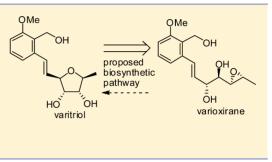
Total Synthesis of Varitriol, Varioxirane, and Enantiomer of the Proposed Biosynthetic Precursor

Gangarajula Sudhakar* and Jakka Raghavaiah

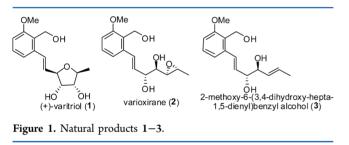
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S Supporting Information

ABSTRACT: The first stereoselective total synthesis of varioxirane was accomplished, and the proposed biosynthetic pathway was supported by converting varioxirane to (+)-varitriol. The first total synthesis of enantiomer of the proposed biosynthetic precursor, (1E,3S,4R,5E)-1-(2-(hydroxymethyl))-3-methoxyphenyl)hepta-1,5-diene-3,4-diol, was also achieved by utilizing the unreacted allylic alcohol obtained during the Sharpless kinetic resolution step. Other key steps include the Horner–Wadsworth–Emmons reaction and the diastereoselective reduction of α,β -unsaturated ketone to its corresponding alcohol.



Varitriol (1) and varioxirane (2) (Figure 1) were isolated from a marine-derived strain of the fungus *Emericella*



variecolor in 2002.¹ Varitriol became a very attractive target for several synthetic organic chemistry groups after its isolation. To date, as many as 15 total syntheses² (among them, two are unnatural varitriol (ent-1),³ one formal synthesis⁴ and two analogues synthesis⁵) have been reported. This is because of the simple structure associated with high levels of biological activity toward renal, CNS, and breast cancer cell lines in the tested NCI's 60-cell panel.^{1,6} Most of the syntheses relied on a chiral pool starting material such as D-ribose, ${}^{2a,b,f-h,3}\gamma$ -D-ribonolactone, 2d,i D-mannitol, 2k dimethyl L-tartrate, 2l and methyl α -D-mannopyranoside.^{2m} Although several synthetic approaches are available for 1, neither the synthesis nor the absolute configuration was reported for varioxirane (2). However, Barrero and co-workers¹ proposed the relative configuration of 2 based on a hypothetical biogenetic relationship between 1 and 2. The same group also proposed that the biosynthesis of 1 might be from 2-methoxy-6-(3,4dihydroxyhepta-1,5- dienyl)benzyl alcohol (3),7 which is a common metabolite of Aspergillus variecolor (the imperfect state of *E. variecolor*), via varioxirane (2).¹ The biosynthetic hypothesis is that enzymatic epoxidation of the corresponding double bond of 3 may yield 2 and intramolecular $S_N 2$ epoxide opening of 2 would yield 1.¹

As part of our interest in developing synthetic strategies for bioactive natural products based on plausible biosynthetic pathways,⁸ we envisaged that the total synthesis of varioxirane (2) would not only provide a new synthetic route to varitriol (1) but also provide support to the proposed biosynthetic pathway. Herein we report the short and efficient synthesis of varitriol (1) from varioxirane (2) and enantiomer of 3 (*ent-3*) by a developing new synthetic strategy that does not rely on chiral pool starting materials.

We anticipated that the functionality embedded frameworks like 4 (Figure 2) can easily provide varioxirane (2), whereas frameworks like 5 can provide 3 by simple transformations such as stereoselective reduction of α,β -unsaturated ketone and global deprotection of protecting groups. Enones 4 and 5 could be derived by reacting aromatic aldehyde 6 with β ketophosphonates 7 and 8, respectively, under Horner– Wadsworth–Emmons reaction conditions. These chiral β ketophosphonates 7 and 8 could easily be obtained from a common starting material, α -hydroxy ester (±)-9 in three steps such as the Sharpless asymmetric epoxidation/kinetic resolution, masking the free secondary hydroxyl group, and introducing phosphonate.

The synthesis began from the Sharpless kinetic resolution⁹ of allylic alcohol (\pm) -9¹⁰ using of (-)-DIPT, Ti(ⁱOPr)₄, and TBHP in CH₂Cl₂ to give the epoxy alcohol (+)-10¹¹ (er 98.89:1.11) in 42% yield (Scheme 1). Silylation of the secondary hydroxyl in 10 furnished 11, which was treated with lithiated methyl phosphonate to afford β -ketophosphonate¹² 7 (R = TBS) in 89% yield (two steps). Coupling of ketophosphonate 7 under optimized reaction conditions (KHMDS, THF, -78 °C to rt) with the aromatic aldehyde 6 (R₁ = Ac), prepared following the literature procedure,¹³

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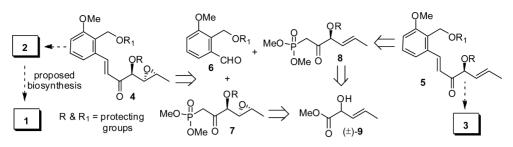
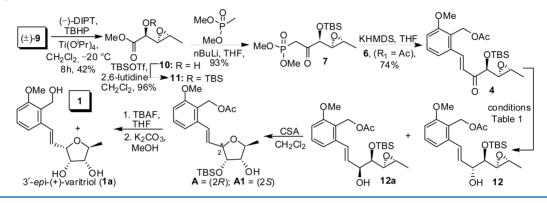


Figure 2. Synthetic plan for 1-3 and retrosynthetic plan for 4 and 5.

Scheme 1. Synthesis of 12, 12a and 1, 1a



afforded the $\alpha_{,\beta}$ -unsaturated ketone **4** (R = TBS, R₁ = Ac) in 74% yield.

We were now ready to investigate the key 1,2-reduction of ketone 4 to install the desired hydroxyl chiral center. Efforts to increase the *anti* selectivity under various reducing reagents yielded the undesired *syn* isomer as a major or exclusive product (Table 1). Reduction with $Zn(BH_4)_2$ at -20 °C¹⁴ as

