*, 3 H, H-25), 1.06–0.98 (*m*, 7 H, H-20 + H-30 + H-23), 0.98–0.92 (*m*, 1 H, H-5), 0.95 (*d*, $J = 6.3$ Hz, 3 H, H-29), 0.88 (*s*, 3 H, H-24), 0.73 (*s*, 3 H, H-26) ppm; ^{13}C NMR (125 MHz, CDCl_3): $\delta = 176.8$ (C-28), 170.9 (C-33), 170.4 (C-31), 150.9 (C-38), 148.8 (C-37), 140.4 (C-13), 139.7 (C-35), 135.9 (C-42), 128.6 (C-40), 126.3 (C-12), 125.5 (C-41), 121.1 (C-43), 120.1 (C-39), 116.9 (C-36), 78.0 (C-3), 69.6 (C-2), 55.2 (C-5), 54.5 (C-18), 49.0 (C-17), 48.2 (C-9), 43.0 (C-14), 42.2 (C-1), 40.0 (C-19), 39.9 (C-8), 39.3 (C-20), 37.4 (C-4), 37.2 (C-22), 36.8 (C-10), 32.8 (C-7), 31.0 (C-21), 29.3 (C-24), 28.0 (C-15), 25.3 (C-16), 23.8 (C-11), 23.5 (C-27), 21.4 (C-32), 21.3 (C-30), 21.0 (C-34), 18.0 (C-6), 17.8 (C-23), 17.5 (C-29), 17.1 (C-26), 16.4 (C-25) ppm; MS (ESI): m/z (%) = 683.6 ($[\text{M}+\text{H}]^+$, 100), 1365.5 ($[\text{2M}+\text{H}]^+$, 20); analysis calculated for $\text{C}_{43}\text{H}_{58}\text{N}_2\text{O}_5$ (682.93): C 75.62, H 8.56, N 4.10; found: C 75.47, H 8.73, N 3.92.

4.2.17. 2 β , 3 β -Diacetyloxy-olean-12-en-28-oic acid 8-quinolinyl amide (**19**)

As described for **5**, compound **19** (136 mg, 90%) was obtained from **3** and 8-aminoquinoline as a white solid; m.p. 140–145 °C; $R_F = 0.86$ (silica gel, chloroform/ethyl acetate, 1:1); $[\alpha]_D^{25} = +24.59^\circ$ ($c = 0.36$, CHCl_3); UV-vis (CHCl_3): λ_{max} (log ϵ) = 247 nm (4.52), 324 nm (3.70); IR (KBr): $\nu = 3332m, 2950vs, 2866s, 1744vs, 1672s, 1528vs, 1486vs, 1464s, 1424s, 1364s, 1326s, 1250vs, 1192s, 1162s, 1056s, 1030s, 1010s, 944m, 826s, 792s, 756m, 684m, 602m$ cm^{-1} ; ^1H NMR (500 MHz, CDCl_3): $\delta = 10.37$ (*s*, 1 H, NH), 8.85 (*dd*, $J = 7.5, 1.4$ Hz, 1 H, H-43), 8.80 (*dd*, $J = 4.2, 1.6$ Hz, 1 H, H-37), 8.14 (*dd*, $J = 8.3, 1.6$ Hz, 1 H, H-39), 7.53–7.43 (*m*, 3 H, H-42 + H-41 + H-38), 5.71 (*dd*, $J = 3.5, 3.5$ Hz, 1 H, H-12), 5.30 (*ddd*, $J = 3.7, 3.7, 3.7$ Hz, 1 H, H-2), 4.60 (*d*, $J = 3.9$ Hz, 1 H, H-3), 2.99 (*dd*, $J = 13.0, 3.7$ Hz, 1 H, H-18), 2.16 (*ddd*, $J = 13.8, 13.8, 3.9$ Hz, 1 H, H-16a), 2.01 (*s*, 6 H, H-32 + H-34), 2.00–1.68 (*m*, 8 H, H-1a + H-11a + H-11b + H-16b + H-19a + H-22a + H-22b + H-15a), 1.59–1.39 (*m*, 4 H, H-9 + H-6a + H-7a + H-21a), 1.38–1.19 (*m*, 5 H, H-6b + H-19b + H-1b + H-21b + H-7b), 1.20 (*s*, 3 H, H-27), 1.10 (*ddd*, $J = 13.6, 3.3, 3.3$ Hz, 1H, H-15b), 0.98 (*s*, 6 H, H-25 + H-29), 0.97 (*s*, 3 H, H-23), 0.96 (*s*, 3 H, H-30), 0.95–0.91 (*m*, 1 H, H-5), 0.86 (*s*, 3 H, H-24), 0.51 (*s*, 3 H, H-26) ppm; ^{13}C NMR (125 MHz, CDCl_3): $\delta = 177.1$ (C-28), 170.8 (C-33), 170.4 (C-31), 148.0 (C-37), 143.4 (C-13), 139.2 (C-36), 136.3 (C-39), 135.1 (C-35), 128.1 (C-

40), 127.7 (C-42), 124.0 (C-12), 121.6 (C-38), 121.2 (C-41), 116.6 (C-43), 78.1 (C-3), 69.8 (C-2), 55.3 (C-5), 48.2 (C-17), 48.2 (C-9), 46.8 (C-19), 42.3 (C-18), 42.2 (C-14), 42.1 (C-1), 39.7 (C-8), 37.4 (C-4), 36.7 (C-10), 34.5 (C-21), 33.2 (C-30), 33.1 (C-22), 32.5 (C-7), 30.9 (C-20), 29.2 (C-24), 27.6 (C-15), 26.1 (C-27), 24.3 (C-16), 23.8 (C-29), 23.7 (C-11), 21.4 (C-32), 21.0 (C-34), 18.0 (C-6), 17.7 (C-23), 16.6 (C-26), 16.0 (C-25) ppm; MS (ESI): m/z (%) = 683.3 ($[M+H]^+$, 100); analysis calculated for $C_{43}H_{58}N_2O_5$ (682.93): C 75.62, H 8.56, N 4.10; found: C 75.52, H 8.68, N 3.94.

