

Min Zhang,* 🔟 Ting Li, Min Qian, Kailu Li, Yukun Qin, Ting Zhao, and Liu-Qing Yang*

School of Chemistry and Chemical Engineering, Jiangsu University, Zhenjiang 212013, Jiangsu, China *E-mail: zhangmin@ujs.edu.cn; yangliuqing@ujs.edu.cn Received January 31, 2018 DOI 10.1002/ihet.3190

Published online 00 Month 2018 in Wiley Online Library (wileyonlinelibrary.com).



A series of 1-azaaurone derivatives were designed and synthesized from 3,5-dimethoxyaniline and 2-chloroacetonitrile. Their structures were characterized by melting point, ¹H NMR, IR, and elemental analysis, as well as ¹³C NMR. The target compounds were evaluated for antitumor activities against human hepatocellular liver carcinoma cell line (HepG-2) and human cervix carcinoma cell line (Hela) using methyl thiazolyl tetrazolium method. The results revealed that several 1-azaaurones exhibited strong proliferation inhibition efficacy against HepG-2 and Hela with an IC₅₀ range of 5.6–8.8 µg/mL without damaging normal cell.

J. Heterocyclic Chem., 00, 00 (2018).

INTRODUCTION

Aurones, a subclass of naturally occurring flavonoids, are met in vegetables, flowers, and fruits where they contribute to the yellow or orange color [1]. They were also used by plants to resist pathogens [2], pests [3], and competitive weeds [4,5]. Recently, increasing attention has been focused on aurones because both natural aurones and synthetic analogs exhibited excellent antitumor [6,7], antioxidant [8,9], antibacterial, and anti-inflammatory [10,11] activities, as well as acted as potent multifunctional defense agents against Alzheimer's disease [12,13]. In addition, aurones have been proved to be valuable inhibitors of human tyrosinase to prevent hyperpigmentation-related abnormality [14,15]. These studies indicate that aurone has evolved to be an excellent leading scaffold for the development of various analogs with significant biological activity.

1-Azaaurone, an isosteric equivalent replacement of aurone by replacing intracyclic O atom at 1-position with NH group (Fig. 1), has been found to display important bioactivity [7,16,17]. Moreover, some azaaurones exhibited much stronger bioactivity than the parent aurones. For example, Souard *et al.* reported that azaaurone analogs demonstrated better antiplasmodial activity in comparison with corresponding aurones [16]. In one of our previous studies [18], it was found that 6substituted aurones possessed excellent antitumor effects against HepG-2, Hela, and MCF-7 cancer cell lines, and the variations of substituents at the A-ring and/or B-ring could cause remarkable changes in the activity. On the other hand, some examples support that the presence of methoxy moieties at C-4 and C-6 positions of aurones may lead to a substantial increase in the activity [4,19,20]. Collectively, it is of interest to synthesize 1azaaurones bearing methoxyl groups at C-4 and C-6 positions and possessing diverse substituents at the B-ring, to evaluate their preliminary antitumor activity and to explore the structure–activity relationship.

RESULTS AND DISCUSSION

Synthesis. The synthetic pathway of title compounds was summarized in Scheme 1. 1-(2-Amino-4,6-dimethoxyphenyl)-2-chloroethan-1-one (1) was synthesized from 3,5-dimethoxyaniline and 2-chloroacetonitrile in the presence of trichloroborane and dry aluminum trichloride, followed by hydrolysis according to the reported procedure [21]. Treatment of 1 with acetic anhydride in the presence of potassium carbonate produced *N*-acetyl-4,6-dimethoxyindolin-3-one (2) in



Figure 1. Basic structures of aurone and 1-azaaurone.





good yield. The target 1-azaaurone derivatives 3a-3m were obtained by the condensation of 2 with various arylaldehydes in H₂O/EtOH under alkali condition. Six of them (3d, 3e, 3g, and 3k-3m) are new compounds. The target compounds were confirmed by ¹H NMR, IR, and elemental analysis, as well as ¹³C NMR. These 1identified azaaurones were exclusively as Ζ stereoisomers on the basis of the ¹H chemical shifts of exocyclic vinyl proton (=CH-), which appeared at δ 6.38–6.84 ppm in ¹H NMR, corresponding to the known values (ca. 6.70 ppm) for (Z)-aurones [4,8,12].