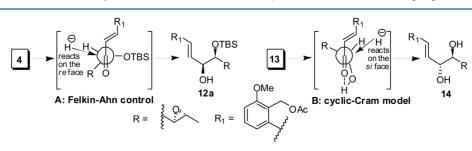
Table 1. Screening Reducing Agents for the Anti-SelectiveReduction of 4 To Give 12

entry	conditions	yield ^{a} (%)	12:12a ^b		
1	$Zn(BH_4)_2$, THF, -20 °C	87	1:2		
2	$Zn(BH_4)_2$, THF, 0 °C	85	1:3.1		
3	LiEt₃BH, THF, −78 °C	76	1:2.3		
4	LiEt ₃ BH, THF, 0 °C	61	1:1		
5	K-Selectride, THF, -78 °C	84	1:1.6		
6	DIBAL-H, CH ₂ Cl ₂ , –78 °C	80	1:2		
7	NaBH ₄ /CeCl ₃ , MeOH, 0 °C	82	only 12a		
8	NaBH ₄ /CeCl ₃ , MeOH, –50 °C	90	only 12a		
^{<i>a</i>} Isolated yields. ^{<i>b</i>} The ratio was determined by ¹ H NMR analysis					

well as at 0 °C resulted in undesired isomer 12a as a major product (entries 1 and 2). Conversion of the chromatographically inseparable 12 and 12a to separable A and A1 (ratio of 1:2 and 92% combined yield) and subsequent deprotections of silyl and acetyl yielded **1** and 3'-epi-(+)-varitriol (**1a**), respectively. Identities of the major and minor products were determined by comparison of **1** with the literature.^{1–3} A similar result was obtained with LiEt₃BH at $-78 \,^{\circ}C^{15}$ (1:2.3, entry 3), whereas at 0 $\,^{\circ}C$ there was no selectivity and only moderate yield (entry 4). K-Selectride and DIBAL-H¹⁶ could not improve the desired selectivity (entries 5 and 6). Reduction with NaBH₄/CeCl₃ in MeOH¹⁷ at 0 $\,^{\circ}C$ as well as at $-50 \,^{\circ}C$ gave exclusively undesired isomer **12a** (entries 7 and 8).

We then reasoned that the reduction of 4 with various reducing agents yielded *syn* isomer 12a as a major product or exclusively because of the Felkin–Ahn control where a dihedral angle of ~90° between the OTBS and carbonyl groups maximizes stereoelectronic interactions in the transition state (Figure 3A). It was expected that freeing a secondary hydroxyl group within 4 to 13 (scheme 2) might induce the stereoselectivity by the cyclic-Cram model (Figure 3B),¹⁸ in which a proton of the hydroxyl group is hydrogen bonded to the carbonyl oxygen, enforcing a syn-periplanar relationship between the hydroxyl and carbonyl groups and leading to the desired *anti* isomer 14.

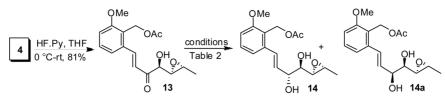
Accordingly, a silyl group of 4 was removed by using HFpyridine to give the epoxy alcohol 13 (Scheme 2), which was subjected to various reducing agents, and the results are



Note

Figure 3. Felkin-Ahn and cyclic-Cram modes of stereocontrol.

Scheme 2. Synthesis of 14 and 14a



illustrated in Table 2. In contrast to 4 (Table 1, entries 7 and 8), a reduction of 13 with $NaBH_4/CeCl_3$ in MeOH at 0 °C as

 Table 2. Screening Reducing Agents for the Anti-Selective

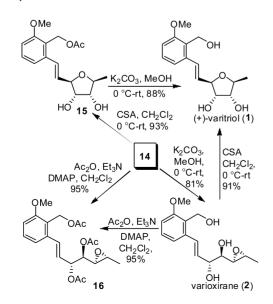
 Reduction of 13 To Give 14

entry	conditions	yield ^{a} (%)	14:14a ^b	
1	NaBH ₄ /CeCl ₃ , MeOH, 0 °C	77	1.1:1	
2	NaBH ₄ /CeCl ₃ , MeOH, -50 °C	86	1.1:1	
3	$Zn(BH_4)_2$, THF, 0 °C	76	1.3:1	
4	$Zn(BH_4)_2$, THF, -78 °C	80	1:1	
5	K-Selectride, -78 °C	70	1.2:1	
6	LiEt ₃ BH, THF, –78 °C	73	11.5:1	
7	LiEt ₃ BH, ether, –78 °C	82	32.3:1	
^{<i>a</i>} Isolated yield. ^{<i>b</i>} The ratio was determined by ¹ H NMR analysis.				

well as at -50 °C found no selectivity (Table 2, entries 1 and 2). With Zn(BH₄)₂ in THF at 0 °C, the desired *anti* diol 14 was slightly improved (1.3:1, entry 3); however, at -78 °C with the same reagent the selectivity was completely lost (entry 4). With K-Selectride the level of selectivity was also poor (1.2:1, entry 5). Gratifyingly, the excellent selectivity of *anti* diol 14 (11.5:1, entry 6) was obtained with LiEt₃BH in THF at -78 °C, and when the solvent was changed from THF to ether the selectivity and yield were further increased (32.3:1, entry 7).

With the desired *anti* diol 14 in hand, to accomplish the target molecules two steps were needed to obtain varitriol (1) and one step to obtain varioxirane (2). Treatment of 14 with camphorsulfonic acid (CSA) in CH_2Cl_2 at 0 °C to rt effected intramolecular epoxide opening, as expected, to provide tetrahydrofuran moiety 15 in excellent yield (Scheme 3).

Scheme 3. Synthesis of Varitriol (1), Varioxirane (2), and Peracetylated Derivative of Varioxirane 16

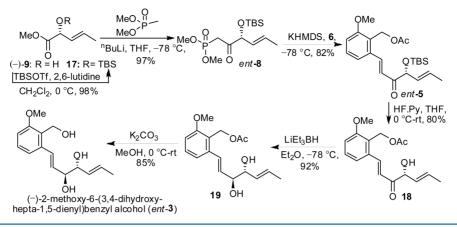


Finally, deacetylation with K₂CO₃ in MeOH furnished varitriol (1) in 88% yield. The spectral data and specific rotation ($[\alpha]^{30}_{D}$ = +23.8 (*c* 0.26, CH₃OH); lit.¹ $[\alpha]^{25}_{D}$ = +18.5 (*c* 2.30, CH₃OH)) of 1 agree well with the literature data.¹⁻³

We then turned our attention toward the total synthesis of varioxirane and converting it to varitriol to support the proposed biosynthetic pathway. The direct deacetylation of **14** provided varioxirane (**2**) in 81% yield. Treatment of **2** with Ac₂O (4 equiv) and Et₃N (6 equiv) furnished the peracetylated derivative of varioxirane **16**, which was also alternatively obtained from **14** using Ac₂O (2.5 equiv) and Et₃N (4 equiv). Spectral data and specific rotation ($[\alpha]^{30}_{D} = -32.9 (c 0.19, CHCl_3)$; lit.¹ $[\alpha]^{25}_{D} = -28.0 (c 0.31, CHCl_3)$) of peracetylated derivative of varioxirane **16** agrees well with the data reported.¹ Upon treatment of varioxirane (**2**) with CSA in CH₂Cl₂ produced varitriol (**1**), which agrees with the proposed biosynthetic pathway.¹

