

Efficient Synthesis of Trisimidazole and Glutaric Acid Bearing Porphyrins: Ligands for Active-Site Models of Bacterial Nitric Oxide Reductase

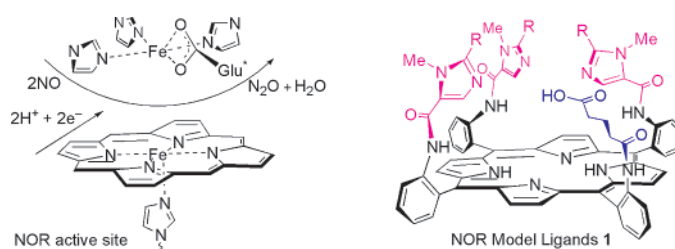
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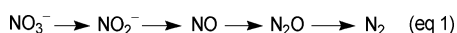
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ABSTRACT



Ligands (1) for active-site models of bacterial nitric oxide reductase (NOR) have been efficiently synthesized. These compounds (1) feature three imidazolyl moieties and one carboxylic acid residue at the Fe_B site, which represent the closest available synthetic model ligands of NOR active center. The stereo conformations of these ligands are established on the basis of steric effects and ¹H NMR chemical shifts under the ring current effect of the porphyrin.

Biological denitrification is a four-step process that reduces nitrate to dinitrogen (eq 1). It represents an alternative to aerobic respiration where the nitric oxides are used as terminal electron acceptors.¹ Denitrification is not only an important process for a variety of bacteria in anaerobic energy harvest but also the only process that returns a huge amount of fixed nitrogen to atmosphere, thereby finishing the inorganic terrestrial nitrogen cycle. Nitric oxide reductase (NOR) is a membrane-bound enzyme that catalyzes the third step of denitrification that reduces nitric oxide to nitrous oxide (eq 2).



The active site of NORs is bimetallic with an imidazole ligated heme iron and a non-heme iron (Fe_B) (Figure 1).¹ The imidazole coordinated to heme iron on the proximal side is reported to disassociate from heme iron after binding of NO.² The Fe_B site is coordinated by three histidyl nitrogen

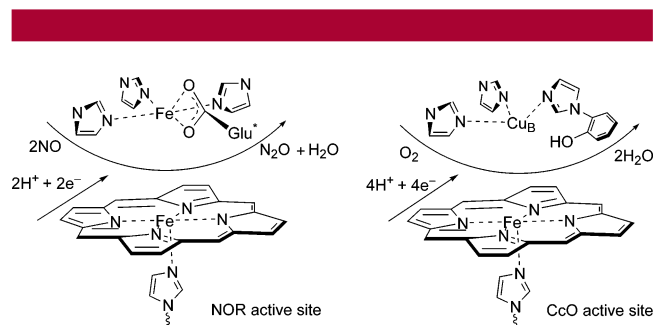


Figure 1. Schematic presentation of NOR and CcO active sites.

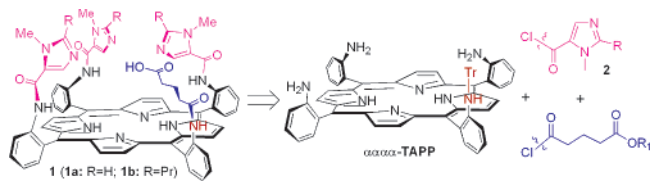
atoms and a postulated ligated glutamic acid residue.³ Comparing NORs with their evolutionary genetic relative

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Cytochrome *c* oxidases (CcOs), the dinuclear catalytic center in NORs is Fe_B-heme-Fe rather than Cu_B-heme-Fe in CcOs.¹ The glutamic acid residues are conserved in NORs, whereas they are absent in CcOs. Additionally, in most CcOs, a redox active phenol group from a tyrosine is coupled to one of the three Cu_B coordinating histidine moieties.³ The glutamic acid residues have been reported to be essential for normal levels of NOR activity.^{3a} Possible functions of these glutamic acids include increased selective binding of the distal non-heme Fe_B over Cu, charge regulation directing to the modulation of redox potential of the catalytic center, and mediation of the proton uptake for NO reduction.

