

Journal of Carbohydrate Chemistry

Publication details, including instructions for authors and subscription information:

<http://www.tandfonline.com/loi/lcar20>

SYNTHESIS OF SIALYL- AND SULFO-Le^x/Le^a ANALOGS CONTAINING N-ALKYL-1-DEOXYNOJIRIMYCIN AS POTENTIAL SELECTIN BLOCKERS

Hiroyasu Furui ^a, Keiko Ando-Furui ^a, Haruko Inagaki ^a, Takayuki Ando ^a, Hideharu Ishida ^a & Makoto Kiso ^a

^a Department of Applied Bioorganic Chemistry, Gifu University, Gifu, 501-1193, Japan
Published online: 19 Aug 2006.

To cite this article: Hiroyasu Furui, Keiko Ando-Furui, Haruko Inagaki, Takayuki Ando, Hideharu Ishida & Makoto Kiso (2001) SYNTHESIS OF SIALYL- AND SULFO-Le^x/Le^a ANALOGS CONTAINING N-ALKYL-1-DEOXYNOJIRIMYCIN AS POTENTIAL SELECTIN BLOCKERS, *Journal of Carbohydrate Chemistry*, 20:9, 789-812

To link to this article: <http://dx.doi.org/10.1081/CAR-100108657>

PLEASE SCROLL DOWN FOR ARTICLE

Taylor & Francis makes every effort to ensure the accuracy of all the information (the "Content") contained in the publications on our platform. However, Taylor & Francis, our agents, and our licensors make no representations or warranties whatsoever as to the accuracy, completeness, or suitability for any purpose of the Content. Any opinions and views expressed in this publication are the opinions and views of the authors, and are not the views of or endorsed by Taylor & Francis. The accuracy of the Content should not be relied upon and should be independently verified with primary sources of information. Taylor and Francis shall not be liable for any losses, actions, claims, proceedings, demands, costs, expenses, damages, and other liabilities whatsoever or howsoever caused arising directly or indirectly in connection with, in relation to or arising out of the use of the Content.

This article may be used for research, teaching, and private study purposes. Any substantial or systematic reproduction, redistribution, reselling, loan, sub-licensing, systematic supply, or distribution in any form to anyone is expressly forbidden. Terms & Conditions of access and use can be found at <http://www.tandfonline.com/page/terms-and-conditions>

SYNTHESIS OF SIALYL- AND SULFO- Le^x/Le^a ANALOGS CONTAINING *N*-ALKYL-1-DEOXYNOJIRIMYCIN AS POTENTIAL SELECTIN BLOCKERS¹

Hiroyasu Furui,² Keiko Ando-Furui, Haruko Inagaki,
Takayuki Ando, Hideharu Ishida,* and Makoto Kiso*

Department of Applied Bioorganic Chemistry,
Gifu University, Gifu 501-1193, Japan

ABSTRACT

A series of novel sialyl- and sulfo- Le^x/Le^a oligosaccharides containing *N*-alkyl-1-deoxynojirimycin as potential selectin blockers have systematically been synthesized *via* the suitably protected intermediates containing *N*-benzyloxycarbonyl-1-deoxynojirimycin. Some of the synthetic oligosaccharides strongly inhibited the adhesion of HL60 cells to IL-1 β -stimulated HUVECs.

INTRODUCTION

The carbohydrate determinants, sialyl Lewis x (sLe^x) and sialyl Lewis a (sLe^a), which are frequently expressed on cancer cells,^{3,4} serve as the ligands for selectin,⁵ a family of cell adhesion molecules implicated in leukocyte traffic and recruitment to sites of inflammation. These carbohydrate determinants, therefore, have been thought to be involved in hematogenous metastasis of some cancer cells.^{6,7}

In the course of synthetic studies on sialoglycoconjugates,^{8,9} we have succeeded not only in the total syntheses^{10,11} of these carbohydrate determinants but also in the synthesis of a variety of their analogs^{12,13} and mimetics.^{14–16} In this

*Corresponding authors.

course, it has been found¹⁷ that the glucosamine residue in these determinants could be replaced by glucose and 1-deoxynojirimycin without loss of the selectin binding activity. It has also been strongly suggested that the sialic acid residue may be substituted by the sulfate group.^{18,19} This paper describes the systematic synthesis of a series of novel sialyl- and sulfo-Le^x/Le^a oligosaccharides containing *N*-alkyl-1-deoxynojirimycin instead of the *N*-acetylglucosamine residue.

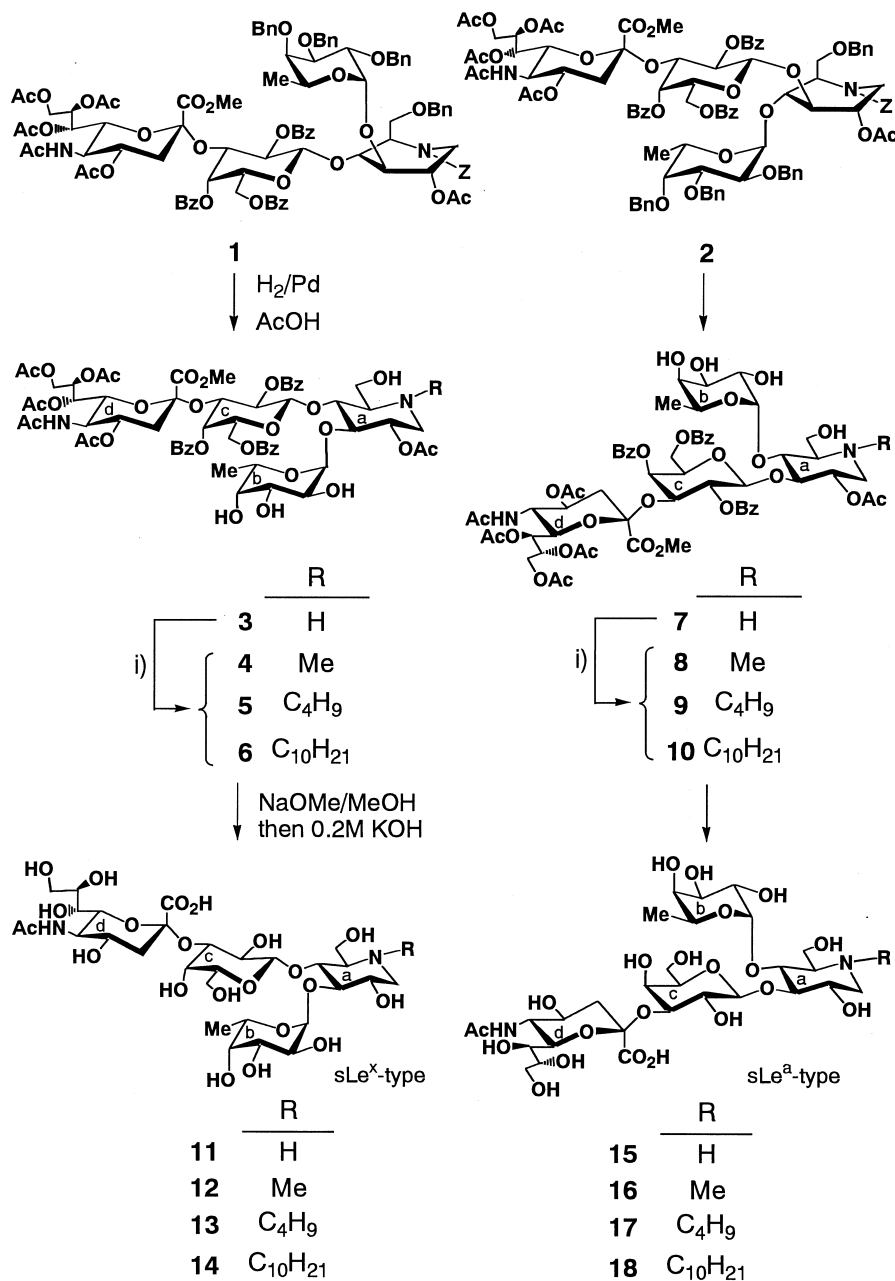
RESULTS AND DISCUSSION

The sialyl Lewis x (sLe^x) (**11–14**) and sialyl Lewis a (sLe^a) (**15–18**) analogs containing 1-deoxynojirimycin or *N*-alkyl-1-deoxynojirimycin were synthesized from the corresponding, suitably protected tetrasaccharides¹⁴ (**1** and **2**), respectively (Scheme 1). Hydrogenolytic removal of the benzyloxycarbonyl (Z) and benzyl (Bn) groups in **1** and **2** were carried out in the presence of palladium chloride in AcOH, to give **3** and **7**, which, upon treatment with formalin,¹⁴ butyraldehyde or decyl aldehyde over palladium hydroxide on carbon in a hydrogen atmosphere, gave **4–6** and **8–10** in good yields. Complete *O*-deacylation of **3–6** and **7–10**, followed by saponification of the methyl ester group, afforded a series of sLe^x-type (**11–14**) and sLe^a-type (**15–18**) tetrasaccharides in high yields (Scheme 1).

For the synthesis of the sulfo Le^x analogs (**27, 37, 38**), (2,3,4-tri-*O*-benzyl- α -L-fucopyranosyl)-(1 \rightarrow 3)-2-*O*-acetyl-6-*O*-benzyl-*N*-benzyloxycarbonyl-1,5-dideoxy-1,5-imino-D-glucitol¹⁴ (**19**) was glycosylated with the galactose donor **20**²⁰ to give trisaccharide **21** (70%), which was converted stepwise, by hydrolytic removal of the benzylidene group, benzylation to **23** and selective removal of the levulinoyl group, to **24** in high yield (Scheme 2). Treatment of **24** with pyridine sulfur trioxide complex in *N,N*-dimethylformamide gave **25** in 96% yield, which underwent reductive *N*-methylation as described for **12** and **16**, to afford **26** in 83% yield. Significant signals in the ¹H NMR spectrum of **26** were a three proton singlet at δ 2.12 (N—Me) and a one proton doublet of doublets at δ 4.72 ($J_{2,3} = 10.3$, $J_{3,4} = 3.7$ Hz, H-3c), showing the desired structure. Compound **27** was prepared by *O*-deacylation of **26**, quantitatively. In the ion-spray MS of **27**, the molecular ion was clearly detected at m/z 564.0 (M—Na—H)[–] and m/z 566.2 (M—Na+H)⁺ in negative and positive ion modes,²¹ respectively, indicating the desired structure unambiguously.

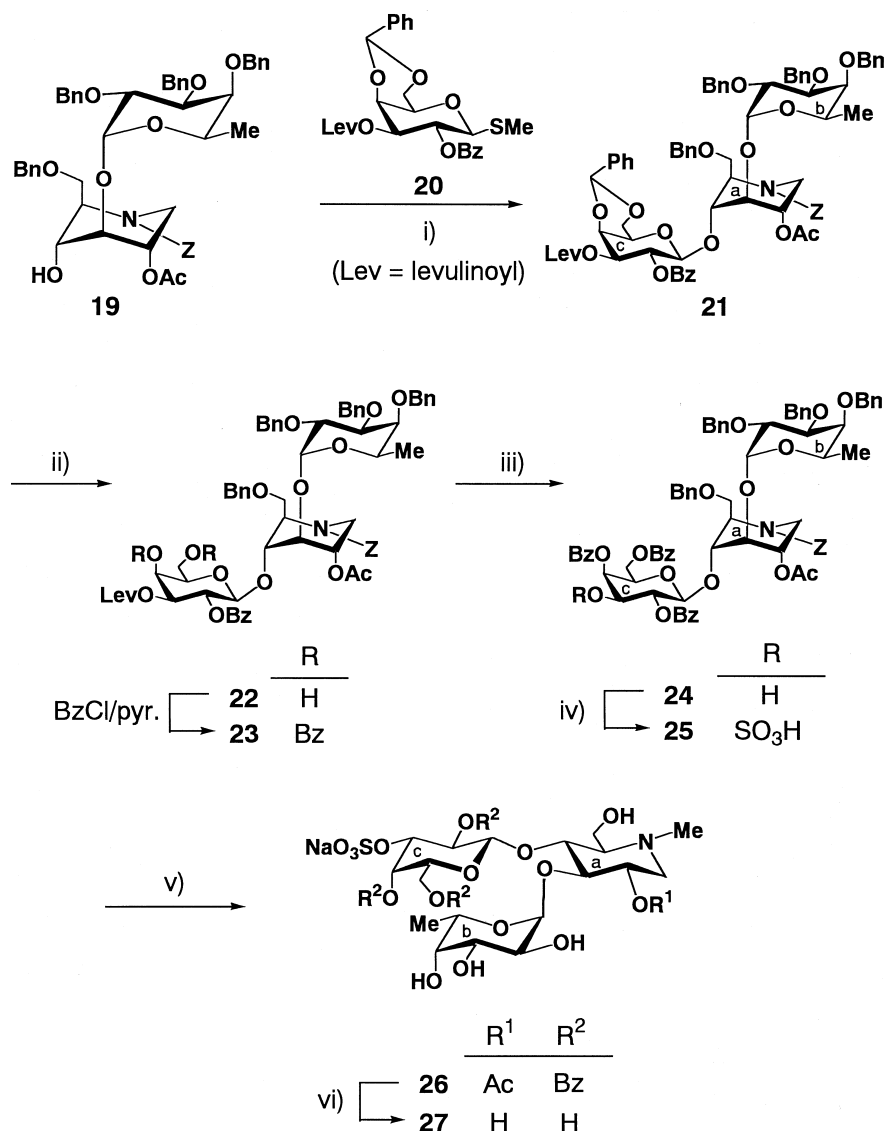
Hydrogenation of **22** over palladium black in AcOH gave **28** (79%), which was then treated with butyraldehyde or decyl aldehyde in the presence of palladium hydroxide on carbon in a hydrogen atmosphere, to give **29** (79%) or **30** (80%), respectively (Scheme 3). The remaining hydroxyls were acetylated, and then the levulinoyl group was cleaved by treatment with hydrazine acetate in EtOH to afford **33** and **35**. Sulfation of **33** and **35** was performed as described for **25** and the resulting **34** (81%) and **36** (88%) were treated with methanolic sodium methoxide to give **37** and **38**, quantitatively. In the ¹H NMR spectra of **34** and **36**, H-3c at the sulfated position appeared at δ 4.68 (dd, $J_{2,3} = 10.3$, $J_{3,4} = 2.9$ Hz) for **34** and δ 4.70 (dd, $J_{2,3} = 10.3$, $J_{3,4} = 3.3$ Hz) for **36**, respectively. In the FAB-MS of **37** and





Scheme 1. i) R'CHO, H₂, Pd(OH)₂-C, R'CH₂OH/AcOH/H₂O R' = H, C₃H₇, C₉H₁₉.

38, a pair of molecular ions were each clearly detected at m/z 628.3 (M—H)[−] and 606.3 (M—Na—H)[−] for **37**, and m/z 712.32 (M—H)[−] and 690.38 (M—Na—H)[−] for **38**, accompanied with a significant fragment ion (M—Na—H—Fuc)[−] at m/z 460.2 for **37** and 544.3 for **38**, respectively, providing the distinct evidence for the desired structures.