Compound (+)-9 was needed for the synthesis of natural 2methoxy-6-(3,4-dihydroxyhepta-1,5-dienyl)benzyl alcohol (3). Compound (+)-9 could be derived from racemic (\pm) -9 by using (+)-DIPT in the Sharpless kinetic resolution step as shown in the retrosynthetic analysis (Figure 2). However, the enantiomer of 3 could be derived from the unreacted allylic alcohol (-)-9 obtained in the Sharpless kinetic resolution step of Scheme 1. Thus, compound (-)-9¹¹ (er 98.6:1.4, HPLC)¹⁹ was silvlated by using TBSOTf to give 17, which was treated with lithiated methyl phosphonate to give β -ketophosphonate ent-8 (R = TBS) in 95% yield over two steps (Scheme 4). Under HWE reaction conditions, ent-8 was coupled with aldehyde 6 to yield the conjugated ketone ent-5 in 82% yield. Desilylation of ent-5 furnished 18 under the optimized conditions (LiEt₃BH, Et₂O, -78 °C) shown in Table 2 and provided anti-diol 19 as a major product (32:1) in favor of the desired isomer with 92% yield. Acetyl deprotection of 19 provided the anticipated ent-3 in 85% yield. The spectral data and specific rotation of (-)-3 ($[\alpha]_{D}^{30} = -36.0$ (*c* 0.05, CHCl₃); natural product (+)-3:⁷ $[\alpha]^{24}_{D}$ = +42.7 (*c* 0.22, CHCl₃)) are in good agreement with the data reported.

In conclusion, a new synthetic route has been developed for varitriol based on the proposed biosynthetic pathway. En route to varitriol (1), the first total synthesis of varioxirane (2) was achieved. Additionally, the first total synthesis of an enantiomer of the proposed biosynthetic precursor, (1E,3S,4R,5E)-1-(2-(hydroxymethyl)-3-methoxyphenyl)hepta-1,5-diene-3,4-diol (*ent-3*), was also accomplished utilizing the same strategy and starting material. The first total synthesis reported herein for 2 and *ent-3* confirmed the absolute configuration of natural varioxirane and (1E,3R,4S,5E)-1-(2-(hydroxymethyl)-3-methoxyphenyl)hepta-1,5-diene-3,4-diol. Application of this strategy to other bioactive molecules and their analogues, in particular compounds related to 3, is in progress in our laboratory, and it will be reported in due course.



EXPERIMENTAL SECTION

(*E*)-Methyl 2-Hydroxypent-3-enoate ((±)-9).¹⁰ Compound (*E*)-2-hydroxypent-3-enenitrile (3.535 g, 30%) was prepared as a colorless oil from (*E*)-but-2-enal (10.0 mL, 8.51 g, 121.5 mmol):^{10a} R_f = 0.38 (20% EtOAc/hexanes); ¹H NMR (300 MHz, CDCl₃) δ 6.03 (m, 1H), 5.58 (m, 1H), 4.90 (t, *J* = 6.0 Hz, 1H), 4.0 (br, 1H), 1.76 (d, *J* = 6.8 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 132.5, 124.7, 118.5, 61.3, 17.2; IR (neat) ν_{max} 3421, 2922, 2248, 1670, 1445, 1262, 1085 cm⁻¹; HRMS (EI) m/z [M]⁺ calcd for C₅H₇N₁O₁ 97.0527, found 97.0527.

(*E*)-Methyl 2-hydroxypent-3-enoate ((±)-9) (4.3 g, 92%) was prepared from the above cyanohydrin (3.5 g, 36 mmol) as a colorless oil: ^{10b} $R_f = 0.3$ (20% EtOAc/hexanes); ¹H NMR (300 MHz, CDCl₃) δ 5.85 (m, 1H), 5.49 (m, 1H), 4.56 (bs, 1H), 3.75 (s, 1H), 3.12 (bs, 1H), 1.69 (d, J = 6.8 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 173.9, 129.4, 127.1, 71.2, 52.4, 17.4; IR (neat): ν_{max} 3450, 2922, 1737, 1440, 1214, 1139, 1049, 966 cm⁻¹; HRMS (ESI) m/z [M + H]⁺ calcd for C₆H₁₁O₃ 131.0703, found 131.0702.

(S)-Methyl 2-Hydroxy-2-((2R,3R)-3-methyloxiran-2-yl)acetate (10). To a suspension of activated powdered 4A molecular sieves (1.3 g) in CH₂Cl₂ (70 mL) were added sequentially Ti(OⁱPr)₄ (4.92 mL, 16.5 mmol) and (-)-DIPT (4.08 mL, 19.8 mmol) at -20 °C. After the mixture was stirred for 30 min, TBHP (12.2 mL, 2.7 M in toluene, 33 mmol) was added and stirring continued for another 30 min at the same temperature. To the above solution was added compound (\pm)-9 (4.3 g, 33 mmol) in CH₂Cl₂ (16 mL) and the resulting solution stirred for 8 h at -20 °C. The reaction mixture was quenched with water, warmed to rt, stirred for overnight, filtered, and concentrated under reduced pressure. The compound was extracted with EtOAc (2 × 200 mL), washed with brine, and dried over Na_2SO_4 . The residue was purified by column chromatography (20% EtOAc/ hexanes) to afford 10 (2.02 g, 42%) as a colorless oil: $R_f = 0.3$ (30% EtOAc/hexanes); $[\alpha]^{28}_{D} = +45.8$ (c 0.27, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 4.24 (t, J = 4.3 Hz, 1H), 3.82 (s, 3H), 3.11 (dq, J = 5.3, 2.0 Hz, 1H), 3.04 (d, J = 4.3 Hz, 1H), 2.97 (dd, J = 5.3, 2.0 Hz, 1H), 1.31 (d, J = 5.3 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 172.1, 69.5, 58.7, 52.6, 51.5, 16.8; IR (neat) $\nu_{\rm max}$ 3448, 2854, 1726, 1464, 1253, 1135, 1098, 1024, 835 cm⁻¹; HRMS (ESI) m/z [M + H]⁺ calcd for C₆H₁₁O₄ 147.0651, found 147.0651.

(S)-Methyl 2-(*tert*-Butyldimethylsilyloxy)-2-((25,3*R*)-3-methyloxiran-2-yl)acetate (11). Compound 11 (3.42 g, 96%) was synthesized as a colorless oil from alcohol 10 (2 g, 13.7 mmol) following the general procedure for TBS protection described in ref 8a: $R_f = 0.6$ (10% EtOAc/hexanes); $[\alpha]^{28}_{D} = +3.59$ (*c* 0.67, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 4.27 (d, J = 3.7 Hz, 1H), 3.77 (s, 3H), 3.08 (qd, J = 5.3, 2.2 Hz, 1H), 3.01 (dd, J = 3.7, 2.2 Hz, 1H), 1.30 (d, J = 5.3 Hz, 3H), 0.89 (s, 9H), 0.08 (s, 3H), 0.06 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 171.4, 70.9, 59.1, 52.1, 50.9, 25.5, 18.2, 16.8, -5.3, -5.4; IR (neat) ν_{max} 2956, 2932, 2858, 1760, 1468, 1253, 1162, 839 cm⁻¹; HRMS (ESI) m/z [M + H]⁺ calcd for C₁₂H₂₅O₄Si 261.1516, found 261.1516.

