Synthesis of Chiral Pyrimidin-2(1*H*)-ones from *N*-Carbamoyl Amino Acids

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A series of previously unknown pyrimidin-2(1H)-ones containing chiral amino acid fragments was synthesized from 1,1,3,3-tetramethoxypropane and N-carbamoyl derivatives of amino acids under acidic conditions.

Key words: Heterocycles, Amino Acids, Condensation, Chirality, Pyrimidines

Introduction

Amino acid derivatives of pyrimidinones are nucleoside analogs and are interesting as biologically active compounds or their precursors [1]. It is known that pyrimidin-2(1H)-ones **1** containing peptide fragments reveal anti-inflammatory activity [2]. Their aza analogs -1.2.4-triazin-3(2H)-ones **2** - do not show or show only low anti-inflammatory activity, however these compounds have been proved to show strong analgesic properties [3]. Pyrimidinones **3** are useful in the treatment or prevention of the diseases mediated by the activation of the β 3-adrenoceptor [4, 5].

The published methods [1-4] for the synthesis of compounds 1-3 lead to products with racemic amino

acid fragments, but it is known that chirality is important for bioactivity. To the best of our knowledge only few methods of synthesis of chiral α -amino acid derivatives of pyrimidines are known [6–8]. One method of synthesis of pyrimidines with chiral α -amino acid fragments at the heterocyclic ring has been reported. The alkylation of cytosine with esters of (S)-lactic acid leads to alkyl derivatives of cytosine (de = 56) [8].

Results and Discussion

In this work the synthesis of pyrimidin-2(1H)ones with amino acid residues is described. It is known that N-carbamovl amino acids easily cyclize to give hydantoins under acidic conditions [9, 10]. The condensation of 1,1,3,3-tetramethoxypropane with Ncarbamoyl derivatives of amino acids in the presence of hydrochloric acid in methanol gives methyl esters of pyrimidinones 5–11 (Scheme 1, Fig. 1). The change of solvent from methanol to ethanol, 2-propanol or water leads to the disappearance of pyrimidinones in the products of the reaction. In the reactions with Ncarbamoyl derivatives of other amino acids such as Lvaline, L-leucine, L-isoleucine, D,L-tryptophan, or Lserine we failed to obtain the corresponding pyrimidinones. Acid hydrolysis of the esters 5 and 6 leads to carboxylic acids 12 (67%) and 13 (76%), respectively (Scheme 1).

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R	Configuration	Product	Yield (%)
CH ₃	(S)	5	43
CH ₃	(R)	6	21
CH ₂ CH ₂ SCH ₃	(S)	7	55
CH ₂ CH ₂ SCH ₃	(R)	8	15
CH ₂ CH ₂ SCH ₃	(R,S)	9	34
CH ₂ Ph	(R,S)	10	40
CH ₂ (<i>p</i> -C ₆ H ₄ OH)	(S)	11	52

Scheme 1. Synthesis of pyrimidin-2(1H)-one derivatives.

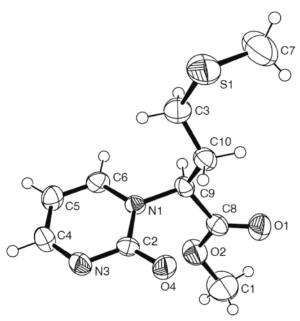


Fig. 1. ORTEP diagram of 7.

In addition we tried to prepare 1-substituted pyrimidin-2-ones from 2-phenylmalonaldehyde **14**. When a mixture of 2-phenylmalonaldehyde and a substituted urea was refluxed in toluene in the presence of *p*-TsOH [11] hydantoins were formed as the sole products from the *N*-carbamoyl derivatives of L-methionine, L-lysine, L-leucine, L-isoleucine, and L-tyrosine. Using different reaction times in the reaction with L-valine led to the products **15** and **16** besides the corresponding hydantoin derivative (Scheme **2**).

Obviously the first step of the reaction leads to the condensation product **15** (Fig. 2). Under forced conditions **15** is cyclized and subsequently decarboxylated to give **16** (Fig. 3). We could not find conditions which allowed the isolation of an intermediate. When benzene or p-xylene were used as solvents, hydantoins were the sole reaction products.

Experimental Section

All commercially obtained solvents and reagents were used as received; N-carbamoyl amino acids [12] and 2-

Scheme 2.