4.2.18. 2β , 3β -Diacetyloxy-ursan-12-en-28-oic acid 8-quinolinyl amide (**20**)

As described for **5**, compound **20** (83 mg, 90%) was obtained from **4** and 8-aminoquinoline as a white solid; m.p. 145–147 °C; R_F = 0.58 (silica gel, hexane/ethyl acetate, 7:3); $[\alpha]_D = -5.00^\circ$ ($c = 0.29$, $CHCl_3$); UV-vis ($CHCl_3$): λ_{max} (log ϵ) = 247 nm (4.51), 324 nm (3.71); IR (KBr): $\nu = 3440m$, $3364m$, $2948s$, $2870m$, $1744vs$, $1672m$, $1596w$, $1578w$, $1526vs$, $1486s$, $1456m$, $1424m$, $1378m$, $1324m$, $1252vs$, $1234s$, $1194m$, $1156m$, $1132w$, $1116w$, $1102w$, $1084w$, $1058m$, $1030m$, $826m$, $792m$, $756m$ cm^{-1} ; 1H NMR (500 MHz, $CDCl_3$): $\delta = 10.30$ (s , 1 H, NH), 8.84–8.82 (m , 1 H, H-43), 8.81–8.79 (m , 1 H, H-37), 8.15 (dd , $J = 8.3, 1.6$ Hz, 1 H, H-39), 7.53–7.49 (m , 1 H, H-42), 7.48–7.44 (m , 2 H, H-41 + H-38), 5.69 (dd , $J = 3.6, 3.6$ Hz, 1 H, H-12), 5.29 (ddd , $J = 3.8, 3.8, 3.5$ Hz, 1 H, H-2), 4.59 (d , $J = 3.9$ Hz, 1 H, H-3), 2.33 (d , $J = 10.8$ Hz, 1 H, H-18), 2.20–2.12 (m , 1 H, H-16a), 2.04–1.80 (m , 6 H, H-1a + H-11a + H-11b + H-15a + H-16b + H-22a), 2.01 (s , 3 H, H-32), 2.00 (s , 3 H, H-36), 1.73–1.65 (m , 1 H, H-22b), 1.62–1.38 (m , 6 H, H-6a + H-7a + H-9 + H-19 + H-21a + H-21b), 1.38–1.28 (m , 2 H, H-1b + H-6b), 1.28–1.22 (m , 1 H, H-7b), 1.15–1.03 (m , 2 H, H-15b + H-20), 1.14 (s , 3 H, H-27), 1.01–0.98 (m , 6 H, H-29 + H-30), 0.96 (s , 6 H, H-23) + H-25), 0.95–0.90 (m , 1 H, H-5), 0.86 (s , 3 H, H-24), 0.51 (s , 3 H, H-26) ppm; ^{13}C NMR (125 MHz, $CDCl_3$): $\delta = 177.0$ (C-28), 170.8 (C-33), 170.4 (C-31), 147.9 (C-37), 139.2 (C-36), 138.0 (C-13), 136.4 (C-39), 135.2 (C-35), 128.1 (C-40), 127.7 (C-42), 127.2 (C-12), 121.5 (C-38), 121.2 (C-41), 116.6 (C-43), 78.1 (C-3), 69.8 (C-2), 55.3 (C-5), 53.9 (C-18), 49.6 (C-17), 48.2 (C-9), 42.5 (C-14), 42.2 (C-1), 40.0 (C-19), 39.9 (C-8), 39.2 (C-20), 37.6 (C-22), 37.4 (C-4), 36.7 (C-10), 32.7 (C-7), 31.2 (C-21), 29.2 (C-24), 28.0 (C-15), 25.3 (C-16), 23.8 (C-27), 23.5 (C-11), 21.4 (C-30 + C-32), 21.0 (C-34), 18.0 (C-6), 17.8 (C-23), 17.5 (C-29), 16.6 (C-26), 16.2 (C-25) ppm; MS (ESI): m/z (%) = 683.7 ($[M+H]^+$, 100); analysis calculated for $C_{43}H_{58}N_2O_5$ (682.93): C 75.62, H 8.56, N 4.10; found: C 75.46, H 8.65, N 3.81.