2-Acetoxymethyl-3-methoxybenzaldehyde (6):¹³ $R_f = 0.4$ (20% EtOAc/hexanes); ¹H NMR (300 MHz, CDCl₃) δ 10.23 (s, 1H), 7.46 (m, 2H), 7.14 (m, 1H), 5.51 (s, 2H), 3.86 (s, 3H), 2.01 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 191.7, 170.6, 158.3, 135.8, 130.1, 124.8, 122.7, 116.1, 56.0, 55.7, 20.7; IR (neat) ν_{max} 2922, 2851, 1730, 1691, 1586, 1468, 1362, 1223, 1023, 965 cm⁻¹; HRMS (ESI) m/z [M + Na]⁺ calcd for C₁₁H₁₂O₄Na 231.0625, found 231.0627.

Dimethyl (*S*)-3-(*tert*-Butyldimethylsilyloxy)-3-((*2S*,*3R*)-3methyloxiran-2-yl)-2-oxopropylphosphonate (7). Compound 7 (4.28 g, 93%) was synthesized from ester 11 (3.4 g, 13.1 mmol) as a highly viscous colorless oil following the procedure reported for βketophosphonate in ref 12: $R_f = 0.3$ (80% EtOAc/hexanes); $[\alpha]^{28}_{D} =$ +S3.5 (*c* 0.71, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 4.18 (d, J =4.2 Hz, 1H), 3.79 (s, 3H), 3.76 (s, 3H), 3.45 (dd, J = 21.8, 15.1 Hz, 1H), 3.18 (dd, J = 21.8, 15.1 Hz, 1H), 3.0 (qd, J = 5.0, 2.5 Hz, 1H), 2.83 (dd, J = 4.2, 2.5 Hz, 1H), 1.30 (d, J = 5.0 Hz, 1H), 0.90 (s, 9H), 0.09 (s, 3H), 0.08 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 202.0, 77.5, 77.5, 58.7, 52.9, 52.8, 51.8, 36.7, 35.3, 25.6, 18.2, 16.9, -4.9, -5.1; IR (neat) ν_{max} 2855, 1727, 1465, 1254, 1153, 1029, 946, 839 cm⁻¹; HRMS (ESI) *m*/*z* [M + H]⁺ calcd for C₁₄H₃₀O₆PSi 353.1541, found 353.1543.

2-((S,E)-4-(tert-Butyldimethylsilyloxy)-4-((2S,3R)-3-methyloxiran-2-yl)-3-oxobut-1-enyl)-6-methoxybenzyl Acetate (4). β -Ketophosphonate 7 (3.4 g, 9.65 mmol) was dissolved in THF (50 mL) and cooled to -78 °C. KHMDS (19.3 mL, 0.5 M in THF, 9.65 mmol) was added slowly. After 30 min at -78 °C, aldehyde 6 (3.01 g, 14.48 mmol) which dissolved in THF (30 mL) was added. After 1 h, the reaction mixture was warmed to rt and stirred for 24 h, the reaction was quenched by addition of aq NH4Cl solution, and the product was extracted with EtOAc. The organic layer was washed with brine, dried over MgSO₄, and concentrated in vacuo. The reaction mixture was purified by column chromatography on silica gel (8% EtOAc in hexanes) to give 4 (3.1 g, 74%) as a colorless oil: $R_f = 0.5$ $(20\% \text{ EtOAc/hexanes}); [\alpha]^{28}_{D} = -4.6 (c \ 0.86, \text{ CHCl}_3); {}^{1}\text{H NMR} (300)$ MHz, CDCl₃) δ 8.03 (d, J = 16.0 Hz, 1H), 7.37 (t, J = 7.9 Hz, 1H), 7.28 (d, J = 7.9 Hz, 1H), 7.18 (d, J = 16.0 Hz, 1H), 6.98 (d, J = 7.9 Hz, 1H), 5.33 (s, 2H), 4.36 (d, J = 3.4 Hz, 1H), 3.86 (s, 3H), 3.08 (qd, J = 5.3, 2.2 Hz, 1H), 2.95 (dd, J = 3.4, 2.2 Hz, 1H), 2.06 (s, 3H), 1.31 (d, J = 5.3 Hz, 3H), 0.93 (s, 9H), 0.12 (s, 3H), 0.10 (s, 3H); 13 C NMR (125 MHz, CDCl₃) δ 198.2, 170.5, 158.3, 140.7, 136.3, 129.9, 123.6, 123.5, 118.7, 112.3, 76.7, 59.3, 56.9, 55.7, 50.8, 25.5, 20.7, 18.1, 16.8, –5.1, –5.2; IR (neat) $\nu_{\rm max}$ 2925, 2853, 1737, 1690, 1609, 1576, 1470, 1379, 1361, 1258, 1224, 1130, 1097, 1023, 951, 838, 673 cm⁻¹; HRMS (ESI) $m/z [M + H]^+$ calcd for C₂₃H₃₅O₆Si 435.2198, found 435.2197.

2-((3*S*,4*R*,*E*)-4-(*tert*-Butyldimethylsilyloxy)-3-hydroxy-4-((2*S*,3*R*)-3-methyloxiran-2-yl)but-1-enyl)-6-methoxybenzyl acetate (12a). Compound 12a (45 mg, 90%) was synthesized as a colorless oil from compound 4 (50 mg, 0.115 mmol) following the procedure described in ref 17: R_f = 0.3 (30% EtOAc/hexanes); [α]²⁸_D = +21.4 (*c* 0.28, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.29 (t, *J* = 8.3 Hz, 1H), 7.10 (d, *J* = 8.3 Hz, 1H), 6.95 (d, *J* = 15.8 Hz, 1H), 6.83 (d, *J* = 8.3 Hz, 1H), 6.19 (dd, *J* = 15.8,5.7 Hz, 1H), 5.27 (s, 2H), 4.36 (ddd, *J* = 5.7, 3.8, 1.5 Hz, 1H), 3.83 (s, 3H), 3.51 (dd, *J* = 5.8, 3.8 Hz, 1H), 2.96 (dd, *J* = 5.3, 2.1 Hz, 1H), 2.82 (dd, *J* = 5.8, 2.1 Hz, 1H), 2.06 (s, 3H), 1.33 (d, *J* = 5.3 Hz, 3H), 0.90 (s, 9H), 0.09 (s, 3H), 0.06 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 171.1, 158.4, 138.9, 132.5, 129.8, 127.9, 121.1, 118.5, 109.8, 74.8, 73.8, 59.1, 57.6, 55.7, 53.3, 25.7, 21.0, 18.1, 17.2, -4.4, -4.8; IR (neat) ν_{max} 3451, 2924, 2853, 1734, 1579, 1470, 1380, 1360, 1251, 1100, 1023, 836 cm⁻¹; HRMS (ESI) *m*/*z* [M + Na]⁺ calcd for C₂₃H₃₆O₆NaSi 459.2157, found 459.2173.