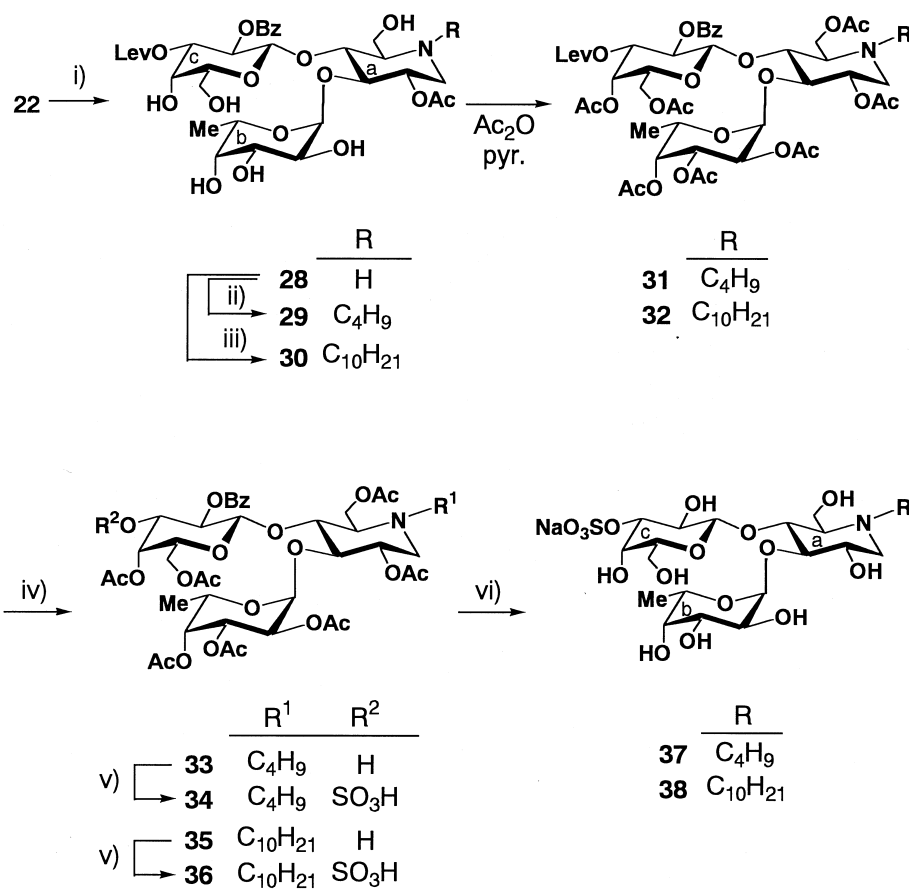
Scheme 2. i) NIS, TMSOTf, CH_2Cl_2 , $0^\circ\text{C} \rightarrow \text{rt}$ (70%); ii) 80% aq AcOH, 45°C (89%); iii) $\text{NH}_2\text{NH}_2 \cdot \text{AcOH}$, EtOH, rt (100%); iv) $\text{SO}_3 \cdot \text{pyr}$, DMF, rt (96%); v) Dowex- Na^+ , then HCHO, H_2/Pd , MeOH (83%); vi) NaOMe, MeOH.

In the synthetic route to the sulfo Le^a analogs (**50**, **60**, **61**), 2-*O*-acetyl-4,6-*O*-benzylidene-*N*-benzyloxycarbonyl-1,5-dideoxy-1,5-imino-D-glucitol (**39**)¹⁴ was first glycosylated with the galactose donor **40** to give **41** (87%) (Scheme 4). Hydrolytic removal of the benzylidene group in **41** and the following regioselective protection of 6-OH by a *tert*-butyldimethylsilyl (TBDMS) group gave **43** in high yield. The remaining 4-OH was then fucosylated with methyl 2,3,4-tri-*O*-benzyl-1-thio- β -L-fucopyranoside¹⁴ in the presence of dimethyl(methylthio)sulfonium tri-



flate (DMTST) in benzene to afford **44** (55%). This was converted to **46** by replacement of the TBDMS group with the benzoyl group, and then the levulinoyl group in **46** was selectively cleaved to give **47**. Sulfation (99%) of **47** as described for **25**, and reductive *N*-methylation (92%), followed by the complete *O*-deacetylation, afforded **50** as the sodium salt (Scheme 4). In the ion-spray MS of **50**, the molecular ion was detected at m/z 564.1 ($M-Na-H$)⁻, from which two major daughter ions were formed and detected at m/z 417.6 ($M-Na-Fuc$)⁻ and m/z 96.8 (HSO_4^-), respectively.

Hydrogenolysis of **46** in AcOH and the reductive *N*-alkylation as described for **29** and **30**, gave **52** (64%) and **53** (75%), respectively, which were then fully acetylated. Selective removal of the levulinoyl group in **54** and **55**, followed by *O*-sulfation as described for **34** and **36**, afforded **57** and **59**, which, upon treatment with methanolic sodium methoxide, gave the sulfo Le^a analogs containing *N*-butyl- and *N*-decyl-1-deoxynojirimycin (**60** and **61**), respectively. In the FAB-MS, the

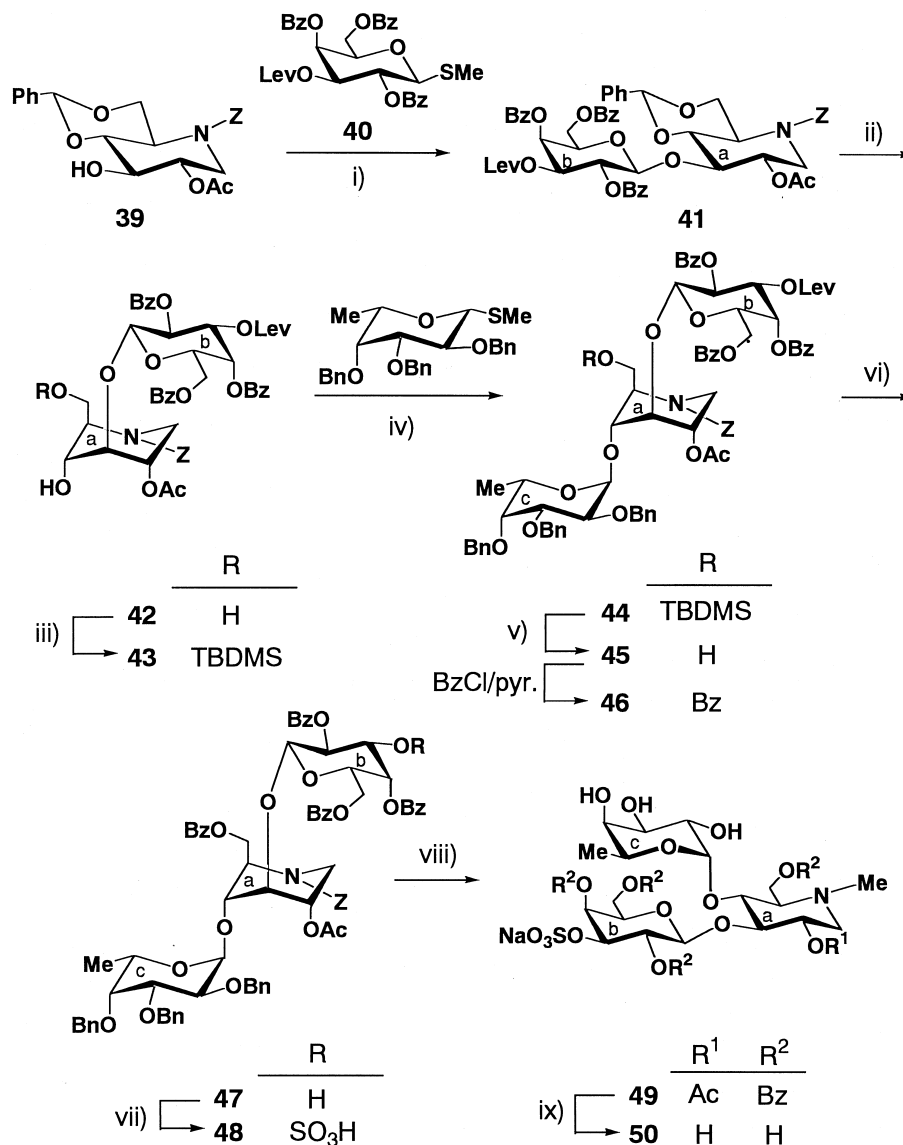


Scheme 3. i) H₂/Pd, AcOH; ii) C₃H₇CHO, 1-butanol, H₂, Pd(OH)₂-C (79%); iii) C₉H₁₉CHO, EtOAc/THF, H₂, Pd(OH)₂-C (80%); iv) NH₂NH₂·AcOH, EtOH, rt; v) SO₃·pyr, DMF, **33** → **34** (81%), **35** → **36** (88%); vi) NaOMe, MeOH.



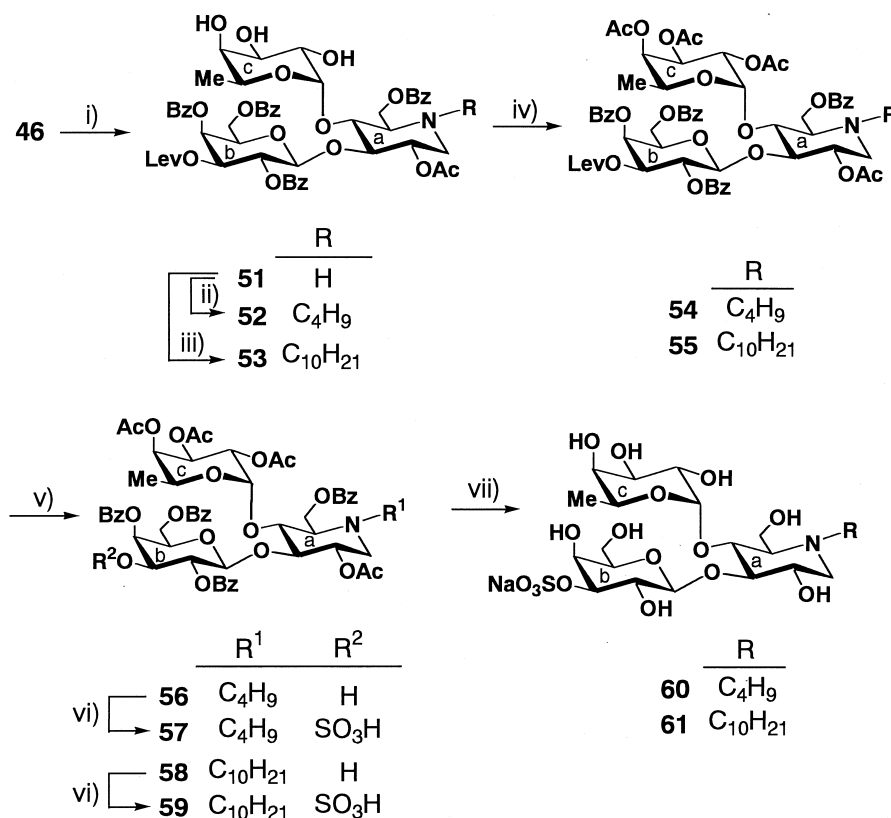
molecular ions were clearly detected at m/z 606.30 ($M-Na$)⁻ for **60**, and m/z 712.26 ($M-H$)⁻ and 690.31 ($M-Na$)⁻ for **61**, respectively, showing the structure assigned.

The preferred conformation of the 1-deoxynojirimycin residue in the protected oligosaccharide intermediates, such as **1**, **2**, **19**, **21–25** and **42–48**, has been



Scheme 4. i) NIS, TMSOTf, CH₂Cl₂, 0°C → rt (87%); ii) 80% aq AcOH, 45°C (86%); iii) TBDMSCl, pyr, CH₂Cl₂, 0°C → rt (92%); iv) DMTST, benzene, 7°C (55%); v) 80% aq AcOH, rt (88%); vi) NH₂NH₂·AcOH, EtOH, rt (93%); vii) SO₃·pyr, DMF, rt (99%); viii) Dowex-Na⁺, then HCHO, H₂/Pd, MeOH (92%); ix) NaOMe, MeOH.





Scheme 5. i) H_2/Pd , AcOH; ii) $\text{C}_3\text{H}_7\text{CHO}$, 1-butanol, H_2 , $\text{Pd}(\text{OH})_2\text{-C}$ (64%); iii) $\text{C}_8\text{H}_{19}\text{CHO}$, H_2 , $\text{Pd}(\text{OH})_2\text{-C}$, EtOAc (75%); iv) Ac_2O , pyr (quant); v) $\text{NH}_2\text{NH}_2\cdot\text{AcOH}$, EtOH, (**56**, 91%; **58**, 93%); vi) $\text{SO}_3\cdot\text{pyr}$, DMF (~90%); vii) NaOMe, MeOH (quant).

suggested to be a flexible ${}^1\text{C}_4$ conformation based on ${}^1\text{H}$ NMR data^{14,22} and X-ray crystallographic analysis.²³ A dramatic conformational change (${}^4\text{C}_1 \rightarrow {}^1\text{C}_4$) was observed in the reductive ring opening of the 4,6-*O*-benzylidene group (**41**→**42**). On the contrary, in the *N*-debenzyloxycarbonylation step (**1**→**3**, **2**→**7**, **25**→**26**, **22**→**28**, **48**→**49**, and **46**→**51**), the ${}^1\text{C}_4$ conformation changed to ${}^4\text{C}_1$ as shown in Scheme 1–5.

The inhibitory effect of the synthetic oligosaccharides on the adhesion of fixed HL-60 cells to IL- β -stimulated HUVECs was examined. Among a series of sialyl Le^x/Le^a analogs (**11**–**18**, Scheme 1), the s Le^a -type analogs (**15**–**18**) expressed stronger activity than the corresponding s Le^x -type analogs (**11**–**14**). The hierarchy of inhibition potency due to the *N*-alkyl group in both s Le^x and s Le^a series were *N*-butyl > *N*-decyl > *N*-methyl, and the activity of *N*-butyl s Le^a derivative (**17**) was most potent. Similarly, a series of sulfo Le^a -type analogs (**50**, **60**, **61**) showed stronger inhibitory activity than the corresponding sulfo Le^x -type analogs (**27**, **37**, **38**). The details of the biological study will be described elsewhere.



In conclusion, a series of novel sialyl- and sulfo-Le^x/Le^a oligosaccharides containing *N*-alkyl-1-deoxynojirimycin have systematically been synthesized as potential selectin blockers.^{17,24–26}

EXPERIMENTAL

General Methods. Optical rotations were determined with a Union PM-201 polarimeter at 25°C, and IR spectra were recorded with a Jasco IRA-100 spectrophotometer. ¹H NMR spectra were recorded on a JEOL JNM-GX 270 (270 MHz) or Varian Unity Inova 400 (400 MHz) spectrometer. FAB-MS were recorded on a JEOL JMS-SX 120A mass spectrometer/JMA-DA 7000 data system.⁹ Ion-spray MS were recorded on an API-III triple quadrupole mass spectrometer (Perkin-Elmer Sciex Instruments) fitted with an atmosphere pressure ionization source.²¹ All reactions were monitored by TLC (Merck silica gel aluminum plate 60F-254) and preparative column chromatography was performed on silica gel (Wako Chemical Co., 200 or 300 mesh) with the solvent systems specified. Concentrations were conducted *in vacuo*.

