Accepted Manuscript

Formal synthesis of Cladospolide C & epi-Cladospolide C using R-(+)- γ -valerolactone as a chiral synthon

Rajender Datrika, Srinivasa Reddy Kallam, Siddaiah Vidavalur, Nagaraju Rajanna, T.V. Pratap

PII: S0040-4020(19)30391-6

DOI: https://doi.org/10.1016/j.tet.2019.04.001

Reference: TET 30246

To appear in: Tetrahedron

Received Date: 5 February 2019
Revised Date: 28 March 2019
Accepted Date: 1 April 2019

Please cite this article as: Datrika R, Kallam SR, Vidavalur S, Rajanna N, Pratap TV, Formal synthesis of Cladospolide C & *epi*-Cladospolide C using *R*-(+)-γ-valerolactone as a chiral synthon, *Tetrahedron* (2019), doi: https://doi.org/10.1016/j.tet.2019.04.001.

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.



Graphical Abstract

Formal synthesis of cladospolide C & *epi*-cladospolide C using R-(+)- γ -valerolactone as a chiral synthon.

Leave this area blank for abstract info.

Rajender Datrika,* Srinivasa Reddy Kallam, Nagaraju Rajanna, Siddaiah Vidavalur, Pratap T. V. *

R-(+)- γ -valerolactone

Cladospolide C



journal homepage: www.elsevier.com



Formal synthesis of Cladospolide C & epi-Cladospolide C using $R-(+)-\gamma$ valerolactone as a chiral synthon.

Rajender Datrika^{a,b*}, Srinivasa Reddy Kallam^a, Siddaiah Vidavalur^b, Nagaraju Rajanna^a, Pratap T.V.^{a,*}

^a Technology Development Centre, Custom Pharmaceutical Services, Dr. Reddy's Laboratories Ltd, Hyderabad 500 049, Telangana, India

ARTICLE INFO

ABSTRACT

Article history: Received Received in revised form Accepted Available online

Keywords: Cladospolide

Gamma valerolactone Wittig reaction

Yammaguchi lactonization

The formal synthesis of Cladospolide-C and its analog is achieved by using enantiopure (R)- γ -valerolactone 10. The significant points of this synthesis are the stereoselective dihydroxylation of α , β -unsaturated ester 16 using Sharpless protocol, Wittig olefination of γ -valerolactol 6 with triphenylphosphonium iodide salt 7, one pot selective oxidation of 22 and subsequent C2-homologation with good E/Z ratio.

2009 Elsevier Ltd. All rights reserved.

Introduction

Cladospolides A–D and iso-Cladospolide B comprise a family of fungal secondary metabolites isolated from the fermented broth of cultures that were obtained from either marine or soil fungi. 1-⁴ Among these, cladospolides A (1) and B (2) were first isolated from the culture filtrate of the fungus Cladosporium cladosporiolides FI-113 by Isogai and co-workers. In 1995, Fukada et al., isolated cladospolide C (4) along with cladospolides A and B from Cladosporium tenuissium.² Later in 2000, Ireland and co-workers isolated 2 from Cladosporium herbarum together with isocladospolide B (3) from the fungal strain I96S215.3 Cladospolide D (5) was isolated in 2001 from the fermentation broth of *Cladosporium* sp. FT0012. 4a Structurally, cladospolides A–C are γ , δ -dihydroxy- α , β -unsaturated 12-membered macrolides, whereas Cladospolide D (5) is a δ -hydroxy- γ -Oxo- α , β -unsaturated 12-membered lactone (**Figure 1**).

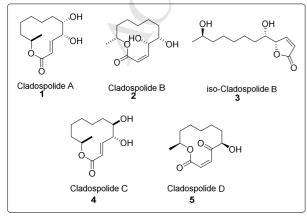


Figure 1.Structures of Cladospolides A–D and iso-Cladospolide B.

Furthermore, Cladospolides A and C possess an (E)-olefin geometry, whereas Cladospolides B and D have a (Z)-olefin geometry. In contrast to Cladospolides A-D, iso-Cladospolide B (3) is a butenolide that is γ -substituted by an aliphatic group. Although the Cladospolides are fungal metabolites, they are plant-growth regulators. For example, Cladospolides A-C inhibit the shoot elongation of rice seedlings.⁴¹

The biological properties of Cladospolide C (4) suggest that it is a gibberellin synthesis inhibitor. Unlike the other members, Cladospolide D (5) shows antimicrobial activity against Mucor recemosus and Pyricularia oryzae. Given their fascinating structural diversities and biological profiles, the Cladospolide family members have generated considerable interest in the synthetic community. Accordingly, various strategies have been developed that utilize diverse key reactions.⁵⁻⁹The absolute and relative stereochemistry for the Cladospolides A, B, and C have been confirmed by several total syntheses from known chiral starting materials.⁵⁻⁹ For instance, Cladospolides A-C have been prepared by Banwell from 1-hepten-2-ol and cis-2, 3-dihydroxychlorobenzene which can be prepared in optically pure form *via* enzymatic resolution and biotransformation, ¹⁰ whereas others have followed chiron approach using carbohydrate sources.1 Synthesis of Cladospolide A was reported by Mori and Maemoto in 1987. Despite their rather intriguing biological profiles, the Cladospolides have only received limited attention as synthetic targets.

In continuation of our pursuit of synthesizing structurally complex molecules beginning with optically pure y-

^b Department of Organic Chemistry & FDW, Andhra University, Visakhapatnam 530 003, Andhra Pradesh, India

Valerolactone, herein we report the formal synthesis of Cladospolide C (4) from γ -Lactone.

 γ -Lactone is very good chiral synthon in the synthesis of natural products. A variety of methods are in practice for the synthesis of optically pure γ -lactones. Though a number of reports available in the literature for the synthesis of optically pure γ -Valerolactone (10), we chose to prepare the latter from D-alanine (11) *via* D-Lactic acid (12).

Retrosynthetic approach:

The retrosynthetic approach for synthesis of Cladospolide C (4) is described in **Scheme 1**. Accordingly Cladospolide C (4) can be achieved by Yamaguchi lactonization of the α , β -unsaturated seco-acid 9. The seco-acid 9 is obtained by selective oxidation, a two carbon homologation using Wittig ylide and saponification of the ester. The seco acid 9is traced to intermediate diol 8 which is formed as a result of hydrogenation of the double bond that formed via Wittig reaction of corresponding triphenylphosphonium salt (7) and lactol (6). Compound 6 can be traced from R-(+)- γ -Valerolactone (10) further compound 7 can be traced from commercial available 1, 3-propanediol (17).

Scheme 1: Retrosynthetic approach for Cladospolide C (4) via Wittig olefination

The retrosynthetic approach for the synthesis of Wittig salt 7 from 1, 3-propanediol (17) is described in the **Scheme 2**. The Wittig salt (7) was obtained by phosphorylation of Iodo alcohol 13. The intermediate iodo alcohol 13 was obtained from hydroxy ester 14 via halogenation under Appel condition followed by NaBH₄ reduction. The hydroxy ester 14 can be

traced from α , β -unsaturated ester **16** by means of Asymmetric Sharpless dihydroxylation. The α , β -unsaturated ester **16** can be traced from 1, 3-propanediol (**17**) by selective mono TBDMS protection followed by C2-homologation.