Synthesis of A and A1. Column-separable **A** and **A1** (1:2) (110 mg, 92%) were prepared from the mixture **12** and **12a** (120 mg, 0.275 mmol) following the same procedure described in ref 8a.

Data of **A** (minor): $R_f = 0.3$ (30% EtOAc/hexanes); $[\alpha]^{26}_{D} = +25.26$ (c 0.77, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.30 (t, J = 7.9 Hz, 1H), 7.13 (d, J = 7.9 Hz, 1H), 6.94 (d, J = 15.7 Hz, 1H), 6.84 (d, J = 7.9 Hz, 1H), 6.09 (dd, J = 15.7, 8.0 Hz, 1H), 5.25 (s, 2H), 4.29 (m, 1H), 3.97 (t, J = 5.5 Hz, 1H), 3.89 (t, J = 5.5 Hz, 1H), 3.84 (s, 3H), 3.63 (m, 1H), 2.67 (d, J = 5.5 Hz, 1H), 2.05 (s, 3H), 1.34 (d, J = 6.4 Hz, 3H), 0.92 (s, 9H), 0.11 (s, 3H), 0.10 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 171.0, 158.4, 138.5, 130.8, 129.9, 129.6, 118.6, 110.0, 84.5, 79.8, 76.7, 76.2, 57.6, 55.8, 25.7, 21.0, 18.9, 18.0, -4.3, -4.9; IR (neat) ν_{max} 3450.9, 2925.9, 2854.4, 1733.3, 1579.9, 1467.7, 1381.6, 1253.9, 1103.4, 1023.0, 964.5, 774.1 cm⁻¹; HRMS (ESI) calcd for C₂₃H₃₆O₆NaSi [M + Na]⁺ 439.2175, found 439.2173.

Data of **A1** (major): $[\alpha]^{28}_{D} = +38.02$ (c 0.40, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.28 (t, J = 7.9 Hz, 1H), 7.17 (d, J = 7.9 Hz, 1H), 6.83 (d, J = 7.9 Hz, 1H), 6.81 (d, J = 15.8 Hz, 1H), 6.28 (dd, J =15.8, 8.0 Hz, 1H), 5.26 (ABq, J = 16.3, 11.6 Hz, 2H), 4.62 (m, 1H), 4.30 (t, J = 5.0 Hz, 1H), 3.98 (m, 1H), 3.84 (s, 3H), 3.74 (m, 1H), 2.34 (d, J = 8.3 Hz, 1H), 2.06 (s, 3H), 1.32 (d, J = 6.4 Hz, 3H), 0.91 (s, 9H), 0.11 (s, 3H), 0.02 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 171.2, 158.4, 138.4, 129.7, 129.5, 121.1, 118.4, 109.8, 81.5, 78.3, 77.9, 74.7, 57.5, 55.8, 25.7, 21.0, 18.8, 18.2, -4.5, -4.9; IR (neat) ν_{max} 3448.6, 2928.0, 2855.5, 1733.6, 1579.9, 1468.8, 1380.9, 1257.4, 1083.4, 967.9, 839 cm⁻¹; HRMS (ESI) calcd for C₂₃H₃₆O₆NaSi [M + Na]⁺ 439.2159, found 439.2173.

Synthesis of 1 and 1a. Independently, 1 (14.5 mg, 82%) and 1a (27.0 mg, 81%) were synthesized from A (37 mg, 0.085 mmol) and A1 (70 mg, 0.160 mmol), respectively, by following the procedures described for deprotection of silyl and acetyl reported in ref 8a.

Data of 1a: $R_f = 0.3$ (EtOAc); $[\alpha]^{28}_{D} = +6.23$ (c 1.38, CH₃OH); ¹H NMR (500 MHz, CD₃CN) δ 7.24 (t, J = 7.9 Hz, 1H), 7.14 (d, J = 7.9 Hz, 1H), 6.96 (d, J = 15.8 Hz, 1H), 6.89 (d, J = 7.9 Hz, 1H), 6.24 (dd, J = 15.8, 7.1 Hz, 1H), 4.66 (d, J = 5.0 Hz, 2H), 4.59 (m, 1H), 4.08 (m, 1H), 3.87 (m, 1H), 3.82 (s, 3H), 3.75 (m, 1H), 3.32 (bd, J = 3.9Hz, 1H), 3.25 (bd, J = 7.3 Hz, 1H) 2.94 (t, J = 3.9 Hz, 1H), 1.23 (d, J = 6.1 Hz, 3H); ¹³C NMR (125 MHz, CD₃CN) δ 158.4, 138.4, 130.1, 129.4, 129.2, 127.0, 119.0, 110.3, 81.3, 78.6, 77.7, 74.1, 55.8, 55.1, 18.7; IR (neat) ν_{max} 3381.5, 2924.7, 2854.7, 1578.5, 1470.8, 1375.7, 1265.1, 1135.0, 1075.2, 1016.4, 975.3, 784.8; HRMS (ESI) calcd for C₁₅H₂₀O₅Na [M + Na]⁺ 303.1197, found 303.1202.

2-((S,É)-4-Hydroxy-4-((2*R***,3***R***)-3-methyloxiran-2-yl)-3-oxobut-1-enyl)-6-methoxybenzyl Acetate (13). Compound 13 (1.84 g, 81%) was synthesized as a solid from 4 (3.1 g, 7.1 mmol) following the TBS deprotection reported in ref 2k: R_f = 0.3 (40% EtOAc/ hexanes); mp 110–112 °C; [\alpha]^{2^8}_{\ D} = +23.3 (***c* **0.51, CHCl₃); ¹H NMR (500 MHz, CDCl₃) \delta 8.17 (d,** *J* **= 16.0 Hz, 1H), 7.37 (t,** *J* **= 7.8, Hz, 1H), 7.32 (d,** *J* **= 7.8 Hz, 1H), 7.00 (d,** *J* **= 16.0 Hz, 1H), 6.99 (d,** *J* **= 7.8 Hz, 1H), 5.35 (ABq,** *J* **= 12 Hz, 2H), 4.07 (d,** *J* **= 7.6 Hz, 1H), 3.87 (s, 3H), 3.17 (qd,** *J* **= 5.3, 2.0 Hz, 1H), 2.70 (dd,** *J* **= 7.6, 2.0 Hz, 1H), 2.06 (s, 3H), 1.38 (d,** *J* **= 5.3 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) \delta 197.9, 170.9, 158.6, 143.1, 135.8, 130.1, 124.0, 123.8, 119.1, 112.9, 76.6, 58.3, 57.0, 55.9, 54.2, 20.9, 17.1; IR (neat) \nu_{max} 3450, 2923, 2851, 1733, 1689, 1610, 1576, 1472, 1440, 1379, 1260, 1099, 1081, 1023, 633 cm⁻¹; HRMS (ESI)** *m***/***z* **[M + Na]⁺ calcd for C₁₇H₂₀O₆Na 343.1150, found 343.1152.**

