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A Highly Selective and Efficient Access to N-9 Alkylated Purine Derivatives via Radical S_{RN}1 Chemistry.

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Abstract: 6-Chloropurine and 2-amino-6-chloropurine were discovered to be efficient nucleophiles in S_{RN1} reactions with various gem halonitroalkane derivatives as substrates to give high yield of N-9 alkylated purine derivatives in a N-9/N-7 ratio greater than 95/5.

N-9 substitued 6-chloropurines are intermediates in the classical methodology for the synthesis of various 6-9 disubstitued purines from the commercially available 6-chloropurine and 2-amino-6-chloropurine¹. Substitution at N-9 is often, but not always selective, so that mixtures of N-9 and of unwanted N-7 substituted derivatives are formed. Therefore the interest for selective methods is constant² and has increased when purine derivatives became needed as potentially useful antiviral drugs³. This prompted us to explore new developments based upon radical SRN1 chemistry which presents original features.

 S_{RN1} reactions in which a substrate $R_1R_2C(NO_2)X$ undergoes replacement of the halide X (Cl, Br) by a nucleophile Nu⁻ are very efficient and generally not (or little) subject to steric hindrance, but in contrast to the great number of carbon-carbon bond forming reactions where carbanions are the nucleophilic reagents⁴, carbon-nitrogen bond formation is much less documented. Besides the preliminary report by N. Kornblum on amines⁵, the only relevant contributions are those of R. Bowman's group on nitroimidazoles⁶ and our's on imidazoles⁷. We report herein our first results on 6-chloropurine 1a and 2amino-6-chloropurine1b as nucleophiles in reactions with variously functionalized substrates (TABLE).

The photostimulated reactions between 6-chloropurine 1a dissolved in a mixture of DMSO/CH₃CN (1:2) (heterogeneous system)^{7,8} and the simple 2-bromo-2-nitropropane 2 or the monofunctionalized substrate⁹ 3 gave a single product 5 or 6 (N.M.R; thin layer chromatography). Such was not the case with 4a or 4b since either of them yielded a mixture of two compounds 7a and 7b which were separated by

^{†.} Deceased 02 August 1991.

Nucleophiles ^a	Substrates		time (min)	Products (yield%) ^b	
		${}^{\text{Br}}_{\text{R}_2} \times {}^{\text{R}_1}_{\text{NO}_2}$		$\mathbf{w}^{\mathbf{Z}} \mathbf{w}^{\mathbf{N}} \mathbf{w}^{\mathbf{N}} \mathbf{w}^{\mathbf{N}} \mathbf{w}^{\mathbf{N}} \mathbf{w}^{\mathbf{N}} \mathbf{w}^{\mathbf{R}_{1}} \mathbf{w}^{\mathbf{R}_{2}} \mathbf{w}^{\mathbf{R}_{2}} \mathbf{w}^{\mathbf{R}_{2}}$	
ia Z=Ci W=H	2 3	$\mathbf{R}_1 = \mathbf{R}_2 = \mathbf{C}\mathbf{H}_3$	180	5	(96)
		R ₁ = CH ₃ , R ₂ = CH ₂ OTHP	120	6 (diastereoisomers)	(91)
		Br H5C6~0~'NO2 4a	120	7a+7b (a/b:90/10) ^C	(60)
		NO2 H5C6 4b	90	7 a+7b (a/b:90/10) ^C	(50)
		Br H5C6~0~~~ NO2 4c	90	7a+7b (a/b:90/10) ^c	(81)
1b Z=Cl W=NH2	2	R ₁ = R ₂ = CH ₃	210	8	(88)
	3	R ₁ = CH ₃ , R ₂ = CH ₂ OTH	P 105	9 (diastereoisomers)	(95)

TABLE

a) 1.5 eq. Nu; 1.5 eq. K₂CO₃; DMSO/CH₃CN (1/2); illumination by HANAU 100W UV lamp. b) isolated pure products. c) ratio determinated from isolated pure products.

colum chromatography. A third experiment using 4c (mixture of cis/trans isomers) gave also a mixture of 7a and 7b in a ratio similar to that determined in reactions from pure 4a or 4b.

The quantitative conversion of 7a to the hypoxanthine derivative 10 (whose structure was unambigously determined by X Ray crystallography, FIG.) (scheme 1) indicates that *the substitution reaction was highly selective on* N-9 and the comparison of the NMR spectra of 6, 7a, 7b and 10unambigously shows that 6 is a diastereoisomeric mixture and that 7a and 7b are conformationally stable isomers.

Comparative examination of NMR data of 7a and 7b indicated that the 6-chloropurine substituent of the minor product 7b has *cis* equatorial configuration. In the chair conformation of the dioxane ring, the NO₂ group of the planar radical intermediate 4 is quasi equatorial and from the product distribution, one can assume that the attack from β face is favored over that from the α face in a ratio (9:1) which reflects the greater steric hindrance to the approach of 1a⁻ caused by two axial hydrogen atoms on carbon C-4 and C-6 (scheme 2).









We then turned our attention to the 2-amino-6-chloropurine 1b as a nucleophile in these reactions, since it is molecule of major importance for the syntheses of guanine derivatives. The S_{RN1} reaction carried out between the derived anion 1b⁻ and the substrates 2 and 3 gave the corresponding N-9 alkylated products 8 and 9 in high yield. Thus, the electron releasing 2-amino group does not affect the nucleophilic behaviour of 1b as compared with 1a.