Fig. 2. ORTEP diagram of 15.

phenylmalonaldehyde [11] were synthesized by known methods. ¹H, ¹³C NMR spectra were recorded on a Bruker DRX-400 spectrometer; tetramethylsilane (TMS) was used as an internal standard. The optical rotation was measured on a Perkin Elmer polarimeter. Mass spectra were recorded on a Bruker Daltonics MicroTOF-Q II mass spectrometer with electrospray ionization.

General procedure for the synthesis of compounds 5–11

1,1,3,3-Tetramethoxypropane (821 μ L, 5.0 mmol) and conc. HCl (12 M, 3 mL) were added to a suspension of the corresponding *N*-carbamoyl amino acid (5.0 mmol) in methanol (10 mL). After a few minutes a colorless suspension resulted, which turned to a clear yellow solution on stirring at ambient temperature for 1 day. Then it was neutralized with saturated aqueous NaHCO₃. After evaporation of methanol the solution was diluted with 40 mL of water and extracted three times with CHCl₃ (50 mL). The organic layer was separated, washed with water and brine and dried over Na₂SO₄. The CHCl₃ was evaporated, and the residue was chromatographed on a silica gel column with EtOAc as an eluent (5–9). Compounds 10, 11 were crystallized from ethanol.

Fig. 3. ORTEP diagram of 16.

(S)-2-(2-Oxopyrimidin-1(2H)-yl)propionic acid methyl ether (5)

Colorless liquid. Yield 0.39 g (43%). – ¹H NMR (400 MHz, [D₆]DMSO): δ = 1.67 (d, J = 7.4 Hz, 3H, CH₃), 3.76 (s, 3H, OCH₃), 5.41 (q, J = 7.4 Hz, 1H, CH), 6.38 (dd, J = 6.7, 4.1 Hz, 1H, Py-H), 7.74 (dd, J = 6.7, 2.8 Hz, 1H, Py-H), 8.60 (dd, J = 3.9, 2.9 Hz, 1H, Py-H). – ¹³C NMR (100 MHz, [D₆]DMSO) δ = 16.5, 53.0, 55.6, 104.2, 145.4, 155.7, 166.1, 170.4. –C₈H₁₀N₂O₃: calcd. C 52.74, H 5.53, N 15.38; found C 52.99, H 5.67, N, 15.45.

(R)-2-(2-Oxopyrimidin-1(2H)-yl)propionic acid methyl ester (6)

Colorless liquid. Yield 0.19 g (21%). - ¹H NMR (400 MHz, [D₆]DMSO) δ = 1.67 (d, J = 7.4 Hz, 3H, CH₃), 3.76 (s, 3H, OCH₃), 5.41 (q, J = 7.4 Hz, 1H, CH), 6.38 (dd, J = 6.7, 4.1 Hz, 1H, Py-H), 7.74 (dd, J = 6.7, 2.8 Hz, 1H, Py-H), 8.60 (dd, J = 3.9, 2.9 Hz, 1H, Py-H). -C₈H₁₀N₂O₃: calcd. C 52.74, H 5.53, N 15.38; found C 52.82, H 5.70, N 15.27.

(S)-4-(Methylthio)-2-(2-oxopyrimidin-1(2H)-yl)butyric acid methyl ester (7)

Colorless crystals. Yield 0.67 g (55%); m. p. 129 °C. – $[\alpha]_D = -100.8$ (c = 1.0, MeOH). – 1 H NMR (400 MHz, $[D_6]$ DMSO): $\delta = 2.01$ (s, 3H, SCH₃), 2.32 – 2.44 (m, 4H,

Table 1. Crystal str	ucture data
for 7 , 15 , and 16 .	