2-((3*R*,4*R*,E)-3,4-Dihydroxy-4-((2*R*,3*R*)-3-methyloxiran-2-yl)but-1-enyl)-6-methoxybenzyl Acetate (14). To a solution of ketone 13 (1.2 g, 3.7 mmol) in ether (20 mL) was slowly added LiEt₃BH (5.6 mL, 1.0 M in THF, 5.6 mmol) at -78 °C. After the

mixture was stirred for 10 min at -78 °C, the reaction mixture was quenched with H₂O. This mixture was extracted with EtOAc, dried over Na₂SO₄₁ and concentrated in vacuo. The residue was purified by flash column chromatography on silica gel (50% EtOAc/hexanes) to afford diol 14 (977 mg, 82%) as colorless oil: $R_f = 0.2$ (70% EtOAc/ hexanes); $[\alpha]_{D}^{28}$ = +10.0 (c 0.24, CHCl₃); ¹H NMR (500 MHz, $CDCl_3$) δ 7.30 (t, J = 8.0 Hz, 1H), 7.11 (d, J = 8.0 Hz, 1H), 7.00 (d, J = 16.0 Hz, 1H), 6.85 (d, J = 8.0 Hz, 1H), 6.22 (dd, J = 16.0, 7.0 Hz, 1H), 5.29 (ABq, J = 12.0 Hz, 2H), 4.49 (dd, J = 7.0, 6.0 Hz, 1H), 3.85 (s, 3H), 3.81 (dd, J = 6.0, 5.0 Hz, 1H), 3.11 (m, 1H), 2.86 (dd, J = 5.0, 3.0 Hz, 1H), 2.45 (bs, 1H), 2.38 (bs, 1H), 2.05 (s, 3H), 1.32 (d, J = 5.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 171.3, 158.5, 138.7, 130.6, 130.0, 129.7, 121.3, 118.9, 110.1, 73.9, 72.2, 61.5, 58.6, 57.6, 55.8, 51.9, 21.1, 17.1; IR (neat) $\nu_{\rm max}$ 3451, 3377, 2921, 2852, 1734, 1579, 1461, 1378, 1220, 1023, 965 cm⁻¹; HRMS (ESI) m/z [M + Na]⁺ calcd for C17H22O6Na 345.1309, found 345.1308.

2-((*E*)-**2**-((*2R*, **3***S*, **4***R*, **5***S*)-**3**, **4**-Dihydroxy-**5**-methyltetrahydrofuran-**2**-yl)vinyl)-**6**-methoxybenzyl Acetate (15). Compound 15 (37 mg, 93%) was synthesized as a liquid from epoxide 14 (40 mg, 0.124 mmol) following the same procedure described for epoxide opening in ref 8a: $R_f = 0.3$ (70% EtOAc/hexanes); $[\alpha]^{28}_{D} = +20.8$ (*c* 0.56, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.30 (t, *J* = 8.3 Hz, 1H), 7.11 (d, *J* = 8.3 Hz, 1H), 6.98 (d, *J* = 15.7 Hz, 1H), 6.83 (d, *J* = 8.3 Hz, 1H), 6.10 (dd, *J* = 15.7, 7.0 Hz, 1H), 5.29 (ABq, *J* = 11.5 Hz, 2H), 4.33 (t, *J* = 7.0 Hz, 1H), 3.94 (m, 2H), 3.84 (s, 3H), 3.79 (t, *J* = 5.5 Hz, 1H), 2.05 (s, 3H), 1.35 (d, *J* = 6.2 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 158.5, 138.8, 131.2, 129.9, 129.5, 121.2, 118.9, 110.0, 84.0, 80.0, 76.2, 75.4, 57.6, 55.8, 21.1, 19.1; IR (neat) ν_{max} 3428, 2921, 2851, 1733, 1580, 1462, 1379, 1266, 1078, 966 cm⁻¹; HRMS (ESI) *m*/*z* [M + Na]⁺ calcd for C₁₇H₂₂O₆Na 345.1308, found 345.1308.

Varitriol (1). (+)-Varitriol (1) (15.3 mg, 88%) was synthesized as a solid from compound **15** (20 mg, 0.062 mmol) following the acetyl deprotection reported in ref 8a: $R_f = 0.3$ (EtOAc); $[\alpha]^{30}{}_D = +23.8$ (c 0.26, CH₃OH); ¹H NMR (500 MHz, CD₃COCD₃) δ 7.23 (t, J = 8.0 Hz, 1H), 7.13 (d, J = 15.8 Hz, 1H), 7.12 (d, J = 8.0 Hz, 1H), 6.90 (d, J = 8.0 Hz, 1H), 6.21 (dd, J = 15.8, 6.7 Hz, 1H), 4.72 (s, 2H), 4.29 (t, J = 6.7 Hz, 1H), 4.23 (brs, 1H), 4.03 (brs, 1H), 3.91 (m, 1H), 3.84 (m, 1H), 3.83 (s, 3H), 3.70 (m, 1H), 3.65 (brs, 1H), 1.28 (d, J = 6.2 Hz, 3H); ¹³C NMR (125 MHz, CD₃COCD₃) δ 158.9, 139.0, 132.4, 129.4, 129.3, 128.0, 119.3, 110.6, 85.3, 80.0, 77.2, 76.5, 56.0, 55.5, 19.5; IR (neat) ν_{max} 3355, 2920, 1578, 1354, 1219, 1088; HRMS (ESI) m/z [M + Na]⁺ calcd for C₁₅H₂₀O₅Na 303.1199, found 303.1202.