Biological activity. The preliminary antitumor activities of all synthetic 1-azaaurones against HepG-2 and Hela were evaluated using methyl thiazolyl tetrazolium (MTT) assay [18]. As demonstrated in Table 1, most of the compounds were active toward HepG-2 and Hela at the concentration of 10 μ g/mL. For example, compounds **3f**,

31, and 3m displayed an inhibition rate of higher than 60% against both HepG-2 and Hela. Regarding antitumor activity against HepG-2, the structure-activity relationship (Table 1) confirmed that the presence of electronwithdrawing group at the B-ring led to less active compounds (3c, 3d, and 3k vs 3a), while those with a donating-electron group at the B-ring showed a little better inhibitory effect (3e, 3f vs 3a). Further, two electron-donating groups at the B-ring led to a dramatic increase in the activity (31, 3m vs 3a). It was also found that hydroxyl group at C-4' position was highly beneficial when 31 was compared with 3m. With regard to the activity against Hela, the presence of electron-donating group at the B-ring was highly advantageous (3f, 3h-3j vs 3a-3d, 3k); further, two methoxyl groups at C-3' and C-4' positions led to the most potent compound (31 vs 3i, **3i**). However, when the methoxy moiety at C-4' position was replaced by hydroxyl group, the activity decreased slightly (3m vs 3l). Taken together, possessing the electron-donating groups at the B-ring is beneficial to the antitumor activity of 1-azaaurones but is disadvantageous to that of aurones [18], which is probably correlated to the difference of O atom and NH group at 1-position.

On the basis of the screening results, compounds 3f, 3j, **31**, and **3m** were selected to further evaluate their IC_{50} values against cancer cell lines HepG-2 and Hela, as well as normal human hepatic cell line (WRL-68) with Aurone A as a reference control, which is the most active aurone derivative in our previous work [18]. As shown in Table 2, each of them was active with IC_{50} value below 10 µg/mL. The most active compound was **3m** with IC₅₀ values of 5.6 and 6.3 µg/mL against HepG-2 and Hela comparative to IC_{50} values of Aurone A (8.4 and 3.3 µg/mL), respectively. More interestingly, compounds 31 and 3m were proved to be ineffective against WRL-68 with IC₅₀ values of 74.1 and 62.4 μ g/ mL, respectively. These results implied that 1-azaaurone derivatives selectively inhibited cancer cell lines without damaging normal cell. Moreover, these 1-azaaurones displayed much higher inhibition toward Hela than the aurones described in the literature [22], which might also

Compound	HepG-2	Hela	Compound	HepG-2	Hela
3a	55.4	NA	3h	51.6	54.8
3b	55.3	31.8	3i	46.7	54.4
3c	43.6	8.75	3j	50.5	60.9
3d	16.8	NA	3k	36.3	16.5
3e	60.3	17.7	31	63.7	80.0
3f	63.2	65.5	3m	82.6	71.7
3g	50.5	22.7			

 Table 1

 Antiproliferative activities of 1-azaaurones 3a–3m (inhibition% at 10 μg/mL).

NA, no activity.

In vitro antitumor activities of some target compounds (IC_{50}, $\mu g/mL).$								
Compound	HepG-2	Hela	WRL-68					
3f	7.5	7.8	ND					

Table 2

Aurone A ^a	8.4	3.3	ND
3m	5.6	6.3	62.4
31	7.4	8.8	74.1
3ј	8.0	6.7	ND
3f	7.5	7.8	ND



be attributed to the replacement of the intracyclic O atom by the NH group.

In conclusion, a series of 1-azaaurones were designed and synthesized, and their antitumor activities against cancer cell lines HepG-2, Hela, and normal cell WRL-68 were evaluated. The preliminary bioassay demonstrated that some 1-azaaurones selectively inhibited cancer cell lines without damaging normal cell. The structureactivity relationship revealed that the substituents at the B-ring played an important role on the activity. The present investigation brings essential elements, which will help to design and synthesize more potent azaaurones.