Scheme 2: Retrosynthesis of 7 (Wittig salt) from 1, 3-Propanediol (17).

Results and discussion:

The synthesis began with reduction of R- γ -Valerolactone (10) to corresponding R- γ -Valerolactol (6) and this key intermediate was used in synthesis of cladospolides as shown in **Scheme 3.** The synthesis of R- γ -Valerolactone (10) was described in literature.

Scheme 3: Synthesis of $R-\gamma$ -valerolactol (6) from $R-\gamma$ -Valerolactone (10).

Reagents and conditions: a) NaNO₂, H₂SO₄; 2 h, 58 %; b) (i) MeOH, H₂SO₄, rt, 24 h;(ii) TBDMSCl/Imidazole, DCM, 25-35 °C, 12 h, 95 %; (iii) DiBAL-H, DCM, -70 \pm 5 °C, 2 h; (iv) C2- Wittig (**21**), 72 %;(v) H₂, Pd/C, Ethyl acetate 40 Psi, 4 h, 95 % d) *p*TSA, MeOH, 25-35 °C, 24 h, 90 %; c) DIBAL-H, DCM, -78 °C, 0.5 h.

We began synthesis of Wittig salt **7** starting from 1,3 propanediol (**17**) as described in **Scheme 4.** The 1,3 propanediol (**17**) was selectively protected as silyl ether using NaH to afford mono protected diol **18** with good yield. The primary alcohol was then oxidized to aldehyde using PCC which was homologated further with C2- Wittig salt **21** to afford α , β unsaturated ester **16** with > 95% trans configuration as indicated by H NMR.

ACCEPTED MANUSCRIPT

Scheme 4: Synthesis of Wittig salt (7a) & (7b) starting from 1, 3 propanediol (17)

Reagents and conditions: a) TBDMSCl, NaH, THF, 0 to 25 °C, 2 h, 90 %; b) PCC/NaOAc, DCM, 25 °C, 4 h; C2-wittig (**21**), 24 h, 82 %; c) (DHQ)₂PHAL (**15a**) or (DHQD)₂PHAL (**15b**) K₃Fe(CN)₆, K₂OsO₄.2H₂O, K₂CO₃, NaHCO₃, MeSO₂NH₂, *t*-Butanol: H₂O (2:3), 24 h, 5 °C, 94 %; d) P*p*TS, Acetone, 2, 2-dimethoxy propane, 24 h, 25 °C, 90 %; e) TBAF, THF, 0-5 °C, 12 h, 95 %; f) I₂, TPP, imidazole, DCM, rt, 2 h, 82 %; g) NaBH₄ MeOH: THF (1: 4), 5 °C, 4 h, 89 %; h) TPP, ACN, reflux, 12 h, 88 %.

The α , β unsaturated ester **16** was then converted into a chiral diol *via* Sharpless asymmetric dihydroxylation. Both the enantiomers **15a** and **15b** were synthesized by using a suitable reagent system. Thus obtained stereogenic centers as secondary diols in compound **15a** and **15b** were protected separately using 2,2 dimethoxy propane in an acetone and catalytic amount of PpTS to afford **14a** in good yields. The compound **14a** was desilylated with TBAF to afford hydroxy ester **19a**. The

The hydroxy ester **19a** was subjected to Mitsunobu mediated halogenation in presence of iodine and triphenylphosphine to afford the corresponding iodo ester **20a**. ¹⁹ The iodo ester **20a** was then treated with NaBH₄ in a MeOH:THF (1:4) to afford the corresponding iodo alcohol **13a**. The iodo alcohol **13a** was finally converted to

corresponding Wittig salt **7a** and its antipode **13b** was traced from **7b** in similar fashion.

Now the stage is set to synthesize the title compound Cladospolide C (4) using Wittig salt 7 and Lactol 6 *via* Wittig olefination as mentioned in **Scheme 5** The obtained lactol 6 (**Scheme 3**) was treated with Wittig salt 7a (**Scheme 4**) under Wittig olefination condition²⁰ to afford corresponding enediol **22a**. Ene-diol **22a** was then subjected to catalytic hydrogenation at 40 psi using Pd(OH)₂ as catalyst at ambient temperature to afford the corresponding saturated diol **8a**. ²¹

$$R$$
-(+)-γ-valerolactol 7a: 4S,5S 22a 4S,5S 22b 4R,5R $\frac{1}{2}$ $\frac{1}{2}$

Scheme 5: Wittig olefination of Lactol 6 & Wittig salt 7.

Reagents and conditions: a) NaHMDS, THF, -78 °C, 86 %; b) (i) H₂-Pd(OH)₂, 40 psi, ethylacetate, rt, 98 %; (ii) BAIB/TEMPO, DCM, 2 h, C2-Wittig (21), rt, 94 %.

ACCEPTED MANUSCRIPT

This step afforded the ideal structural frame work to expedite the final molecule formal synthesis via selective oxidation. Thus compound $\bf 8a$ was subjected to one pot selective oxidation and concomitant C2-ylide extension under BAIB/TEMPO²² conditions to afford $\bf 9a$ with good $\bf E$ -selectivity and yield. Diastereomer $\bf 9b$ traced from $\bf 8b$ in similar fashion with $\bf E:\bf Z$ ratio (96.2 : 3.8) estimated by $\bf GC.^{23}$

The *epi*-cladospolide C (**23**) was synthesized formally from Compound **9a** and similarly titled Cladospolide-C (**4**) was synthesized formally from **9b** (**Scheme 6**) using the reported procedure. ²⁴ Cladospolide B (**2**) can be synthesized formally from *epi*-Cladospolide-C (**23**) using reported procedure. ²⁵

Conclusion:

In summary, formal synthesis of Cladospolide C (4) and *epi*-Cladospolide C (23) has been developed with good yields. The route of synthesis developed for these macrolides utilize fairly inexpensive reagents and operationally friendly processes. This strategy can be utilized for the stereoselective synthesis of other macrolides. The application of this methodology for the synthesis of other biologically active complex macrolides is currently underway.

Scheme 6: Formal synthesis of Cladospolide C (4) & epi-Cladospolide C (23)

Experimental section:

General information:

All reagents were used as received from commercial sources without further purification or prepared as described in the literature. Reactions were monitored by thin layer chromatography (TLC) performed on Merck TLC silica gel 60 F254 aluminium plates. Visualization of the spots on the TLC plates was achieved by exposure to UV radiation (254 nm) or by using an appropriate TLC staining reagent (such as PMA, anisaldehyde and ninhydrin). Chromatographic purification of products was carried out by flash column chromatography on silica gel (60-120 mesh or 100-200 mesh or 230-400 mesh as the case required). Melting points were determined using a Differential Scanning Calorimeter (DSC, Q-2000, TA) apparatus. Infrared spectra were recorded on a Perkin-Elmer 1650 Fourier transform spectrometer. ¹H and ¹³C NMR spectra were recorded on a Varian 400 MHz spectrometer. Chemical shifts (δ) in ppm are reported relative to Me₄Si (= 0 ppm) by using residual solvent signals as internal reference [CDCl₃: δ = 7.26 ppm (1 H NMR) and 77.0 ppm (13 C NMR); CD₃OD: δ = 3.31 ppm (1 H NMR) and 49.2 ppm (13 C NMR); DMSO- d_6 : δ = 2.50 ppm (¹H NMR) and 39.5 ppm (¹³C NMR)]. LRMS data were recorded on an Agilent 1200 Series liquid chromatography module hyphenated to a 6430 Triple Quad LC/MS system. HRMS spectra were recorded on Micromass LCT Premier mass spectrometer equipped with an ESI lockspray source for accurate mass values.