	7	15	16
Empirical formula	C ₁₀ H ₁₄ N ₂ O ₃ S	C ₁₅ H ₁₈ N ₂ O ₄	C ₁₄ H ₁₆ N ₂ O
$M_{\rm r}$	242.29	290.31	228.29
Crystal size, mm ³	$0.25 \times 0.13 \times 0.07$	$0.24 \times 0.13 \times 0.08$	$0.44 \times 0.21 \times 0.07$
Crystal system	orthorhombic	monoclinic	triclinic
Space group	$P2_12_12_1$	$P2_1$	PĪ
a, Å	6.1061(3)	6.5984(3)	6.2046(12)
b, Å	13.8993(13)	11.9696(5)	9.368(3)
c, Å	` '	` '	* *
	14.5862(10) 90.0	9.9658(4) 90.0	11.194(2)
α , deg			93.098(19)
β , deg	90.0	93.980(3)	103.775(17)
γ, deg	90.0	90.0	102.96(2)
V, Å ³	1237.94(16)	785.20(6)	611.9(2)
Z	4	2	2
$D_{\rm calcd.}$, g cm ⁻³	1.30	1.23	1.24
$\mu(\text{Mo}K_{\alpha}), \text{cm}^{-1}$	0.3	0.1	0.1
F(000), e	512	308	244
hkl range	$+8 \to -6, \pm 19, \pm 20$	$\pm 8, \pm 14, +12 \rightarrow -11$	$+7 \rightarrow -6, \pm 11, \pm 13$
θ range, deg	2.79 - 30.52	2.66 – 26.36	2.76 - 26.37
Refl. measured/unique		1669/1283	2481 /1184
$R_{\rm int}$	0.0165	0.0161	0.0236
Param, refined	147	206	154
$R(F)/wR(F^2)^a$ (all refl.)		0.0368/0.0557	0.0870/0.0666
GoF $(F^2)^b$	1.002	1.006	1.007
$\Delta \rho_{\text{fin}} \text{ (max/min), e Å}^{-3}$		-0.11/0.09	-0.13/0.10

^a $R(F) = ||F_0| - |F_c|| / \Sigma |F_0|, wR(F^2) = [\Sigma w (F_0^2 - F_c^2)^2 / \Sigma w (F_0^2)^2]^{1/2}, w = [\sigma^2 (F_0^2) + (AP)^2 + BP]^{-1},$ where $P = (\text{Max}(F_0^2, 0) + 2F_c^2)/3$; ^b $\text{GoF} = [\Sigma w (F_0^2 - F_c^2)^2 / (n_{\text{obs}} - n_{\text{param}})]^{1/2}.$

CH₂), 3.65 (s, 3H, OCH₃), 5.12 (t, J = 6.9 Hz, 1H, CH), 6.51 (dd, J = 6.5, 4.1 Hz, 1H, Py-H), 8.22 (dd, J = 6.5, 2.8 Hz, 1H, Py-H), 8.60 (dd, J = 4.1, 2.8 Hz, 1H, Py-H). $-^{13}$ C NMR (100 MHz, [D₆]DMSO): $\delta = 14.9$, 28.0, 29.8, 53.0, 61.9, 104.7, 149.9, 155.6, 167.3, 169.7. $- C_{10}H_{14}N_{2}O_{3}S$: calcd. C 49.57, H 5.82, N 11.56; found C 49.52, H 5.88, N 11.89.

(R)-4-(Methylthio)-2-(2-oxopyrimidin-1(2H)- $yl)butyric\ acid\ methyl\ ester\ (8)$

Colorless crystals. Yield 0.18 g (15%); m. p. 133 °C. – $[\alpha]_D = +94.3$ (c = 1.0, MeOH). – 1 H NMR (400 MHz, [D₆]DMSO): $\delta = 2.04$ (s, 3H, SCH₃), 2.29 – 2.44 (m, 4H, CH₂), 3.69 (s, 3H, OCH₃), 5.05 – 5.08 (m, 1H, CH), 6.43 (dd, J = 6.5, 4.1 Hz, 1H, Py-H), 8.15 (dd, J = 6.5, 2.8 Hz, 1H, Py-H), 8.55 (dd, J = 4.0, 2.8 Hz, 1H, Py-H). – $C_{10}H_{14}N_2O_3S$: calcd. C 49.57, H 5.82, N 11.56; found C 49.64, H 5.93, N 11.70

(R,S)-4-(Methylthio)-2-(2-oxopyrimidin-1(2H)-yl)butyric acid methyl ester (9)

Colorless crystals. Yield 0.41 g (34%); m. p. 105 °C. $^{-1}$ H NMR (400 MHz, [D₆]DMSO): $\delta = 2.04$ (s, 3H, SCH₃), 2.32-2.43 (m, 4H, CH₂), 3.69 (s, 3H, OCH₃), 4.97 – 5.16 (m, 1H, CH), 6.43 (dd, J = 6.5, 4.1 Hz, Py-H), 8.15 (dd, J = 6.5, 2.8 Hz, Py-H), 8.56 (dd, J = 4.1, 2.8 Hz, Py-H). $^{-13}$ C NMR (100 MHz, [D₆]DMSO): $\delta = 14.9, 28.1, 29.9, 53.0, 61.9, 104.6, 149.8, 155.6, 167.3, 169.7. – HRMS ((+)-ESI-$

MS): m/z = 243.0848 (calcd. 243.0798 for $C_{10}H_{15}N_2O_3S$, $[M+H]^+$). $-C_{10}H_{14}N_2O_3S$: calcd. C 49.57, H 5.82, N 11.56; found C 49.54, H 5.86, N 11.60.