EXPERIMENTAL

IR spectroscopy were recorded on a Nicolet Nexus 470 spectrometer (Nicolet Instrument Corporation, Madison, WI) as KBr pellets. Melting points were uncorrected and obtained using an X-4 digital melting point apparatus. NMR spectra were recorded on a Bruker AVANCEII 400 MHz instrument (400 MHz for ¹H, 100 MHz for ¹³C). Elemental analysis was carried out with a FLASH1112A analyzer (CE Instruments Ltd, Milan, Italy). 1. 1-(2-Amino-4,6-Synthesis of compound dimethoxyphenyl)-2-chloroethan-1-one (1) was prepared basing on the procedure [21] from the reaction of 3,5dimethoxyaniline (3.06 g, 20.0 mmol) and 2chloroacetonitrile (23.6 mmol) in dry dichloromethane (20 mL), in the presence of trichloroborane (23.6 mmol) and anhydrous aluminum trichloride (2.93 g, 22.0 mmol) at 0°C. After working up, the reaction solution was treated with hydrochloric acid solution (2 mol/L), followed by heating at 80°C for 0.5 h. The resulting solution was extracted three times with CH₂Cl₂, washed with water, dried with Na₂SO₄, and followed by removing the solvent to give a yellow solid, yield 70%, mp 90-92°C. 2. N-Acetyl-4,6-**Synthesis** compound of dimethoxyindolin-3-one (2) was prepared according to a

reported method [21]. A solution of intermediate 1 (1.15 g, 5.0 mmol), K₂CO₃ (0.83 g, 6.0 mmol) in acetone (30 mL) was stirred at room temperature till all solid was dissolved. Then, the solution was treated with acetic anhydride (1.08 g, 10.6 mmol) and heated under reflux until completion of the reaction (monitored via thin-layer chromatography). The resulting solution was poured into ice water after cooling. The formed solid was filtered off, washed with water, dried, and recrystallized from EtOH to afford product 2 in 60% yield, mp 93–95°C.

General synthetic procedure for 1-azaaurones 3a-3m. Oxindole 2 (0.24 g, 1.0 mmol) was dissolved in EtOH (5 mL) then treated with KOH solution (10%, 5 mL). The corresponding arylaldehyde (1.2 mmol) was added to the mixture. Then, the solution was heated at 50°C for 3-5 h. EtOH was evaporated after the reaction was finished by thin-layer chromatography monitoring. Ice water was added to the residue, followed by neutralizing with hydrochloric acid solution (2 mol/L). The precipitate was collected by filtration, washed with water three times, and purified by recrystallization from EtOH to yield compounds 3a-3m.

(Z)-2-Benzylidene-4,6-dimethoxy-2,3-dihydro-1H-indol-3one (3a). Compound 3a has been reported previously [23,24]. Yellow solid, yield 83.0%, mp 134–136°C. ¹H NMR (400 MHz, CDCl₃) *δ*: 3.86 (s, 3H, OCH₃), 3.93 (s, 3H, OCH₃), 5.93 (d, J = 1.6 Hz, 1H, ArH), 6.06 (d, J = 2.0 Hz, 1H, ArH), 6.75 (s, 1H, =CH), 6.84 (br s, 1H, NH), 7.29-7.34 (m, 1H, ArH), 7.39-7.45 (m, 2H, ArH), 7.49-7.53 (m, 1H, ArH). IR (KBr) v: 3404, 3068, 1672, 1624, 1597, 1585, 1464, 1451, 1398, 1364, 1263, 1223, 1211, 1159, 1126, 1050 cm⁻¹. Anal. Calcd for C₁₇H₁₅NO₃: C 72.58, H 5.37, N 4.98. Found: C 72.26, H 5.53, N 5.20.