2.1 Experimental procedures

3-(tert-butyldimethylsilyloxy) Synthesis of **propan-1-ol** (18): To a stirred solution of 1, 3-propane diol (17) (3.0 g, 39.42 mmol) in dry THF (40 mL) at 0 °C was added oil free NaH (0.95 g, 39.42 mmol, 1.0 equiv.) in portions over 15 min. The reaction mixture was stirred at room temperature for 30 min, then cooled to 0 °C after which TBDMSCl (5.94 g, 39.42 mmol, 1.0 equiv.) was added. The reaction mixture was stirred at room temperature for 2 h. It was then quenched with ice cold water (10 mL) and the solution extracted with EtOAc (3 X 30 mL). The combined organic layers were washed with water, brine, dried (Na₂SO₄), and concentrated. The residue was purified by silica gel column chromatography to give 18 (6.75 g, 90%). Analytical data: Colorless oil.; IR (CHCl₃, cm⁻¹): 3373, 2955, 2930, 2858, 1472, 1256, 1087, 965, 836, 774.; ¹H NMR (400 MHz, CDCl₃): δ 0.078 (s, 6H), 0.90 (s, 9H), 1.75 (m, 2H), 2.55 (t, J = 4.9 Hz, 1H), 3.78 (m, 4H).; 13 C NMR (100 MHz, CDCl₃): δ –5.53 (2C), 18.13, 25.63, 25.83, 25.90, 34.19, 62.27, 62.79.; HRMS (ESI+): Calcd. for $C_9H_{22}O_2Si$ +H: 191.1466 found: 191.1467.

2.1.2. **Synthesis** of butyldimethylsilyloxy) pent-2-enoate (16): To a mixture of PCC (5.1 g, 23.64 mmole, 1.5 equiv.) and NaOAc (1.94 g, 23.64 mmole, 1.5 equiv.) in dry CH₂Cl₂ (60 mL) was added a solution of alcohol (18) (3.0 g, 5.76 mmole) in CH₂Cl₂ (15 mL) at 0 °C. The reaction mixture was stirred for 4 h at rt and then diluted with petroleum ether/EtOAc (7: 3, 80 mL). The slurry was stirred and filtered through a pad of silica gel and Celite. The residue was washed 3 to 4 times with petroleum ether/EtOAc (7: 3) and filtered. The filtrate was concentrated to give virtually pure aldehyde, which was used directly in the next reaction. To a stirred solution of aldehyde (4.05 g, 18.07 mmole) in dry DCM (60 mL) at 0 °C was added C2-Wittig (6.04 g, 1.1 equivalents ylide). The mixture was stirred at room temperature for 24 h at rt, solvent was removed under vacuum and crude residue was purified by silica gel column chromatography using petroleum ether/EtOAc (9: 1) as eluent to afford 16 (3.34 g, 82% overall two stages). Colorless oil.; IR (CHCl₃, cm⁻¹): 3054, 2930, 2897, 2856, 1724, 1655, 1472, 1464, 1448, 1388, 1367, 1313, 1257, 1176, 1097, 1043, 980, 867, 837, 776, 668.; ¹H NMR (400 MHz, CDCl₃): δ 0.04 (s, 6H), 0.89 (s, 9H), 1.26 (t, J = 7.3Hz, 3H), 2.38-2.43 (m, 2H), 3.70 (t, J = 6.8 Hz, 2H), 4.15(q, J = 7.3 Hz, 2H), 5.85 (dt, J = 15.6 & 1.5 Hz, 1H), 6.91(dt, J = 15.6 & 1.5 Hz, 1H).; ¹³C NMR (100 MHz, CDCl₃): δ -5.35, 14.23, 25.63, 25.85, 35.67, 60.14, 61.55, 122.93, 145.83, 166.48.; HRMS (ESI+): Calcd. for C₁₃H₂₆O₃Si +H: 259.1729 found: 259.1721.

2.1.3. **Synthesis** of (2S,3R)-Ethyl5-(tertbutyldimethylsilyloxy)-2, 3-dihydroxypentanoate (15a): To a mixture of K_3 Fe(CN) ₆ (11.46 g, 34.82 mmole, 3.0 equiv.), K₂CO₃ (4.81 g, 34.82 mmole, 3.0 equiv.), MeSO₂NH₂ (1.10 g, 11.60 mmole, 1.0 equiv.), NaHCO₃ (2.92 g, 34.82 mmole, 3.0 equiv.), (DHQ)₂PHAL (90.4 mg, 0.116 mmole, 1 mol %), K₂OsO₄ .2H₂O (17.1 mg, 46.4 mmole, 0.4 mol %), t-BuOH (40 mL), and water (60 mL) were added. The mixture was stirred for 5 min and cooled to 0 °C in an ice bath. To the cooled mixture, a solution of the α , β -unsaturated ester **16** (3.0 g, 11.60 mmol) in t-BuOH (20 mL) was added. The reaction mixture was stirred at 0 °C for 24 h. It was then quenched with solid Na₂SO₃ (6 g) and stirred for 30 min. The solution was extracted with EtOAc (3 X 50 mL). The combined organic layers were washed with brine, dried (Na₂SO₄), and concentrated. The residue was purified by silica gel column chromatography using petroleum ether/EtOAc (9: 1 to 3: 2) as eluent to afford 15a (3.2 g, 94%). Similarly antipode 15b synthesized from 16 using above procedure using (DHQD)₂PHAL (2.9 g, 85%). **Compound 15a:** Viscous oil.; $[\alpha]_D^{20} = +3.00$ (c 0.5, CHCl₃).; IR (CHCl₃, cm⁻¹): 3472, 3020, 2957, 2931, 2859, 1737, 1472, 1388, 1258, 1217, 1129, 1097, 1026, 838, 759, 669.; ¹H NMR (400 MHz, CDCl₃): δ 0.074 (s, 6H), 0.89 (s, 9H), 1.29 (t, J = 7.1 Hz, 3H), 1.71–1.77 (m, 1H), 1.90– 1.99 (m, 1H), 3.20 (d, J = 7.1 Hz, 1H, OH), 3.25 (d, J = 4.9Hz, 1H, OH), 3.81-3.93 (m, 2H), 4.05 (dd, J = 3.5, 1.9 Hz, 1H), 4.16–4.20 (m, 1H), 4.25 (q, J = 7.1 Hz, 2H).; ¹³C NMR (100 MHz, CDCl₃): δ –5.56 (2C), 14.14, 18.14, 25.82 (3C), 35.26, 61.59, 61.81, 72.09, 73.67, 173.16.; HRMS (ESI+): Calcd. for $C_{13}H_{28}O_5Si + H$: 293.1784 found: 293.1770.