The biomimetic approach to studying the structural-functional relationships of metalloenzyme activities has proven to be quite successful.^{3c,4} Simulation and variation of synthetic models allows for the easy and rapid comparison of coordination environments, spectroscopic properties, and functional capabilities of the metalloenzyme active site. Such information may be difficult to obtain from wild type enzymes or their mutants due to their restricted availability and difficulty in handling. Previously, only a few synthetic models have been developed to study NOR.^{1a,5} Recently, Karlin and co-workers reported structural models of NOR active site featuring a porphyrin covalently linked to a tetradentate chelate (TMPA, tris(2-pyridylmethyl)amine).⁵ In these models, the TMPA moiety was intended to mimic the trisimidazole ligand environment of Fe_B site in NORs. A carboxylic group that mimics the glutamic acid residue was not included in these models. A more faithful ligand environment is desirable for modeling of metalloenzymes, as a more exact mimic should shed light on metal binding affinity, spectroscopic properties, and redox potential of the active site of the native enzyme. Our interest on NOR modeling originates from our long standing effort in CcO mimicry and investigation of the chemistry occurring at the bimetallic Fe/Cu active site of CcO using synthetic bioinorganic models.⁶ Herein we report the syntheses of NOR model ligands **1** (**1a**, **1b**) featuring a porphyrin bearing trisimidazole pickets and a carboxylic acid residue (Scheme 1). These ligands represent the best available NOR model ligands

Scheme 1. Retrosynthetic Scheme of NOR Model Compound **1**

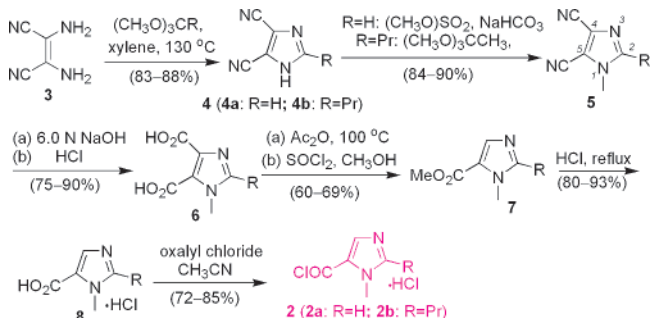


closely replicating the ligand environment of the active site of native NORs.

The retro-synthetic scheme of compounds **1** is shown in Scheme 1. The four amino groups of ααα-TAPP (tetrakis-5,10,15,20-(*o*-aminophenyl)porphyrin) provide excellent linkages for installation of imidazole and carboxylic acid moieties by forming amide bonds with corresponding imidazolecarboxylic acid chloride and glutaric acid chloride.⁷ One amino group for introduction of the carboxylic acid residue is selectively discriminated from other three amino groups for imidazole moieties by protection with a trityl group.

Preparation of imidazolecarboxylic acid chloride **2** is shown in Scheme 2. Synthesis of 5-imidazolecarboxylic acids

Scheme 2. Synthetic Scheme of Acid Chlorides **2**



8 followed a literature procedure incorporated with our own modification.⁸ Diaminomaleonitrile **3** was condensed with trimethyl orthoformate and trimethyl orthobutyrates to give dicyanoimidazole **4a** and **4b** in yields 83–88%, respectively. **4a** was methylated with dimethyl sulfate in NaHCO₃ aqueous solution providing **5a** in 90% yield. Due to the lower solubility of **4b** in water, an alternative method was employed for methylation of **4b**. Refluxing **4b** in trimethyl orthoacetate provided 1-methylated 4,5-dicyanoimidazole **5b** in 84% yield. The *N*-methyl group in intermediates **5–8** not only results in higher solubility of this series of imidazole intermediates in organic solvents but also leads to stereo

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(4) (a) Holm, R. H.; Solomon, E. I. *Chem. Rev.* **2004**, *104*, 347. (b) Collman, J. P.; Boulatov, R.; Sunderland, C. J.; Fu, L. *Chem. Rev.* **2004**, *104*, 561. (c) Solomon, E. I.; Szilagyi, R. K.; George, S. D.; Basumallick, L. *Chem. Rev.* **2004**, *104*, 419. (d) Rao, P. V.; Holm, R. H. *Chem. Rev.* **2004**, *104*, 527. (e) Mirica, L. M.; Ottenwaelde, X.; Stack, T. D. P. *Chem. Rev.* **2004**, *104*, 1013.

(5) (a) Wasser, I. M.; Huang, H.-W.; Moenne-Loccoz, P.; Karlin, K. D. *J. Am. Chem. Soc.* **2005**, *127*, 3310. (b) Wasser, I. M.; Martens, C. F.; Verani, C. N.; Rentschler, E.; Huang, H.-W.; Moenne-Loccoz, P.; Zakharov, L. N.; Rheingold, A. L.; Karlin, K. D. *Inorg. Chem.* **2004**, *43*, 651.

