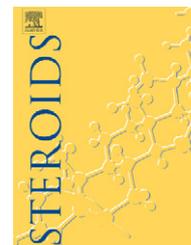


available at www.sciencedirect.comjournal homepage: www.elsevier.com/locate/steroids

Synthesis of an isomeric mixture (24RS,25RS) of sodium scymnol sulfate

Donald W. Harney, Theodore A. Macrides*

Natural Products Research Group, School of Medical Sciences, RMIT University, PO Box 71, Bundoora 3083, Victoria, Australia

ARTICLE INFO

Article history:

Received 30 August 2007

Received in revised form

29 November 2007

Accepted 6 December 2007

Published on line 15 December 2007

Keywords:

Sodium scymnol sulfate

Cholic acid

Shark bile

Chemical synthesis

ABSTRACT

This is the first reported multistep synthesis of the shark bile sterol sodium scymnol sulfate epimeric at the C-24 hydroxyl and C-27 sulfate positions. The starting cholic acid was protected as the tetrahydropyran ether (THP) derivative, reduced to the C-24 alcohol and oxidized to the protected aldehyde. This aldehyde was then coupled with methyl 3-hydroxypropionate using 2 equiv. of lithium diethylamide at -65°C to produce methyl (24RS,25RS)-24,27-dihydroxy-3 α ,7 α ,12 α ,tris[(tetrahydropyran-2-yl)oxy]-5 β -cholestan-26-oate. After protecting the 24 and 27 hydroxyls as the THP derivatives, this fully protected ester was then reduced to the monoalcohol. The monoalcohol was sulfated using the sulfur trioxide–triethylamine complex in dimethylformamide. The protective THP groups were removed with methanolic HCl and the sulfate was converted to the sodium salt with sodium ethoxide in methanol. This general synthetic scheme has application to produce a range of monosulfated sterols.

© 2007 Elsevier Inc. All rights reserved.

1. Introduction

The natural sodium scymnol sulfate has been purified by our group from shark bile by preparative HPLC for use in commercial skin care products [1]. We have investigated many of the wide-ranging biological properties of scymnol and its sulfate including its potent antioxidant [2,3] and hepatoprotective activities [4,5]. Ishida et al. have also investigated its protective properties against vascular endothelial cell injury [6–8].

The sodium scymnol sulfate isolated from shark bile contains the 24R alcohol substituent and either or both of the 27R and 27S sulfate esters [9] (see Fig. 1). Thus, as a further progression in the potential replacement of the natural product with a synthetic product, we designed the synthetic scheme shown in Fig. 2 to allow monosulfation at the C-27 position. This scheme leads to the (24RS,25RS) epimeric mixture of alcohols whereas the natural product comprises the (24R,24R)

and/or (24R,25S) epimers. Our group had synthesized epimeric unsulfated (24RS) scymnol [10] previously. (24R) scymnol was synthesized subsequently by Adhikari et al. [11]. However a sulfated form of scymnol had not been previously synthesized.

2. Experimental

2.1. General

^1H NMR were carried out on samples purified from the crude reaction products by silica gel column chromatography using Bruker 300 and 400 MHz Avance Spectrometers using deuteriochloroform (or other indicated solvents) as the reference or an internal deuterium lock. The multiplicity of the signals is indicated as s=singlets, m=multiplet and br=broad. ^{13}C NMR spectra were recorded on an Avance 400 MHz instrument using an internal deuterium lock and

* Corresponding author. Tel.: +61 3 99257070; fax: +61 3 99257063.

E-mail address: macrides@rmit.edu.au (T.A. Macrides).

0039-128X/\$ – see front matter © 2007 Elsevier Inc. All rights reserved.

doi:10.1016/j.steroids.2007.12.006

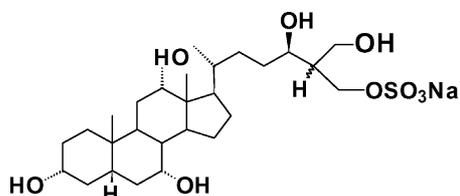


Fig. 1 – (24R,25RS) sodium scymnol sulfate.