Compound 15b: Viscous oil.; $[\alpha]_D^{25} = -3.24$ (c 0.5, CHCl₃).; IR (CHCl₃, cm⁻¹): 3479, 3019, 2956, 2930, 2885, 1736, 1471, 1369, 1250, 1210, 1096, 837, 758, 667.; ¹H

(E)-Ethyl PT5-(tert) A NMR (CDCl₃, 400 MHz): δ 0.067 (s, 6H), 0.88 (s, 9H), pate (16): To a mixture quiv.) and NaOAc (1.94 (m, 1H), 3.19-3.26 (m, 1H), 3.81-3.91 (m, 2H), 4.04-4.06 (m, 1H), 4.16-4.18 (m, 1H), 4.24-4.30 (q, J = 7.4 & 14.5 (3.0 g, 5.76 mmole) in ion mixture was stirred petroleum ether/EtOAc d and filtered through a 1.28-1.31 (t, J = 7.0 Hz, 3H), 1.70-1.75 (m, 1H), 1.91-1.97 (m, 1H), 3.81-3.91 (m, 2H), 4.04-4.06 (m, 1H), 4.16-4.18 (m, 1H), 4.24-4.30 (q, J = 7.4 & 14.5 Hz, 2H); 13 C NMR (CDCl₃, 100 MHz): δ-5.54, -5.52, 14.14, 18.14, 25.82, 35.24, 61.61, 61.82, 72.11, 73.67, 173.18.; HRMS (ESI+): Calcd. for $C_{13}H_{28}O_5Si + Hi$: 293.1784 found: 293.1775.

2.1.4. Synthesis of ethyl (4*R*,5*S*)-5-(2-((*tert*-butyldimethylsilyl)oxy)ethyl)-2,2-dimethyl-1,3-

dioxolane-4-carboxylate (14a): To a solution of diol 15a (3.0 g, 10.26 mmole) in dry acetone (50 mL) was added 2, 2-dimethoxy propane (3.78 mL, 30.77 mmole, 3.0 equiv.) followed by p-TsOH (5 mg). The reaction mixture was stirred at room temperature for 12 h. Solid NaHCO₃ (0.5 g) was then added and stirred for 15 min. The solution was filtered through a pad of silica gel and washed with EtOAc and concentrated. The residue was purified by silica gel column chromatography using petroleum ether/EtOAc (9: 1 to 4: 1) as eluent to afford **14a** (3.07 g, 90%). Similarly antipode 15b was synthesized from 14b using above procedure to afford (2.71 g. 88%). Compound 14a: Colorless oil.; $[\alpha]_D^{20} = -1.0$ (c 0.5, CHCl₃).; IR (CHCl₃, cm ¹): 2988, 2955, 2930, 2858, 1758, 1472, 1382, 1372, 1257, 1217, 1192, 1168, 1102, 1036, 939, 837, 776, 759, 667.; ¹H NMR (400 MHz, CDCl₃): δ 0.039 (s, 6H), 0.871 (s, 9H), 1.26 (t, J = 7.1 Hz, 3H), 1.42 (s, 3H), 1.44 (s, 3H), 1.80–1.89 (m, 1H), 1.96–2.06 (m, 1H), 3.72–3.82 (m, 2H), 4.17–4.29 (m, 4H).; ¹³C NMR (100 MHz, CDCl₃): δ –5.45, 14.12, 18.20, 25.8, 27.13, 36.46, 59.38, 61.22, 75.98, 78.96, 110.66, 170.69.; HRMS (ESI+): Calcd. for $C_{16}H_{32}O_5Si + H$: 293.1784 found: 293.1788.

Compound 14b: Colorless oil.; $[\alpha]_D^{25} = +1.41$ (c 0.5, CHCl₃).; IR (CHCl₃, cm⁻¹): 3021, 2991, 2931, 2884, 1752, 1472, 1484, 1383, 1372, 1256, 1216, 110, 811, 837, 758, 667; ¹H NMR (CDCl₃, 400MHz): δ 0.041 (s, 6H), 0.87 (s, 9H), 1.26-1.29 (t, J = 7.0 Hz, 3H), 1.42 (s, 3H), 1.45 (s, 3H), 1.80-1.87 (m, 1H), 1.97-2.03 (m, 1H), 3.70-3.80 (m, 2H), 4.17-4.28 (m, 4H).; ¹³C NMR (CDCl₃, 100MHz): δ 5.44, -5.39, 14.15, 18.23, 25.64, 25.85, 27.15, 36.48, 59.40, 61.27, 75.98, 78.96, 110.68, 170.73.; HRMS (ESI+): Calcd. for C₁₆H₃₂O₅Si + H: 293.1784 found: 293.1786.

2.1.5. Synthesis of ethyl(4S, 5R)-5-(2-hydroxyethyl)-2, 2-dimethyl-1, 3-dioxolane-4-carboxylate (19a): To a solution of **14a** (8.0 g, 0.024 mole) in THF (40 mL) at 0 °C was added TBAF (1.0 M solution in THF, 28.91 mL, 0.028 mole) drop wise. The resulting brown solution was stirred for 12 h at 0-5 °C. up on consumption of starting material as indicated by TLC, the solvent was removed in vacuum, and the crude residue was purified by flash column chromatography (hexanes/EtOAc) to afford diol 19a (5.0g, 95%). Similarly antipode 19b was synthesized from 14b using above procedure to afford (4.71 g. 94%). **Compound 19a:** Colorless oil.; $[\alpha]_D^{25} = -1.76$ (c 0.9, CHCl₃).; IR (CHCl₃ cm⁻¹): 3501, 2939, 2988, 1755, 1466, 1373, 1214, 1098, 880, 872.; ¹H NMR (400 MHz, CDCl₃): $\delta 1.26$ (t, J = 7.12 Hz, 3H), 1.42 (s, 3H), 1.46 (s, 3H), 1.89-2.18 (m 2H), 3.8 (t, J = 5.76 Hz, 2H), 4.19 (m, 4H).;¹³C NMR (100 MHz, CDCl₃): δ 14.09, 25.50, 27.00, 35.76, 59.97, 61.52, 77.72, 78.92, 110.97, 170.73.; HRMS (ESI+): Calcd. for $C_{10}H_{18}O_5+H$: 219.1232 found: 219.1243.

Compound 19b: Colorless oil.; $[\alpha]_0^{25} = +1.27$ (c $[0.5]_A$ A $[0.5]_A$ SHRMS (ESI+): Calcd. for $C_8H_{15}O_3I$ + H: 287.0144 found: CHCl₃).; IR (CHCl₃, cm⁻¹):3520, 3018, 2992, 2939, 1749, 287.0146. 1456, 1383, 1216, 1099, 755, 667.; H NMR (CDCl₃, 400 MHz): δ 1.25-1.29 (t, J = 7.0 Hz, 3H), 1.41 (s, 3H), 1.44 (s, 3H), 1.89-1.95 (m, 1H), 1.96-2.07 (m, 1H), 2.39 (bs, 1H), 3.77-3.81 (q, J = 5.3 & 10.1 Hz, 2H), 4.18-4.28 (m, 4H).; 13 C NMR (CDCl₃, 100 MHz): δ 14.12, 25.52, 27.02, 35.79, 59.95, 61.56, 76.7, 78.94, 110.99, 170.77.; HRMS (ESI+): Calcd. for $C_{10}H_{18}O_5 + H$: 219.1232 found: 219.1240.