(Z)-2-(2-Fluorobenzvlidene)-4.6-dimethoxv-2.3-dihvdro-1Hindol-3-one (3b). Compound **3b** has been reported in the literature [19]. Orange solid, yield 68.4%, mp 110-113°C. ¹H NMR (400 MHz, DMSO- d_6) δ : 3.83 (s, 3H, OCH_3), 3.85 (s, 3H, OCH_3), 6.01 (d, J = 2.0 Hz, 1H, ArH), 6.19 (d, *J* = 2.0 Hz, 1H, ArH), 6.44 (s, 1H, =CH), 7.25–7.33 (m, 2H, ArH), 7.35–7.41 (m, 1H, ArH), 7.76– 7.82 (m, 1H, ArH), 9.76 (br s, 1H, NH). IR (KBr) v: 3362, 3088, 2978, 1667, 1626, 1577, 1516, 1513, 1467, 1252, 1219, 1198, 1438, 1372, 1296, 1153, 1119,1017 cm⁻¹. Anal. Calcd for C₁₇H₁₄FNO₃: C 68.22, H 4.71, N 4.68. Found: C 68.46, H 4.53, N, 5.05.

(Z)-2-(3-Fluorobenzylidene)-4,6-dimethoxy-2,3-dihydro-1Hindol-3-one (3c). Compound 3c has been reported in the literature [25]. Orange solid, yield 66.3%, mp 95-96°C. ¹H NMR (400 MHz, DMSO- d_6) δ : 3.82 (s, 3H, OCH₃), 3.85 (s, 3H, OCH₃), 6.01 (d, J = 1.6 Hz, 1H, ArH), 6.20 (d, J = 1.6 Hz, 1H, ArH), 6.38 (s, 1H, =CH), 7.11–7.18 (m, 1H, ArH), 7.43–7.54 (m, 3H, ArH), 9.79 (br s, 1H, NH). IR (KBr) v: 3412, 3036, 1627, 1590, 1511, 1458, 1380, 1242, 1218, 1156, 1120, 1050, 1009 cm⁻¹. Anal. Calcd for C₁₇H₁₄FNO₃: C 68.22, H 4.71, N 4.68. Found: C 67.91, H 4.93, N 4.97.

(Z)-2-(4-Fluorobenzylidene)-4,6-dimethoxy-2,3-dihydro-1Hindol-3-one (3d). Orange solid, yield 68.9%, mp 108– 110°C. ¹H NMR (400 MHz, DMSO- d_6) δ : 3.82 (s, 3H, OCH₃), 3.84 (s, 3H, OCH₃), 5.99 (d, J = 1.6 Hz, 1H, ArH), 6.19 (d, J = 1.6 Hz, 1H, ArH), 6.40 (s, 1H, =CH), 7.24– 7.31 (m, 2H, ArH), 7.67–7.75 (m, 2H, ArH), 9.73 (br s, 1H, NH). ¹³C NMR (100 MHz, DMSO- d_6) δ : 55.4, 55.6, 88.6, 90.6, 103.0, 105.9, 115.7 (d, J = 21.4 Hz, 2C), 130.9 (d, J = 3.2 Hz), 131.4 (d, J = 8.0 Hz, 2C), 135.1, 157.0, 159.6, 161.3 (d, J = 245.3 Hz), 167.9, 181.3. IR (KBr) v: 3420, 3032, 1626, 1595, 1507, 1462, 1380, 1262, 1217, 1162, 1128 cm⁻¹. Anal. Calcd for C₁₇H₁₄FNO₃: C 68.22, H 4.71, N 4.68. Found: C 68.01, H 4.90, N 4.89.

(Z)-2-(2-Methylbenzylidene)-4,6-dimethoxy-2,3-dihydro-1Hindol-3-one (3e). Yellow solid, yield 82.6%, mp 213– 214°C. ¹H NMR (400 MHz, CDCl₃) δ : 2.38 (s, 3H, CH₃), 3.84 (s, 3H, OCH₃), 3.92 (s, 3H, OCH₃), 5.90 (d, J = 2.0 Hz, 1H, ArH), 6.00 (d, J = 1.6 Hz, 1H, ArH), 6.81 (br s, 1H, NH), 6.84 (s, 1H, =CH), 7.17–7.25 (m, 3H, ArH), 7.47–7.51 (m, 1H, ArH). ¹³C NMR (100 MHz, CDCl₃) δ : 20.0, 55.7, 55.8, 88.1, 91.0, 105.1, 107.5, 126.2, 127.9, 128.0, 130.7, 133.8, 136.8, 138.1, 156.5, 160.4, 168.5, 181.9. IR (KBr) v: 3420, 3095, 2966, 1674, 1607, 1517, 1462, 1370, 1257, 1217, 1200, 1154, 1123, 1033, 1001 cm⁻¹. Anal. Calcd for C₁₈H₁₇NO₃: C 73.20, H 5.80, N 4.74. Found: C 73.01, H 5.95, N 4.50.