4.2.19. *2β, 3β-Diacetyloxy-olean-12-en-28-oic acid 4-isoquinolinyl amide (21)*

As described for **5**, compound **21** (71 mg, 58%) was obtained from **3** and 4-aminoisoquinoline as a white solid; m.p. 176–180 °C; $R_F = 0.69$ (silica gel, chloroform/ethyl acetate, 1:1); $[\alpha]_D = +39.41^\circ$ ($c = 0.35$, CHCl_3); UV-vis (CHCl_3): λ_{max} ($\log \epsilon$) = 290 nm (3.75), 302 nm (3.75), 324 nm (3.79); IR (KBr): $\nu = 2948s, 2878m, 1744vs, 1684m, 1648w, 1522m, 1490m, 1474m, 1458m, 1398m, 1366m, 1328w, 1252vs, 1236s, 1194m, 1162w, 1056m, 1030m \text{ cm}^{-1}$; $^1\text{H NMR}$ (500 MHz, CDCl_3): $\delta = 9.13$ (*s*, 1 H, H-36), 9.08 (*s*, 1 H, H-37), 8.02–7.99 (*m*, 2 H, H-39 + NH), 7.79–7.73 (*m*, 2 H, H-42 + H-40), 7.64 (*ddd*, $J = 8.0, 5.8, 2.1$ Hz, 1 H, H-41), 5.63 (*dd*, $J = 3.4, 3.4$ Hz, 1 H, H-12), 5.30 (*ddd*, $J = 3.8, 3.8, 3.8$ Hz, 1 H, H-2), 4.61 (*d*, $J = 3.9$ Hz, 1 H, H-3), 2.88 (*dd*, $J = 12.4, 3.4$ Hz, 1 H, H-18), 2.15 (*ddd*, $J = 13.7, 13.7, 3.9$ Hz, 1 H, H-16a), 2.02 (*s*, 3 H, H-32), 2.02 (*s*, 3 H, H-34), 2.01–1.81 (*m*, 7 H, H-11a + H-11b + H-16b + H-22a + H-22b + H-1a + H-19a), 1.76 (*ddd*, $J = 13.8, 13.8, 3.9$ Hz, 1 H, H-15a), 1.61–1.37 (*m*, 5 H, H-6a + H-6b + H-7a + H-21a + H-9), 1.37–1.27 (*m*, 4 H, H-7b + H-21b + H-1b + H-19b), 1.23 (*s*, 3 H, H-27), 1.19–1.13 (*m*, 1 H, H-15b), 1.07 (*s*, 3 H, H-25), 1.01 (*s*, 6 H, H-23 + H-29), 0.99–0.94 (*m*, 1 H, H-5), 0.98 (*s*, 3 H, H-30), 0.88 (*s*, 3 H, H-24), 0.71 (*s*, 3 H, H-26) ppm; $^{13}\text{C NMR}$ (125 MHz, CDCl_3): $\delta = 177.0$ (C-28), 170.9 (C-33), 170.4 (C-31), 149.4 (C-37), 145.2 (C-13), 137.2 (C-36), 130.6 (C-40), 129.8 (C-35), 128.8 (C-38), 128.5 (C-39), 128.4 (C-43), 127.4 (C-41), 123.6 (C-12), 120.0 (C-42), 78.0 (C-3), 69.7 (C-2), 55.3 (C-5), 48.1 (C-9), 48.1 (C-17), 46.8 (C-19), 43.1 (C-18), 42.6 (C-14), 42.0 (C-1), 39.7 (C-8), 37.5 (C-4), 36.8 (C-10), 34.3 (C-21), 33.1 (C-30), 33.1 (C-22), 32.5 (C-7), 30.9 (C-20), 29.2 (C-24), 27.5 (C-15), 26.1 (C-27), 24.2 (C-16), 23.8 (C-29), 23.7 (C-11), 21.4 (C-32), 21.0 (C-34), 18.0 (C-6), 17.7 (C-23), 17.4 (C-26), 16.1 (C-25) ppm; MS (ESI): m/z (%) = 683.5 ($[\text{M}+\text{H}]^+$, 100), 1365.2 ($[\text{2M}+\text{H}]^+$, 46); analysis calculated for $\text{C}_{43}\text{H}_{58}\text{N}_2\text{O}_5$ (682.93): C 75.62, H 8.56, N 4.10; found: C 75.43, H 8.65, N 3.97.

4.2.20. *3β-Acetyloxy-urs-12-en-28-oic acid (24)*

Compound **24** (192 mg, 88%) was synthesized as described for compound **3** starting from **22**. The spectroscopic data were in accordance with the data from the literature.

4.2.21. *2α, 3β, 24-Triacetyloxy-urs-12-en-28-oic acid (25)*

Compound **25** (209 mg, 83%) was synthesized as described for **3** starting from **23** and obtained as a white solid; m.p. 153–159 °C (lit.[29]: 151.2–154.6 °C); $R_F = 0.23$ (silica gel, hexane/ethyl acetate, 7:3); $[\alpha]_D = +34.71^\circ$ ($c = 0.35$, CHCl_3); IR (KBr): $\nu = 3298m, 2950s, 2872m, 1748vs, 1698s, 1456s, 1370s, 1234vs, 1156m, 1144s, 964m, 918m \text{ cm}^{-1}$; $^1\text{H NMR}$