Varioxirane (2). Compound 2 (70 mg, 81%) was synthesized as a liquid from compound 14 (100 mg, 0.31 mmol) following the same procedure described in ref 8a: $R_f = 0.3$ (EtOAc); $[\alpha]^{30}_{D} = -24.61$ (*c* 0.13, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.26 (d, J = 8.4 Hz, 1H), 7.25 (d, J = 15.9 Hz, 1H), 7.07 (d, J = 8.4 Hz, 1H), 6.84 (d, J = 8.4 Hz, 1H), 6.21 (dd, J = 15.9, 6.7 Hz, 1H), 4.80 (s, 2H), 4.50 (dd, J = 6.7, 4.2 Hz, 1H), 3.86 (s, 3H), 3.81 (t, J = 4.2 Hz, 1H), 3.10 (qd, J = 5.0, 2.5 Hz, 1H), 2.86 (dd, J = 4.2, 2.5 Hz, 1H), 2.55 (brs, 1H), 1.70 (bs, 2H), 1.31 (d, J = 5.0 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 157.7, 137.7, 130.5, 129.3, 128.9, 125.9, 119.1, 109.7, 73.8, 72.7, 58.8, 55.6, 52.3, 17.1; IR (neat) ν_{max} 3454, 3376, 2923, 2853, 1577, 1469, 1260, 1074, 998 cm⁻¹; HRMS (ESI) m/z [M + Na]⁺ calcd for C₁₅H₂₀O₅Na 303.1204, found 303.1202.

Triacetylvarioxirane (16). Compound **16** (42 mg, 95%) was synthesized as a colorless oil from compound **2** (30 mg, 0.11 mmol), Ac₂O (0.04 mL, 0.42 mmol), and Et₃N (0.09 mL, 0.64 mmol) following the same procedure described for acetyl in ref 8a: $R_f = 0.5$ (30% EtOAc/hexanes); $[\alpha]^{30}_{D} = -32.9$ (*c* 0.19, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.31 (t, J = 8.3 Hz, 1H), 7.11 (d, J = 8.3 Hz, 1H), 6.98 (d, J = 15.8 Hz, 1H), 6.86 (d, J = 8.3 Hz, 1H), 6.14 (dd, J = 15.8, 7.5 Hz, 1H), 5.70 (ddd, J = 7.5, 3.7, 1.5 Hz, 1H), 5.26 (s, 2H), 4.90 (dd, J = 6.0, 3.7 Hz, 1H), 3.84 (s, 3H), 3.03 (dq, J = 5.3, 2.2 Hz, 1H), 2.85 (dd, J = 6.0, 2.2 Hz, 1H), 2.12 (s, 3H), 2.09 (s, 3H), 2.07 (s, 3H), 1.30 (d, J = 5.3 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 171.0, 169.8, 169.7, 158.4, 138.2, 131.9, 131.7, 129.9, 125.8, 118.8, 110.4, 73.7, 73.1, 57.5, 55.9, 55.8, 52.9, 21.0, 20.9, 20.8, 17.1; IR (neat) ν_{max} 2922, 2851, 1741, 1579, 1469,1373,1224,1024 cm⁻¹; HRMS (ESI) *m*/*z* [M + Na]⁺ calcd for C₂₁H₂₆O₈Na 429.1503, found 429.1519.

Varitriol (1). Varitriol (1) (18.2 mg, 91%) was synthesized from varioxirane (2) (20 mg, 0.07 mmol) following the same procedure described in ref 8a.

(*R*,*E*)-Methyl 2-Hydroxypent-3-enoate ((–)-9). Compound (–)-9 (1.2 g, 24%) was obtained from (±)-9 (5 g, 38.46 mmol) following the same procedure described for 10: $R_f = 0.6$ (30% EtOAc/hexanes); $[\alpha]^{30}_{D} = -96.4$ (*c* 0.09, CHCl₃);

(*R*,*E*)-Methyl 2-(*tert*-Butyldimethylsilyloxy)pent-3-enoate (17). Compound 17 (3.677 g, 98%) was synthesized as a colorless oil from (-)-9 (2 g, 15.38 mmol) following the same procedure described in ref 8a: $R_f = 0.7$ (10% EtOAc/hexanes); [α]²⁸_D = +3.9 (*c* 0.35, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 5.82 (m, 1H), 5.56 (qdd, J = 15.2, 5.7, 1.6 Hz, 1H), 4.64 (m, 1H), 3.71 (s, 3H), 1.70 (dt, J= 6.6, 1.6 Hz, 3H), 0.90 (s, 9H), 0.08 (s, 3H), 0.06 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 172.6, 128.2, 128.2, 73.1, 51.9, 25.6, 18.3, 17.5, -5.1, -5.2; IR (neat) ν_{max} 2929, 2857, 1758, 1436, 1253, 1153, 1062, 965, 835 cm⁻¹; HRMS (ESI) m/z [M + H]⁺ calcd for C₁₂H₂₅O₃Si 245.1562, found 245.1567.

(*R*,*E*)-Dimethyl 3-(*tert*-Butyldimethylsilyloxy)-2-oxohex-4enylphosphonate *ent*-(8). Compound *ent*-8 (4.673 g, 97%) was synthesized as a highly viscous colorless oil from ester 17 (3.5 g, 14.34 mmol) following the same procedure described in ref 12: $R_f = 0.3$ (70% EtOAc/hexanes); $[\alpha]^{28}_{\rm D} = +4.97$ (*c* 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 5.83 (m, 1H), 5.38 (ddd, *J* = 15.0, 6.3,1.6 Hz, 1H), 4.53(d, *J* = 6.3 Hz, 1H), 3.75 (s, 3H), 3.72 (s, 3H), 3.32–3.10 (m, 2H), 1.69 (d, *J* = 6.3 Hz, 3H), 0.87 (s, 9H), 0.04 (s, 3H), 0.02 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 202.1, 201.2, 130.1, 127.4, 80.0, 79.9, 52.9, 52.8, 52.7, 35.3, 33.5, 25.6, 18.1, 17.7, -4.7, -5.1; IR (neat) ν_{max} 2955, 2857, 1727, 1682, 1467, 1389, 1257, 1034, 968, 839 cm⁻¹; HRMS (ESI) *m*/*z* [M + H]⁺ calcd for C₁₄H₃₀O₅ PSi 337.1586, found 337.1594.

2-((*R***, 1***E***, 5***E***)-4-(***tert***-Butyldimethylsilyloxy)-3-oxohepta-1,5dienyl)-6-methoxybenzyl Acetate** *ent***-(5). Compound** *ent***-5 (4.589 g, 82%) was synthesized as a solid from \beta-ketophosphonate** *ent***-8 (4.5 g, 13.39 mmol) and aldehyde 6 (4.178 g, 20.08 mmol) following the same procedure described for 4: R_f = 0.5 (20% EtOAc/hexanes); mp 70-72 °C; [\alpha]^{31}_{D} = -93.7 (***c* **0.17, CHCl₃); ¹H NMR (500 MHz, CDCl₃) \delta 7.98 (d,** *J* **= 15.9 Hz, 1H), 7.34 (t,** *J* **= 8.2 Hz, 1H), 7.25 (d,** *J* **= 8.2 Hz, 1H), 7.08 (d,** *J* **= 15.9 Hz, 1H), 6.94 (d,** *J* **= 8.2 Hz, 1H), 5.90 (dqd,** *J* **= 15.2, 6.7, 1.6 Hz, 1H), 5.50 (qdd,** *J* **= 15.2, 5.5, 1.6 Hz, 1H), 5.30 (ABq,** *J* **= 12.0 Hz, 2H), 4.65 (m, 1H), 3.84 (s, 3H), 2.03 (s, 3H), 1.71 (td,** *J* **= 6.7, 1.6 Hz, 3H), 0.92 (s, 9H), 0.08 (s, 3H), 0.06 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) \delta 198.6, 170.7, 158.4, 140.3, 136.8, 129.9, 128.6, 128.4, 123.8, 123.4, 118.9, 112.2, 79.5, 57.1, 55.8, 25.7, 20.8, 18.2, 17.7, -4.8, -5.0; IR (neat) \nu_{max} 2929, 2855, 1737, 1693, 1575, 1470, 1379, 1257, 1096, 964, 884, 674 cm⁻¹; HRMS (ESI) m/z [M + H]⁺ calcd for C₂₃H₃₅O₅Si 419.2235, found 419.2248.**