(Z)-2-(3-Methylbenzylidene)-4,6-dimethoxy-2,3-dihydro-1Hindol-3-one (3f). Compound 3f has been reported previously [24]. Yellow solid, yield 65.1%, mp 168– 170°C. ¹H NMR (400 MHz, CDCl₃) δ : 2.38 (s, 3H, CH₃), 3.85 (s, 3H, OCH₃), 3.90 (s, 3H, OCH₃), 5.91 (d, J = 1.6 Hz, 1H, ArH), 6.08 (d, J = 2.0 Hz, 1H, ArH), 6.71 (s, 1H, =CH), 7.01 (br s, 1H, NH), 7.08–7.12 (m, 1H, ArH), 7.28–7.33 (m, 3H, ArH). IR (KBr) v: 3244, 2970, 2848, 1675, 1605, 1511, 1458, 1430, 1370, 1262, 1216, 1158, 1119, 1046 cm⁻¹. Anal. Calcd for C₁₈H₁₇NO₃: C 73.20, H 5.80, N 4.74. Found: C 73.54, H 5.62, N 4.90.

(Z)-2-(4-Methylbenzylidene)-4,6-dimethoxy-2,3-dihydro-1Hindol-3-one (3g). Yellow solid, yield 84.2%, mp 187– 189°C. ¹H NMR (400 MHz, CDCl₃) δ : 2.37 (s, 3H, CH₃), 3.85 (s, 3H, OCH₃), 3.91 (s, 3H, OCH₃), 5.91 (d, J = 1.6 Hz, 1H, ArH), 6.07 (d, J = 2.0 Hz, 1H, ArH), 6.74 (s, 1H, =CH), 6.91 (br s, 1H, NH), 7.21 (d, J = 8.0 Hz, 2H, ArH), 7.40 (d, J = 8.0 Hz, 2H, ArH). ¹³C NMR (100 MHz, CDCl₃) δ : 21.3, 55.7(2C), 88.3, 91.1, 105.0, 109.9, 129.2(2C), 129.7(2C), 132.1, 135.8, 138.2, 156.5, 160.3, 168.4, 182.4. IR (KBr) v: 3285, 3100, 2998, 1683, 1630, 1610, 1590, 1508, 1463, 1383, 1319, 1254, 1214, 1158, 1115, 1042 cm⁻¹. Anal. Calcd for C₁₈H₁₇NO₃: C 73.20, H 5.80, N 4.74. Found: C 72.95, H 6.11, N 4.94.

(Z)-2-(2-Methoxylbenzylidene)-4,6-dimethoxy-2,3-dihydro-1H-indol-3-one (3h). Compound **3h** has been reported in the literature [25]. Yellow solid, yield 67.1%, mp 214–216°C. ¹H NMR (400 MHz, CDCl₃) δ : 3.85 (s, 3H, OCH₃), 3.91 (s, 3H, OCH₃), 3.92 (s, 3H, OCH₃), 5.88 (d, J = 1.6 Hz, 1H, ArH), 6.00 (d, J = 2.0 Hz, 1H, ArH), 6.84 (s, 1H, =CH), 6.93–7.03 (m, 2H, ArH), 7.27–7.34 (m, 2H, ArH + NH), 7.46 (dd, J = 7.6, 1.2 Hz, 1H, ArH). IR (KBr) v: 3377, 2933, 2840, 1671, 1630, 1589, 1511, 1489, 1463, 1434, 1381, 1264, 1246, 1218, 1157, 1117, 1029 cm⁻¹. Anal. Calcd for C₁₈H₁₇NO₄: C 69.44, H 5.50, N 4.50. Found: C 69.65, H 5.44, N, 4.24.