(500 MHz, CDCl₃): δ = 5.24 (*dd*, J = 3.5, 3.5 Hz, 1 H, H-12), 5.16 (*ddd*, J = 10.9, 10.9, 4.6 Hz, 1 H, H-2), 5.08 (*d*, J = 10.3 Hz, 1 H, H-3), 3.85 (*d*, J = 11.7 Hz, 1 H, H-24a), 3.58 (*d*, J = 11.8 Hz, 1 H, H-24b), 2.19 (*d*, J = 11.3 Hz, 1 H, H-18), 2.11–1.90 (*m*, 4 H, H-11a + H-11b + H-1a + H-16a), 2.08 (*s*, 3 H, H-36), 2.02 (*s*, 3 H, H-34), 1.98 (*s*, 3 H, H-32), 1.84 (*ddd*, J = 13.4, 13.4, 4.1 Hz, 1 H, H-15a), 1.75–1.59 (*m*, 4 H, H-16b + H-22a + H-22b + H-9), 1.54–1.25 (*m*, 8 H, H-6a + H-6b + H-21a + H-21b + H-7a + H-7b + H-5 + H-19), 1.17–0.96 (*m*, 3 H, H-15b + H-1b + H-20), 1.10 (*s*, 3 H, H-25), 1.07 (*s*, 3 H, H-27), 0.94 (*d*, J = 6.3 Hz, 3 H, H-30), 0.87 (*s*, 3 H, H-23), 0.84 (*d*, J = 6.7 Hz, 3 H, H-29), 0.76 (*s*, 3 H, H-26) ppm; ¹³C NMR (125 MHz, CDCl₃): δ = 183.6 (C-28), 171.0 (C-35), 170.6 (C-31), 170.5 (C-33), 138.2 (C-13), 125.4 (C-12), 75.0 (C-3), 70.1 (C-2), 65.4 (C-24), 52.7 (C-18), 48.1 (C-17), 47.8 (C-9), 47.7 (C-5), 43.9 (C-1), 42.1 (C-14), 42.1 (C-4), 39.7 (C-8), 39.2 (C-19), 38.9 (C-20), 38.0 (C-10), 36.8 (C-22), 32.6 (C-7), 30.7 (C-21), 28.0 (C-15), 24.1 (C-16), 23.6 (C-27), 23.5 (C-11), 21.3 (C-30), 21.2 (C-32), 21.0 (C-36), 20.9 (C-34), 18.0 (C-6), 17.2 (C-25), 17.1 (C-29), 17.1 (C-26), 14.1 (C-23) ppm; MS (ESI): m/z (%) = 615.1 ([M+H]⁺, 14), 632.2 ([M+NH₄]⁺, 40), 637.4 ([M+Na]⁺, 100); analysis calculated for C₃₆H₅₄O₈ (614.81): C 70.33, H 8.85; found: C 70.02, H 8.97.

4.2.22. 3 β -Acetyloxy-ursan-12-en-28-oic acid 5-quinolinyl amide (**26**)

As described for **5**, compound **26** (109 mg, 87%) was obtained from **24** and 5-aminoquinoline as a white solid; m.p. 163–165 °C; R_F = 0.53 (silica gel, chloroform/ethyl acetate, 1:1); $[\alpha]_D^{25}$ = +2.98° (c = 0.38, CHCl₃); UV-vis (CHCl₃): λ_{max} (log ϵ) = 241 nm (4.44), 308 nm (3.72), 317 nm (3.72); IR (KBr): ν = 3260 s , 2968 vs , 2926 vs , 2870 s , 1736 vs , 1684 s , 1650 vs , 1596 s , 1520 s , 1490 vs , 1370 s , 1320 s , 1246 vs , 1204 s , 1146 m , 1026 s , 1006 s , 986 s , 968 s , 802 s cm⁻¹; ¹H NMR (500 MHz, CDCl₃): 8.91 (*dd*, J = 4.2, 1.5 Hz, 1 H, H-38), 8.14 (*d*, J = 8.4 Hz, 1 H, H-40), 8.08 (*s*, 1 H, NH), 7.99 (*d*, J = 7.4 Hz, 1 H, H-34), 7.92 (*d*, J = 8.5 Hz, 1 H, H-36), 7.67 (*dd*, J = 8.5, 7.7 Hz, 1 H, H-35), 7.41 (*dd*, J = 8.6, 4.2 Hz, 1 H, H-39), 5.54 (*dd*, J = 3.4, 3.4 Hz, 1 H, H-12), 4.47 (*dd*, J = 11.0, 4.8 Hz, 1 H, H-3), 2.19 (*d*, J = 10.4 Hz, 1 H, H-18), 2.13 (*ddd*, J = 13.6, 13.6, 4.1 Hz, 1 H, H-16a), 2.06–1.98 (*m*, 2 H, H-2a + H-22a), 2.02 (*s*, 3 H, H-32), 1.95–1.79 (*m*, 3 H, H-2b + H-16b + H-15a), 1.72 (*ddd*, J = 13.6, 13.6, 4.2 Hz, 1 H, H-22b), 1.65–1.23 (*m*, 11 H, H-11a + H-11b + H-1a + H-21a + H-21b + H-9 + H-19 + H-7a + H-7b + H-6a + H-6b), 1.17–0.98 (*m*, 1 H, H-15b + H-1b + H-19 + H-20), 1.15 (*s*, 3 H, H-27), 1.01 (*d*, J = 6.2 Hz, 3 H, H-30), 0.96 (*d*, J = 6.5 Hz, 3 H, H-29), 0.84 (*s*, 3 H, H-24), 0.83 (*s*, 3 H, H-25), 0.83–0.78 (*m*, 1 H, H-5), 0.81 (*s*, 3 H, H-23), 0.69 (*s*, 3 H, H-26) ppm; ¹³C NMR (125 MHz, CDCl₃): δ = 176.9 (C-28), 171.1 (C-31), 150.1 (C-38), 148.5 (C-37), 140.2 (C-

13), 133.1 (C-33), 130.0 (C-40), 129.7 (C-35), 126.5 (C-36), 126.4 (C-12), 122.6 (C-41), 120.8 (C-34), 120.7 (C-39), 80.9 (C-3), 55.4 (C-5), 54.6 (C-18), 49.5 (C-17), 47.6 (C-9), 42.8 (C-14), 40.1 (C-19), 39.7 (C-8), 39.3 (C-20), 38.4 (C-1), 37.8 (C-4), 37.7 (C-22), 36.9 (C-10), 32.9 (C-7), 31.0 (C-21), 28.2 (C-24), 28.0 (C-15), 25.2 (C-16), 23.6 (C-11), 23.6 (C-27), 23.5 (C-2), 21.4 (C-30), 21.3 (C-32), 18.2 (C-6), 17.4 (C-29), 17.4 (C-26), 16.8 (C-23), 15.6 (C-25) ppm; MS (ESI): m/z (%) = 625.5 ($[M+H]^+$, 100), 1249.3 ($[2M+H]^+$, 26); analysis calculated for $C_{41}H_{56}N_2O_3$ (624.89): C 78.80, H 9.03, N 4.48; found: C 78.64, H 8.75, N 4.29.