proton decoupling. The chemical shifts are given as δ values (ppm) and referenced to tetramethylsilane for deuteriochloroform and 2,2,3,3-d(4)-3-(trimethylsilyl)propionic acid sodium salt (TSP) for deuterium oxide (D_2O) and deuteromethanol (CD_3OD) as solvent. High resolution mass spectroscopy was carried out using a Bruker BioApex II FTICR electrospray in positive ion mode using ammonium acetate in acetonitrile and water mixture. Microanalysis was performed by the University of Otago, Dunedin New Zealand. Analytical TLC was carried out using precoated Merck Kieselgel 60 F₂₅₄ silica gel aluminium plates from E. Merck (Darmstadt, Germany). Visualization was performed by spraying with phosphomolybdic acid (5%, w/v) and sulfuric acid in glacial acetic acid (7.5%, w/v) followed by heating for 5 min at 110°C. Column chromatography was performed with Merck Kieselgel 60 (230–400 mesh). Melting points are uncorrected. Reagents were purified by standard techniques. Tetrahydrofuran (THF) and diethyl ether (ether) were dried with sodium using benzophenone ketyl as the indicator. Petroleum spirits refers to the fraction boiling in the range 40–60°C. The 3,4-dihydro-2H-pyran (DHP) was freshly distilled before

use. All reagents were analytical grade and purchased from regular commercial sources. Cholic acid (3 α ,7 α ,12 α -5 β -cholan-24-oic acid) $\geq 99\%$ was obtained from Fluka BioChemika (Buchs, Switzerland). The sulfur trioxide–triethylamine complex was prepared by the method of Tserng and Klein [12].

2.2. Synthesis

2.2.1. Tetrahydropyran-2-yl-

3 α ,7 α ,12 α ,tris[(tetrahydropyran-2-yl)oxy]-5 β -cholan-24-oate (compound 2, Fig. 2)

A mixture of cholic acid (16.42 g, 40 mmol) and *p*-toluenesulfonic acid (40 mg) in DCM (100 ml) was magnetically stirred for 10 min at 22°C and then 3,4-dihydro-2H-pyran (17.5 g, 208 mmol, 1.3 equiv.) added in stages over 30 min. An exotherm raised the temperature to 39.5°C and the insoluble cholic acid dissolved during the reaction. The reaction returned to 22°C and after a further 3 h of stirring the crude product 2 was concentrated in a rotary evaporator under vacuum to yield a colourless oily residue (37.1 g) that was used in the next reaction after 1.5 h; R_f 0.5 (ethyl acetate/petroleum spirits = 1/3). The product on standing yielded a second main spot at R_f 0.15—presumably due to equilibration between the eight possible tetrahydropyran (THP) acetals. ¹H NMR ($CDCl_3$) 0.63–0.75 (3H, m, H-18), 0.86–1.07 (6H, m, H-19, H-21), 3.32–4.22 (11H, m, H-3 β , H-7 β , H-12 β , 4CH₂O), 4.54–5.01 (4H, m, 4CHO). Positive ion high resolution MS: calculated for C₄₄H₇₂O₉Na [$M + Na^+$]: 767.5074. Found: 767.5077.