(R,S)-2-(2-Oxopyrimidin-1(2H)-yl)-3-phenylpropionic acid methyl ester <math>(10)

Colorless crystals. Yield 0.52 g (40%); m.p. 162 °C. – $[\alpha]_D = 0$ (c = 1.0, MeOH). – 1 H NMR (400 MHz, [D₆]DMSO): $\delta = 3.36 - 3.46$ (m, 2H, CH₂), 3.71 (s, 3H, OCH₃), 5.19 (dd, J = 10.1, 5.7 Hz, 1H), 6.16 (dd, J = 6.5, 4.1 Hz, 1H), 7.04 – 7.11 (m, 2H), 7.12 – 7.21 (m, 3H), 7.74 (dd, J = 6.5, 2.8 Hz, 1H), 8.42 (dd, J = 4.1, 2.8 Hz, 1H). – 13 C NMR (100 MHz, [D₆]DMSO): $\delta = 33.5$, 52.4, 64.0, 103.5, 126.7, 128.4, 128.9, 136.1, 149.5, 154.9, 166.6, 168.7. – HRMS ((+)-ESI-MS): m/z = 259.1094 (calcd. 259.1077 for C₁₄H₁₅N₂O₃, [M+H]⁺).

(S)-3-(4-Hydroxyphenyl)-2-(2-oxopyrimidin-1(2H)-yl)propionic acid methyl ester (11)

Colorless crystals. Yield 0.71 g (52%); m. p. 166 °C. – $[\alpha]_{\rm D} = -242.9$ (c = 1.0, MeOH). – $^{1}{\rm H}$ NMR (400 MHz, $[{\rm D}_{6}]{\rm DMSO}$): $\delta = 3.22 - 3.33$ (m, 2H), 3.67 (s, 3H, OCH₃), 5.16 (dd, J = 10.2, 5.6 Hz), 6.28 (dd, J = 6.5, 4.2 Hz), 6.59 (d, J = 8.3 Hz, 2H, 4-OH-Ph), 6.85 (d, J = 8.3 Hz, 2H, 4-OH-Ph), 7.81 (dd, J = 6.5, 2.7 Hz, 1H), 8.47 – 8.49 (m, 1H), 9.22 (s, 1H). – $^{13}{\rm C}$ NMR (100 MHz, $[{\rm D}_{6}]{\rm DMSO}$): $\delta = 32.7$, 52.3, 64.1, 103.5, 115.2, 125.9, 129.8, 149.5,

154.9, 156.0, 166.5, 168.8. – HRMS ((+)-ESI-MS): m/z = 275.1044 (calcd. 275.1026 for $C_{14}H_{15}N_{2}O_{4}$, $[M+H]^{+}$).

Procedure for the synthesis of 12 and 13

Conc. HCl (3 mL) was added to the solution of pyrimidinone **5** or **6** (200 μ g) in dioxane (10 mL). The reaction mixture was stirred for 1 day, and then it was evaporated in a Petri dish to dryness and crystallized from 2-propanol.

(S)-2-(2-Oxopyrimidin-1(2H)-yl)propionic acid (12)

Colorless crystals. Yield 0.12 g (67%); m. p. 179 °C. – $[\alpha]_D = -39.4$ (c = 1.0, MeOH). – ¹H NMR (400 MHz, [D₆]DMSO): $\delta = 1.66$ (d, J = 7.3 Hz, 3H, CH₃), 5.21 (q, J = 7.2 Hz, 1H), 6.72 (dd, J = 6.4, 5.0 Hz, 1H), 8.63 – 8.70 (m, 1H), 8.73 (dd, J = 4.7, 2.6 Hz, 1H). – ¹³C NMR (100 MHz, [D₆]DMSO): $\delta = 15.4$, 59.0, 104.7, 150.5, 156.0, 162.3, 170.6. – HRMS ((+)-ESI-MS): m/z = 169.0616 (calcd. 169.0608 for C₇H₉N₂O₃, [M+H]⁺). – C₇H₈N₂O₃: calcd. C 50.00, H 4.80, N 16.66; found C 50.12, H 4.93, N 16.77.