4.2.23. 2 α , 3 β , 24-Triacetyloxy-ursan-12-en-28-oic acid 5-quinolinyl amide (**27**)

As described for **5**, compound **27** (63 mg, 52%) was obtained from **25** and 5-aminoquinoline as a white solid; m.p. 168–172 °C; R_F = 0.49 (silica gel, chloroform/ethyl acetate, 1:1); $[\alpha]_D^{25} = +6.08^\circ$ ($c = 0.31$, $CHCl_3$); UV-vis ($CHCl_3$): λ_{max} ($\log \epsilon$) = 241 nm (4.26), 306 nm (3.53), 317 nm (3.52); IR (KBr): $\nu = 3388w$, 2950s, 2926s, 2870m, 1746vs, 1682m, 1596m, 1572w, 1522m, 1486s, 1370s, 1234vs, 1044s, 800 cm^{-1} ; 1H NMR (500 MHz, $CDCl_3$): $\delta = 8.93$ (dd, $J = 4.2, 1.5$ Hz, 1 H, H-42), 8.12 (d, $J = 8.3$ Hz, 1 H, H-44), 8.02 (d, $J = 7.1$ Hz, 1 H, H-38), 7.98 (s, 1 H, NH), 7.93 (d, $J = 8.5$ Hz, 1 H, H-40), 7.68 (dd, $J = 8.5, 7.7$ Hz, 1 H, H-39), 7.42 (dd, $J = 8.6, 4.2$ Hz, 1 H, H-43), 5.55 (dd, $J = 3.4, 3.4$ Hz, 1 H, H-12), 5.13 (ddd, $J = 10.7, 10.7, 4.5$ Hz, 1 H, H-2), 5.07 (d, $J = 10.3$ Hz, 1 H, H-3), 3.82 (d, $J = 11.8$ Hz, 1 H, H-24a), 3.57 (d, $J = 11.9$ Hz, 1 H, H-24b), 2.21 (d, $J = 10.4$ Hz, 1 H, H-18), 2.14 (ddd, $J = 13.7, 13.7, 4.2$ Hz, 1 H, H-16a), 2.10–1.89 (m, 5 H, H-1a + H-11a + H-11b + H-22a + H-16b), 2.08 (s, 3 H, H-36), 2.01 (s, 3 H, H-34), 1.98 (s, 3 H, H-32), 1.85 (ddd, $J = 14.5, 14.5, 4.4$ Hz, 1 H, H-15a), 1.79–1.66 (m, 2 H, H-22b + H-9), 1.65–1.51 (m, 2 H, H-19 + H-21a), 1.50–1.28 (m, 6 H, H-7a + H-7b + H-21b + H-6a + H-6b + H-5), 1.18–1.12 (m, 3 H, H-15b + H-1b + H-20), 1.17 (s, 3 H, H-27), 1.01 (d, $J = 6.3$ Hz, 3 H, H-30), 1.00 (s, 3 H, H-25), 0.96 (d, $J = 6.5$ Hz, 3 H, H-29), 0.84 (s, 3 H, H-23), 0.70 (s, 3 H, H-26) ppm; ^{13}C NMR (125 MHz, $CDCl_3$): $\delta = 176.7$ (C-28), 170.9 (C-35), 170.6 (C-31), 170.5 (C-33), 150.4 (C-42), 148.8 (C-31), 140.3 (C-13), 132.9 (C-37), 129.6 (C-39), 129.4 (C-44), 126.8 (C-40), 125.9 (C-12), 122.5 (C-45), 120.8 (C-43), 120.6 (C-38), 74.8 (C-3), 70.0 (C-2), 65.4 (C-24), 54.6 (C-18), 49.5 (C-17), 47.7 (C-5), 47.7 (C-9), 43.9 (C-1), 42.8 (C-14), 42.0 (C-4), 40.1 (C-19), 39.8 (C-8), 39.3 (C-20), 37.9 (C-10), 37.8 (C-22), 32.5 (C-7), 31.0 (C-21), 28.0 (C-15), 25.2 (C-16), 23.6 (C-11), 23.5 (C-27), 21.3 (C-30), 21.2 (C-32), 21.0 (C-36), 20.9 (C-34), 17.9 (C-6), 17.4 (C-26), 17.3 (C-29), 17.2 (C-25), 14.0 (C-23) ppm; MS (ESI): m/z (%)

= 741.5 ($[M+H]^+$, 100), 1482.1 ($[2M+H]^+$, 24); analysis calculated for $C_{45}H_{60}N_2O_7$ (740.97): C 72.94, H 8.16, N 3.78; found: C 72.73, H 7.97, N 3.51.