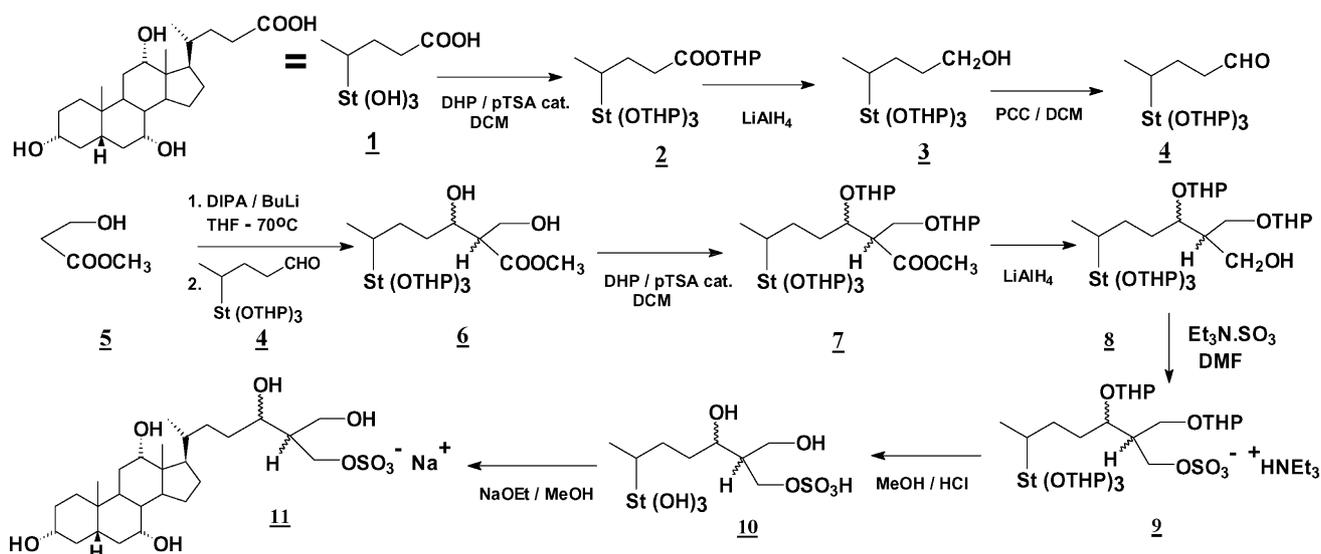


Fig. 2 – (1) Cholic acid; (2) tetrahydropyran-2-yl-3 α ,7 α ,12 α ,tris[(tetrahydropyran-2-yl)oxy]-5 β -cholan-24-oate; (3) 3 α ,7 α ,12 α ,tris[(tetrahydropyran-2-yl)oxy]-5 β -cholan-24-ol; (4) 3 α ,7 α ,12 α ,tris[(tetrahydropyran-2-yl)oxy]-5 β -cholan-24-al; (5) methyl 3-hydroxypropionate; (6) methyl 24,26-dihydroxy-3 α ,7 α ,12 α ,tris[(tetrahydropyran-2-yl)oxy]-5 β -cholestan-26-oate; (7) methyl 3 α ,7 α ,12 α ,24,26-penta[(tetrahydropyran-2-yl)oxy]-5 β -cholestan-26-oate; (8) 3 α ,7 α ,12 α ,24,26-penta[(tetrahydropyran-2-yl)oxy]-5 β -cholestan-26-ol; (9) (24RS,25RS)-3 α ,7 α ,12 α ,24,26-penta[(tetrahydropyran-2-yl)oxy]-5 β -cholestan-27-yl triethylammonium sulfate; (10) (24RS,25RS)-3 α ,7 α ,12 α ,24,26-pentahydroxy-5 β -cholestan-27-yl sulfonic acid; (11) (24RS,25RS)-3 α ,7 α ,12 α ,24,26-pentahydroxy-5 β -cholestan-27-yl sodium sulfate.

2.2.2. $3\alpha,7\alpha,12\alpha$,Tris[(tetrahydropyran-2-yl)oxy]-5 β -cholan-24-ol (compound 3,

Fig. 2)

A dry flask was charged with dry ether (120 ml) and dry THF (25 ml) containing crude compound 2 (37.1 g). The mixture at 22 °C was mechanically stirred under dry nitrogen and LiAlH₄ (50 ml, 1 M in ether) added over 10 min. During this time the spontaneous reflux was moderated with room temperature water bath cooling. After further reflux heating of 1.5 h the resulting clear solution was cooled and ethyl acetate (17.5 ml) was added over 7 min. Reflux heating was recommenced for a further 15 min, after which water (3.5 ml) was added dropwise. The reaction was cooled to room temperature and the resulting precipitate was stirred for another 30 min. The reaction mixture was filtered through a bed of Celite and washed with ether (300 ml). The crude product was concentrated in a rotary evaporator under vacuum, with toluene (6 ml) being added towards the end to azeotrope out residual water. The crude title compound 3 was isolated as a colourless oil (27.2 g). *R*_f 0.4 (ethyl acetate/petroleum spirits = 4/6); ¹H NMR (CDCl₃) 0.62–0.75 (3H, m, H-18), 0.84–1.05 (6H, m, H-19, H-21), 3.33–4.23 (11H, m, H-3 β , H-7 β , H-12 β , 2H-24, 3CH₂O), 4.55–4.97 (3H, m, OCHO).

2.2.3. $3\alpha,7\alpha,12\alpha$,Tris[(tetrahydropyran-2-yl)oxy]-5 β -cholan-24-al (compound 4,

Fig. 2)

A mixture of of pyridinium chlorochromate (15.09 g, 70 mmol) and anhydrous sodium acetate (1.15 g, 14 mmol) in DCM (80 ml) was mechanically stirred at 22 °C while crude oil 3 (27.2 g) in DCM (10 ml) was added in one lot. After an initial exotherm, the reaction returned to 22 °C and after 3.5 h dry ether (80 ml) was added, stirring continued for another 30 min and finally the solvent was decanted. The resulting tarry residue was triturated with dry ether (2 × 20 ml) and the solvent washes decanted. The reaction mixture was concentrated in a rotary evaporator under vacuum to yield the crude product (31.4 g). Purification of the crude product by column chromatography with silica gel using stepwise elution with ether in petroleum spirits increasing from 5 to 50%, yielded a colourless oily residue of compound 4 (11.12 g, 17.24 mmol) after rotary evaporation under vacuum. This product represented a yield of 46.6% from cholic acid. *R*_f 0.5 (ethyl acetate/petroleum spirits = 1/3); ¹H NMR (CDCl₃) 0.61–0.72 (3H, m, H-18), 0.81–1.05 (6H, m, H-19, H-21), 3.34–4.20 (9H, m, H-3 β , H-7 β , H-12 β , 3CH₂O), 4.59–4.81 (3H, m, OCHO), 9.75 (1H, s, H-24).