(R)-2-(2-Oxopyrimidin-1(2H)-yl)propionic acid (13)

Colorless crystals. Yield 0.14 g (76%); m. p. $183-184\,^{\circ}\text{C.}$ - $[\alpha]_{\text{D}}=+37.1$ (c=1.0, MeOH). - ^{1}H NMR (400 MHz, [D₆]DMSO): $\delta=1.65$ (d, J=7.3 Hz, 3H, CH₃), 5.20 (q, J=7.3 Hz, 1H, CH), 6.70 (dd, J=6.3, 5.0 Hz, 1H, Py-H), 8.66 (dd, J=6.4, 2.5 Hz, 1H, Py-H), 8.72 (dd, J=4.8, 2.6 Hz, 1H, Py-H). - $C_{7}H_{8}N_{2}O_{3}$: calcd. C 50.00, H 4.80, N 16.66; found C 49.59, H 5.02, N 16.72.

Procedure for the synthesis of compounds 15 and 16

2-Phenylmalonaldehyde (1.0 g, 6.75 mmol) and N-carbamoyl L-valine (1.62 g, 10.12 mmol) were mixed with toluene (60 mL). The mixture was refluxed with a Dean-Stark trap. Then it was cooled to 40 °C, the toluene solution was decanted, and the resin residue was crystallized from ethanol.

(2S)-3-Methyl-2-((E)-3-(3-oxo-2-phenylpropylidene)-ureido)butyric acid (15)

Colorless crystals. Yield 0.15 g (8%); m. p. 203 – 204 °C. – $^1{\rm H}$ NMR (400 MHz, [D₆]DMSO): $\delta=0.85$ (d, J=6.8 Hz, 3H, CH₃), 0.92 (d, J=6.8 Hz, 3H, CH₃), 2.04 – 2.21 (m,

1H, CH), 4.17 – 4.20 (m, 1H, CH), 7.21 – 7.23 (m, 2H, Ph), 7.34 – 7.46 (m, 4H, Ph), 7.77 (d, J = 12.4 Hz, 1H), 9.14 (d, J = 12.4 Hz, 1H), 9.31 (s, 1H, COOH), 12.63 (br.s, 1H, NH). – HRMS ((+)-ESI-MS): m/z = 291.1320 (calcd. 291.1339 for $C_{15}H_{19}N_2O_4$, $[M+H]^+$).

1-Isobutyl-5-phenylpyrimidin-2(1H)-one (16)

Yellow crystals. Yield 0.20 g (13%); m. p. 177 °C. – 1 H NMR (400 MHz, [D₆]DMSO): δ = 0.95 (d, J = 6.7 Hz, 6H, 2 × CH₃), 2.10 – 2.28 (m, 1H, CH), 3.77 (d, J = 7.4 Hz, 1H, CH₂), 7.29 – 7.33 (m, 1H, Ph), 7.40 – 7.44 (m, 2H, Ph), 7.55 – 7.57 (m, 2H, Ph), 8.48 (d, J = 3.4 Hz, 1H), 8.87 (d, J = 3.4 Hz, 1H). – 13 C NMR (100 MHz, [D₆]DMSO): δ = 164.8, 155.6, 147.4, 133.7, 129.5, 127.9, 125.8, 116.6, 57.9, 27.4, 19.9. – HRMS ((+)-ESI-MS): m/z = 229.1352 (calcd. 229.1335 for C₁₄H₁₇N₂O, [M+H]⁺).

Crystal structure determinations

X-Ray analyses were carried out on an Oxford Diffraction Xcalibur S CCD diffractometer using the software package CRYSALISPRO [13]. X-Ray data collection was carried out at 295(2) K with graphite-monochromatized $\text{Mo}K_{\alpha}$ radiation ($\lambda=0.71073\,\text{Å}$). The structures were solved with Direct Methods using SHELXS-97 [14] and refined by full-matrix least-squares procedures on F^2 with SHELXL-97 [15]. Non-H atoms were refined anisotropically, hydrogen atoms were placed in idealized positions and were constrained to ride on their parent atoms. Due to the absence of heavy atoms in the non-centrosymmetric crystal structures of 7 and 15, Friedel pairs were merged, and Flack parameters were not refined. Table 1 summarizes the crystal data and numbers pertinent to data collection and structure refinement.

CCDC 873468 (7), CCDC 936498 (15) and CCDC 936499 (16) contain the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre *via* www.ccdc. cam.ac.uk/data_request/cif.

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