2.1.6. Synthesis of ethyl (4R, 5S)-5-(2-iodoethyl)-2, 2dimethyl-1, 3-dioxolane-4-carboxylate (20a): To the solution of PPh3 (7.07g, 0.027mole) in anhydrous CH2Cl2 (22.5 ml) under N_2 , was added 1H-imidazole $(1.82g,\, 0.027$ mole) and then with I_2 (6.78g, 0.027 mole). A solution of **19a** (4.5 g, 0.020mole) in anhydrous CH₂Cl₂ (22.5 ml) was added, and the mixture was stirred at r.t. for 2 h. Evaporation of the solvent gave a crude product, which was filtered through a short silica-gel column (Hexane/Ethylacetate 9 : 1; R_f: 0.5) to give pure **20a** (5.5g, 82%). Similarly antipode 20b synthesized from 19b using above procedure (5.8 g. 83%). Compound 20a: Yellow oil.; $[\alpha]_D^{25} = -3.08$ (c 0.5, CHCl₃).; IR (CHCl₃, cm⁻¹): 2988, 2908, 2937, 1759, 1731, 1445, 1352, 1200, 1036, 854.; ¹H NMR(400 MHz, CDCl₃): δ 1.289 (t, J = 7.16 Hz, 3H), 1.43 (s, 3H), 1.45 (s, 3H), 2.13-2.21 (m 1H), 2.27-2.35 (m, 1H), 3.21-3.35 (m, 2H), 4.12-4.28 (m, 4H).; ¹³C NMR(100 MHz, CDCl₃): δ 0.51, 14.19, 25.60, 27.06, 37.74, 61.53, 78.37, 78.68, 111.25, 170.33.; HRMS (ESI+): Calcd. for $C_{10}H_{17}O_4I + H$: 329.0250 found: 329.0235.

Compound 20b: Yellow oil.; $[\alpha]_D^{25} = +3.52$ (c 0.51, CHCl₃).; IR (CHCl₃, cm⁻¹): 3020, 2989, 2938, 2906, 1756, 1732, 1445, 1372, 1238, 1035, 853, 757, 667.; ¹H NMR (CDCl₃, 400 MHz): δ 1.28-1.31 (t, J = 7.0 Hz, 3H), 1.42 (s, 3H), 1.44 (s, 3H), 2.14-2.19 (m, 1H), 2.20-2.32 (m, 1H), 3.20-3.34 (m, 2H), 4.11-4.20 (m, 2H), 4.20-4.27 (m, 2H).; ¹³C NMR (CDCl₃, 100 MHz): δ 0.59, 14.22, 25.62, 27.08, 37.74, 61.56, 78.37, 78.68, 111.26, 170.35.; HRMS (ESI+): Calcd. for $C_{10}H_{17}O_4I$ + H: 329.0250 found: 329.0257.

2.1.7. Synthesis of ((4S, 5S)-5-(2-iodoethyl)-2, 2-iodoethyl)dimethyl-1, 3-dioxolan-4-yl) methanol 13a: To the solution of **20a** (5.0 g, 0.015 mole) in MeOH (10.0 mL) and THF (40.0 mL) at 0-10 °C was added NaBH₄ (0.855g, 0.022 moles) lot wise (H₂ evolution) under inert atmosphere. After complete addition temperature was allowed to come to 25-35 °C and maintained for 1-2 hours by TLC monitoring. Upon absence of starting material, reaction mass was cooled to 0-10 °C and quenched with saturated ammonium solution and product was extracted with 2X65 mL ethyl acetate, combined organic layers evaporated and the residue was purified by silica gel column chromatography using petroleum ether/EtOAc (9: 1 to 7: 3) as eluent to afford **13a** (3.9 g, 89 %). Similarly antipode 13b synthesized from 20b using above procedure (3.71 g. 84%).

Compound 13a:Pale yellow oil.; $[\alpha]_D^{25} = -7.99$ (c 0.5, CHCl₃).; IR (CHCl₃, cm⁻¹): 3436, 2985, 2933, 2876, 1437, 1371, 1218, 1090, 847.; ¹H NMR (400 MHz, CDCl₃): δ 1.40 (s, 6H), 1.92 (bs, 1H), 2.07-2.12 (m, 2H), 3.20-3.26 (m, 1H), 3.29-3.35 (m, 1H), 3.61-3.64 (m, 1H), 3.75-3.81 (m, 2H), 39.9-3.98 (m, 1H),; ¹³C NMR (100 MHz, CDCl₃): δ 1.00, 26.99, 27.28, 37.43, 61.75, 80.60, 81.12, 109.19.;

Compound 13b: Pale yellow oil.; $[\alpha]_D^{25} = +5.06$ (c 0.5, CHCl₃).; IR (CHCl₃, cm⁻¹): 3435, 2985, 2933, 2876, 1437, 1371, 1218, 1049, 1089, 847.; H NMR (CDCl₃, 400 MHz): δ 1.39 (s, 3H), 2.03-2.12 (m, 3H), 3.19-3.34 (m, 2H), 3.61-3.63 (m, 1H), 3.74-3.80 (m, 2H), 3.80-3.97 (m, 2H).; ¹³C NMR (CDCl₃, 100 MHz): δ 1.09, 26.99, 27.28, 37.40, 61.75, 77.34, 80.63, 109.20.; HRMS (ESI+): Calcd. for $C_8H_{15}O_3I + H: 287.0144$ found: 287.0147.

2.1.8. Synthesis of ((4S, 5S)-5-(2-(iodotriphenyl-15phosphanyl) ethyl)-2, 2-dimethyl-1, 3-dioxolan-4-yl) methanol (7a): To the solution of 13a (3.5 g, 0.012mole) in Acetonitrile (35 mL) was added TPP (6.41 g, 0.024 mole) and reflux for 12 h. solvent was evaporated and obtained crude residue was purified by column chromatography by using 5% MeOH/ DCM to afford 7a (5.9g, 88%). Similarly antipode 7b synthesized from 13b using above procedure to afford (6.1 g, 90%). Compound **7a:** Waxy solid.; $[\alpha]_D^{20} = -1.66$ (c 0.5, CHCl₃).; IR (CHCl₃, cm⁻¹): 3377, 2987, 2934, 2880, 2247, 2198, 1587, 1483, 1376, 1216, 1110, 1053, 909, 721, 687, 642.; ¹H NMR (400 MHz, CDCl₃): δ1.32 (s, 3H), 1.34 (s, 3H), 1.81-1.92 (m 1H), 2.22-2.32 (m, 1H), 3.56-3.67 (m, 1H), 3.73-3.75 (m, 1H), 3.83-3.95 (m, 4H), 4.38-4.42 (m, 1H), 7.68-7.83 (m, 15H).; ¹³C NMR (100 MHz, CDCl₃): δ 19.34, 19.86, 26.58, 26.62, 26.87, 27.10, 62.13, 78.73, 78.89, 79.98, 108.68, 117.46, 118.32, 130.51, 130.63, 133.59, 133.69, 135.19, 135.22.; HRMS (ESI+): Calcd. for C₂₆H₃₀O₃P-I: 421.1933 found: 421.1930.