(Methyl 5-Acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-*D*-glycero- α -*D*-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-(2,4,6-tri-*O*-benzoyl- β -*D*-galactopyranosyl)-(1 \rightarrow 4)-[α -*L*-fucopyranosyl-(1 \rightarrow 3)]-2-*O*-acetyl-1,5-dideoxy-1,5-imino-*D*-glucitol (3). Compound **1** (113 mg) in acetic acid (10 mL) was hydrogenolyzed in the presence of palladium chloride (200 mg) for 2 days at rt. The catalyst was filtered off and washed with MeOH. The combined filtrate and washings were concentrated. Column chromatography (15:1 CH₂Cl₂—MeOH) of the residue on silica gel gave **3** (37.6 mg, 75%): [α]_D -4° (*c* 0.5, CH₂Cl₂); ¹H NMR (CDCl₃) δ 1.39 (d, 3H, *J*_{5,6} = 6.6 Hz, H-6b), 1.56 (s, 3H, AcN), 1.65 (t, 1H, *J*_{gem} = *J*_{3ax,4} = 12.5 Hz, H-3dax), 1.77, 1.90, 1.98, 2.11, 2.25 (5s, 15H, 5AcO), 2.41 (dd, 1H, *J*_{gem} = 12.5, *J*_{3eq,4} = 5.1 Hz, H-3deq), 3.09 (dd, 1H, *J*_{gem} = 12.3, *J*_{1eq,2} = 5.1 Hz, H-1aeq), 3.82 (s, 3H, COOMe), 4.91 (d, 1H, *J*_{1,2} = 8.42 Hz, H-1c), 5.12 (d, 1H, *J*_{1,2} = 3.85 Hz, H-1b), 5.14 (dd, 1H, *J*_{6,7} = 2.57 Hz, H-7d), 5.22 (d, 1H, *J*_{3,4} = 3.1 Hz, H-4c), 5.48 (dd, 1H, *J*_{2,3} = 9.9 Hz, H-2c), 5.80 (m, 1H, H-8d), 7.48–8.22 (m, 15H, 3Ph).

Anal. Calcd for C₆₁H₇₄N₂O₂₉ (1229.25): C, 56.39; H, 5.74; N, 2.16. Found: C, 56.12; H, 5.51; N, 2.08.

(Methyl 5-Acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-*D*-glycero- α -*D*-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-(2,4,6-tri-*O*-benzoyl- β -*D*-galactopyranosyl)-(1 \rightarrow 4)-[α -*L*-fucopyranosyl-(1 \rightarrow 3)]-2-*O*-acetyl-*N*-butyl-1,5-dideoxy-1,5-imino-*D*-glucitol (5). A mixture of **3** (61 mg), butyraldehyde (1 mL), 1-butanol (10 mL), acetic acid (0.5 mL) and water (0.5 mL) was vigorously stirred with palladium hydroxide on carbon (60 mg) in a hydrogen atmosphere overnight at rt. The catalyst was filtered off and washed with MeOH. The combined filtrate and washings were concentrated. Column chromatography (15:1 CH₂Cl₂—MeOH)



of the residue on silica gel gave **5** (50 mg, 79%): $[\alpha]_D -11^\circ$ (c 1, CH₂Cl₂); ¹H NMR (CDCl₃) δ 0.87 (t, 3H, J = 6.4 Hz, Me), 1.24 (m, 4H, —C₂H₄Me), 1.38 (d, 3H, J_{5,6} = 6.6 Hz, H-6b), 1.56 (s, 3H, AcN), 1.65 (t, 1H, J_{gem} = J_{3ax,4} = 12.5 Hz, H-3dax), 1.76, 1.90, 1.97, 2.11, 2.25 (5s, 15H, 5AcO), 2.40 (dd, 1H, J_{gem} = 12.6, J_{3eq,4} = 4.4 Hz, H-3deq), 2.51, 2.65 (2m, 2H, N—CH₂—C₃H₇), 3.01 (dd, 1H, J_{gem} = 10.8, J_{1eq,2} = 5.1 Hz, H-1aeq), 3.54 (dd, 1H, J_{2,3} = 9.9, J_{3,4} = 3.7 Hz, H-3c), 3.83 (s, 3H, COOMe), 5.11 (dd, 1H, H-7d), 5.12 (d, 1H, J_{1,2} = 8.8 Hz, H-1c), 5.17 (d, 1H, J_{1,2} = 2.8 Hz, H-1b), 5.19 (d, 1H, J_{3,4} = 3.7 Hz, H-4c), 5.50 (~t, 1H, J_{1,2} = 8.8, J_{2,3} = 9.9 Hz, H-2c), 5.80 (m, 1H, H-8d), 7.47–8.24 (m, 15H, 3Ph).

Anal. Calcd for C₆₅H₈₂N₂O₂₉ (1355.36): C, 57.60; H, 6.10; N, 2.07. Found: C, 57.52; H, 6.00; N, 1.83.

(Methyl 5-Acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylate)-(2→3)-(2,4,6-tri-O-benzoyl- β -D-galactopyranosyl)-(1→4)-[α -L-fucopyranosyl-(1→3)]-2-O-acetyl-N-decyl-1,5-dideoxy-1,5-imino-D-glucitol (6**).** A mixture of **3** (30 mg), decyl aldehyde (0.5 mL), ethyl acetate (5 mL), acetic acid (0.1 mL) and water (0.05 mL) was vigorously stirred with palladium hydroxide on carbon (30 mg) for 10 h at rt. Work-up and column chromatography (20:1 CH₂Cl₂—MeOH) on silica gel gave **6** (50 mg, 60%): $[\alpha]_D +4.3^\circ$ (c 0.4, CH₂Cl₂); ¹H NMR (CDCl₃) δ 0.88 (t, 3H, J = 6.2 Hz, Me), 1.25 (m, 16H, —C₈H₁₆Me), 1.38 (d, 3H, J_{5,6} = 6.6 Hz, H-6b), 1.51 (s, 3H, AcN), 1.66 (t, 1H, J_{gem} = J_{3ax,4} = 12.3 Hz, H-3dax), 1.77, 1.91, 1.97, 2.11, 2.26 (5s, 15H, 5AcO), 2.40 (dd, 1H, J_{gem} = 12.3, J_{3eq,4} = 4.2 Hz, H-3deq), 3.01 (dd, 1H, J_{gem} = 10.8, J_{1eq,2} = 4.4 Hz, H-1aeq), 3.54 (dd, 1H, J_{2,3} = 9.9, J_{3,4} = 3.5 Hz, H-3c), 3.81 (s, 3H, COOMe), 5.12 (d, 1H, J_{1,2} = 8.6 Hz, H-1c), 5.17 (dd, 1H, H-7d), 5.19 (d, 1H, J_{3,4} = 3.5 Hz, H-4c), 5.50 (~t, 1H, H-2c), 5.81 (m, 1H, H-8d), 7.52–8.35 (m, 15H, 3Ph).

Anal. Calcd for C₇₁H₉₄N₂O₂₉ (1439.52): C, 59.24; H, 6.58; N, 1.95. Found: C, 59.11; H, 6.45; N, 1.85.

(Methyl 5-Acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylate)-(2→3)-(2,4,6-tri-O-benzoyl- β -D-galactopyranosyl)-(1→3)-[α -L-fucopyranosyl-(1→4)]-2-O-acetyl-1,5-dideoxy-1,5-imino-D-glucitol (7**).** Compound **2** (179 mg) in acetic acid (10 mL) was hydrogenolyzed as described for **3**. Work-up and column chromatography (10:1 CH₂Cl₂—MeOH) of the product on silica gel afforded **7** (92 mg, 71%): $[\alpha]_D -24.1^\circ$ (c 1.46, CH₂Cl₂); ¹H NMR (CDCl₃) δ 1.27 (d, 3H, J_{5,6} = 6.6 Hz, H-6b), 1.56 (s, 3H, AcN), 1.65 (t, 1H, J_{gem} = J_{3ax,4} = 12.6 Hz, H-3dax), 1.78, 1.91, 2.06, 2.16, 2.18 (5s, 15H, 5AcO), 2.42 (dd, 1H, J_{gem} = 12.6, J_{3eq,4} = 4.4 Hz, H-3deq), 3.09 (dd, 1H, H-1aeq), 3.79 (s, 3H, COOMe), 4.74 (m, 1H, H-2a), 4.81 (m, 1H, H-4d), 4.95 (d, 1H, H-1b), 5.14 (d, 1H, J_{1,2} = 8.3 Hz, H-1c), 5.25 (d, 1H, J_{3,4} = 2.8 Hz, H-4c), 5.30 (dd, 1H, H-7d), 5.36 (t, 1H, H-2c), 5.56 (m, 1H, H-8), 7.41–8.09 (m, 15H, 3Ph).

Anal. Calcd for C₆₁H₇₄N₂O₂₉ (1299.25): C, 56.39; H, 5.74; N, 2.16. Found: C, 56.25; H, 5.55; N, 1.96.



(Methyl 5-Acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-(2,4,6-tri-*O*-benzoyl- β -D-galactopyranosyl)-(1 \rightarrow 3)-[α -L-fucopyranosyl-(1 \rightarrow 4)]-2-*O*-acetyl-*N*-butyl-1,5-dideoxy-1,5-imino-D-glucitol (9). A mixture of **7** (50 mg), butyraldehyde (1 mL), 1-butanol (10 mL), acetic acid (0.5 mL) and water (0.5 mL) was hydrogenated as described for **5**. Work-up and column chromatography (15:1 CH₂Cl₂—MeOH) of the product on silica gel gave **9** (34 mg, 65%): [α]_D -12° (*c* 0.74, CH₂Cl₂); ¹H NMR (CDCl₃) δ 0.86 (t, 3H, *J* = 6.6 Hz, Me), 1.24 (m, 4H, —C₂H₄Me), 1.25 (d, 3H, *J*_{5,6} = 6.1 Hz, H-6b), 1.53 (s, 3H, AcN), 1.65 (t, 1H, *J*_{gem} = *J*_{3ax,4} = 12.8 Hz, H-3dax), 1.77, 1.91, 2.07, 2.12, 2.18 (5s, 15H, 5AcO), 2.43 (dd, 1H, *J*_{gem} = 13.4, *J*_{3eq,4} = 4.9 Hz, H-3deq), 2.45 (m, 2H, N—CH₂—C₃H₇), 2.91 (dd, 1H, H-1aeq), 3.81 (s, 3H, COOMe), 4.68 (m, 1H, H-2a), 4.82 (m, 1H, H-4d), 5.03 (d, 1H, *J*_{1,2} = 3.3 Hz, H-1b), 5.16 (d, 1H, *J*_{1,2} = 8.1 Hz, H-1c), 5.26 (dd, 1H, *J*_{7,8} = 9.5 Hz, H-7), 5.33 (dd, 1H, *J*_{3,4} = 3.5 Hz, H-4c), 5.36 (t, 1H, H-2c), 5.56 (m, 1H, H-8d), 7.41–8.09 (m, 15H, 3Ph).

Anal. Calcd for C₆₅H₈₂N₂O₂₉ (1355.36): C, 57.60; H, 6.01; N, 2.07. Found: C, 57.47; H, 6.02; N, 1.92.

(Methyl 5-Acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-(2,4,6-tri-*O*-benzoyl- β -D-galactopyranosyl)-(1 \rightarrow 3)-[α -L-fucopyranosyl-(1 \rightarrow 4)]-2-*O*-acetyl-*N*-decyl-1,5-dideoxy-1,5-imino-D-glucitol (10). A mixture of compound **7** (20 mg), decyl aldehyde (0.5 mL), ethyl acetate (5 mL), acetic acid (0.5 mL) and water (0.1 mL) was hydrogenated as described for **6**. Work-up and column chromatography (20:1 CH₂Cl₂—MeOH) on silica gel gave **10** (11 mg, 56%): [α]_D $+2.5^{\circ}$ (*c* 0.24, CH₂Cl₂); ¹H NMR (CDCl₃) δ 0.88 (t, 3H, *J* = 6.2 Hz, Me), 1.25 (m, 16H, —C₈H₁₆Me), 1.25 (d, 3H, H-6b), 1.52 (s, 3H, AcN), 1.67 (t, 1H, *J*_{gem} = *J*_{3ax,4} = 12.6 Hz, H-3dax), 1.78, 1.91, 2.07, 2.13, 2.20 (5s, 15H, 5AcO), 2.44 (dd, 1H, *J*_{gem} = 12.5, *J*_{3eq,4} = 4.2 Hz, H-3deq), 2.45 (m, 2H, NCH₂), 2.95 (dd, 1H, *J*_{gem} = 11.7, *J*_{1eq,2} = 4 Hz, H-1aeq), 3.82 (s, 3H, COOMe), 4.72 (m, 1H, H-2a), 4.83 (m, 1H, H-4d), 5.03 (d, 1H, *J*_{1,2} = 3.5 Hz, H-1b), 5.16 (d, 1H, H-1c), 5.26 (dd, 1H, *J*_{6,7} = 2.6, *J*_{7,8} = 9.5 Hz, H-7d), 5.31 (d, 1H, *J*_{3,4} = 4.2 Hz, H-4c), 5.36 (dd, 1H, *J*_{1,2} = 8.24, *J*_{2,3} = 10.3 Hz, H-2c), 5.56 (m, 1H, H-8d), 7.42–8.10 (m, 15H, 3Ph).

Anal. Calcd for C₇₁H₉₄N₂O₂₉ (1439.52): C, 59.24; H, 6.58; N, 1.95. Found: C, 59.18; H, 6.44; N, 1.93.

(5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonic acid)-(2 \rightarrow 3)-(β -D-galactopyranosyl)-(1 \rightarrow 4)-[α -L-fucopyranosyl-(1 \rightarrow 3)]-1,5-dideoxy-1,5-imino-D-glucitol (11). Compound **3** (30.3 mg) was treated with a catalytic amount of NaOMe in dry MeOH overnight. One mL of 0.2 M KOH was added and the mixture was stirred overnight at rt. The solution was neutralized with Amberlite IR-120 (H⁺), then filtered and concentrated. The product was purified by column chromatography (1:1 MeOH—H₂O) on Sephadex LH-20 to give **11** (17.8 mg) in quantitative yield: [α]_D -19° (*c* 0.5, 1:1 MeOH—H₂O); ¹H NMR (D₂O) δ 1.18 (d, 3H, H-6b), 1.78 (t, 3H, *J* = 12 Hz, H-3dax), 2.03 (s, 3H, AcN),



2.76 (dd, 1H, $J_{\text{gem}} = 12$, $J_{3\text{eq},4} = 4.4$ Hz, H-3 deq), 3.03 (dd, 1H, $J_{\text{gem}} = 12.5$ Hz, H-1 aeq), 5.39 (d, 1H, $J_{1,2} = 4$ Hz, H-1b), 5.54 (d, 1H, $J_{1,2} = 7.7$ Hz, H-1c): FAB-MS (negative ion mode): m/z 761.35 ($\text{M}-\text{H}$)⁻ for $\text{C}_{29}\text{H}_{49}\text{N}_2\text{O}_{21}$ (Exact mass: 761.2828).