2.2.4. Methyl-(24RS,25RS)-24,27-dihydroxy- $3\alpha,7\alpha,12\alpha$,tris[(tetrahydropyran-2-yl)oxy]-5 β -cholestan-26-oate (compound 6,

Fig. 2)

A solution of diisopropylamine (3.83 g, 37.8 mmol) in dry THF (100 ml) was mechanically stirred under dry nitrogen. After cooling to –65 °C in dry ice/ethanol, 1.6 M butyl lithium (23.7 ml, 37.8 mmol) in hexane was added over 3 min. The reaction was warmed to –20 °C and held for 20 min before recooling to –65 °C. Methyl-3-hydroxypropionate 5 (1.975 g, 1.82 mmol) in THF (3 ml) was added over 12 min. The reaction was warmed to –20 °C and held for 10 min before recool-

ing to –65 °C. Toluene (5 ml) was added to the protected aldehyde 4 (11.12 g, 17.2 mmol) and distilled in a rotary evaporator under vacuum in order to remove any residual water or DCM, which would react with lithium diethylamide. The protected aldehyde 4 in THF (12 ml) was added to the lithiated methyl-3-hydroxypropionate solution over a period of 30 min at –65 °C. After 2 h the reaction mixture was removed from the cooling bath and ether (55 ml) and 20% aqueous ammonium chloride (35 ml) were added. The mixture was warmed to room temperature and the organic layer was separated. The aqueous layer was washed with ether (60 and 30 ml) and the combined organic layer and ether extracts were dried (Na₂SO₄). The solution was rotary evaporated under vacuum to give 15.9 g of the crude product. Purification of the crude product by silica column chromatography using a stepwise elution with DCM/petroleum spirits (3/1) and increasing in stages to 100% DCM gave the title compound 6. A second chromatography was required on some impure fractions. Following rotary evaporation under vacuum, the title compound 6 (4.515 g, 6.027 mmol) was isolated as a colourless oil. This corresponded to a yield of 35% from purified aldehyde 4 and 16.3% from cholic acid. *R*_f 0.25 (ether/DCM = 3/2); ¹H NMR (CDCl₃) 0.60–0.73 (3H, m, H-18), 0.82–1.04 (6H, m, H-19, H-21), 2.51–2.60 and 2.63–2.70 (1H, dm, HC–COO), 3.35–3.56 (H-3 β , 3OCHax), 3.56–3.67 (1H, br, H-7 β), 3.67–3.81 (4H, H-12 β , CH₃) 3.81–4.20 (6H, m, 3OCHeq, CH₂OH, CHO), 4.57–4.83 (3H, m, OCHO). MS: calculated for C₄₃H₇₆O₁₀N [M + NH₄⁺]: 766.5464. Found: 766.5501.

2.2.5. Methyl-(24RS,25RS)- $3\alpha,7\alpha,12\alpha,24,27$ -penta[(tetrahydropyran-2-yl)oxy]-5 β -cholestan-26-oate (compound 7, Fig. 2)

Compound 6 (7.46 g, 8.13 mmol) and *p*-toluenesulfonic acid (45 mg) in DCM (70 ml) was magnetically stirred while DHP (2.04 g, 24.2 mmol) was added over 5 min resulting in a 5 °C exotherm. The reaction was allowed to proceed for a further 3 h after which the reaction mixture was concentrated in a rotary evaporator under vacuum to give the title compound 7 (9.69 g) as a crude colourless oil. *R*_f 0.75 (ether/DCM = 1/4); ¹H NMR (CDCl₃) 0.60–0.73 (3H, m, H-18), 0.82–1.04 (6H, m, H-19, H-21), 2.20–2.40 (1H, m, H-25), 3.36–3.58 (6H, m, H-3 β , 5OCHax), 3.58–3.66 (1H, m H-7 β), 3.66–3.80 (4H, m, H-12 β , CH₃), 3.80–4.18 (8H, CHOTHP, CH₂OTHP, 5OCHeq), 4.54–4.84 (5H, m, OCHO). MS: calculated for C₅₃H₉₂O₁₂N [M + NH₄⁺]: 934.6614. Found: 934.6590.