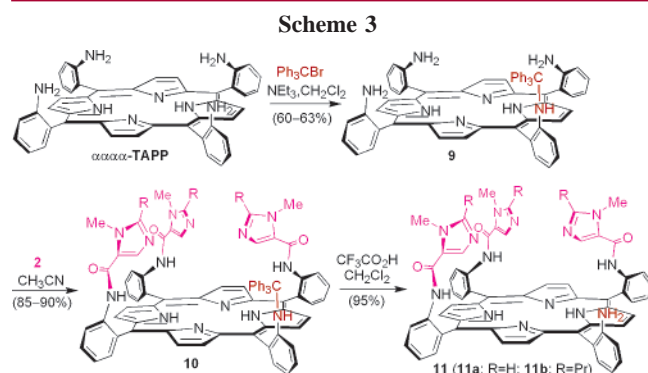
(6) (a) Boulatov, R.; Collman, J. P.; Shiryayeva, I. M.; Sunderland, C. J. *J. Am. Chem. Soc.* **2002**, *124*, 11923. (b) Collman, J. P.; Boulatov, R. *Angew. Chem., Int. Ed.* **2002**, *41*, 3487. (c) Collman, J. P.; Sunderland, C. J.; Boulatov, R. *Inorg. Chem.* **2002**, *41*, 2282. (d) Collman, J. P.; Boulatov, R.; Shiryayeva, I. M.; Sunderland, C. J. *Angew. Chem., Int. Ed.* **2002**, *41*, 4139.

(7) For some examples of ααα-TAPP as a platform in synthesis of biomimetic models, see: (a) Collman, J. P.; Gagne, R. R.; Reed, C. A.; Halbert, T. R.; Lang, G.; Robinson, W. T. *J. Am. Chem. Soc.* **1975**, *97*, 1427. (b) Wuenschell, G. E.; Tetreau, C.; Lavalette, D.; Reed, C. A. *J. Am. Chem. Soc.* **1992**, *114*, 3346. (c) Collman, J. P.; Yan, Y.-L.; Eberspacher, T.; Xie, X.; Solomon, E. I. *Inorg. Chem.* **2005**, *44*, 9628.

(8) O'Connell, J. F.; Parquette, J.; Yelle, W. E.; Wang, W.; Rapoport, H. *Synthesis* **1988**, 767.

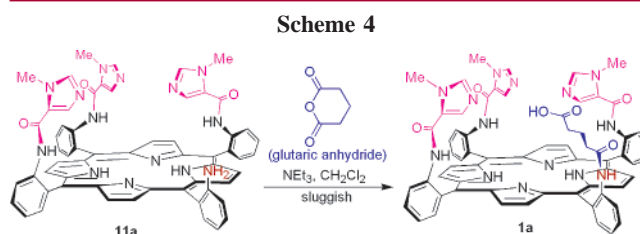
conformational control of model compounds **1** as discussed later. Hydrolysis of **5a** and **5b** in refluxing sodium hydroxide following by acidification with hydrochloric acid gives imidazoledicarboxylic acids **6a** and **6b** in 75–90% yield, respectively. Decarboxylation of imidazoledicarboxylic acids **6** (**6a**, **6b**) at 95–100 °C in acetic anhydride provides 5-imidazoleacryboxylic acids **8** (**8a**, **8b**) as the major product. To remove side product 4-imidazoledicarboxylic acid (5–25%), the decarboxylated reaction mixture was transformed to methyl esters **7** (**7a**, **7b**). 5-Esters **7a**, **7b** were isolated in 60–69% yield by column chromatography on silica gel. Pure 5-imidazoledicarboxylic acids **8a** and **8b** were obtained in 80–93% yield by refluxing their ester precursors in concentrated hydrochloric acid. Treatment of imidazole acids **8** (**8a**, **8b**) with oxalyl chloride in acetonitrile give imidazole acid chlorides **2** (**2a**, **2b**) in 72–85% yield.