Anal. Calcd for $\text{C}_{29}\text{H}_{50}\text{N}_2\text{O}_{21}$ (762.71): C, 45.67; H, 6.61; N, 3.67. Found: C, 45.38; H, 6.42; N, 3.49.

(5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonic acid)-(2 \rightarrow 3)-(β -D-galactopyranosyl)-(1 \rightarrow 4)-[α -L-fucopyranosyl-(1 \rightarrow 3)]-N-butyl-1,5-dideoxy-1,5-imino-D-glucitol (13). Compound **5** (50 mg) was treated with a catalytic amount of NaOMe, and then 0.2 M KOH as described for **11**. Work-up and column chromatography (1:1 MeOH—H₂O) on Sephadex LH-20 gave **13** (30 mg, quant): $[\alpha]_{\text{D}} -46^\circ$ (c 0.5, 1:1 MeOH—H₂O); ¹H NMR (D₂O) δ 0.95 (t, 3H, $J = 7.3$ Hz, Me), 1.19 (d, 3H, $J_{5,6} = 6.6$ Hz, H-6b), 1.40, 1.74 (m, 4H, C₂H₄Me), 1.79 (t, 3H, $J = 12$ Hz, H-3 dax), 2.03 (s, 3H, AcN), 2.77 (dd, 1H, $J_{\text{gem}} = 12.5$, $J_{3\text{eq},4} = 4.4$ Hz, H-3 deq), 3.29 (m, 2H, N—CH₂—), 5.36 (d, 1H, H-1b).

Anal. Calcd for $\text{C}_{33}\text{H}_{58}\text{N}_2\text{O}_{21}$ (818.82): C, 48.41; H, 7.14; N, 3.42. Found: C, 48.26; H, 6.88; N, 3.18.

(5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonic acid)-(2 \rightarrow 3)-(β -D-galactopyranosyl)-(1 \rightarrow 4)-[α -L-fucopyranosyl-(1 \rightarrow 3)]-N-decyl-1,5-dideoxy-1,5-imino-D-glucitol (14). The title compound was prepared from **6** (65 mg) as described for **13** and purified by column chromatography (2:1 MeOH—H₂O) on Sephadex LH-20 to afford **14** (41 mg, quant): $[\alpha]_{\text{D}} -43^\circ$ (c 0.8, 2:1 MeOH—H₂O); ¹H NMR (D₂O) δ 0.87 (t, 3H, Me), 1.18 (d, 3H, $J_{5,6} = 6$ Hz, H-6b), 1.28, 1.59 (m, 16H, C₈H₁₆Me), 1.80 (t, 3H, H-3 dax), 1.90 (s, 3H, AcN), 5.41 (d, 1H, H-1b).

Anal. Calcd for $\text{C}_{39}\text{H}_{70}\text{N}_2\text{O}_{21}$ (902.98): C, 51.88; H, 7.81; N, 3.10. Found: C, 51.74; H, 7.67; N, 3.00.

(5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonic acid)-(2 \rightarrow 3)-(β -D-galactopyranosyl)-(1 \rightarrow 3)-[α -L-fucopyranosyl-(1 \rightarrow 4)]-1,5-dideoxy-1,5-imino-D-glucitol (15). The title compound was prepared from **7** (62 mg) as described for **11** to give **15** (36.4 mg, quant): $[\alpha]_{\text{D}} -30^\circ$ (c 0.74, 1:1 MeOH—H₂O); ¹H NMR (D₂O) δ 1.19 (d, 3H, H-6b), 1.80 (t, 1H, $J = 12$ Hz, H-3 dax), 2.03 (s, 3H, AcN), 2.77 (dd, 1H, $J_{\text{gem}} = 12$, $J_{3\text{eq},4} = 4.4$ Hz, H-3 deq), 2.99 (dd, 1H, H-1 aeq), 4.83 (d, 1H, $J_{1,2} = 8.1$ Hz, H-1c), 5.02 (d, 1H, $J_{1,2} = 3.5$ Hz, H-1b): FAB-MS (negative ion mode): m/z 761.32 ($\text{M}-\text{H}$)⁻ for $\text{C}_{29}\text{H}_{49}\text{N}_2\text{O}_{21}$ (Exact mass: 761.2828).

Anal. Calcd for $\text{C}_{29}\text{H}_{50}\text{N}_2\text{O}_{21}$ (762.71): C, 45.67; H, 6.61; N, 3.67. Found: C, 45.42; H, 6.60; N, 3.60.

(5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonic acid)-(2 \rightarrow 3)-(β -D-galactopyranosyl)-(1 \rightarrow 3)-[α -L-fucopyranosyl-(1 \rightarrow 4)]-N-butyl-1,5-dideoxy-1,5-imino-D-glucitol (17). Deprotection of **9** (34 mg) and



column chromatography as described for gave the title compound **17** (20 mg, quant): $[\alpha]_D -52^\circ$ (*c* 0.45, 1:1 MeOH—H₂O); ¹H NMR (D₂O) δ 0.96 (t, 3H, *J* = 7.3 Hz, Me), 1.22 (d, 3H, *J*_{5,6} = 6.6 Hz, H-6b), 1.40, 1.72 (m, 4H, C₂H₄Me), 1.81 (t, 1H, *J* = 12.5 Hz, H-3*dax*), 2.05 (s, 3H, AcN), 2.78 (dd, 1H, *J*_{gem} = 12.5, *J*_{3eq,4} = 4.6 Hz, H-3*deq*), 5.11 (d, 1H, H-1b).

Anal. Calcd for C₃₃H₅₈N₂O₂₁ (818.82): C, 48.41; H, 7.14; N, 3.42. Found: C, 48.11; H, 7.04; N, 3.27.

(5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonic acid)-(2 \rightarrow 3)-(β -D-galactopyranosyl)-(1 \rightarrow 3)-[α -L-fucopyranosyl-(1 \rightarrow 4)]-N-decyl-1,5-dideoxy-1,5-imino-D-glucitol (18**). The title compound was prepared from **10** (52 mg) as described for **14** to afford **18** (33 mg, quant): $[\alpha]_D -35^\circ$ (*c* 0.62, 2:1 MeOH—H₂O); ¹H NMR (D₂O) δ 0.87 (t, 3H, Me), 1.18 (d, 3H, *J* = 6 Hz, H-6b), 1.28, 1.55 (m, 16H, C₈H₁₆Me), 1.80 (t, 3H, H-3*dax*), 1.90 (s, 3H, AcN), 5.15 (d, 1H, H-1b).**

Anal. Calcd for C₃₉H₇₀N₂O₂₁ (902.98): C, 51.88; H, 7.81; N, 3.10. Found: C, 51.69; H, 7.68; N, 2.93.

(2-O-Benzoyl-4,6-O-benzylidene-3-O-levulinoyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-[2,3,4-tri-O-benzyl- α -L-fucopyranosyl-(1 \rightarrow 3)]-2-O-acetyl-6-O-benzyl-N-benzylloxycarbonyl-1,5-dideoxy-1,5-imino-D-glucitol (21**). To a stirred mixture of **19** (154 mg, 1 equiv), methyl 2-O-benzoyl-4,6-O-benzylidene-3-O-levulinoyl-1-thio- β -D-galactopyranoside²⁰ (**20**, 182 mg, 2 equiv), molecular sieves 4Å (300 mg) and CH₂Cl₂ (5 mL), were added *N*-iodosuccinimide (NIS, 164 mg, 4 equiv) and trimethylsilyl trifluoromethanesulfonate (TMSOTf, 14 μ L, 0.4 equiv) at 0°C, and stirring was continued overnight at 0°C~rt. The solids were filtered off and the filtrate was successively washed with M Na₂CO₃, Na₂S₂O₃ and water, dried (Na₂SO₄), and concentrated. The residue was chromatographed (1:2 EtOAc-hexane) on a column of silica gel to give **21** (164 mg, 70%): $[\alpha]_D -30^\circ$ (*c* 1.33, CH₂Cl₂); ¹H NMR (CDCl₃) δ 1.10 (d, 3H, *J*_{5,6} = 6.6 Hz, H-6b), 1.63 (s, 3H, AcO), 1.92 (s, 3H, Lev—Me), 2.43–2.61 (m, 4H, Lev—C₂H₄), 3.28 (dd, 1H, *J*_{gem} = 11.9 Hz, H-1a), 4.44 (d, 1H, *J*_{1,2} = 8.2 Hz, H-1c), 4.56–4.70 (4d, 4H, 2PhCH₂O), 4.89 (d, 1H, H-1b), 4.90 (dd, 1H, H-3c), 4.89, 4.97 (2d, 2H, *J*_{gem} = 12.5 Hz, PhCH₂OCO), 5.52 (t, 1H, *J*_{1,2} = *J*_{2,3} = 8.24 Hz, H-2c), 5.53 (s, 1H, benzylidene CH), 7.17–7.94 (m, 35H, 7Ph).**

Anal. Calcd for C₇₅H₇₉NO₁₉ (1298.44): C, 69.38; H, 6.13; N, 1.08. Found: C, 69.13; H, 6.06; N, 1.03.

(2-O-Benzoyl-3-O-levulinoyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-[2,3,4-tri-O-benzyl- α -L-fucopyranosyl-(1 \rightarrow 3)]-2-O-acetyl-6-O-benzyl-N-benzylloxycarbonyl-1,5-dideoxy-1,5-imino-D-glucitol (22**). A solution of **21** (164 mg) in 80% aq AcOH (10 mL) was stirred overnight at 45°C and concentrated. The residue was chromatographed (3:1 EtOAc-hexane) on a column of silica gel to give **22** (136 mg, 89%): $[\alpha]_D -39^\circ$ (*c* 1, CH₂Cl₂); ¹H NMR (CDCl₃) δ 1.01 (d, 3H, *J*_{5,6} = 6 Hz, H-6b), 1.92 (s, 3H, AcO), 2.10 (s, 3H, Lev—Me), 2.37–2.69 (m, 4H,**



Lev—C₂H₄), 3.30 (dd, 1H, H-1a), 4.92 (br s, 1H, H-1b), 5.50 (t, 1H, J_{1,2} = J_{2,3} = 9.9 Hz, H-2c), 7.19–7.93 (m, 30H, 4Ph).

Anal. Calcd for C₆₈H₇₅NO₁₉ (1210.34): C, 67.48; H, 6.25; N, 1.16. Found: C, 67.42; H, 6.18; N, 0.98.

(2,4,6-Tri-*O*-benzoyl-3-*O*-levulinoyl-β-D-galactopyranosyl)-(1→4)-[(2,3,4-tri-*O*-benzyl-α-L-fucopyranosyl)-(1→3)]-2-*O*-acetyl-6-*O*-benzyl-*N*-benzyloxycarbonyl-1,5-dideoxy-1,5-imino-D-glucitol (23) and (2,4,6-Tri-*O*-benzoyl-β-D-galactopyranosyl)-(1→4)-[(2,3,4-tri-*O*-benzyl-α-L-fucopyranosyl)-(1→3)]-2-*O*-acetyl-6-*O*-benzyl-*N*-benzyloxycarbonyl-1,5-dideoxy-1,5-imino-D-glucitol (24). To a solution of **22** (48 mg) in pyridine (5 mL) was added benzoyl chloride (18 μL, 4 equiv) and the mixture was stirred overnight at rt. Work-up and column chromatography (2:3 EtOAc-hexane) gave **23** (48 mg, 86%): [α]_D −37° (c 1, CH₂Cl₂); ¹H NMR (CDCl₃) δ 0.99 (d, 3H, H-6b), 1.64 (s, 3H, AcO), 1.88 (s, 3H, Lev—Me), 2.32–2.59 (m, 4H, Lev—C₂H₄), 3.23 (near d, 1H, J_{gem} = 14.7 Hz, H-1a), 3.62 (dd, 1H, J_{2,3} = 10.3, J_{3,4} = 2.2 Hz, H-3b), 4.53 (d, 1H, J_{1,2} = 8.8 Hz, H-1c), 4.58, 4.69 (2d, 2H, J_{gem} = 12.3 Hz, PhCH₂OCO), 4.94 (d, 1H, J_{1,2} = 3.7 Hz, H-1b), 5.28 (dd, 1H, J_{2,3} = 10.3 Hz, H-3c), 5.52 (dd, 1H, H-2c), 5.76 (d, 1H, J_{3,4} = 3.3 Hz, H-4c), 7.21–8.12 (m, 40H, 8Ph).

To a solution of **23** (131 mg) in EtOH (20 mL) was added hydrazine acetate (10 mg, 1.2 equiv) and the mixture was stirred for 1 h at rt, and then concentrated. The residue was chromatographed (125:1 CH₂Cl₂—MeOH) on a column of silica gel to afford **24** (122 mg, quant): [α]_D −50° (c 2.4, CH₂Cl₂); ¹H NMR (CDCl₃) δ 0.98 (d, 3H, J_{5,6} = 6 Hz, H-6b), 1.81 (s, 3H, AcO), 3.62 (near d, 1H, J_{gem} = 14.7 Hz, H-1a), 3.95 (dd, 1H, J_{2,3} = 10 Hz, H-3c), 4.94 (s, 1H, J_{1,2} = 3.7 Hz, H-1b), 5.72 (d, 1H, J_{3,4} = 3.5 Hz, H-4c), 7.13–8.14 (m, 40H, 8Ph).

Anal. Calcd for C₇₇H₇₇NO₁₉ (1320.45): C, 70.04; H, 5.88; N, 1.06. Found: C, 69.88; H, 5.75; N, 1.01.