(Z)-2-(3-Methoxylbenzylidene)-4,6-dimethoxy-2,3-dihydro-1H-indol-3-one (3i). Compound **3i** has been reported previously [24]. Orange-yellow solid, yield 85.0%, mp 187–189°C. ¹H NMR (400 MHz, CDCl₃) δ : 3.83 (s, 3H, OCH₃), 3.85 (s, 3H, OCH₃), 3.90 (s, 3H, OCH₃), 5.91 (d, J = 1.6 Hz, 1H, ArH), 6.05 (d, J = 2.0 Hz, 1H, ArH), 6.69 (s, 1H, =CH), 6.84 (dd, J = 8.0, 2.0 Hz, 1H, ArH), 6.99 (br s, 1H, NH), 7.00–7.03 (m, 1H, ArH), 7.07–7.12 (m, 1H, ArH), 7.29–7.35 (m, 1H, ArH). IR (KBr) v: 3426, 3077, 2942, 1667, 1626, 1592, 1516, 1463, 1385, 1256, 1216, 1158, 1120, 1051 cm⁻¹. Anal. Calcd for C₁₈H₁₇NO₄: C 69.44, H 5.50, N 4.50. Found: C 69.67, H 5.26, N 4.74.

(Z)-2-(4-Methoxylbenzylidene)-4,6-dimethoxy-2,3-dihydro-1H-indol-3-one (3j). Compound **3j** has also been reported in the literature [24]. Yellow solid, yield 87.3%, mp 213– 215°C. ¹H NMR (400 MHz, CDCl₃) δ : 3.83 (s, 3H, OCH₃), 3.84 (s, 3H, OCH₃), 3.89 (s, 3H, OCH₃), 5.90 (d, J = 1.6 Hz, 1H, ArH), 6.08 (d, J = 1.6 Hz, 1H, ArH), 6.73 (s, 1H, =CH), 6.93 (d, J = 8.8 Hz, 2H, ArH), 6.97 (br s, 1H, NH), 7.46 (d, J = 8.8 Hz, 2H, ArH). IR (KBr) v: 3421, 3089, 2929, 1630, 1591, 1509, 1467, 1366, 1299, 1253, 1217, 1176, 1160, 1119, 1029 cm⁻¹. Anal. Calcd for C₁₈H₁₇NO₄: C, 69.44; H 5.50, N 4.50. Found: C 69.35, H 5.66, N 4.35.

(Z)-2-(3-Trifluoromethylbenzylidene)-4,6-dimethoxy-2,3-

dihydro-1H-indol-3-one (3k). Yellow solid, yield 77.4%, mp 160–162°C. ¹H NMR (400 MHz, CDCl₃) δ : 3.86 (s, 3H, OCH₃), 3.90 (s, 3H, OCH₃), 5.92 (d, J = 0.8 Hz, 1H, ArH), 6.11 (d, J = 0.8 Hz 1H, ArH), 6.69 (s, 1H, =CH), 7.10 (br s, 1H, NH), 7.49–7.54 (m, 2H, ArH), 7.64–7.69 (m, 1H, ArH), 7.70–7.73 (m, 1H, ArH). IR (KBr) v: 3444, 3036, 1683, 1599, 1471, 1386, 1328, 1279, 1219, 1160, 1117, 1070 cm⁻¹. *Anal.* Calcd for C₁₈H₁₄F₃NO₃: C 61.89, H 4.04, N 4.01. Found: C 61.65, H 4.31, N 4.24.