2.2.6. (24RS,25RS)- $3\alpha,7\alpha,12\alpha,24,27$ -Penta[(tetrahydropyran-2-yl)oxy]-5 β -cholestan-26-ol (compound 8, Fig. 2)

Crude fully protected ester 7 (9.69 g, 8.13 mmol) in dry ether (150 ml) was mechanically stirred under dry nitrogen and LiAlH₄ (12.9 ml of 1 M in ether) was added over 3 min and the reaction mixture exotherm resulted in mild reflux. After a further 2 h reflux the solution was cooled to room temperature and ethyl acetate (5 ml) was added over a 2-min period. The reaction mixture was refluxed for a further 10 min, and cooled again to room temperature after which water (1.3 ml) was added dropwise and a precipitate formed. The reaction mixture was stirred for a further 30 min and filtered through a bed of Celite, washing with ether (200 ml). The combined

ether solution was concentrated in a rotary evaporator under vacuum to give the crude title compound **8** (9.4 g). Purification by silica column chromatography using a stepwise elution with DCM/petroleum spirits (1/1–0/1) immediately followed by ether/petroleum spirits (15/75 and 1/3) gave pure **8** (6.33 g, 6.90 mmol) as a colourless oil. This represented a yield of 77.6% based on compound **6**. R_f 0.3 (ether/DCM = 3/7); $^1\text{H NMR}$ (CDCl_3) 0.60–0.73 (3H, m, H-18), 0.81–1.08 (6H, m, H-19, H-21), 3.34–3.58 (H-3 β , 5OCHax), 3.58–3.69 (1H br, H-7 β), 3.69–3.81 (3H H-12 β , CH_2OH), 3.81–4.18 (8H, m, CHOTHP, CH_2OTHP , 5OCHeq), 4.44–4.85 (5H, m, OCHO). MS: calculated for $\text{C}_{52}\text{H}_{92}\text{O}_{11}\text{N}$ [$\text{M} + \text{NH}_4^+$]: 906.6665. Found: 906.6625.

2.2.7. (24RS,25RS)-3 α ,7 α ,12 α ,24,26-Penta[(tetrahydropyran-2-yl)oxy]-5 β -cholestan-27-yl triethylammonium sulfate (compound **9**, Fig. 2)

Compound **8** (4.40 g, 4.80 mmol) in dry dimethylformamide (DMF) (15 ml) was warmed to obtain solution. The solution was cooled to room temperature and sulfur trioxide–triethylamine complex (1.28 g, 7.06 mmol, 1.47 equiv.) was added and magnetically stirred for 3.5 h after which time methanol (1.5 ml) was added and the reaction allowed to proceed with stirring for a further 30 min to react excess reagent. The solution was concentrated in a rotary evaporator under vacuum to remove DMF and the resulting crude oil of title compound **9** (6.35 g) was used immediately in the next step. An attempt to purify this compound in a previous preparation led to partial deprotection as indicated by a reduction of the acetal protons by NMR analysis. It is proposed that the THP-protected triethylammonium alkyl sulfate **9** acted as a mild acidic catalyst during chromatography with methanol and DCM causing partial deprotection, whereas an NMR of the crude compound **9** showed the required five acetal protons indicating no deprotection. MS on the crude product: found mass of 986.6160 which corresponded to [$\text{M} - \text{Et}_3\text{N} + \text{NH}_4^+$] calc. 986.6239; R_f 0.75 (BuOH/ $\text{CH}_3\text{COOH}/\text{H}_2\text{O} = 8/1/1$).

2.2.8. (24RS,25RS)-3 α ,7 α ,12 α ,24,26-Pentahydroxy-5 β -cholestan-27-yl sodium sulfate (compound **11**, Fig. 2)

2.2.8.1. Deprotection to (24RS,25RS)-3 α ,7 α ,12 α ,24,26-pentahydroxy-5 β -cholestan-27-yl sulfonic acid (compound **10**, Fig. 1). The crude compound **9** (6.35 g) was dissolved in methanol (100 ml) and stirred magnetically at room temperature. One molar HCl was freshly prepared from 8.3 ml of 38% aqueous HCl and made up to 100 ml with methanol. Addition of the 1 M HCl solution (17 ml, 17 mmol) was completed over a 5-min period. The reaction was allowed to proceed at room temperature and the deprotection was carefully monitored by TLC. The deprotection was complete after 3 h and showed one main spot on TLC at R_f 0.4 (BuOH/ $\text{CH}_3\text{COOH}/\text{H}_2\text{O} = 8/1/1$). The resulting deprotected sulfonic acid compound **10** as shown in Fig. 2 was then converted at room temperature to the sodium salt by the addition of sodium ethoxide (1.80 g, 26.4 mmol) in methanol (15 ml) that was added over a 2-min reaction period with constant stirring. A small drop of the reaction mixture added to a drop of water indicated a pH 10 had been reached. Gaseous CO_2 was bubbled through the reaction mixture for 15 min and this resulted in a drop to pH 8. The reaction mixture was concentrated in a rotary evaporator under vacuum