4.2.24. 2α , 3β , 24-Triacetyloxy-ursan-12-en-28-oic acid 4-isoquinolinyl amide (**28**)

As described for **5**, compound **28** (53 mg, 44%) was obtained from **25** and 4-aminoisoquinoline as a white solid; m.p. 173–177 °C; R_F = 0.40 (silica gel, chloroform/ethyl acetate, 1:1); $[\alpha]_D = -0.85^\circ$ ($c = 0.33$, $CHCl_3$); UV-vis ($CHCl_3$): λ_{max} ($\log \epsilon$) = 289 nm (3.68), 301 nm (3.67), 324 nm (3.72); IR (KBr): $\nu = 3396m$, 2948m, 2926m, 2870m, 1746vs, 1684m, 1626w, 1586w, 1522m, 1488m, 1456m, 1370s, 1252vs, 1044s cm^{-1} ; 1H NMR (500 MHz, $CDCl_3$): $\delta = 9.08$ (s, 1 H, H-38), 9.03 (s, 1 H, H-39), 8.00 (d, $J = 8.2$ Hz, 1 H, H-41), 7.91 (m, 1 H, NH), 7.77–7.72 (m, 2 H, H-44 + H-42), 7.63 (ddd, $J = 8.0, 5.9, 2.0$ Hz, 1 H, H-43), 5.56 (dd, $J = 3.4, 3.4$ Hz, 1 H, H-12), 5.14 (ddd, $J = 10.5, 10.5, 4.5$ Hz, 1 H, H-2), 5.07 (d, $J = 10.3$ Hz, 1 H, H-3), 3.82 (d, $J = 11.8$ Hz, 1 H, H-24a), 3.57 (d, $J = 11.8$ Hz, 1 H, H-24b), 2.22 (d, $J = 10.1$ Hz, 1 H, H-18), 2.15 (ddd, $J = 13.6, 13.6, 4.2$ Hz, 1 H, H-16a), 2.10–1.91 (m, 5 H, H-1a + H-11a + H-11b + H-22a + H-16b), 2.08 (s, 3 H, H-36), 2.01 (s, 3 H, H-34), 1.98 (s, 3 H, H-32), 1.86 (ddd, $J = 14.4, 14.4, 4.4$ Hz, 1 H, H-15a), 1.77 (ddd, $J = 13.6, 13.6, 4.3$ Hz, 1 H, H-22b), 1.69 (dd, $J = 11.0, 6.6$ Hz, 1 H, H-9), 1.62 (ddd, $J = 13.3, 6.6, 3.2$ Hz, 1 H, H-21a), 1.59–1.52 (m, 1 H, H-19), 1.50–1.28 (m, 6 H, H-7a + H-7b + H-21b + H-6a + H-6b + H-5), 1.20–1.04 (m, 3 H, H-15b + H-1b + H-20), 1.17 (s, 3 H, H-27), 1.02 (d, $J = 6.3$ Hz, 3 H, H-30), 1.00 (s, 3 H, H-25), 0.97 (d, $J = 6.5$ Hz, 3 H, H-29), 0.85 (s, 3 H, H-23), 0.73 (s, 3 H, H-26) ppm; ^{13}C NMR (125 MHz, $CDCl_3$): $\delta = 176.8$ (C-28), 170.9 (C-35), 170.6 (C-31), 170.5 (C-33), 149.5 (C-39), 140.2 (C-13), 137.6 (C-38), 130.5 (C-42), 130.0 (C-37), 128.9 (C-40), 128.5 (C-41), 128.3 (C-45), 127.4 (C-43), 126.0 (C-12), 120.1 (C-44), 74.9 (C-3), 70.0 (C-2), 65.4 (C-24), 54.7 (C-18), 49.5 (C-17), 47.7 (C-5), 47.7 (C-9), 43.9 (C-1), 42.8 (C-14), 42.1 (C-4), 40.1 (C-19), 39.8 (C-8), 39.3 (C-20), 37.9 (C-10), 37.8 (C-22), 32.5 (C-7), 31.0 (C-21), 28.0 (C-15), 25.2 (C-16), 23.6 (C-11), 23.5 (C-27), 21.3 (C-30), 21.2 (C-32), 21.0 (C-36), 20.9 (C-34), 17.9 (C-6), 17.4 (C-26), 17.4 (C-29), 17.2 (C-25), 14.0 (C-23) ppm; MS (ESI): m/z (%) = 741.5 ($[M+H]^+$, 100), 763.3 ($[M+Na]^+$, 4), 1482.3 ($[2M+H]^+$, 92); analysis calculated for $C_{45}H_{60}N_2O_7$ (740.97): C 72.94, H 8.16, N 3.78; found: C 72.75, H 8.33, N 3.52.

Acknowledgments

Thanks are due to Dr. R. Kluge for measuring the ESI-MS spectra, and to Dr. D. Ströhl and his team for many NMR spectra. We like to thank J. Wiese, MSc, for measuring optical rotations, UV-vis and IR spectra, and Ms J. Pech for the microanalyses. The cell lines were kindly provided by Dr. Thomas Müller (Dept. of Haematology/Oncology, Martin-Luther Universität Halle-Wittenberg). Support by the “WissenschaftsCampus Halle WCH” is gratefully recognized.