(2,4,6-Tri-*O*-benzoyl-3-*O*-sulfo-β-D-galactopyranosyl)-(1→4)-[(2,3,4-tri-*O*-benzyl-α-L-fucopyranosyl)-(1→3)]-2-*O*-acetyl-6-*O*-benzyl-*N*-benzyloxycarbonyl-1,5-dideoxy-1,5-imino-D-glucitol (25) and (2,4,6-Tri-*O*-benzoyl-3-*O*-sulfo-β-D-galactopyranosyl)-(1→4)-[α-L-fucopyranosyl)-(1→3)]-2-*O*-acetyl-1,5-dideoxy-1,5-imino-*N*-methyl-D-glucitol sodium salt (26). To a solution of **24** (119 mg) in *N,N*-dimethylformamide (0.5 mL) was added pyridine sulfur trioxide complex (115 mg, 8 equiv) and the mixture was stirred for 1 h at rt, then cooled to 0°C. MeOH (1 mL) was added and the mixture was stirred for 30 min at 0°C and concentrated. Column chromatography (15:1 CH₂Cl₂—MeOH) of the residue on silica gel gave **25** (121 mg, 96%): [α]_D −16.6° (c 2.4, CH₂Cl₂). Compound **25** (114 mg) in MeOH was treated with cation-exchange resin Dowex-Na⁺, and then hydrogenolyzed in the presence of formalin (0.5 mL) and HCl-free palladium black (300 mg). Work-up and column chromatography (10:1 CH₂Cl₂—MeOH) afforded **26** (62 mg, 83%): [α]_D −30.5° (c 0.89, MeOH); ¹H NMR (DMSO-*d*₆, 50°C) δ 1.18 (d, 3H, J_{5,6} = 6.6 Hz, H-6b), 1.99 (s, 3H, AcO), 2.04 (t, 1H, J_{1ax,2} = 10.4 Hz, H-1aax), 2.12 (s, 3H, *N*—Me), 2.79 (dd, 1H, J_{gem} = 10.8, J_{1eq,2} = 4.8 Hz, H-1aeq),



4.10, 4.51 (2dd, 2H, $J_{\text{gem}} = 10.8$, $J_{5,6} = 8.3$, 4.8 Hz, H-6c,6'c), 4.72 (dd, 1H, $J_{2,3} = 10.3$, $J_{3,4} = 3.7$ Hz, H-3c), 4.94 (d, 1H, $J_{1,2} = 3.7$ Hz, H-1b), 5.05 (d, 1H, $J_{1,2} = 8.2$ Hz, H-1c), 5.41 (dd, 1H, $J_{2,3} = 10.3$ Hz, H-2c), 5.82 (d, 1H, $J_{3,4} = 3.7$ Hz, H-4c), 7.49–8.12 (m, 15H, 3Ph).

Anal. Calcd for $\text{C}_{42}\text{H}_{48}\text{NO}_{20}\text{SNa}$ (941.89): C, 53.56; H, 5.14; N, 1.49. Found: C, 53.46; H, 4.97; N, 1.44.

(3-*O*-Sulfo- β -D-galactopyranosyl)-(1 \rightarrow 4)-[α -L-fucopyranosyl-(1 \rightarrow 3)]-1,5-dideoxy-1,5-imino-*N*-methyl-D-glucitol sodium salt (27). A mixture of **26** (49 mg) and a small amount of NaOMe in MeOH was stirred overnight at rt and concentrated. The residue was chromatographed (1:1 MeOH–H₂O) on a column of Sephadex LH-20 to give **27** (30 mg, quant) as an amorphous mass: ¹H NMR (DMSO-*d*₆, 55°C) δ 1.01 (d, 3H, $J_{5,6} = 6.4$ Hz, H-6b), 2.00 (t, 1H, $J_{\text{gem}} = J_{1\text{ax},2} = 10.7$ Hz, H-1aax), 2.23 (s, 3H, *N*–Me), 2.77 (dd, 1H, $J_{\text{gem}} = 11$, $J_{1\text{eq},2} = 5.1$ Hz, H-1aeq), 4.49 (d, 1H, $J_{1,2} = 8.3$ Hz, H-1c), 4.60 (q, 1H, $J_{5,6} = 6.4$ Hz, H-5b), 5.11 (d, 1H, $J_{1,2} = 3.6$ Hz, H-1b). LRMS (ion-spray MS, negative ion mode) *m/z* 564.0 (*M*–Na–H)[–], (ion-spray MS, positive ion mode) *m/z* 566.2 (*M*–Na+H)⁺. HRMS Calcd for $\text{C}_{19}\text{H}_{35}\text{NO}_{16}\text{S}$: 565.1677. Found: 565.1675.

(2-*O*-Benzoyl-3-*O*-levulinoyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-[α -L-fucopyranosyl-(1 \rightarrow 3)]-2-*O*-acetyl-1,5-dideoxy-1,5-imino-D-glucitol (28). Compound **22** (457 mg) in AcOH (15 mL) was hydrogenolyzed with palladium black (500 mg). Work-up and column chromatography (5:1 CH₂Cl₂–MeOH) gave **28** (213 mg, 79%): $[\alpha]_{\text{D}} -50^{\circ}$ (*c* 1.1, MeOH); ¹H NMR (CD₃OD) δ 1.34 (d, 3H, H-6b), 2.10 (s, 3H, Lev–Me), 2.43–2.64 (m, 4H, Lev–C₂H₄), 3.42 (dd, 1H, H-1aeq), 5.05 (d, 1H, H-1c), 5.13 (d, 1H, H-1b), 5.55 (d, 1H, $J_{1,2} = J_{2,3} = 9.7$ Hz, H-2c).

Anal. Calcd for $\text{C}_{32}\text{H}_{45}\text{NO}_{17}$ (715.70): C, 53.70; H, 6.34; N, 1.96. Found: C, 53.64; H, 6.06; N, 1.66.

(2-*O*-Benzoyl-3-*O*-levulinoyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-[α -L-fucopyranosyl-(1 \rightarrow 3)]-2-*O*-acetyl-*N*-butyl-1,5-dideoxy-1,5-imino-D-glucitol (29). A mixture of **28** (51.3 mg), 1-butanol (2 mL) and butyraldehyde (98 μ L, 20 equiv) was hydrogenated in the presence of palladium hydroxide on carbon (50 mg). Work-up and column chromatography (10:1 CH₂Cl₂–MeOH) on silica gel afforded **29** (43.7 mg, 79%): $[\alpha]_{\text{D}} -53.6^{\circ}$ (*c* 0.88, CH₂Cl₂); ¹H NMR (CD₃OD) δ 0.85 (t, 3H, Me), 1.15–1.35 (m, 4H, C₂H₄), 1.35 (d, 3H, H-6b), 1.97 (s, 3H, AcO), 2.05 (s, 3H, Lev–Me), 2.19 (t, 1H, $J = 10.6$ Hz, H-1aax), 2.35–2.70 (m, 6H, Lev–C₂H₄, *N*–CH₂), 3.02 (dd, 1H, $J_{\text{gem}} = 10.8$, $J_{1\text{eq},2} = 4.8$ Hz, H-1aeq), 4.09 (d, 1H, $J_{3,4} = 3.0$ Hz, H-4c), 4.90 (m, 1H, H-2a), 4.99 (d, 1H, $J_{1,2} = 7.9$ Hz, H-1c), 5.05 (dd, 1H, $J_{2,3} = 10$, $J_{3,4} = 3.0$ Hz, H-3c), 5.14 (d, 1H, $J_{1,2} = 3.8$ Hz, H-1b), 5.54 (dd, 1H, $J_{1,2} = 8$, $J_{2,3} = 10$ Hz, H-2c), 7.47–8.10 (m, 5H, Ph).

Anal. Calcd for $\text{C}_{36}\text{H}_{53}\text{NO}_{17}$ (771.81): C, 56.02; H, 6.92; N, 1.81. Found: C, 55.90; H, 6.84; N, 1.79.



(2-*O*-Benzoyl-3-*O*-levulinoyl-β-D-galactopyranosyl)-(1→4)-[α-L-fucopyranosyl-(1→3)]-2-*O*-acetyl-*N*-decyl-1,5-dideoxy-1,5-imino-D-glucitol (30). A mixture of **28** (73 mg), decyl aldehyde (296 μL, 20 equiv) and 1:1 EtOAc-THF (2 mL) was hydrogenated in the presence of palladium hydroxide on carbon (55 mg). Work-up and column chromatography as described for **29** gave **30** (70 mg, 80%): $[\alpha]_D -47.6^\circ$ (*c* 1.4, CH₂Cl₂); ¹H NMR (CD₃OD) δ 0.89 (t, 3H, Me), 1.15–1.35 (m, 16H, C₈H₁₆), 1.36 (d, 3H, H-6b), 1.98 (s, 3H, AcO), 2.05 (s, 3H, Lev—Me), 2.19 (t, 1H, *J* = 10.6 Hz, H-1_{ax}), 2.35–2.70 (m, 6H, Lev—C₂H₄, N—CH₂), 3.02 (dd, 1H, *J*_{gem} = 10.8, *J*_{1eq,2} = 4.8 Hz, H-1_{aeq}), others are same as those of **29**.

Anal. Calcd for C₄₂H₆₅NO₁₇ (855.97): C, 58.93; H, 7.65; N, 1.64 Found: C, 58.93; H, 7.53; N, 1.60.

(4,6-Di-*O*-acetyl-2-*O*-benzoyl-3-*O*-levulinoyl-β-D-galactopyranosyl)-(1→4)-[(2,3,4-tri-*O*-acetyl-α-L-fucopyranosyl)-(1→3)]-2,6-di-*O*-acetyl-*N*-butyl-1,5-dideoxy-1,5-imino-D-glucitol (31) and (4,6-Di-*O*-acetyl-2-*O*-benzoyl-β-D-galactopyranosyl)-(1→4)-[(2,3,4-tri-*O*-acetyl-α-L-fucopyranosyl)-(1→3)]-2,6-di-*O*-acetyl-*N*-butyl-1,5-dideoxy-1,5-imino-D-glucitol (33). Compound **29** (45 mg) in pyridine (1 mL) was treated with Ac₂O (66 μL) overnight at rt. Work-up and column chromatography (50:1 CH₂Cl₂—MeOH) on silica gel gave **31** (60 mg, quant): $[\alpha]_D -62^\circ$ (*c* 1.12, CH₂Cl₂); ¹H NMR (CD₃OD) δ 0.83 (t, 3H, Me), 1.10–1.30 (m, 4H, C₂H₄), 1.34 (d, 3H, H-6b), 1.98–2.24 (8s, 24H, 7AcO, Lev—Me), 2.30–2.70 (m, 6H, Lev—C₂H₄, N—CH₂), 3.02 (dd, 1H, *J*_{gem} = 11.2, *J*_{1eq,2} = 5.3 Hz, H-1_{aeq}), 3.74, 3.85 (2t, 2H, *J* = 9 Hz, H-3_{a,4a}), 3.90 (t, 1H, *J* = 7 Hz, H-5c), 4.19, 4.28 (2dd, 2H, H-6_{a,6'a}), 4.34, 4.57 (2dd, 2H, *J*_{gem} = 11.4, *J*_{5,6} = 7.7, *J*_{5,6'} = 6.6 Hz, H-6_{c,6'c}), 4.61 (d, 1H, *J*_{1,2} = 8.2 Hz, H-1c), 4.90 (m, 1H, H-2a), 5.03 (q, 1H, *J* = 6 Hz, H-5b), 5.07 (dd, *J*_{1,2} = 4, *J*_{2,3} = 10.8 Hz, H-2b), 5.11 (dd, 1H, *J*_{2,3} = 10.3, *J*_{3,4} = 3.5 Hz, H-3c), 5.22 (dd, 1H, *J*_{2,3} = 10.8, *J*_{3,4} = 3.3 Hz, H-3b), 5.39 (dd, 1H, *J*_{1,2} = 8.2, *J*_{2,3} = 10.3 Hz, H-2c), 5.45–5.50 (m, 3H, H-1b, H-4b, H-4c), 7.45–8.10 (m, 5H, Ph).

A mixture of **31** (55.2 mg) and hydrazine acetate (6.1 mg, 1.2 equiv) in ethanol was stirred for 1.5 h at rt. Work-up and column chromatography (100:1 CH₂Cl₂—MeOH) afforded **33** (46.5 mg, 93%): $[\alpha]_D -74.4^\circ$ (*c* 0.93, CH₂Cl₂); ¹H NMR (CD₃OD) δ 1.98–2.24 (7s, 21H, 7AcO), 2.87 (br s, 1H, 3-OH of Gal), 3.95 (br dd, 1H, *J*_{2,3} = 10.4, *J*_{3,4} = 4 Hz, H-3c), 5.20 (dd, 1H, *J*_{1,2} = 8.2, *J*_{2,3} = 10.4 Hz, H-2c), 5.38 (dd, 1H, *J*_{3,4} = 3.7 Hz, H-4c), 5.44 (d, 1H, *J*_{3,4} = 2.9 Hz, H-4b), 5.47 (d, 1H, *J*_{1,2} = 4 Hz, H-1b), other peaks are similar to those of **31**.

Anal. Calcd for C₄₃H₅₉NO₂₁ (925.93): C, 55.78; H, 6.42; N, 1.51. Found: C, 55.53; H, 6.37; N, 1.25.

(4,6-Di-*O*-acetyl-2-*O*-benzoyl-3-*O*-levulinoyl-β-D-galactopyranosyl)-(1→4)-[(2,3,4-tri-*O*-acetyl-α-L-fucopyranosyl)-(1→3)]-2,6-di-*O*-acetyl-*N*-decyl-1,5-dideoxy-1,5-imino-D-glucitol (32) and (4,6-Di-*O*-acetyl-2-*O*-benzoyl-β-D-galactopyranosyl)-(1→4)-[(2,3,4-tri-*O*-acetyl-α-L-fucopyranosyl)-(1→3)]-2,6-di-*O*-acetyl-*N*-decyl-1,5-dideoxy-1,5-imino-D-glucitol (35). Acetylation of **30** (70 mg) was performed as described for **31**. Column chromatography (100:1



CH₂Cl₂—MeOH) on silica gel gave **32** (91 mg, quant): [α]_D −58.4° (c 1.64, CH₂Cl₂); ¹H NMR (CD₃OD) δ 0.88 (t, 3H, Me), 1.10–1.30 (m, 16H, C₈H₁₆), 1.98–2.24 (8s, 24H, 7AcO, Lev—Me), 5.11 (dd, 1H, J_{2,3} = 10, J_{3,4} = 3.5 Hz, H-3c), other peaks are similar to those of **31**.

The levulinoyl group of **32** (81.7 mg) was selectively removed by treatment with hydrazine acetate (8.4 mg, 1.2 equiv) as described for **33** to afford **35** (71 mg, 95%): [α]_D −67.4° (c 1.42, CH₂Cl₂); ¹H NMR (CD₃OD) δ 1.98–2.24 (7s, 21H, 7AcO), 2.86 (br s, 1H, 3-OH of Gal), 3.01 (dd, 1H, J_{gem} = 11.2, J_{1eq,2} = 5.3 Hz, H-1aeq), 3.73, 3.82 (2t, 2H, J_{3,4} = J_{4,5} = 9.2 Hz, H-3a, H-4a), 3.85 (t, 1H, H-5c), 3.95 (dd, 1H, J_{2,3} = 10.4, J_{3,4} = 3.7 Hz, H-3c), 4.25, 4.37 (2dd, 2H, H-6a, H-6'a), 4.34, 4.54 (2dd, 2H, H-6c, H-6'c), 4.50 (d, 1H, J_{1,2} = 8 Hz, H-1c), 4.89 (m, 1H, J_{1ax,2} = J_{2,3} = 10, J_{1eq,2} = 5 Hz, H-2a), 5.03 (q, 1H, J_{5,6} = 6 Hz, H-5b), 5.07 (dd, 1H, J_{1,2} = 4, J_{2,3} = 10.8 Hz, H-2b), 5.20 (dd, 1H, J_{1,2} = 8, J_{2,3} = 10.4 Hz, H-2c), 5.21 (dd, 1H, J_{2,3} = 10.8, J_{3,4} = 3 Hz, H-3b), 5.38 (d, 1H, H-4c), 5.44 (d, 1H, H-4b), 5.47 (d, 1H, H-1b), 7.45–8.15 (m, 5H, Ph).