(Z)-2-(3,4-Dimethoxylbenzylidene)-4,6-dimethoxy-2,3dihydro-1H-indol-3-one (31). Yellow solid, yield 83.6%, mp 137–139°C. ¹H NMR (400 MHz, CDCl₃) δ : 3.83 (s, 3H, OCH₃), 3.88 (s, 3H, OCH₃), 3.89 (s, 3H, OCH₃), 3.91 (s, 3H, OCH₃), 5.90 (d, J = 1.6 Hz, 1H, ArH), 6.08 (d, J = 1.6 Hz, 1H, ArH), 6.68 (s, 1H, =CH), 6.87 (d, J = 8.0 Hz, 1H), 7.00 (d, J = 1.6 Hz, 1H, ArH), 7.06 (br s, 1H, NH), 7.12 (dd, J = 8.4, 1.6 Hz, 1H, ArH). ¹³C NMR (100 MHz, DMSO- d_6) δ : 55.4, 55.5, 55.6(2C), 88.5, 90.5, 103.3, 108.1, 111.9, 113.3, 122.6, 127.1, 134.2, 148.8, 148.9, 156.9, 159.4, 167.6, 181.3. IR (KBr) v: 3432, 3020, 1667, 1627, 1590, 1510, 1465, 1369, 1271, 1249, 1216, 1160, 1116, 1025 cm⁻¹. *Anal.* Calcd for C₁₉H₁₉NO₅: C 66.85, H 5.61, N 4.10. Found: C 66.95, H 5.33, N 3.86.

(Z)-2-(4-Hydroxy-3-methoxybenzylidene)-4,6-dimethoxy-2,3dihydro-1H-indol-3-one (3m). Yellow solid, yield 85.3%, mp 259–260°C. ¹H NMR (400 MHz, DMSO- d_6) δ : 3.81 (s, 3H, OCH₃), 3.84 (s, 3H, OCH₃), 3.85 (s, 3H, OCH₃), 5.97 (d, J = 2.0 Hz, 1H, ArH), 6.20 (d, J = 2.0 Hz, 1H, ArH), 6.39 (s, 1H, =CH), 6.84 (d, J = 8.4 Hz, 1H, ArH), 7.13–7.21 (m, 2H, ArH), 9.42 (s, 1H, OH), 9.51 (br s, 1H, NH). IR (KBr) v: 3363, 3015, 2921, 1665, 1625, 1577, 1513, 1470, 1445, 1372, 1296, 1252, 1219, 1198, 1153, 1119, 1042, 1019 cm⁻¹. Anal. Calcd for C₁₈H₁₇NO₅: C 66.05, H 5.23, N 4.28. Found: C 66.30, H 5.21, N 4.44.

Biological screening. The potential toxicity of the title compounds against HepG-2, Hela, and WRL-68 cells were performed using MTT assay [18] as follows: Cells were seeded into 96-multiwell plate (4 \times 10³ cells/well) and incubated for 24 h. Different concentrations of each test compound were added to the cells triplicate wells. The cells without test compounds were used as negative controls. Following the exposure of 48 h to the test compound, MTT solution (100 µL/well, 1 mg/mL) was added to each well and left for another 4 h. After removing MTT, dimethyl sulfoxide was added to the cell to dissolve the precipitate, and the absorbance (Abs) of each well was measured at 570 nm using the ELISA reader (Spectra MAX 190, Molecular Devices Corporation, USA). The inhibitory effect was calculated as follows: Inhibition $(\%) = (1 - Abs_{compound}/Abs_{control}) \times 100. \text{ IC}_{50} \text{ was}$ obtained for each test compound using SPSS 16.0 version (SPSS Inc., Chicago, USA).

Acknowledgments. We are very grateful for the financial support from the National Natural Science Foundation of China (no. 31501686), the Natural Science Foundation of Jiangsu Province (BK20140536), and the Jiangsu Government Scholarship for Overseas Studies (JS-2016-100).

REFERENCES AND NOTES

[1] Ono, E.; Fukuchi-Mizutani, M.; Nakamura, N.; Fukui, Y.; Yonekura-Sakakibara, K.; Yamaguchi, M.; Nakayama, T.; Tanaka, T.; Kusumi, T.; Tanaka, Y. Proc Natl Acad Sci U S A 2006, 103, 11075. [2] Kong, C.-H.; Xu, X.-H.; Zhang, M.; Zhang, S.-Z. Pest Manag Sci 2010, 66, 1018.

[3] Morimoto, M.; Fukumoto, H.; Nozoe, T.; Hagiwara, A.; Komai, K. J Agric Food Chem 2007, 55, 700.

[4] Zhang, M.; Xu, X.-H.; Cui, Y.; Xie, L.-G.; Kong, C.-H. Pest Manag Sci 2012, 68, 1512.