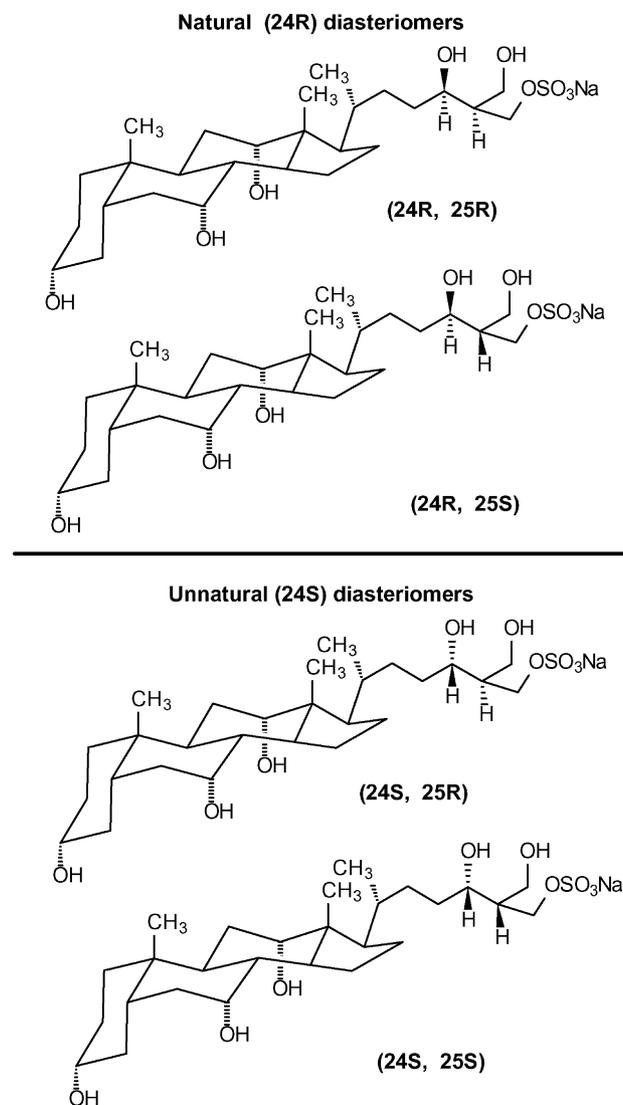


Fig. 3 – The four stereoisomers for the synthetic compound **11** of Fig. 2.

to a low volume and silica gel (25 ml) was added. Further evaporation of methanol gave a dry powder, which was added to the top of a large chromatographic column of silica gel in methanol (5%, v/v) in DCM. Fractions were collected while increasing the methanol in a stepwise manner from 5 to 10, 15 to 20, 25 and finally 30 and 50% in DCM. The title compound **11** (2.13 g, 3.74 mmol) was collected and the yield was 78% based on the purified protected monoalcohol compound **8**. R_f 0.3 (BuOH/ $\text{CH}_3\text{COOH}/\text{H}_2\text{O} = 8/1/1$); calculated for $\text{C}_{27}\text{H}_{47}\text{O}_9\text{SNa}$: C, 56.8; H, 8.3; S, 5.6; Na, 4.0. Found: C, 55.6; H, 8.3; S, 5.6; Na, 3.9; $^1\text{H NMR}$ (CD_3OD) 0.71 (3H, s, H-18), 0.92 (3H, m, H-19), 1.0–1.06 (3H, m, H-21), 2.18–2.36 (2H, m, H-4 and 9), 3.27–3.43 (m, H-3 β + CH_3OH), 3.43–3.66 (3H, m, H-24, H-26), 3.79 (1H, m, H-7 β), 3.97 (1H, m, H-12 β), 4.07–4.17 (1.5H, m, H-27), 4.17–4.22 (0.5H, m, H-27); $^{13}\text{C NMR}$ (CD_3OD) 15.28 (C18), 20.07, 20.19 (C21), 25.36 (C19), 26.10 (C15), 29.24 (C9), 30.46 (C16), 30.86 (C11), 32.15, 32.77, 32.96 (C23), 33.21 (C2), 34.46, 34.68 (C22), 37.00 (C6), 37.41 (C10), 38.13 (C1), 38.75 (C20), 41.24 (C4), 42.53 (C8), 44.23 (C5), 47.99, 48.36 (C25), 48.66 (C13), 49.04 (C14),

49.31 (C17), 61.75, 62.44, 62.52 (C26), 68.94, 69.80, 69.96 (C27), 70.88 (C7), 73.06, 73.29, 73.54 (C24), 74.34 (C3), 75.75 (C12); MS: calculated for $C_{27}H_{52}O_9NS$ [$M^- + NH_4^+ + H^+$]: 566.3339. Found: 566.3357; mp: 173 °C and after rechromatography 185–186 °C.

3. Results and discussion

The protective group chosen for this synthesis was the THP group rather than the silyl ether for the following reasons. It is more amenable to commercial synthesis, and can be removed under mild conditions without causing desulfation. However, the NMR of the protected intermediates is more difficult to interpret due to the chirality of each THP group.