References

- [1] C. Fitzmaurice, D. Dicker, A. Pain, H. Hamavid, M. Moradi-Lakeh, M.F. MacIntyre, C. Allen, G. Hansen, R. Woodbrook, C. Wolfe, R.R. Hamadeh, A. Moore, A. Werdecker, B.D. Gessner, B. Te Ao, B. McMahon, C. Karimkhani, C. Yu, G.S. Cooke, D.C. Schwebel, D.O. Carpenter, D.M. Pereira, D. Nash, D.S. Kazi, D. De Leo, D. Plass, K.N. Ukwaja, G.D. Thurston, K. Yun Jin, E.P. Simard, E. Mills, E.-K. Park, F. Catala-Lopez, G. deVeber, C. Gotay, G. Khan, H.D. Hosgood, 3rd, I.S. Santos, J.L. Leasher, J. Singh, J. Leigh, J. Jonas, J. Sanabria, J. Beardsley, K.H. Jacobsen, K. Takahashi, R.C. Franklin, L. Ronfani, M. Montico, L. Naldi, M. Tonelli, J. Geleijnse, M. Petzold, M.G. Shrimel, M. Younis, N. Yonemoto, N. Breitborde, P. Yip, F. Pourmalek, P.A. Lotufo, A. Esteghamati, G.J. Hankey, R. Ali, R. Lunevicius, R. Malekzadeh, R. Dellavalle, R. Weintraub, R. Lucas, R. Hay, D. Rojas-Rueda, R. Westerman, S.G. Sepanlou, S. Nolte, S. Patten, S. Weichenthal, S.F. Abera, S.-M. Fereshtehnejad, I. Shiue, T. Driscoll, T. Vasankari, U. Alsharif, V. Rahimi-Movaghar, V.V. Vlassov, W.S. Marcenés, W. Mekonnen, Y.A. Melaku, Y. Yano, A. Artaman, I. Campos, J. MacLachlan, U. Mueller, D. Kim, M. Trillini, B. Eshrati, H.C. Williams, K. Shibuya, R. Dandona, K. Murthy, B. Cowie, A.T. Amare, C.A. Antonio, C. Castaneda-Orjuela, C.H. van Gool, F. Violante, I.-H. Oh, K. Deribe, K. Soreide, L. Knibbs, M. Kereselidze, M. Green, R. Cardenas, N. Roy, T. Tillman, Y. Li, H. Krueger, L. Monasta, S. Dey, S. Sheikhabaei, N. Hafezi-Nejad, G.A. Kumar, C.T. Sreeramareddy, L. Dandona, H. Wang, S.E. Vollset, A. Mokdad, J.A. Salomon, R. Lozano, T. Vos, M. Forouzanfar, A. Lopez, C. Murray, M. Naghavi, The Global Burden of Cancer 2013, *JAMA Oncol.* 1 (2015) 505-527.
- [2] J. Reedijk, New clues for platinum antitumor chemistry: kinetically controlled metal binding to DNA, *Proc. Natl. Acad. Sci. USA* 100 (2003) 3611-3616.

- [3] E. Wong, C.M. Giandomenico, Current status of platinum-based antitumor drugs, *Chem. Rev.* 99 (1999) 2451-2466.
- [4] J. Rautio, H. Kumpulainen, T. Heimbach, R. Oliyai, D. Oh, T. Jarvinen, J. Savolainen, Prodrugs: design and clinical applications, *Nature reviews. Drug discovery* 7 (2008) 255-270.
- [5] G. Villaverde, A. Baeza, G.J. Melen, A. Alfranca, M. Ramirez, M. Vallet-Regi, A new targeting agent for the selective drug delivery of nanocarriers for treating neuroblastoma, *J. Mater. Chem. B* 3 (2015) 4831-4842.
- [6] F. Marcucci, F. Lefoulon, Active targeting with particulate drug carriers in tumor therapy: fundamentals and recent progress, *Drug Discov. Today*. 9 (2004) 219-228.
- [7] C. Mari, V. Pierroz, S. Ferrari, G. Gasser, Combination of Ru(II) complexes and light: new frontiers in cancer therapy, *Chem. Sci.* 6 (2015) 2660-2686.
- [8] A. Gandioso, E. Shaili, A. Massaguer, G. Artigas, A. Gonzalez-Canto, J.A. Woods, P.J. Sadler, V. Marchan, An integrin-targeted photoactivatable Pt(IV) complex as a selective anticancer pro-drug: synthesis and photoactivation studies, *Chem. Commun.* 51 (2015) 9169-9172.
- [9] S. Moktan, C. Ryppa, F. Kratz, D. Raucher, A thermally responsive biopolymer conjugated to an acid-sensitive derivative of paclitaxel stabilizes microtubules, arrests cell cycle, and induces apoptosis, *Invest. New Drugs* 30 (2012) 236-248.
- [10] N.S. Abadeer, C.J. Murphy, Recent Progress in Cancer Thermal Therapy Using Gold Nanoparticles, *J. Phys. Chem. C* 120 (2016) 4691-4716.
- [11] Y.-J. Li, M. Dong, F.-M. Kong, J.-P. Zhou, Folate-decorated anticancer drug and magnetic nanoparticles encapsulated polymeric carrier for liver cancer therapeutics, *Int. J. Pharm.* 489 (2015) 83-90.
- [12] W. Kai, X. Xiaojun, P. Ximing, H. Zhenqing, Z. Qiqing, Cytotoxic effects and the mechanism of three types of magnetic nanoparticles on human hepatoma BEL-7402 cells, *Nanoscale Res. Lett.* 6 (2011) 480.
- [13] P. Skehan, R. Storeng, D. Scudiero, A. Monks, J. McMahon, D. Vistica, J.T. Warren, H. Bokesch, S. Kenney, M.R. Boyd, New colorimetric cytotoxicity assay for anticancer-drug screening, *J. Natl. Cancer Inst.* 82 (1990) 1107-1112.
- [14] B. Siewert, E. Pianowski, A. Obernauer, R. Csuk, Towards cytotoxic and selective derivatives of maslinic acid, *Bioorg. Med. Chem.* 22 (2014) 594-615.