Compound 7b: Waxy solid.; $[\alpha]_D^{25} = + 4.92$ (c 0.5, CHCl₃).; IR (CHCl₃ cm⁻¹): 3368, 3059, 3014, 2966, 2937, 1588, 1438, 1216, 1113, 1060, 762, 724, 662.; ¹H NMR (CDCl₃, 400 MHz): δ 1.30 (s, 3H), 1.32 (s, 3H), 1.80-1.88 (m, 1H), 2.01-2.27 (m, 1H), 3.41-3.60 (m, 1H), 3.67-3.87 (m, 3H), 4.33-4.38 (m, 1H), 7.67-7.82 (m, 15H).; 13 C NMR (CDCl₃, 100 MHz): 19.32, 19.85, 26.54, 26.59, 26.89, 27.12, 61.95, 78.53, 78.69, 80.03, 108.73, 117.35, 118.21, 130.57, 130.70, 133.57, 133.67, 135.27, 135.30.; HRMS (ESI+): Calcd. for $C_{26}H_{30}O_3P - I$: 421.1933 found: 421.1928.

2.1.9. Synthesis of (R, Z)-7-((4S, 5S)-5-(hydroxymethyl)-2, 2-dimethyl-1, 3-dioxolan-4-yl) hept-5-en-2-ol (22a): To the solution of 7a (5.3 g, 9.7 mmole) in THF (25 mL) was added 1.0 molar NaHMDS (35.29 mL, 35.2 mmole) at -30 °C under N₂ atmosphere (yellow suspension to orange colored solution). for 15-30 min. The solution of lactol 6 (0.9g, 8.8 mmole) was added as a solution in THF (10 mL) at -30 °C to above orange colored reaction mass, maintained addition 1-2 h, then quenched with sat. NH₄Cl (45 mL) and allowed to come to rt and stirred for 1-2 h. The product was extracted with 3 X 45 mL of ethyl acetate solvent was evaporated and obtained crude residue was purified by column chromatography to afford 22a(1.85 g, 86%). Similarly antipode 22bwas synthesized from 7b using above procedure to afford (1.75g, 82%).

Compound 22a: Viscous oil.; $[\alpha]_D^{20} = -3.5$ (c 0.2, CHCl₃).; IR (CHCl₃, cm⁻¹): 3413, 2984, 2927, 2250, 1650, 1455, 1375, 1217, 1164, 1055, 904, 842, 725, 648.; ¹H NMR (400 MHz, CDCl₃): δ 1.17-1.18 (d, J = 6.2 Hz, 3H), 1.39 (s, 3H), 1.41 (s, 3H), 1.47-1.53 (m 2H), 2.06-2.28 (m, 2H), 2.29-2.49 (m, 3H), 3.46 (bs, 1H), 3.61-3.64 (m, 1H), 3.73-3.82 (m, 3H), 3.89-3.95 (m, 1H), 5.43-5.56 (m, 2H).; ¹³C NMR (100 MHz, CDCl₃): δ 132.28, 124.63, 108.70,

80.86, 66.77, 62.07, 38.45, 27.17, 26.95, 23.70, 23.52; $A \times U S$ (400 MHz, CDCl₃): δ 6.82-6.88 (dd, J = 5.7, 15.8 Hz, 1H), GC-MS (ES): 229 [M⁺-15]. 6.08-6.12 (dd, J = 1.4 Hz, 15.6 Hz, 1H), 4.11-4.23 (m,

Compound 22b: Viscous oil.; $[a]_D^{20} = +3.0$ (c 0.2, CHCl₃).; IR (CHCl₃, cm⁻¹): 3429, 3015, 2989, 2932, 1634, 1373, 1216, 1057, 843, 755, 667 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 1.17-1.19 (d, J = 6.2 Hz, 3H), 1.40 (s, 3H), 1.42 (s, 3H), 1.47-1.53 (m 2H), 2.06-2.51 (m, 5H), 3.58-3.65 (m, 2H), 3.73-3.80 (m, 3H), 3.89-3.96 (m, 1H), 5.43-5.56 (m, 2H).; ¹³C NMR (100 MHz, CDCl₃): δ 132.31, 124.67, 108.72, 80.85, 66.75, 62.02, 38.44, 27.19, 26.96, 23.77, 23.53.; GC-MS (ES): 229 [M⁺-15].

(R)-7-((4S,5S)-5-4.3.1.10. **Synthesis** of (hydroxymethyl)-2, 2-dimethyl-1, 3-dioxolan-4-yl) **heptan-2-ol (8a):** To a solution of **22a**(1.8 g, 7.3 mmole) in ethyl acetate (36 mL) at rt added 20% Pd(OH)₂ by wt.and hydrogenated for 4 h at 40psi, upon completion of reaction catalyst was removed by filtration, thus obtained filtrate was evaporated to afford 8a (1.75 g, 96 %). Similarly antipode 8b was synthesized from 22busing above procedure to afford (1.88g, 98%). Compound 8a: Colorless oil.; $[\alpha]_D^{20} = +2.33$ (c 0.2, CHCl₃).; IR (CHCl₃, cm⁻¹): 3450, 2988, 2965, 2933, 2860, 1438, 1457, 1379, 1243, 1050, 756, 666.; ¹H NMR (400 MHz, CDCl₃): δ 1.16-1.18 (d, J = 6.2 Hz, 3H), 1.33-2.12 (m, 18H), 3.56-3.60 (m, 1H), 3.69-3.89 (m 4H).; ¹³C NMR (100 MHz, CDCl₃): 8 23.49, 25.55, 25.57, 25.95, 27.01, 27.37, 29.59, 32.93, 39.16, 61.97, 68.05, 81.43, 108.57.; HRMS (ESI+): Calcd. for C₁₃H₂₇O4+H: 247.1909 found: 247.1899.

Compound 8b: Colorless oil.; $[\alpha]_D^{20} = -3.11$ (c 0.2, CHCl₃).; IR (CHCl₃ cm⁻¹): 3435, 3018, 2990, 2934, 2860, 1458, 1372, 1215, 770, 669.; ¹H NMR (400 MHz, CDCl₃): δ 1.16-1.17 (d, J = 6.2 Hz, 3H), 1.33-1.79 (m, 18H), 3.56-3.60 (m, 1H), 3.69-3.80 (m 3H), 3.84-3.87 (m, 1H).; ¹³C NMR (100 MHz, CDCl₃): δ 23.47, 25.55, 25.94, 27.01, 27.36, 29.56, 32.91, 61.99, 68.05, 81.46, 108.58.; GC-MS (ES): 231 [M⁺-15].

2.1.11. Synthesis of Ethyl (E)-3- $\{(4S, 5S)$ -5-[(R)-6-Hydroxyheptyl]-2, 2-dimethyl-1, 3-dioxolan-4ylacrylate (9a): To diol 8a (175 mg, 0.71 mmol) in CH₂Cl₂ (3 mL) at 0 °C were added BAIB (263 mg, 0.81 mmol) and TEMPO (10 mg, 0.06 mmol). After stirring at room temperature for 2 h, the mixture was cooled to 0 °C, and (ethoxycarbonylmethylene) triphenylphosphorane (21) (320 mg, 0.92 mmol) was added. The stirring was continued for another 2 h at room temperature. After completion of the reaction, the mixture was concentrated under reduced pressure. The residue was purified by column chromatography (hexanes/EtOAc) to afford α , β unsaturated ester **9a** (210 mg, 94%). Similarly diastereomer 9b synthesized from 8b using above procedure to afford (195 mg, 88%). Compound 9a: Pale yellow liquid.; $\left[\alpha\right]_{D}^{20} = +10$ (c 0.1, CHCl₃).; IR (CHCl₃. cm⁻¹): 3504, 2984, 2933, 2860, 2250, 1716, 1660, 1459, 1373, 1240, 1168, 1103, 1036, 907, 725, 647.; ¹H NMR (400 MHz, CDCl₃): δ 6.82-6.87 (dd, J = 5.8, 15.6 Hz, 1H), 6.08-6.12 (d, J = 15.6 Hz, 1H), 4.16-4.23 (m, 2H), 4.11-4.15 (m, 1H), 3.67–3.82 (m, 2H), 1.27–1.62 (m, 19H), 1.16-1.18 (d, J = 6.16Hz, 3H).; ¹³C NMR (100 MHz, CDCl₃): 8 166.02, 144.11, 122.73, 109.34, 80.21, 68.03, 60.59, 39.15, 31.96, 29.54, 27.24, 26.63, 25.91, 25.54, 23.49, 14.18.; GC-MS (ES): 299 [M⁺-15].