This first reported synthesis of sodium scymnol sulfate commences with cholic acid. This reacted with DHP and the resulting THP protected THP cholate ester **2** was reduced with lithium aluminium hydride and worked up by addition of a small amount of water to produce a precipitate that was filtered off. The ester had been stripped of the DCM solvent before reduction. Investigation of an alternative reduction with Red Al[®] in DCM resulted in problematic emulsions and was not pursued further. The alcohol **3** was oxidized to the aldehyde **4** with pyridinium chlorochromate in the presence of anhydrous sodium acetate [13] and purified by column chromatography. Methyl-3-hydroxypropionate (**5**) was prepared from 3-hydroxypropionitrile and methanolic HCl containing a little water [14]. The pure aldehyde **4** was coupled using 2 equiv. of lithium diethylamide in THF at –65 °C according to the method of Sunazuka et al. [15]. The reaction produced compound **6** which was chromatographed and then protected as the THP derivative **7**. The methyl ester group at C-26 was reduced to the monohydroxyl compound **8**. The triethylamine salt of the C-27 monosulfate **9** was prepared by reaction of the monohydroxyl compound **8** and $Et_3N \cdot SO_3$ in DMF according to the method of Tserng and Klein [12]. After stripping the DMF under high vacuum, the THP protective groups were removed in methanolic HCl to produce compound **10**. Care was taken to prevent desulfation by excessive methanolic HCl treatment [16]. Sodium ethoxide was dissolved in methanol and gradually added to the compound **10** reaction mixture, changing the pH to the range of 9–10. The pH was then adjusted to between pH 7 and 8 and the crude mixture was stripped of solvent, absorbed onto a portion of silica gel and chromatographed to yield pure (24RS,25RS) sodium scymnol sulfate **11**. This was characterised using ^{13}C and 1H NMR and compared with the two [24R,25R] and [24R,25S] stereoisomers of natural sodium scymnol sulfate using the results of Ishida et al. [9]. Our ^{13}C NMR results comparison is summarised in Table 1. Extra chemical shifts were observed for the unnatural isomers (24S,25R) and (24S,25S) produced in our product (24RS,25RS) sodium scymnol sulfate **11** at positions 21, 23, 26 and 27 as shown in Fig. 4. Fig. 3 shows the steric orientations at the 24 and 25 positions for the four stereoisomers of our synthetic compound **11**.

We believe that the observed protective effect of scymnol is due to its free radical inhibitory effect in biological systems [2]. If this is so, the 24R and 24S stereoisomers may be equivalent in activity. We have also produced ^{14}C -radiolabelled 24RS,25RS

Table 1 – ^{13}C NMR data of the two natural scymnol sulfate and four synthetic stereoisomers

Position	Isheda et al. [9]		Present work (24R,25R, 24R,25S (24S,25R; 24S,25R)
	24R,25R	24R,25S	
20	38.6	38.6	38.74
21	19.9	19.9	20.17, 20.19
22	34.5	34.6	34.46, 34.67
23	32.9	33.1	32.15, 32.77, 32.96
24	73.1	72.9	73.05, 73.27
25	48.3	48.5	47.98, 48.35
26	61.7	62.3	61.74, 62.42, 62.51
27	69.7	68.9	68.93, 69.78, 69.95

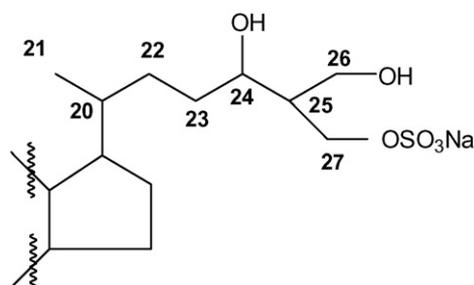


Fig. 4 – Positions in the sodium scymnol sulfate side-chain.

sodium scymnol sulfate for use in mouse pharmacokinetic studies [17] using the synthetic method in this paper.

The synthetic sodium scymnol sulfate was also confirmed by high resolution MS and microanalysis.

In conclusion, we have successfully completed the synthesis of (24RS,25RS) sodium scymnol sulfate and the general synthetic scheme presented in this paper may be useful in the synthesis of a range of sterol monosulfated products commencing from various bile acids.