Introduction of the three imidazole pickets to $\alpha\alpha\alpha$ -TAPP is shown in Scheme 3. $\alpha\alpha\alpha$ -TAPP was prepared and

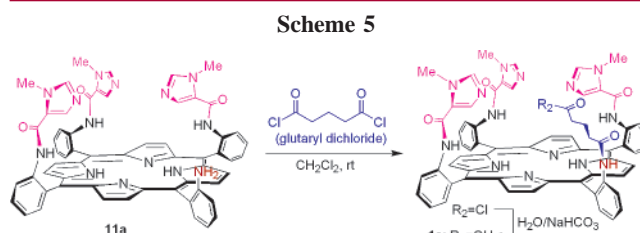


enriched following literature procedures.^{7a,9} After treatment of $\alpha\alpha\alpha$ -TAPP with 1.1 equiv of trityl bromide in CH_2Cl_2 at room temperature, intermediate **9** with one selectively protected amino group was obtained in around 60% yield. Compound **9** is prone to atropisomerize when stored at room temperature for a long time. Reaction of both freshly prepared intermediate **9** and imidazole acid chloride **2** (**2a**, **2b**) provided trisimidazole compounds **10** in 85–90% yields. Using an excess amount of imidazole acid chloride is important for achieving a high yield of the trisimidazole picketed compound **10**. Traces of mono- and bisimidazole side products were removed during column purification. Intermediate **10** is stable; no atropisomerization was observed after storage in a refrigerator for months. Simple treatment of **10** with trifluoroacetic acid provided compound **11** in around 95% yield. This frees the trityl-protected amino group for subsequent introduction of the glutaric acid residue.

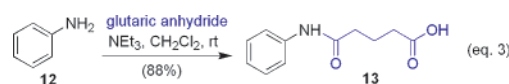
Several strategies were examined in order to find the most efficient method to introduce the glutaric acid residue (Schemes 4–6). Reaction of **11a** with glutaric anhydride was first tested (Scheme 4). The reaction was found to be very sluggish and the product **1a** difficult to isolate from the



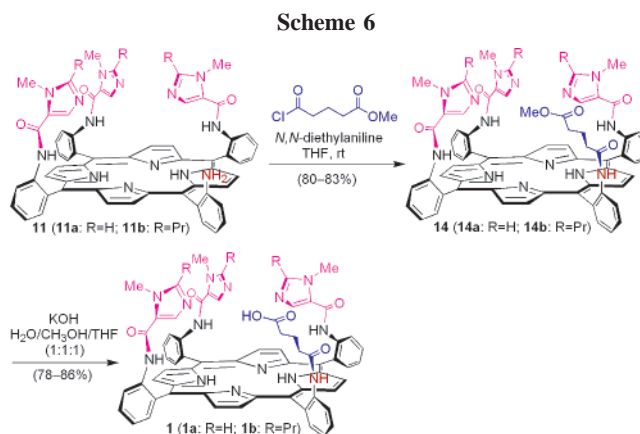
reaction mixture. The outcome is tentatively explained by the unfavorable steric effect from bulky trisimidazole-bearing porphyrin substrate **11a**, as in a model reaction, phenylamine **12** was found to react with glutaric anhydride readily to provide corresponding amidoglutaranic acids **13** in good yield under similar conditions (eq 3). Reaction of **11a** with an excess of glutaryl dichloride followed by hydrolysis of the acid chloride intermediate with water was also examined (Scheme 5). The reaction provided the expected compound



1a, but some side products with polarity similar to that of **1a** were difficult to remove by column purification.



A more practical method for introducing the glutaric acid residue is shown in Scheme 6. Intermediate **11a** was treated with methyl 5-chloro-5-oxoverate to provide glutaric ester



(9) Lindsey, J. J. *Org. Chem.* **1980**, *45*, 5215.

14a in 83% isolated yield. Compared with its glutaric acid counterpart **1a**, ester **14a** is less polar and can be easily purified by column chromatography on silica gel. Hydrolysis of methyl ester group of **14a** is quite selective and efficient in a KOH solution of H₂O/CH₃OH/THF (v/v 1/1/1), providing compound **1a** in 86% yield. An analogous model compound **1b** was synthesized similarly following Scheme 6. Compared with **1a**, due to the propyl groups in imidazole moieties, **1b** is less polar during column purification on silica gel. The identities of NOR model compounds **1a** and **1b** were characterized by ¹H NMR, ¹³C NMR, and high-resolution mass spectroscopy (HRMS).¹⁰

The proposed geometry of the trisimidazole pickets and the glutaric acid residue over the ααα-TAPP platform is shown in Figure 2. The glutaric residue leans inward to the

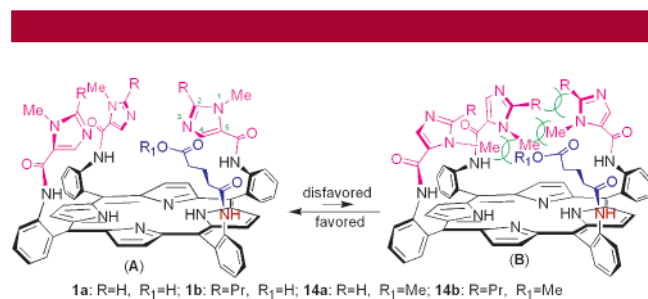


Figure 2. Stereo conformations of compounds **1** and **14**.