Anal. Calcd for C₄₉H₇₁NO₂₁ (1010.09): C, 58.27; H, 7.09; N, 1.39. Found: C, 58.25; H, 6.92; N, 1.29.

(4,6-Di-*O*-acetyl-2-*O*-benzoyl-3-*O*-sulfo-β-D-galactopyranosyl)-(1→4)-[(2,3,4-tri-*O*-acetyl-α-L-fucopyranosyl)-(1→3)]-2,6-di-*O*-acetyl-*N*-butyl-1,5-dideoxy-1,5-imino-D-glucitol (34**) and (3-*O*-Sulfo-β-D-galactopyranosyl)-(1→4)-[α-L-fucopyranosyl-(1→3)]-*N*-butyl-1,5-dideoxy-1,5-imino-D-glucitol sodium salt (**37**).** To a solution of **33** (38.4 mg) in *N,N*-dimethylformamide (2 mL) was added pyridine sulfur trioxide complex (33.7 mg, 5 equiv) and the mixture was stirred for 2 h at rt, then cooled to 0°C. MeOH (1 mL) was added and the mixture was stirred for 30 min at 0°C and concentrated. Column chromatography (10:1 CH₂Cl₂—MeOH) of the residue on silica gel gave **34** (36.4 mg, 81%): [α]_D −42° (c 0.75, CH₂Cl₂); ¹H NMR (DMSO-*d*₆) δ 4.68 (dd, 1H, J_{2,3} = 10.3, J_{3,4} = 2.9 Hz, H-3c), 4.75 (d, 1H, J_{1,2} = 8 Hz, H-1c).

A mixture of **34** (40 mg) and methanolic sodium methoxide (3 mL) was stirred overnight at rt, and then concentrated. Column chromatography (1:1 MeOH—H₂O) of the residue on Sephadex LH-20 afforded **37** (41 mg, quant): [α]_D −40° (c 0.92, 1:1 MeOH—H₂O); ¹H NMR (DMSO-*d*₆) δ 0.86 (t, 3H, Me), 1.00 (d, 3H, J_{5,6} = 6.4 Hz, H-6b), 1.10–1.40 (m, 4H, C₂H₄), 2.08 (t, 1H, J_{gem} = J_{1ax,2} = 11 Hz, H-1aax), 2.79 (dd, 1H, J_{1eq,2} = 5 Hz, H-1aeq), 4.42 (d, 1H, J_{1,2} = 7.7 Hz, H-1c), 4.52 (q, 1H, H-5b), 5.04 (d, 1H, J_{1,2} = 3.7 Hz, H-1b). LRMS (FAB-MS, negative ion mode) *m/z* 628.27 (M—H)[−], 606.27 (M—Na—H)[−]. HRMS Calcd for C₂₂H₃₉NO₁₆SN_a: 628.1887. Found: 628.1886.

(4,6-Di-*O*-acetyl-2-*O*-benzoyl-3-*O*-sulfo-β-D-galactopyranosyl)-(1→4)-[(2,3,4-tri-*O*-acetyl-α-L-fucopyranosyl)-(1→3)]-2,6-di-*O*-acetyl-*N*-decyl-1,5-dideoxy-1,5-imino-D-glucitol (36**) and (3-*O*-Sulfo-β-D-galactopyranosyl)-(1→4)-[α-L-fucopyranosyl-(1→3)]-*N*-decyl-1,5-dideoxy-1,5-imino-D-glucitol sodium salt (**38**).** To a solution of **35** (46.4 mg) in DMF (2 mL) was added pyridine sulfur trioxide complex (74.6 mg, 10 equiv) and the mixture was stirred for



1.5 h at rt. Work-up and column chromatography (20:1 CH₂Cl₂—MeOH) as described for **34** gave **36** (47.5 mg, 88%): $[\alpha]_D -35.7^\circ$ (*c* 0.85, CH₂Cl₂); ¹H NMR (DMSO-*d*₆) δ 4.70 (dd, 1H, *J*_{2,3} = 10.3, *J*_{3,4} = 3.3 Hz, H-3c), 4.76 (d, 1H, *J*_{1,2} = 8.6 Hz, H-1c).

A mixture of **36** (45 mg) in methanolic sodium methoxide (3 mL) was treated as described for **37** to afford **38** (46 mg, quant): $[\alpha]_D -51^\circ$ (*c* 1, 1:1 MeOH—H₂O); ¹H NMR (DMSO-*d*₆) δ 0.86 (t, 3H, Me), 1.00 (d, 3H, *J*_{5,6} = 6.4 Hz, H-6b), 1.10–1.40 (m, 16H, C₈H₁₆), 2.08 (t, 1H, *J* = 10 Hz, H-1aax), 2.79 (dd, 1H, *J*_{gem} = 11, *J*_{1eq,2} = 5 Hz, H-1aeq), 4.43 (d, 1H, *J*_{1,2} = 7.5 Hz, H-1c), 4.53 (q, 1H, H-5b), 5.06 (d, 1H, *J*_{1,2} = 3.7 Hz, H-1b). LRMS (FAB-MS, negative ion mode) *m/z* 712.32 (M—H)[−], 690.38 (M—Na—H)[−]. HRMS Calcd for C₂₈H₅₁NO₁₆SN_a: 712.2826. Found: 712.2824.

Methyl 2,4,6-Tri-*O*-benzoyl-3-*O*-levulinoyl-1-thio-β-D-galactopyranoside (40). To a stirred solution of methyl 2,6-di-*O*-benzoyl-1-thio-β-D-galactopyranoside²⁷ (740 mg) in 1:1 CH₂Cl₂–pyridine (40 mL) was added dropwise a solution of levulinic anhydride (570 mg, 1.5 equiv) in CH₂Cl₂ (10 mL) at −50°C. The mixture was stirred for 20 min at −50°C and washed successively with 2 M HCl and water, dried (Na₂SO₄), and concentrated. The residue was chromatographed (1:2 EtOAc–hexane) on silica gel to give methyl 2,6-di-*O*-benzoyl-3-*O*-levulinoyl-1-thio-β-D-galactopyranoside (760 mg, 84%): $[\alpha]_D +29^\circ$ (*c* 1, CH₂Cl₂); ¹H NMR (CDCl₃) δ 2.11 (s, 3H, Lev—Me), 2.23 (s, 3H, SMe), 2.43–2.73 (m, 4H, Lev—C₂H₄), 3.08 (d, 1H, *J* = 4.2 Hz, 4-OH), 4.03 (t, 1H, *J*_{5,6} = *J*_{5,6'} = 6.3 Hz, H-5), 4.30 (br t, 1H, H-4), 4.55 (d, 1H, *J*_{1,2} = 9.9 Hz, H-1), 4.62, 4.67 (2dd, 2H, *J*_{5,6} = 6.0, *J*_{5,6'} = 6.6 Hz, H-6,6'), 5.13 (dd, 1H, *J*_{2,3} = 10, *J*_{3,4} = 3.1 Hz, H-3), 5.68 (t, 1H, *J*_{1,2} = *J*_{2,3} = 10 Hz, H-2), 7.40–8.10 (m, 10H, 2Ph). This compound was benzoylated with benzoyl chloride in pyridine to afford the title compound **40**: $[\alpha]_D +50^\circ$ (*c* 0.74, CH₂Cl₂); ¹H NMR (CDCl₃) δ 5.88 (dd, 1H, *J*_{3,4} = 3.3, *J*_{4,5} = 0.73 Hz, H-4).

Anal. Calcd for C₃₃H₃₂O₁₀S (620.68): C, 63.86; H, 5.20. Found: C, 63.85; H, 5.04.

(2,4,6-Tri-*O*-benzoyl-3-*O*-levulinoyl-β-D-galactopyranosyl)-(1→3)-2-*O*-acetyl-4,6-*O*-benzylidene-*N*-benzyloxycarbonyl-1,5-dideoxy-1,5-imino-D-glucitol (41). A mixture of **39** (53 mg), **40** (92 mg, 1.2 equiv), molecular sieves 4Å (200 mg) and CH₂Cl₂ (4 mL) was stirred overnight at rt, then cooled to 0°C. NIS (67 mg, 2.4 equiv) and TMSOTf (6 μL, 0.24 equiv) were added, and the mixture was stirred overnight at 0°C~rt as described for **21**. Work-up and column chromatography (500:1 CH₂Cl₂—MeOH) gave **41** (108 mg, 87%): $[\alpha]_D -14.4^\circ$ (*c* 2.17, CH₂Cl₂); ¹H NMR (CDCl₃) δ 1.69 (s, 3H, AcO), 1.91 (s, 3H, Lev—Me), 2.27–2.62 (m, 4H, Lev—C₂H₄), 3.33 (dd, 1H, *J*_{gem} = 14, *J*_{1ax,2} = 6.4 Hz, H-1aax), 3.42 (m, 1H, *J*_{4,5} = *J*_{5,6} = 10, *J*_{5,6'} = 4.3 Hz, H-5a), 3.78 (dd, 1H, *J*_{gem} = 14, *J*_{1eq,2} = 2.4 Hz, H-1aeq), 4.32, 4.46 (2dd, 2H, *J*_{gem} = 11.3, *J*_{5,6} = 7.6, *J*_{5,6'} = 5.8 Hz, H-6b,6'b), 4.77 (m, 1H, *J*_{2,3} = 6.7 Hz, H-2a), 4.85 (dd, 1H, *J*_{gem} = 11, *J*_{5,6'} = 4.3 Hz, H-6'a), 5.05 (d, 1H, *J*_{1,2} = 7.9 Hz, H-1b), 5.10 (2d, 2H, Z—CH₂), 5.33 (dd, 1H,



$J_{2,3} = 10$, $J_{3,4} = 3.4$ Hz, H-3b), 5.55 (dd, 1H, H-2b), 5.60 (s, 1H, benzylidene CH), 5.78 (d, 1H, $J_{3,4} = 3.4$ Hz, H-4b), 7.20–8.20 (m, 25H, 5Ph).

Anal. Calcd for $C_{55}H_{53}NO_{17}$ (1000.02): C, 66.06; H, 5.34; N, 1.40. Found: C, 65.93; H, 5.11; N, 1.37.

(2,4,6-Tri-*O*-benzoyl-3-*O*-levulinoyl- β -D-galactopyranosyl)-(1 \rightarrow 3)-2-*O*-acetyl-*N*-benzyloxycarbonyl-1,5-dideoxy-1,5-imino-D-glucitol (42) and **(2,4,6-Tri-*O*-benzoyl-3-*O*-levulinoyl- β -D-galactopyranosyl)-(1 \rightarrow 3)-2-*O*-acetyl-*N*-benzyloxycarbonyl-6-*O*-*tert*-butyldimethylsilyl-1,5-dideoxy-1,5-imino-D-glucitol (43)**. A mixture of **41** (323 mg) and 80% aq AcOH (20 mL) was stirred overnight at 45°C, then concentrated. Column chromatography (125:1 CH_2Cl_2 —MeOH) of the residue on silica gel gave **42** (252 mg, 86%): $[\alpha]_D -39^\circ$ (*c* 0.66, CH_2Cl_2); 1H NMR ($CDCl_3$) δ 1.43 (s, 3H, AcO), 1.89 (s, 3H, Lev—Me), 4.67 (m, 1H, $J = 4\sim 5$ Hz, H-2a), 4.88 (d, 1H, $J_{1,2} = 7.9$ Hz, H-1b), 5.02, 5.15 (2d, 2H, $J_{gem} = 12.5$ Hz, Z—CH₂), 5.42 (dd, 1H, $J_{2,3} = 10.6$, $J_{3,4} = 3.1$ Hz, H-3b), 5.61 (dd, 1H, H-2b), 5.83 (d, 1H, H-4b).

To a cooled solution of **42** (213 mg) in CH_2Cl_2 (10 mL) and pyridine (5 mL) was added *tert*-butyldimethylsilyl chloride (176 mg, 3 equiv), and the mixture was stirred overnight at rt. Work-up and column chromatography (250:1 CH_2Cl_2 —MeOH) on silica gel afforded **43** (221 mg, 92%): $[\alpha]_D +34.6^\circ$ (*c* 1.66, CH_2Cl_2); 1H NMR ($CDCl_3$) δ 0.86 (s, 9H, *t*-butyl), 1.27 (s, 3H, AcO), 1.92 (s, 3H, Lev—Me), 2.30–2.60 (m, 4H, Lev—C₂H₄), 3.30 (dd, 1H, $J_{gem} = 12.8$ Hz, H-1a), 4.69 (narrow m, 1H, H-2a), 4.85 (d, 1H, $J_{1,2} = 7.9$ Hz, H-1b), 5.03, 5.16 (2d, 2H, $J_{gem} = 12.3$ Hz, Z—CH₂), 5.40 (dd, 1H, $J_{2,3} = 10.4$, $J_{3,4} = 3$ Hz, H-3b), 5.63 (dd, 1H, H-2), 5.84 (d, 1H, $J_{3,4} = 3$ Hz, H-4b), 7.20–8.20 (m, 20H, 4Ph).

Anal. Calcd for $C_{54}H_{63}NO_{17}Si$ (1026.17): C, 63.21; H, 6.19; N, 1.36. Found: C, 63.17; H, 6.00; N, 1.21.

(2,4,6-Tri-*O*-benzoyl-3-*O*-levulinoyl- β -D-galactopyranosyl)-(1 \rightarrow 3)-[(2,3,4-tri-*O*-benzyl- α -L-fucopyranosyl)-(1 \rightarrow 4)]-2-*O*-acetyl-*N*-benzyloxycarbonyl-6-*O*-*tert*-butyldimethylsilyl-1,5-dideoxy-1,5-imino-D-glucitol (44). To a stirred mixture of **43** (47 mg), methyl 2,3,4-tri-*O*-benzyl-1-thio- β -L-fucopyranoside¹⁴ (26 mg, 1.2 equiv) and molecular sieves 4Å (100 mg) in benzene was added dimethyl(methylthio)sulfonium triflate (DMTST, 63 mg, 4 equiv) at 7°C, and stirring was continued for 1.5 h at 7°C. Work-up and column chromatography (1:2 EtOAc-hexane) gave **44** (36 mg, 55%): $[\alpha]_D -43^\circ$ (*c* 0.5, CH_2Cl_2); 1H NMR ($CDCl_3$) δ 0.87 (s, 9H, *t*-butyl), 1.07 (d, 3H, $J_{5,6} = 6.4$ Hz, H-6c), 1.81 (s, 3H, AcO), 1.91 (s, 3H, Lev—Me), 5.09 (d, 1H, $J_{1,2} = 3.7$ Hz, H-1c), 7.10–8.15 (m, 35H, 7Ph).

Anal. Calcd for $C_{81}H_{91}NO_{21}Si$ (1442.69): C, 67.44; H, 6.36; N, 0.97. Found: C, 67.25; H, 6.19; N, 0.72.