Compound 9b: Pale yellow liquid., $[\alpha]_D^{20} = -7.9$ (c 0.1, CHCl₃).; IR (CHCl₃, cm⁻¹): 3528, 3018, 2988, 2936, 2881, 1717, 1662, 1459, 1371, 1216, 1035, 756, 668.; ¹H NMR

(400 MHz, CDCl₃): δ 6.82-6.88 (dd, J = 5.7, 15.8 Hz, 1H), 6.08-6.12 (dd, J = 1.4 Hz, 15.6 Hz, 1H), 4.11-4.23 (m, 3H), 3.69–3.80 (m, 2H), 1.40–1.64 (m, 19H), 1.16-1.18 (d, J = 6.2 Hz, 3H).; ¹³C NMR (100 MHz, CDCl₃): δ 166.05, 144.11, 122.74, 109.336, 80.58, 80.56, 68.04, 60.62, 39.16, 31.95, 29.55, 27.26, 25.94, 25.54, 23.49, 14.21.; GC-MS (ES): 299 [M⁺-15].

Acknowledgments

The authors would like to thank Dr. H. Rama Mohan and Dr. Reddy's Laboratories for their support and encouragement. We also thank analytical department of Dr. Reddy's Laboratories Ltd for providing analytical support.

References and notes

- (a) Hirota, A.; Isogai, A.; Sakai, H. Agric. Biol. Chem. 1981, 45, 799; (b) Hirota, A.; Sakai, H.; Isogai, A.; Kitano, Y.; Ashida, T.; Hirota, H.; Takahashi, T. Agric. Biol. Chem. 1985, 49, 903; (c) Hirota, H.; Hirota, A.; Sakai, H.; Isogai, A.; Takahashi, T. Bull. Chem. Soc. Jpn. 1985, 58, 2147; (d) Hirota, A.; Sakai, H.; Isogai, A. Agric. Biol. Chem. 1985, 49, 731.
- Fujii, Y.; Fukuda, A.; Hamasaki, T.; Ichimoto, I.; Nakajima, H. Phytochemistry 1995, 40, 1443.
- Smith, C. J.; Abbanat, D.; Bernan, V. S.; Maiese, W. M.; Greenstein, M.; Jampa, J.; Tahir, A.; Ireland, C. M. J. Nat. Prod. 2000, 63, 142.
- (a) Zhang, H.; Tomoda, H.; Tabata, N.; Miura, H.; Namikoshi, M.; Yamaguchi, Y.; Masuma, R.; Omura, S. *J. Antibiot.* 2001, 54, 635. (b) Chada, R, R.; Devatha, S.; Nagavaram, N, R.; Eur. J. Org. Chem. 2013, 18, 3786-3796
- (a) Si, D.; Sekar, N. M.; Kaliappan, K. P.; Org. Biomol. Chem. 2011, 9, 6988; (b) Rajesh, K.; Suresh, V.; Selvam, J. J. P.; Rao, C. B.; Venkateswarlu, Y. Synthesis, 2010, 1381; (c) Kaliappan, K. P.; Si, D. Synlett, 2009, 2441; (d) Banwell, M. G.; Loong, D. T. J. Org. Biomol. Chem. 2004, 2, 2050; (e) Banwell, M. G.; Jolliffe, K. A.; Loong, D. T. J.; McRae, K. J.; Vounatsos, F.J. Chem. Soc. Perkin Trans. 1 2002, 22; (f) Solladié, G.; Almario, A.; Tetrahedron: Asymmetry 1995, 6, 559; g) Solladié, G.; Antonio, A.; Pure Appl. Chem. 1994, 66, 2159; (h) Ichimoto, I.; Sato, M.; Kirihata, M.; Ueda, H.; Chem. Express, 1987, 2, 495; (i) Maemoto, S.; Mori, K. Chem. Lett. 1987, 109; (j) Mori, K.; Maemoto, S. Liebigs Ann. Chem. 1987, 863.
- (a) Yadav, J. S.; Mandal, S. S. Synlett, 2011, 19, 2803;
 (b) Prasad, K. R.; Gandi, V. R. Tetrahedron: Asymmetry, 2011, 22, 499;
 (c) Xing, Y.; O'Doherty, G. A. Org. Lett. 2009, 11, 1107;
 d) Wang, W.-K.; Zhang, J.-Y.; He, J.-M.; Tang, S.-B.; Wang, X.-L.; She, X.-G.; Pan, X.-F. Chin. J. Chem. 2008, 26, 1109;
 (e) Sharma, G. V. M.; Reddy, J. J. R.; Reddy, K. L. Tetrahedron Lett. 2006, 47, 6537;
 (f) Sharma, G. V. M.; Reddy, J. J. R.; Reddy, K. L. Tetrahedron Lett. 2006, 47, 6531;
 (g) Pandey, S. K.; Kumar, P. Tetrahedron Lett. 2005, 46, 6625;
 (h) Austin, K. A. B.; Banwell, M. G.; Loong, D. T. J.; Rae, D.; Willis, A. C. Org. Biomol. Chem. 2005, 3, 1081.
- (a) Reddy, C. R.; Rao, N. N.; Sujitha, P.; Kumar, C. G. Synthesis, 2012, 44, 1663; (b) Trost, B. M.; Aponick, A. J. Am. Chem. Soc. 2006, 128, 3931; (c) Srihari, P.; Bhasker, E. V.; Harshavardhan, S. J.; Yadav, J. S. Synthesis, 2006, 4041; d) Franck, X.; Araujo, M. E. V.;

Tetrahedron Lett. 2001, 42, 2801; (e) Prasad, K. R.; Gandi, V. R. Tetrahedron: Asymmetry, 2010, 21, 275; f) Ferrie, L.; Reymond, S.; Capdevielle, P.; Cossy, J. Synlett, 2007, 2891.