porphyrin center. The 1-NMe groups on imidazole pickets point outward leaving the 3-imidazole nitrogen atoms closer to the porphyrin center (conformation **A**). A reversed stereo orientation of the 1-NMe groups directed toward the porphyrin center is unfavorable due to a strong steric repulsion among the appended groups (conformation **B**). Such spatial orientation is consistent with the ¹H NMR spectra of the product (Figures 3 and 4).

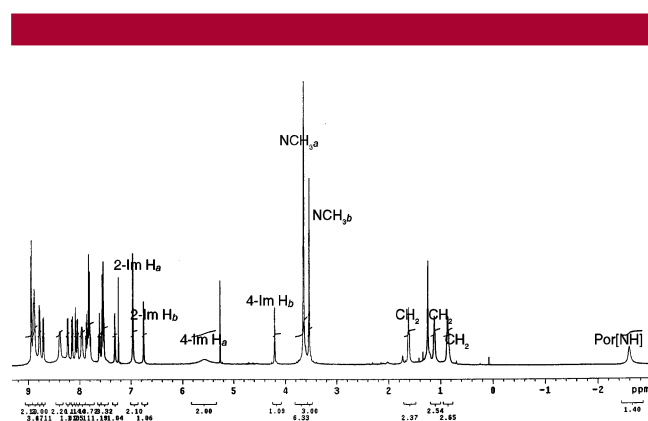


Figure 3. ¹H NMR spectrum of compound **1a** in CDCl₃.

¹H NMR spectra is a characteristic tool to probe the stereorelation between the porphyrin ring and appended

(10) See the Supporting Information for details.

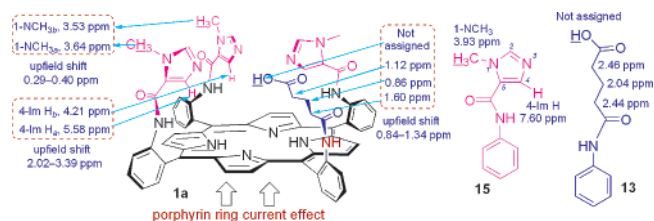


Figure 4. ¹H NMR chemical shifts of **1a**, **13**, and **15** in CDCl₃.

substituents. Compared to those of non-porphyrin analogues, the chemical shifts of substituents closer to the porphyrin ring center are strongly shifted upfield due to the ring current effect of the porphyrin ring. The influence of the porphyrin ring current effect on the relation between ¹H NMR chemical shifts of the substituents and their stereo orientation is demonstrated with the ¹H NMR spectrum of compound **1a** (Figures 3 and 4).¹¹ Compared with that of its non-porphyrin analogue **15**,¹² the diagnostic imidazole proton 4-Im H of **1a** strongly shifts upfield 2.02–3.39 ppm, which is much more than that of imidazole 1-NCH₃ (0.29–0.40 ppm), suggesting that 4-Im H is close to the porphyrin center while 1-NMe is located at the porphyrin periphery. The upfield shift (0.84–1.34 ppm) of glutaric acid group on compound **1a** compared to its analogue **13** implies that this glutaric acid moiety is suspended over the porphyrin plane. A similar ¹H NMR shift pattern is also observed in compounds **14a**, **14b**, and **1b** implying a similar stereo orientation of imidazole and glutaric groups in these molecules.¹⁰

In summary, two porphyrin compounds **1a** and **1b** with three imidazoles and one glutaric acid residue suspended over the porphyrin plane have been prepared. These compounds closely resemble the ligand environment of the diiron center of NORs. Metalation of these model compounds, spectroscopic characterization of their metal complexes, and investigation on interaction between the metal complex and NO and O₂ are currently in progress. These studies should provide insightful understanding about the structure features and mechanism of NORs.

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Supporting Information Available: Synthetic procedures, characterization, and ¹H NMR, ¹³C NMR spectra, and high-resolution mass spectra (HRMS) of all new compounds. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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(11) The chemical shifts of the carboxylic acid protons of **1a** and **1b** cannot be assigned due to overlap of ¹H NMR spectra.

(12) ¹H NMR spectra of non-porphyrin analogues **13** and **15** are provided in the Supporting Information.