2-((*R***, 1***E***, 5***E***)-4-Hydroxy-3-oxohepta-1,5-dienyl)-6-methoxybenzyl Acetate (18). Compound 18 (680 mg, 80%) was synthesized as a liquid from compound 5 (1.2 g, 2.8 mmol) following the same procedure described in ref 2k: R_f = 0.4 (40% EtOAc/hexanes); [\alpha]^{30}_{D} = -128.7 (***c* **0.17, CHCl₃); ¹H NMR (500 MHz, CDCl₃) \delta 8.10 (d,** *J* **= 15.8 Hz, 1H), 7.37 (t,** *J* **= 8.0 Hz, 1H), 7.25 (d,** *J* **= 8.0 Hz, 1H), 6.99 (d,** *J* **= 8.0 Hz, 1H), 6.79 (d,** *J* **= 15.8 Hz, 1H), 6.0 (qd,** *J* **= 15.1, 6.7 Hz, 1H), 5.45 (qdd,** *J* **= 15.1, 7.7, 1.6 Hz, 1H), 5.32 (s, 2H), 4.82 (d,** *J* **= 7.7, 1H), 3.87 (s, 3H), 2.06 (s, 3H), 1.78 (dd,** *J* **= 6.7, 1.6 Hz, 3H), ¹³C NMR (125 MHz, CDCl₃) \delta 197.9, 170.7, 158.5, 149.0, 141.6, 136.5, 132.1, 130.0, 127.4, 123.6, 118.9, 112.7, 77.4, 57.0, 55.8, 20.8, 17.9; IR (neat) \nu_{max} 3451, 2928, 2850, 1737, 1692, 1575, 1440, 1250, 1127, 1004, 915, 830; HRMS (ESI) m/z [M + Na]⁺ calcd for C₁₇H₂₀O₅Na 327.1195, found 327.1202.**

2-((1*E*,**3***S*,**4***R*,**5***E*)-**3**,**4**-**Dihydroxyhepta-1**,**5**-dienyl)-6-methoxybenzyl Acetate (19). Compound 19 (182 mg, 92%) was synthesized as a colorless oil from ketone 18 (200 mg, 0.65 mmol) following the same procedure described for 14: $R_f = 0.3$ (60% EtOAc/hexanes); $[\alpha]^{30}_{D} = -96.6$ (*c* 0.06, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.30 (t, *J* = 7.9 Hz, 1H), 7.09 (d, *J* = 7.9 Hz, 1H), 6.93 (d, *J* = 15.8 Hz, 1H), 6.84 (d, *J* = 7.9 Hz, 1H), 6.12 (dd, *J* = 15.8, 6.4 Hz, 1H), 5.82 (qd, *J* = 15.3, 6.4 Hz, 1H), 5.54 (dd, *J* = 15.3, 7.0 Hz 1H), 5.28 (ABq, *J* = 12.0 Hz, 2H), 4.30 (m, 1H), 4.18 (m, 1H), 3.84 (s, 3H), 2.33 (brs, 1H),

2.18 (brs, 1H), 2.06 (s, 1H), 1.74 (d, J = 6.4 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 171.3, 158.5, 139.0, 131.2, 130.1, 129.9, 129.7, 128.9, 121.2, 118.9, 110.0, 75.6, 75.4, 57.7, 55.8, 21.1, 17.9; IR (neat) ν_{max} 3426, 2921, 2852, 1730, 1579, 1470, 1379, 1250, 1079, 1022, 963 cm⁻¹; HRMS (ESI) m/z [M + Na]⁺ calcd for C₁₇H₂₂O₅Na 329.1361, found 329.1359.

(1*E*,3*S*,4*R*,5*E*)-1-(2-(Hydroxymethyl)-3-methoxyphenyl)-hepta-1,5-diene-3,4-diol (*ent*-3). Compound *ent*-3 (58 mg, 85%) was synthesized as a liquid from compound 19 (80 mg, 0.26 mmol) following the same procedure described in ref 8a: $R_f = 0.3$ (EtOAc); $[\alpha]^{30}_{D} = -36.0$ (*c* 0.05, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.18 (t, *J* = 8.0 Hz, 1H), 7.01 (d, *J* = 8.0 Hz, 1H), 6.94 (d, *J* = 16.0 Hz, 1H), 6.76 (d, *J* = 8.0 Hz, 1H), 6.06 (dd, *J* = 16.0, 4.5 Hz, 1H), 5.74 (dq, *J* = 15.2, 6.4 Hz, 1H), 5.48 (dd, *J* = 15.2, 4.5 Hz, 1H), 4.73 (ABq, *J* = 12.0 Hz, 2H), 4.24 (t, *J* = 6.4 Hz, 1H), 4.12 (m, 1H), 3.80 (s, 3H), 2.60 (brs, 2H), 1.67 (d, *J* = 6.4 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 159.2, 139.2, 132.6, 131.2,130.9, 130.4, 130.2, 127.4, 120.6, 110.0, 76.9, 76.7, 57.7, 57.0, 19.2; IR (neat) ν_{max} 3380, 2919, 1577, 1471, 1381, 1261, 1079, 969 cm⁻¹; HRMS (ESI) *m*/*z* [M + Na]⁺ calcd for C₁₅H₂₀O₄Na 287.1249, found 287.1253.

ASSOCIATED CONTENT

Supporting Information

¹H and ¹³C NMR spectra of all new compounds. This material is available free of charge via the Internet at http://pubs.acs.org.

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Notes

The authors declare no competing financial interest.

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(19) Increasing the reaction time from 8 to 10 h increased the er of (-)-9 and decreased the yield. With 8 h reaction time, the er of (-)-9 is 84.7:15.3 (40% yield), whereas with 10 h the ratio is 98.6:1.4 (24% yield) (see the Supporting Information for HPLC chromatograms).