(2,4,6-Tri-*O*-benzoyl-3-*O*-levulinoyl- β -D-galactopyranosyl)-(1 \rightarrow 3)-[(2,3,4-tri-*O*-benzyl- α -L-fucopyranosyl)-(1 \rightarrow 4)]-2-*O*-acetyl-*N*-benzyloxycarbonyl-1,5-dideoxy-1,5-imino-D-glucitol (45) and **(2,4,6-Tri-*O*-benzoyl-3-*O*-lev-**



ulinoyl- β -D-galactopyranosyl)-(1 \rightarrow 3)-[(2,3,4-tri-*O*-benzyl- α -L-fucopyranosyl)-(1 \rightarrow 4)]-2-*O*-acetyl-6-*O*-benzoyl-*N*-benzyloxycarbonyl-1,5-dideoxy-1,5-imino-D-glucitol (46). A mixture of **44** (36 mg) and 80% aq AcOH (10 mL) was stirred for 3 h at rt, then concentrated. Column chromatography (125:1 CH₂Cl₂—MeOH) of the residue on silica gel gave **45** (29 mg, 88%): [α]_D -43° (*c* 1, CH₂Cl₂); ¹H NMR (CDCl₃) δ 1.12 (d, 3H, *J*_{5,6} = 6.2 Hz, H-6c), 1.79 (s, 3H, AcO), 1.93 (s, 3H, Lev—Me), 4.90 (d, 1H, *J*_{gem} = 11.5 Hz, Z—CH₂), 4.93 (d, 1H, *J*_{1,2} = 7.9 Hz, H-1b), 5.38 (dd, 1H, *J*_{2,3} = 10.4, *J*_{3,4} 3 Hz, H-3b), 5.55 (d, 1H, H-2b), 5.83 (d, 1H, H-4b), 7.2–8.2 (m, 35H, 7Ph).

To a solution of **45** (29 mg) in pyridine (10 mL) was added benzoyl chloride (3 μ L, 1.2 equiv) and the mixture was stirred overnight at rt. Work-up and column chromatography (250:1 CH₂Cl₂—MeOH) afforded **46** (31 mg, quant): [α]_D -60° (*c* 2.37, CH₂Cl₂); ¹H NMR (CDCl₃) δ 1.10 (d, 3H, *J*_{5,6} = 6.2 Hz, H-6c), 1.79 (s, 3H, AcO), 1.89 (s, 3H, Lev—Me), 4.93 (d, 1H, *J*_{1,2} = 7.9 Hz, H-1b), 5.40 (d, 1H, H-3b), 5.60 (dd, 1H, *J*_{2,3} = 10.4 Hz, H-2b), 5.82 (d, 1H, *J*_{3,4} = 3.3 Hz, H-4b).

Anal. Calcd for C₈₂H₈₁NO₂₂ (1432.54): C, 68.75; H, 5.70; N, 0.98. Found: C, 68.48; H, 5.42; N, 0.70.

(2,4,6-Tri-*O*-benzoyl-3-*O*-sulfo- β -D-galactopyranosyl)-(1 \rightarrow 3)-[(α -L-fucopyranosyl)-(1 \rightarrow 4)]-2-*O*-acetyl-6-*O*-benzyl-1,5-dideoxy-1,5-imino-*N*-methyl-D-glucitol sodium salt (49) and (3-*O*-sulfo- β -D-galactopyranosyl)-(1 \rightarrow 3)-[(α -L-fucopyranosyl)-(1 \rightarrow 4)]-1,5-dideoxy-1,5-imino-*N*-methyl-D-glucitol sodium salt (50). Treatment of **46** (155 mg) with hydrazine acetate (10 mg) in EtOH, as described for **24**, gave **47** (135 mg, 93%): [α]_D -76° (*c* 2, CH₂Cl₂); ¹H NMR (CDCl₃) δ 1.08 (d, 3H, *J*_{5,6} = 6.4 Hz, H-6c), 1.82 (s, 3H, AcO), 3.23 (bs, 1H, H-1a), 4.87 (d, 1H, *J*_{gem} = 12.1 Hz, CH₂ of Z), 4.92 (d, 1H, *J*_{1,2} = 8.1 Hz, H-1b), 5.08 (bs, 1H, H-1c), 5.40 (dd, 1H, *J*_{2,3} = 10.1 Hz, H-2b), 5.77 (d, 1H, *J*_{3,4} = 3.3 Hz, H-4b). A mixture of **47** (101 mg) and pyridine sulfur trioxide complex (96 mg, 8 equiv) in *N,N*-dimethylformamide (0.5 mL) was stirred for 2 h at rt. Work-up and column chromatography on silica gel, as described for **25**, afforded **48** (109 mg, quant): [α]_D -56° (*c* 2, CH₂Cl₂). Compound **48** (99 mg) in MeOH was treated with cation-exchange resin Dowex-Na⁺, and then hydrogenolyzed in MeOH (5 mL) in the presence of formalin (0.5 mL) and HCl-free palladium black (200 mg) for 4 days at rt. Work-up and column chromatography (10:1 CH₂Cl₂—MeOH) on silica gel gave **49** (66 mg, 92%): [α]_D -30.5° (*c* 0.9, MeOH); ¹H NMR (DMSO-*d*₆, 50°C) δ 1.20 (d, 3H, *J*_{5,6} = 6.4 Hz, H-6c), 1.99 (s, 3H, AcO), 2.05 (t, 1H, *J* = 10.1 Hz, H-1a_{ax}), 2.20 (s, 3H, N—Me), 2.81 (dd, *J*_{gem} = 11, *J*_{1eq,2} = 5.1 Hz, H-1a_{eq}), 4.75 (d, 1H, *J*_{1,2} = 3.7 Hz, H-1c), 4.79 (dd, 1H, H-3b), 5.16 (d, 1H, *J*_{1,2} = 8.2 Hz, H-1b), 5.36 (dd, 1H, *J*_{2,3} = 10.4 Hz, H-2b), 5.86 (d, 1H, *J*_{3,4} = 3.1 Hz, H-4b), 7.44–8.05 (m, 20H, 4Ph).

To a solution of **49** (56 mg) in MeOH (5 mL) was added a small amount of NaOMe (pH = 12), and the mixture was stirred for 2 days at rt. and concentrated. The residue was chromatographed (1:1 MeOH—H₂O) on a column of Sephadex LH-20 to give **50** (31 mg, quant) as an amorphous mass: [α]_D -56° (*c* 0.114, 1:1 MeOH—H₂O); ¹H NMR (DMSO-*d*₆, 50°C) δ 0.99 (d, 3H, H-6c), 2.01 (t, 1H, J



= 10.4 Hz, H-1 $_{aax}$), 2.23 (s, 3H, N—Me), 2.82 (dd, 1H, $J_{gem} = 10.8$, $J_{1eq,2} = 4.8$ Hz, H-1 $_{aeq}$), 4.62 (d, 1H, $J_{1,2} = 7.7$ Hz, H-1b), 4.77 (q, 1H, $J_{5,6} = 6.6$ Hz, H-5c), 4.93 (d, 1H, $J_{1,2} = 3.7$ Hz, H-1c). Ion-spray MS (negative-ion mode) m/z 564.1 (M—Na—H) $^-$; MS/MS ($P = m/z$ 563.9) m/z 417.6 (M—Na—Fuc) $^-$. 96.8 (HSO $_4$) $^-$.

(2,4,6-Tri-*O*-benzoyl-3-*O*-levulinoyl- β -D-galactopyranosyl)-(1 \rightarrow 3)-[(α -L-fucopyranosyl)-(1 \rightarrow 4)]-2-*O*-acetyl-1,5-dideoxy-1,5-imino-D-glucitol (51). Compound **46** (1.34 g) in AcOH (25 mL) was hydrogenolyzed with palladium black (1.30 g) as described for **28** to give **51** (660 mg, 71%): $[\alpha]_D -45^\circ$ (c 1.63, MeOH); 1H NMR (CD $_3$ OD) δ 1.41 (d, 3H, H-6c), 1.82 (s, 3H, AcO), 2.10 (s, 3H, Lev—Me), 2.26–2.54 (m, 4H, Lev—C $_2$ H $_4$), 3.11 (dd, 1H, $J_{gem} = 12.5$ Hz, H-1 $_{aeq}$), 3.64 (t, 1H, $J_{3,4} = 9.7$ Hz, H-3a), 4.56 (m, 1H, $J_{1ax,2} = J_{2,3} = 10.3$, $J_{1eq,2} = 5.3$ Hz, H-2a), 5.05 (d, 1H, $J_{1,2} = 3.9$ Hz, H-1c), 5.28 (d, 1H, $J_{1,2} = 7.7$ Hz, H-1b), 7.43–8.12 (m, 15H, 3Ph).

Anal. Calcd for C $_{46}$ H $_{53}$ NO $_{19}$ (923.92): C, 59.80; H, 5.78; N, 1.52. Found: C, 59.63; H, 5.71; N, 1.41.

(2,4,6-Tri-*O*-benzoyl-3-*O*-levulinoyl- β -D-galactopyranosyl)-(1 \rightarrow 3)-[(α -L-fucopyranosyl)-(1 \rightarrow 4)]-2-*O*-acetyl-1,5-imino-*N*-butyl-D-glucitol (52) and (2,4,6-Tri-*O*-benzoyl-3-*O*-levulinoyl- β -D-galactopyranosyl)-(1 \rightarrow 3)-[(2,3,4-tri-*O*-acetyl- α -L-fucopyranosyl)-(1 \rightarrow 4)]-2,6-di-*O*-acetyl-1,5-dideoxy-1,5-imino-*N*-butyl-D-glucitol (54). A mixture of **51** (67.5 mg), 1-butanol (3 mL) and butyraldehyde (129 μ L, 20 equiv) was hydrogenated with palladium hydroxide on carbon (68 mg) overnight at rt. Work-up and column chromatography (25:1 CH $_2$ Cl $_2$ –MeOH) on silica gel gave **52** (46 mg, 64%): $[\alpha]_D -32.3^\circ$ (c 1.0, CH $_2$ Cl $_2$); 1H NMR (CDCl $_3$) δ 0.85 (t, 3H, Me), 1.15–1.35 (m, 4H, C $_2$ H $_4$), 1.37 (d, 3H, H-6c), 1.91 (s, 3H, AcO), 2.02 (s, 3H, Lev—Me), 2.34–2.60 (m, 4H, Lev—C $_2$ H $_4$), 2.95 (dd, 1H, $J_{gem} = 11.9$, $J_{1eq,2} = 4.2$ Hz, H-1 $_{aeq}$), 5.51 (dd, 1H, $J_{1,2} = 8.1$, $J_{2,3} = 10.4$ Hz, H-2b), 7.30–8.13 (m, 15H, 3Ph). Treatment of **52** (46.7 mg) with Ac $_2$ O (36 μ L, 8 equiv) in pyridine (1 mL) afforded **54**: $[\alpha]_D -53^\circ$ (c 1.0, CH $_2$ Cl $_2$); δ 0.85 (t, 3H, Me), 1.23–1.30 (m, 4H, C $_2$ H $_4$), 1.30 (s, 3H, H-6c), 1.93–2.20 (5s, 15H, 5AcO), 2.06 (s, 3H, Lev—Me), 2.14 (t, 1H, $J_{gem} = J_{1ax,2} = 11.7$ Hz, H-1 $_{aax}$), 2.35–2.63 (m, 4H, Lev—C $_2$ H $_4$), 3.02 (dd, 1H, $J_{gem} = 11.7$, $J_{1eq,2} = 4.8$ Hz, H-1 $_{aeq}$), 5.06 (d, 1H, $J_{1,2} = 3.5$ Hz, H-1c), 5.49 (dd, 1H, $J_{1,2} = 8.1$, $J_{2,3} = 10.4$ Hz, H-2b), 7.40–8.11 (m, 15H, 3Ph).

Anal. Calcd for C $_{58}$ H $_{69}$ NO $_{23}$ (1148.17): C, 60.67; H, 6.06; N, 1.22. Found: C, 60.66; H, 5.83; N, 0.99.

(2,4,6-Tri-*O*-benzoyl-3-*O*-levulinoyl- β -D-galactopyranosyl)-(1 \rightarrow 3)-*O*-[(α -L-fucopyranosyl)-(1 \rightarrow 4)]-2-*O*-acetyl-1,5-dideoxy-1,5-imino-*N*-decyl-D-glucitol (53) and (2,4,6-Tri-*O*-benzoyl-3-*O*-levulinoyl- β -D-galactopyranosyl)-(1 \rightarrow 3)-*O*-[(2,3,4-tri-*O*-acetyl- α -L-fucopyranosyl)-(1 \rightarrow 4)]-2,6-di-*O*-acetyl-1,5-dideoxy-1,5-imino-*N*-decyl-D-glucitol (55). A mixture of **51** (79 mg), EtOAc (1 mL) and decyl aldehyde (320 μ L, 20 equiv) was hydrogenated with palladium hy-



dioxide on carbon (79 mg) for 2 days at rt., and worked up as described for **52** to afford **53** (67.3 mg, 75%): $[\alpha]_D -30.3^\circ$ (*c* 1.36, CH₂Cl₂); ¹H NMR (CDCl₃) δ 0.87 (t, 3H, Me), 1.23 (m, 16H, C₈H₁₆), 1.37 (d, 3H, H-6c), 1.91 (s, 3H, AcO), 2.02 (s, 3H, Lev—Me), 2.34–2.60 (m, 4H, Lev—C₂H₄). Treatment of **53** (66.6 mg) with Ac₂O (47 μ L) and pyridine (1 mL) gave **55** (77 mg): $[\alpha]_D -51^\circ$ (*c* 1.43, CH₂Cl₂); ¹H NMR (CDCl₃) δ 0.88 (t, 3H, Me), 1.24 (m, 16H, C₈H₁₆), 1.31 (s, 3H, H-6c), 1.93–2.21 (5s, 15H, 5AcO), 2.07 (s, 3H, Lev—Me), 2.31–2.63 (m, 4H, Lev—C₂H₄), 3.03 (dd, 1H, $J_{\text{gem}} = 11.7$, $J_{1eq,2} = 5.1$ Hz, H-1aeq), 5.05 (d, 1H, $J_{1,2} = 3.1$ Hz, H-1c), 5.50 (dd, 1H, $J_{1,2} = 8.2$, $J_{2,3} = 10.4$ Hz, H-2b), 7.40–8.11 (m, 15H, 3Ph).

Anal. Calcd for C₆₄H₈₁NO₂₃ (1232.34): C, 62.38; H, 6.63; N, 1.14. Found: C, 62.31; H, 6.62; N, 0.92.