Acknowledgements

The authors wish to thank Ms Nicolette Kalafatis for technical laboratory support and Dr Paul Wright for assistance in manuscript preparation. Mass spectral analyses were performed by Mr Stuart Thomson and Ms Sally Duck of Monash University. We wish to thank Dr Greg Simpson, Deputy Chief of the CSIRO Molecular and Health Science Technologies Clayton, Victoria for technical support, and in particular CSIRO staff Carl Braybrook for MS interpretation and Dr Roger J. Mulder for ^{13}C NMR spectroscopy on the synthetic sodium scymnol sulfate. This work was supported by the Australian Research Council (ARC) Collaborative Research Grant Program (Grant No. LP0219239) and the Industry Partner, McFarlane Marketing (Aust.) Pty Ltd, Melbourne, Vic., Australia.

REFERENCES

- [1] Ketsugo[®] Isolutrol for acne treatment.
- [2] Macrides TA, Shihata A, Kalafatis N, Wright PFA. A comparison of the hydroxyl radical scavenging properties of

- the shark bile steroid 5 beta-scygnol and plant pycnogenols. *Biochem Mol Biol Int* 1997;42(6):1249-60.
- [3] Macrides T (Inventor), J.W. Broadbent Nominees Pty. Ltd., Australia, assignee. Treatment of medical disorders associated with free radical formation. WO 95/10283.
- [4] Macrides TA, Naylor LM, Kalafatis N, Shihata A, Wright PFA. Hepatoprotective effects of the shark bile salt 5 β -scymnol on acetaminophen-induced liver damage in mice. *Fundam Appl Toxicol* 1996;33(1):31-7.
- [5] Slitt AL, Naylor L, Hoivik J, Manatou JE, Macrides T, Cohen SD. The shark bile salt 5 β -scymnol abates acetaminophen toxicity, but not covalent binding. *Toxicology* 2004;203(1-3):109-21.
- [6] Ishida H, Nakayasu H, Nukaya H, Tsuji K. Study of the pharmacological effect of the bile salt sodium scymnol sulfate from *Rhizoprionodon acutus*. II. Prophylactic effect of scymnol on lesion development in a rat peripheral arterial occlusion model. *Biol Pharm Bull* 1998;21(3):240-4.
- [7] Ishida H, Nakayasu H, Nukaya H, Tsuji K. Study of the pharmacological effect of the bile salt, sodium scymnol sulfate, from *Rhizoprionodon acutus*. III. Protective effect of scymnol against vascular endothelial cell damage in a rat peripheral arterial occlusion model. *Biol Pharm Bull* 1999;22(8):822-7.
- [8] Ishida H, Nakayasu H, Tsuji H. Study of the pharmacological effect of the bile salt, sodium scymnol sulfate, from *Rhizoprionodon acutus*. IV. Effects of naturally occurring bile alcohols, bile acids and their conjugates on lesion development and vascular endothelial cell injury in a rat peripheral arterial occlusion model. *Biol Pharm Bull* 1999;22(8):828-35.
- [9] Ishida H, Kinoshita S, Natsuyama R, Nukaya H, Tsuji K, Nagasawa M, et al. Study on the bile salt, sodium scymnol sulfate, from *Lamna ditropis*. III. The structures of a new sodium scymnol sulfate and new anhydroscymnols. *Biol Pharm Bull* 1992;40(4):864-8.
- [10] Amiet GA, Kalafatis N, Macrides T. On the synthesis of scymnol. *Aust J Chem* 1993;46:1347-54.
- [11] Adhikari R, Cundy DJ, Francis CL, Gebara-Coghlan M, Krywult B, Lubin C, et al. A scalable stereoselective synthesis of scymnol. *Aust J Chem* 2005;58:34-8.
- [12] Tserng KY, Klein PD. Synthesis of sulfate esters of lithocholic acid, glycolithocholic acid and tauroolithocholic acid with sulphur trioxide-triethylamine. *J Lipid Res* 1977:491-5.
- [13] Corey EJ, Suggs JW. Pyridinium chlorochromate: an efficient reagent for oxidation of primary and secondary alcohols to carbonyl compounds. *Tetrahedron Lett* 1975;31:2647-50.
- [14] Ogawa T, Nakazato A, Sato M, Hatayama K. Synthesis of 2- and 3-nitroxypropanol by chemoselective reduction of methyl 2- and 3-nitrooxypropionate. *Synthesis* 1990;6:459-60.
- [15] Sunazuka T, Tzuziki K, Kumagai H, Tomada H, Tanaka H, Nagashima H, et al. Synthesis of 1233A analogs and their inhibitory activity against hydroxymethylglutaryl Coenzyme A synthase. *J Antibiot (Tokyo)* 1992;45(7):1139-47.
- [16] Kantor TG, Schubert M. A method for the desulfation of chondroitin sulfate. *J Am Chem Soc* 1957;79:152-3.
- [17] Yendle S, Harney D, Macrides T, Wright P. Pharmacokinetic study of scymnol sulfate in mice. Abstracts of the 15th World Congress of Pharmacology. *Acta Pharmacol Sin Suppl* 2006;1354:300222.