[5] Zhang, M.; Chen, G.-Y.; Li, T.; Liu, B.; Yang, L.-Q.; Xu, X.-H. J Heterocyclic Chem 2015, 52, 1887.

[6] Cheng, H.; Zheng, L.; Liu, Y.; Chen, S.; Cheng, H.; Lu, X.; Zheng, Z.; Zhou, G.-C. Eur J Med Chem 2010, 45, 5950.

[7] Sharma, M. C.; Sharma, S.; Sharma, P.; Kumar, A. Med Chem Res 2014, 23, 181.

[8] Lee, C.-Y.; Chew, E.-H.; Go, M.-L. Eur J Med Chem 2010, 45, 2957.

[9] Nenadis, N.; Sigalas, M. P. Food Res Int 2011, 44, 114.

[10] Bandgar, B. R.; Patil, S. A.; Korbad, B. L.; Biradar, S. C.; Nile, S. N.; Khobragade, C. N. Eur J Med Chem 2010, 45, 3223.

[11] Sutton, C. L.; Taylor, Z. E.; Farone, M. B.; Handy, S. T. Bioorg Med Chem Lett 2017, 27, 901.

[12] Sheng, R.; Xu, Y.; Hu, C.; Zhang, J.; Lin, X.; Li, J.; Yang, B.; He, Q.; Hu, Y. Eur J Med Chem 2009, 44, 7.

[13] Li, Y.; Qiang, X.; Yang, X.; Xiao, G.; Liu, Q.; Ai, J.; Tan, Z.; Deng, Y. Eur J Med Chem 2017, 126, 762.

[14] Okombi, S.; Rival, D.; Bonnet, S.; Mariotte, A. M.; Perrier, E.; Boumendjel, A. J Med Chem 2006, 49, 329.

[15] Haudecoeur, R.; Carotti, M.; Gouron, A.; Maresca, M.; Buitrago, E.; Hardre, R.; Bergantino, E.; Jamet, H.; Belle, C.; Reglier, M.; et al. ACS Med Chem Lett 2017, 8, 55.

[16] Souard, F.; Okombi, S.; Beney, C.; Chevalley, S.; Valentin, A.; Boumendjel, A. Bioorg Med Chem 2010, 18, 5724.

[17] Carrasco, M. P.; Machado, M.; Goncalves, L.; Sharma, M.; Gut, J.; Lukens, A. K.; Wirth, D. F.; Andre, V.; Duarte, M. T.; Guedes,

R. C.; et al. ChemMedChem 2016, 11, 2194.

[18] Bao, Y.-T.; Zhang, M.; Li, T.; Xiao, H.-F.; Zhao, T.; Xu, X.-H.; Yang, L. J Heterocyclic Chem 2016, 53, 637.

[19] Gerby, B.; Boumendjel, A.; Blanc, M.; Bringuier, P. P.; Champelovier, P.; Fortune, A.; Ronot, X.; Boutonnat, J. Bioorg Med Chem Lett 2007, 17, 208.

[20] Li, Y.; Qiang, X.; Luo, L.; Yang, X.; Xiao, G.; Liu, Q.; Ai, J.; Tan, Z.; Deng, Y. Eur J Med Chem 2017, 126, 762.

[21] Sugasawa, T.; Adachi, M.; Sasakura, K.; Kitagawa, A. J Org Chem 1979, 44, 578.

[22] Huang, X.-W.; Wang, Z.; Chen, Q.-L.; Sun, Y.-N.; Wang, C.-L.; Liu, Z.-L.; Liu, J.-L. Chin J Org Chem 2013, 33, 2565 (in Chinese).

[23] Lawson, M. A.; Mariotte, A.-M.; Boumendjel, A. Heterocycl Commun 2003, 9, 149.

[24] Sim, H. M.; Loh, K. Y.; Yeo, W. K.; Lee, C. Y.; Go, M. L. ChemMedChem 2011, 6, 713.

[25] Di Giovanni, S.; Borloz, A.; Urbain, A.; Marston, A.; Hostettmann, K.; Carrupt, P.-A.; Reist, M. Eur J Pharm Sci 2008, 33, 109.

SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section at the end of the article.