- [15] S. Sommerwerk, L. Heller, I. Serbian, R. Csuk, Straightforward partial synthesis of four diastereomeric 2,3-dihydroxy-olean-12-en-28-oic acids from oleanolic acid, *Tetrahedron* 71 (2015) 8528-8534.
- [16] S. Sommerwerk, R. Csuk, Convenient and chromatography-free partial syntheses of maslinic acid and augustic acid, *Tetrahedron Lett.* 55 (2014) 5156-5158.
- [17] X.A. Wen, H.B. Sun, J. Liu, K.G. Cheng, P. Zhang, L.Y. Zhang, J. Hao, L.Y. Zhang, P.Z. Ni, S.E. Zographos, D.D. Leonidas, K.M. Alexacou, T. Gimisis, J.M. Hayes, N.G. Oikonomakos, Naturally occurring pentacyclic triterpenes as inhibitors of glycogen phosphorylase: Synthesis, structure-activity relationships, and X-ray crystallographic studies, *J. Med. Chem.* 51 (2008) 3540-3554.
- [18] X.A. Wen, J. Xia, K.G. Cheng, L.Y. Zhanga, P. Zhanga, J. Liu, L.Y. Zhang, P.Z. Nia, H.B. Sun, Pentacyclic triterpenes. Part 5: Synthesis and SAR study of corosolic acid derivatives as inhibitors of glycogen phosphorylases, *Bioorg. Med. Chem. Lett.* 17 (2007) 5777-5782.
- [19] Y.Y. Ma, W. Li, B. Yu, Regio-selective Dehydrogenation on the D or E Rings of Oleanolic Acid by Pd-Promoted C-H Activation, *Acta Chim. Sinica* 71 (2013) 541-548.
- [20] O.B. Flekhter, E.I. Boreko, L.R. Nigmatullina, E.V. Tret'yakova, N.I. Pavlova, L.A. Baltina, S.N. Nikolaeva, O.V. Savinova, V.F. Eremin, F.Z. Galin, G.A. Tolstikov, Synthesis and antiviral activity of betulonic acid amides and conjugates with amino acids, *Russ. J. Bioorg. Chem.* 30 (2004) 80-88.
- [21] J.W. Shao, Y.C. Dai, J.P. Xue, J.C. Wang, F.P. Lin, Y.H. Guo, In vitro and in vivo anticancer activity evaluation of ursolic acid derivatives, *Eur. J. Med. Chem.* 46 (2011) 2652-2661.
- [22] Y.Q. Meng, Y.L. Song, Z.K. Yan, Y. Xia, Synthesis and in vitro Cytotoxicity of Novel Ursolic Acid Derivatives, *Molecules*, 15 (2010) 4033-4040.
- [23] Y.Q. Meng, D. Liu, L.L. Cai, H. Chen, B. Cao, Y.Z. Wang, The synthesis of ursolic acid derivatives with cytotoxic activity and the investigation of their preliminary mechanism of action, *Bioorg. Med. Chem.* 17 (2009) 848-854.
- [24] C.M. Ma, S.Q. Cai, J.R. Cui, R.Q. Wang, P.F. Tu, M. Hattori, M. Daneshtalab, The cytotoxic activity of ursolic acid derivatives, *Eur. J. Med. Chem.* 40 (2005) 582-589.
- [25] S.X. Hua, R.Z. Huang, M.Y. Ye, Y.M. Pan, G.Y. Yao, Y. Zhang, H.S. Wang, Design, synthesis and in vitro evaluation of novel ursolic acid derivatives as potential anticancer agents, *Eur. J. Med. Chem.* 95 (2015) 435-452.

- [26] X. Yang, Y.F. Li, W. Jiang, M.R. Ou, Y.L. Chen, Y. Xu, Q. Wu, Q. Zheng, F.Q. Wu, L. Wang, W.T. Zou, Y.T.J. Zhang, J.W. Shao, Synthesis and Biological Evaluation of Novel Ursolic acid Derivatives as Potential Anticancer Prodrugs, *Chem. Biol. Drug Des.* 86 (2015) 1397-1404.
- [27] K. Liu, P.C. Liu, R. Liu, X. Wu, Dual AO/EB staining to detect apoptosis in osteosarcoma cells compared with flow cytometry, *Medic. Sci. Mon. Basic Res.* 21 (2015) 15-20.
- [28] M.S. Alam, N. Chopra, M. Ali, M. Niwa, Oleanen and stigmasterol derivatives from *Ambroma augusta*, *Phytochemistry*, 41 (1996) 1197-1200.
- [29] J.F. Li, R.Z. Huang, G.Y. Yao, M.Y. Ye, H.S. Wang, Y.M. Pan, J.T. Xiao, Synthesis and biological evaluation of novel aniline-derived asiatic acid derivatives as potential anticancer agents, *Eur. J. Med. Chem.* 86 (2014) 175-188.

- Amide derivatives of augustic, 2-epi-corosolic and asiatic acid were synthesized
- They were tested for their antitumor activity using human cancer cell lines
- An augustic acid derived 4-isoquinoliny amide showed increased cytotoxicity
- A 4-isoquinoliny derivative of asiatic acid (**28**) gave $EC_{50} = 80$ nM (A2780 cells)
- The compounds act by apoptosis

ACCEPTED MANUSCRIPT