(3-O-Sulfo- β -D-galactopyranosyl)-(1 \rightarrow 3)-[(α -L-fucopyranosyl)-(1 \rightarrow 4)]-1,5-dideoxy-1,5-imino-N-butyl-D-glucitol sodium salt (60). Compound **54** (50.3 mg) was treated with hydrazine acetate (5 mg, 1.2 equiv) in EtOH (1 mL) as described for **24** to give **56** (42 mg, 91%): $[\alpha]_D -64^\circ$ (*c* 0.84, CH₂Cl₂); ¹H NMR (CDCl₃) δ 0.85 (d, 3H, Me), 1.19 (d, 3H, H-6c), 1.18–1.26 (m, 4H, C₂H₄), 1.92–2.18 (5s, 15H, 5AcO), 2.43–2.56 (m, 2H, N—CH₂), 3.05 (dd, 1H, $J_{\text{gem}} = 11.7$ Hz, H-1aeq), 4.12 (dd, 1H, H-3b), 4.53, 4.83 (2dd, 2H, $J_{\text{gem}} = 11.4$, $J_{5,6} = 6.8$, $J_{5,6'} = 5.9$ Hz, H-6b,6'b), 5.08 (d, 1H, $J_{1,2} = 8.4$ Hz, H-1b), 5.77 (d, 1H, $J_{3,4} = 3.7$ Hz, H-4b), 7.41–8.12 (m, 15H, 3Ph), and complete loss of the Lev group. A mixture of **56** (55 mg) and pyridine sulfur trioxide complex (74.6 mg, 10 equiv) in *N,N*-dimethylformamide (2 mL) was stirred for 1.5 h at rt, worked up as described for **48**, and purified on a column of silica gel (20:1 CH₂Cl₂—MeOH) to give **57** (56 mg): $[\alpha]_D -41.3^\circ$ (*c* 0.4, CH₂Cl₂); ¹H NMR (DMSO-*d*₆) δ 0.86 (t, 3H, Me), 1.24 (m, 4H, C₂H₄), 1.99–2.11 (5s, 15H, 5AcO), 5.12 (d, 1H, $J_{1,2} = 9.3$ Hz, H-1b), 5.25 (d, 1H, $J_{1,2} = 3.1$ Hz, H-1c), 5.90 (d, 1H, $J_{3,4} = 3.1$ Hz, H-4b), 7.46–8.13 (m, 15H, 3Ph). *O*-Deacylation of **57** (56 mg) and chromatography on a column of Sephadex LH-20, as described for **50**, afforded **60** (quant) as an amorphous mass: $[\alpha]_D -67^\circ$ (*c* 0.33, 1:1 MeOH—H₂O); ¹H NMR (DMSO-*d*₆) δ 0.86 (t, 3H, Me), 0.99 (d, 3H, H-6c), 1.10–1.40 (m, 4H, C₂H₄), 2.80 (dd, 1H, H-1aeq), 4.60 (d, 1H, $J_{1,2} = 7.7$ Hz, H-1b), 4.90 (d, 1H, $J_{1,2} = 3.5$ Hz, H-1c). FAB-MS (negative ion mode) *m/z* 606.30 (M—Na)[−], (C₂₂H₄₀NO₁₆S: Exact 606.2068).

(3-O-Sulfo- β -D-galactopyranosyl)-(1 \rightarrow 3)-[(α -L-fucopyranosyl)-(1 \rightarrow 4)]-1,5-dideoxy-1,5-imino-N-decyl-D-glucitol sodium salt (61). Treatment of **55** with hydrazine acetate in EtOH, as described for **56**, gave **58**: $[\alpha]_D -58^\circ$ (*c* 1.32, CH₂Cl₂); ¹H NMR (CDCl₃) δ 0.88 (t, 3H, Me), 1.91–1.21 (m, 16H, C₈H₁₆), 1.92–2.18 (5s, 15H, 5AcO), 2.20–2.55 (m, 2H, N—CH₂), 3.06 (dd, 1H, H-1aeq), 4.14 (dd, 1H, H-3b), 4.54, 4.84 (2dd, 2H, $J_{\text{gem}} = 11.5$, $J_{5,6} = J_{5,6'} = 7.0$ Hz, H-6b,6'b), 5.31 (dd, 1H, $J_{1,2} = 8.3$, $J_{2,3} = 3.7$ Hz, H-2b), 5.32 (dd, 1H, $J_{1,2} = 3.5$, $J_{2,3} = 9.0$ Hz, H-2c), 5.76 (d, 1H, $J_{3,4} = 3.7$ Hz, H-4b), and complete loss of the Lev group. Sulfation of **58** (60 mg) was carried out as described for **57** to give **59** (61 mg), which was treated with NaOMe in MeOH to afford **61** (quant) as an amor-



phous mass: ^1H NMR ($\text{DMSO}-d_6$) δ 0.86 (t, 3H, Me), 0.98 (d, 3H, H-6c), 1.25 (m, 16H, C_8H_{16}), 2.15–2.19 (m, 2H, N— CH_2), 2.81 (dd, 1H, $J_{\text{gem}} = 11.4$ Hz, H-1aeq), 4.59 (d, 1H, $J_{1,2} = 7.7$ Hz, H-1b), 4.89 (d, 1H, $J_{1,2} = 3.5$ Hz, H-1c). FAB-MS (negative ion mode) m/z 712.26 ($\text{M}-\text{H}$) $^-$, ($\text{C}_{28}\text{H}_{51}\text{NO}_{16}\text{SNa}$: Exact 712.2826), 690.31 ($\text{M}-\text{Na}$) $^-$ ($\text{C}_{28}\text{H}_{52}\text{NO}_{16}\text{S}$: Exact 690.3007).

ACKNOWLEDGMENTS

This work was supported in part by Grants-in-Aid for Scientific Research from the Ministry of Education, Science, and Culture of Japan. The authors thank Dr. Takao Ikami of Sanwa Kagaku Kenkyusho for FAB-MS measurements, Dr. Akihiro Kondo of Biotechnology Research Laboratories, Takara Shuzo Co., Ltd. for the ion-spray MS analysis, and Drs. Yohji Ezure, Yoshiaki Yoshikuni, and Tadaaki Ohgi of Nippon Shinyaku Co., Ltd. for multiform support including selectin binding assay.

REFERENCES

1. Synthetic Studies on Sialoglycoconjugates, Part 122. For Part 121, see Ando, T.; Ishida, H.; Kiso, M. A highly efficient synthetic route to $\alpha(2\rightarrow3)/\alpha(2\rightarrow6)$ disialyl Lewis x as a cancer associated carbohydrate antigen, *J. Carbohydr. Chem.* **2001**, *20*, in press.
2. Present address: Research Institute, Kagome Co. Ltd., 17 Nishi-Tomiyama, Nishi-Nasuno-machi, Nasu-gun, Tochigi 329–27, Japan.
3. Gunnar, C. H.; David, Z. Biosynthesis of the cancer-associated sialyl-Le^a antigen. *J. Biol. Chem.* **1985**, *260*, 9388–9392.
4. Fukushima, K.; Hirota, M.; Terasaki, P.; Wakisaka, A.; Togashi, H.; Chia, D.; Suyama, N.; Fukushi, Y.; Nudelman, E.; Hakomori, S. Characterization of sialosylated Lewis X as a new tumor-associated antigen. *Cancer Res.* **1984**, *44*, 5279–5285.
5. Varki, A. Selectin ligands. *Proc. Natl. Acad. Sci. USA* **1994**, *91*, 7390–7397.
6. Kannagi, R. Carbohydrate-mediated cell adhesion involved in hematogenous metastasis of cancer. *Glycoconjugate J.* **1997**, *14*, 577–584.
7. Nakamori, S.; Furukawa, H.; Hiratsuka, M.; Iwanaga, T.; Imaoka, S.; Ishikawa, O.; Kabuto, T.; Sasaki, Y.; Kameyama, M.; Ishiguro, S.; Irimura, T. Expression of carbohydrate antigen sialyl Le(a): a new functional prognostic factor in gastric cancer. *J. Clin. Oncol.* **1997**, *15*, 816–825.
8. Sawada, N.; Ito, M.; Ishida, H.; Kiso, M. The first synthesis of glycan parts of lacto-ganglio- and neolactoganglio-series gangliosides. *Tetrahedron Lett.* **2001**, *42*, 1745–1747.
9. Komba, S.; Yamaguchi, M.; Ishida, H.; Kiso, M. 6-*O*-Sulfo de-*N*-acetylsialyl Lewis x as a novel high-affinity ligand for human L-selectin: Total synthesis and structural characterization. *Biol. Chem.* **2001**, *382*, 233–240.
10. Kameyama, A.; Ishida, H.; Kiso, M.; Hasegawa, A. Synthetic Studies on Sialoglycoconjugates 22: Total synthesis of tumor-associated ganglioside, sialyl Lewis X. *J. Carbohydr. Chem.* **1991**, *10*, 549–560.



11. Kameyama, A.; Ishida, H.; Kiso, M.; Hasegawa, A. Synthetic Studies on Sialoglycoconjugates 59: Total synthesis of tumor-associated ganglioside, sialyl-Le^a. *J. Carbohydr. Chem.* **1994**, *13*, 641–654.
12. Hasegawa, A.; Kiso, M. Synthesis of sialyl Lewis X ganglioside and analogs. *Methods Enzymol.* **1994**, *242*, 158–173.
13. Ikami, T.; Ishida, H.; Kiso, M. Synthesis and biological activity of glycolipids, with a focus on gangliosides and sulfatide analogs. *Methods Enzymol.* **2000**, *311*, 547–568.
14. Kiso, M.; Furui, H.; Ando, K.; Ishida, H.; Hasegawa, A. Systematic synthesis of *N*-methyl-1-deoxynojirimycin-containing, Le^x, Le^a, sialyl-Le^x and sialyl-Le^a epitopes recognized by selectins. *Bioorg. Med. Chem.* **1994**, *2*, 1295–1308.
15. Kiso, M.; Furui, H.; Ishida, H.; Hasegawa, A. Studies on selectin binding inhibitors: Synthesis of sialyl-Lewis X and sialyl-Lewis A epitope analogs containing 2-acetamido derivative of *N*-methyl-1-deoxynojirimycin. *J. Carbohydr. Chem.* **1996**, *15*, 1–14.
16. Ogawa, H.; Harada, Y.; Kyotani, Y.; Ueda, T.; Kitazawa, S.; Kandori, K.; Seto, T.; Ishiyama, K.; Kojima, M.; Ohgi, T.; Ezure, Y.; Kise, M. An efficient synthesis of sulfo Lewis x analog containing 1-deoxynojirimycin. *J. Carbohydr. Chem.* **1998**, *17*, 729–738.
17. Hasegawa, A.; Kiso, M. Design and synthesis of cell adhesion carbohydrate ligands/inhibitors. In *Carbohydrates in Drug Design*; Witczak, Z. J.; Nieforth, K. A. Eds.; Marcel Dekker, Inc.; New York, 1997; 137–155.
18. Brandley, B. K.; Kiso, M.; Abbas, S.; Nikrad, P.; Srivasatava, O.; Foxall, C.; Oda, Y.; Hasegawa, A. Structure-function studies on selectin carbohydrate ligands. Modification to fucose, sialic acid and sulphate as a sialic acid replacement. *Glycobiology* **1993**, *3*, 633–639.
19. Yuen, C.-T.; Bezouska, K.; O'Brien, J.; Stoll, M.; Lemonie, R.; Lubineau, A.; Kiso, M.; Hasegawa, A.; Bockovich, J.; Nicolaou, K. C.; Feizi, T. Sulfated blood group Lewis^a, a superior oligosaccharide ligand for human E-selectin. *J. Biol. Chem.* **1994**, *269*, 1595–1598.
20. Yoshida, M.; Suzuki, T.; Ishida, H.; Kiso, M.; Hasegawa, A. Synthetic studies on sialoglycoconjugates 81: Synthesis of positional isomers of sialyl Lewis X epitope containing 1-deoxy-D-glucose in place of *N*-acetylglucosamine, and their inhibitory activity to selectin-mediated adhesion. *J. Carbohydr. Chem.* **1996**, *15*(2), 147–162.
21. Tanahashi, E.; Murase, K.; Shibuya, M.; Igarashi, Y.; Ishida, H.; Hasegawa, A.; Kiso, M. Synthetic studies on selectin ligands/inhibitors: A systematic synthesis of sulfatide and its higher congeners carrying 2-(tetradecyl)hexadecyl group as a ceramide substitute. *J. Carbohydr. Chem.* **1997**, *16*(6), 831–858.
22. Kiso, M.; Katagiri, H.; Furui, H.; Hasegawa, A. Studies on 1-deoxynojirimycin-containing glycans: Synthesis of novel disaccharides related to lactose, lactosamine, and chitobiose. *J. Carbohydr. Chem.* **1992**, *11*, 627–644.
23. Kiso, M.; Ando, K.; Inagaki, H.; Ishida, H.; Hasegawa, A. Synthetic and structural studies of α -sialyl-(2→6) and α -sialyl-(2→3) 1-deoxynojirimycin derivatives potentially useful for biomedical applications. *Carbohydr. Res.* **1995**, *272*, 159–178.
24. Simanek, E. E.; McGarver, G. J.; Jablonowski, J. A.; Wong, C.-H. Selectin-carbohydrate interactions: from natural ligands to designed mimics. *Chem. Rev.* **1998**, *98*, 833–862.
25. Wada, Y.; Saito, T.; Matsuda, N.; Ohmoto, H.; Yoshino, K.; Ohashi, M.; Kondo, H.; Ishida, H.; Kiso, M.; Hasegawa, A. Studies on selectin blockers. 2. Novel selectin



- blocker as potential therapeutics for inflammatory disorders. *J. Med. Chem.* **1996**, *39*, 2055–2059.
26. Tsukida, T.; Moriyama, H.; Kurokawa, K.; Achiha, T.; Inoue, Y.; Kondo, H. Studies on selectin blockers. 7. Structure-activity relationships of sialyl Lewis x mimetics based on modified Ser-Glu dipeptides. *J. Med. Chem.* **1998**, *41*, 4279–4287, and references therein.
 27. Garegg, P. J.; Oscarson, S. A synthesis of 8-methoxycarbonyloct-1-yl *O*- α -D-galactopyranosyl-(1 \rightarrow 3)-*O*- β -D-galactopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranoside. *Carbohydr. Res.* **1985**, *136*, 207–213.

Received May 28, 2001

Accepted August 23, 2001



Request Permission or Order Reprints Instantly!

Interested in copying and sharing this article? In most cases, U.S. Copyright Law requires that you get permission from the article's rightsholder before using copyrighted content.

All information and materials found in this article, including but not limited to text, trademarks, patents, logos, graphics and images (the "Materials"), are the copyrighted works and other forms of intellectual property of Marcel Dekker, Inc., or its licensors. All rights not expressly granted are reserved.

Get permission to lawfully reproduce and distribute the Materials or order reprints quickly and painlessly. Simply click on the "Request Permission/Reprints Here" link below and follow the instructions. Visit the [U.S. Copyright Office](#) for information on Fair Use limitations of U.S. copyright law. Please refer to The Association of American Publishers' (AAP) website for guidelines on [Fair Use in the Classroom](#).

The Materials are for your personal use only and cannot be reformatted, reposted, resold or distributed by electronic means or otherwise without permission from Marcel Dekker, Inc. Marcel Dekker, Inc. grants you the limited right to display the Materials only on your personal computer or personal wireless device, and to copy and download single copies of such Materials provided that any copyright, trademark or other notice appearing on such Materials is also retained by, displayed, copied or downloaded as part of the Materials and is not removed or obscured, and provided you do not edit, modify, alter or enhance the Materials. Please refer to our [Website User Agreement](#) for more details.

[Order now!](#)

Reprints of this article can also be ordered at

<http://www.dekker.com/servlet/product/DOI/101081CAR100108657>