- 8. (a) Reddy, C. R.; Rao, N. N. Tetrahedron Lett. 2009, 50, 2478; (b) Chou, C.-Y.; Hou, D. -R. J. J. Org. Chem. 2006, 71, 9987; (c) Banwell, M. G. Loong, D. T. J.; Willis, A. C. Aust. J. Chem. 2005, 58, 511.
- 9. (a) Si, D.; Kaliappan, K. P. Synlett, 2012, 19, 2822; (b) Lu, K.-J.; Chen, C.-H.; Hou, D.-R. Tetrahedron, 2009, 65, 225; (c) Xing, Y.; Penn, J. H.; O'Doherty, G. A. Synthesis, 2009, 2847; (d) Xing, Y.; O'Doherty, G. A. Org. Lett. 2009, 11, 1107.
- 10. (a) Banwell, M. G.; Loong, D. T. J.; Willis, A. C. Aust. J. Chem. 2005, 58, 511. (b) Hudlicky, T.; Gonzalez, D.; Gibson, D. T. AldrichimicaActa1999, 32, 35.
- 11. (a) Banwell, M. G.; Loong, D. T. J. Org. Biomol. Chem. 2004, 2, 2050; (b) Banwell, M. G.; Jolliffe, K. A.; Loong, D. T. J.; McRae, K. J.; Vounatsos, F. J. Chem. Soc., Perkin Trans. 1 2002, 22; (c) Solladie, G.; Almario, A. Tetrahedron: Asymmetry, 1995, 6, 559; (d) Solladie, G.; Almario, A. Pure Appl. Chem. 1994, 66, 2159; (e) Ichimoto, I.; Sato, M.; Kirihata, M.; Ueda, H. Chem. Express, 1987, 2, 495; (f) Maemoto, S.; Mori, K. Chem. Lett. 1987, 109; (g) Mori, K.; Maemoto, S. Liebigs Ann. Chem. 1987, 863; (h) Pandey, S. K.; Kumar, P. Tetrahedron Lett. 2005, 46, 6625; (i) Austin, K. A. B.; Banwell, M. G.; Loong, D. T. J.; Rae, A. D.; Willis, A. C. Org. Biomol. Chem. 2005, 3, 1081; (j) Banwell, M. G.; Loong, D. T. J.; Willis, A. C. Aust. J. Chem. 2005, 58, 511; (k) Chou, C.-Y.; Hou, D.-R. J. Org. Chem. 2006, 71, 9887; (1) Sharma, G. V. M.; Reddy, K. L.; Reddy, J. J. Tetrahedron Lett. 2006, 47, 6537.
- 12. (a) Mori K.; Maemoto, S. Liebigs Ann. Chem., 1987, 863; (b) Maemoto S.; Mori, K. Chem. Lett. 1987, 109.
- 13. (a) Mandrioli, R.; Mercolini, L.; Saracino, M.A.; Raggi, M. A.; Curr. Med. Chem. 2012, 19, 1846; (b) Ghosh, A. K.; Swanson, L. J. Org. Chem. 2003, 68, 9823; (c) Lipshutz, B. H.; Lower, A.; Kucejko, R. J.; Noson, K. Org. Lett. 2006, 8, 2969; (d) Stangeland, E. L.; Sammakia, T. J.Org. Chem. 2004, 69, 2381; (e) Hilbon. J. W.; Lu, Z. H.; Jurgens, A. R.; Fang, Q. K,; Byers, P.; Wald, S. A.; Senanayake, C. H. Tetrahedron Lett. 2001. 42, 8919; (f) Datrika, R.; Kallam, S. R.; Khobare, S. R.; Gajare, V. S.; Kommi, M.; Mohan, H. R.; Vidavalur, S.; Pratap, T. V. Tetrahedron: Asymmetry, 2016, 27, 603; (g) Datrika, R.; Kallam, S. R.; Gajare, V.; Khobare, S.; Rama, V. S.; Kommi, M.; Hindupur, R. M.; Vidavulur, S.; Tadikonda, P. V. Chemistry Select, 2017, 2, 5828.
- 14. (a) Jacobs, H.; Berryman, K.; Jones, J.; Gopalan, A. Synth. Commun. 1990, 7, 999; (b) Tsuboi, S.; Sakamoto, J.; Kawano, T.; Utaka, M.; Takeda, A. J. Org. Chem. 1991, 56, 7177; (c) White, J. D.; Somers, T. C.; Reddy, G. N. J. Org. Chem. 1992, 57, 4991; (d) Taylor, S. K.; Atkinson, R. F.; Almli, E. P.; Carr, M. D.; Van Huis, T. J.; Whittaker, M. R. Tetrahedron: Asymmetry, 1995, 6, 157; (e) Nair, V.; Prahakaran, J.; George, T. G. Tetrahedron, 1997, 53, 15061; (f) Rodriguez, A. D.; Borzecka, W.; Lavandera, I.; Gotor, V. ACS Catal. 2014, 4, 386.
- 15. Datrika, R.; Kallam, S.; Rambabu, K.; Vidavulur, S.; Pratap, T. V.; Synth. Commun. 2018, 48, 2801-2808.

- Jullian, J.-C.; Hocquemiller, R. Figadere, D. A. N. St. A. V. St., Westbrook, J. A.; Schaus, S. E. J. Am. Chem. Soc. 2004, 126, 11440; (b) Mortensen, M. S.; Osbourn, J. M.; O' Doherty, G. A. Org. Lett. 2007, 9,
 - 17. (a) Li, De-Run.; Tu, Y. Qiang.; Lin, Guo-Qiang.; Zhou, Wei-Shan. Tetrahedron Lett. 2003, 44, 8729; (b) Li, De-Run.; Zhang, Dong-Hui.; Sun, Cai-Yun.; Zhang, Ji-Wen.; Yang, Li.; Chen, Jian.; Liu, Bo.; Su, Ce.; Zhou, Wei-Shan.; Lin, Guo-Qiang. Chem. Eur. J, 2006, 12, 1185; (c) Fernandes, R.A.; Chavan, V. P. Tetrahedron: Asymmetry, 2011, 22, 1312.
 - 18. Corey, E. J.; Venkateswarlu, A. J. Am. Chem. Soc. **2008**, 94, 6190.
 - 19. Badalassi, F.; Klein, G.; Crotti, P.; Reymond, J-L. Helv. Chim. Acta. 2002, 85, 3090.
 - 20. Matsumura, Y.; Mori, N.; Nakano, T.; Sasakura, H.; Matsugi, T.; Hara H.; Morizawa. Y. Tetrahedron Lett. **2004,** 45, 1527.
 - 21. Pearlman, W. M. Tetrahedron Lett. 1967, 8, 1663.
 - 22. (a) Reddy, Ch. R.; Rao, N. N.; Sujitha, P.; Kumar, C. G. Eur. J. Org. Chem. 2012, 1819; (b) Vatele, J. M. Tetrahedron Lett. 2006, 47, 715.
 - 23. GC-Ms Conditions: Column:HP-5, 30 m*0.32 mm*0.25 microns; Oven program: 100 °C (0 min hold); 10 °C/min to 270°C (10 min hold); Flow: 1.0 mL /min, Injector Temperature:220°C; Auxillary temperature: 380 °C; Injection volume: 0.2 microliter; Split :100:1; Diluent: Acetonitrile; Mass conditions: Source: Electron impact; Analyser :Qudrupole; Source temperature:230°C; Qudrupole; Temperature:150°C; Tune file: Auto tune; Solvent delay:3 min; Mass range: 20-700 a.m.u
 - 24. (a) Xing, Y.; O' Doherty, G. A. Org. Lett. 2009, 11, 1107; (b) Reddy, C. R.; Rao, N. N. Tetrahedron Lett. 2009, 50, 2478.
 - 25. Austin, K. A. B.; Banwell, M. G.; Loong, D. T. J.; Rae, D.; Willis, A. C. Org. Biomol. Chem. 2005, 3, 1081.

Supplementary Material

Supplementary data (detailed experimental analysis and spectral analysis including ¹H, ¹³C, and HRMS associated with this article can be found, in the online version, at http://