Since neither of these purine derived anions were used before as nucleophiles in $S_{RN}1$ reactions, we have performed classical experiments supporting the four step mechanism (scheme 3) on a model reaction between 1 and 3.



For the same reaction time (120 min) it was observed that -i) In the dark, no substitution product was obtained, since illumination which initiates the chain by providing the activation energy for the monoelectronic transfer to $R_1R_2C(NO_2)X$ and formation of the radical anion $R_1R_2C(NO_2)X^{-1}$ was suppressed. -ii) Under illumination, but in presence of a catalytic amount of *para* dinitrobenzene (5%) acting as an electron trap towards the two radical anions $R_1R_2C(NO_2)X^{-1}$ and $R_1R_2C(NO_2)Nu^{-1}$ the yield of $R_1R_2C(NO_2)Nu$ was significantly diminished (16% instead of 91%). -iii) Under illumination, but in the presence of galvinoxyle which acts as a radical scavenger towards $R_1R_2C(NO_2)$, no substitution product was formed.

Conclusion. For the first time, 6-chloro **1a** or 2-amino-6-chloropurine **1b** are demonstrated to be efficient nucleophiles for S_{RN1} reaction with gem bromo-nitro derivatives leading to N-9 alkyled purines. Thus, radical S_{RN1} reactions which are highly regioselective and furthermore compatible with chlorine borne on C-6 of purines are promising for the synthesis of various adenine and guanine derivatives. The scope and limitations of this methodology are currently under active investigation.

Experimental Section:

Reagents. All chemical reagents and solvents were purchased from Aldrich (France) and were used without further purification.

General Methods. Melting point was obtained on a Reichert apparatus. The IR spectra were recorded on a Nicolet (205, FT-IR) spectrometer. The UV spectra were obtained on a Perkin-Elmer lambda 5 UV/VIS spectrophotometer. The mass spectra were recorded on a AEI.MS-50 (MS-EI) or EI.MS-9 (MS-CI) spectrometer. The proton and carbon NMR spectra were recorded on either a Bruker spectrometer 4.7 T (200 Mz) or 9.4 T (400 Mz). Chemical schifts are reported in δ units, parts per million (ppm) downfield from TMS for ¹H NMR ($\delta = 0$) and from CDCl₃ for ¹³C NMR ($\delta = 77.14$). Column chromatography utilized silica gel 60 (230-400 mesh) from E. Merck laboratories as the solid phase.

General procedure for the preparation of gem nitro-N-9-(6-chloro or 2-amino-6-chloropurine)alkanes:

A solution of **1a** or **1b** (1.5 eq) in a mixture of dry DMSO/CH₃CN (ratio 1:2 and 10 ml/mmol) was stirred with K_2CO_3 (1.5 eq) about 30 min under argon. The gem bromonitroalkane (1 eq), dissolved in 2 ml of DMSO/CH₃CN, was then added by syringe to the heterogeneous mixture. After exposure to 100W fluorescent light (HANAU), under argon (reaction monitored by TLC silica gel), the mixture was poured onto ice-water and extracted twice by CH₂Cl₂. The organic layer was dried and the solvent was removed. The crude product was purified by silica gel column chromatography (Ether/pentane/methanol: 70/25/5).

6-chloro-9-(2-nitro-propane-2-yl)-purine 5

Oil.

SM (CI): m/z 242, 244 (MH⁺), 195, 197 (MH⁺-HNO₂), 155,157 (6-chloro-purine+H)⁺.
¹H NMR (CDCl₃): δ (ppm) 2.54 (s, 6H), 8.31 (s,1H, H₈ purine), 8.67 (s, 1H, H₂ purine).
¹³C NMR(DMSO-D₆): δ (ppm) 151.8 (C2), 145.7 (C8), 97.1, 24.9.
Anal. Calc. for C₈H₈N₅ClO₂: C: 39.75; H: 3.31. Found: C: 39.91; H: 3.51.

6-chloro-[2-nitro-1-(2-oxy-tetrahydropyrannyl)-propane-2-yl]-purine 6.

1st diastereoisomer:

Oil

IR (CDCl₃): v (cm⁻¹) 1030, 1130, 1200, 1220, 1290, 1340, 1560 (NO₂), 1580, 2940.

SM (CI): m/z 342-344 (MH⁺), 295-297 (MH⁺- HNO₂), 155-157 (6-chloro purine), 85 (THP)⁺, 57. UV (CH₃OH): λ max (nm) 263.

¹H NMR (CDCl₃): δ (ppm) 1.33-1.90 (large, 6H), 2.55 (s, 3H), 3.55 (m, 1H), 3.83 (m, 1H), 4.60 (d, 1H, J=10Hz), 4,63 (broad s, 1H), 4.72 (d, 1H, J=10Hz), 8.66 (s, 1H, H₈ purine), 8.80 (s, 1H, H₂ purine).

¹³C NMR (CDCl₃): δ (ppm) 152.25 (C6), 151.95 (C2), 151.6 (C4), 144.4, (C8), 131.95 (C5), 100.2, 97.65, 69.8, 63.45, 30, 24.9, 21.5, 19.4.

2nd diastereoisomer:

Oil

IR (CDCl₃): v (cm⁻¹) 1030, 1130, 1200, 1220, 1290, 1340, 1560 (NO₂), 1580, 2940.

SM (IC): m/z 342-344 (MH⁺), 295-297 (MH⁺- HNO₂), 155-157 (6-chloro purine), 85 (THP)⁺, 57. UV (CH₃OH): λ max (nm) 263.

¹H NMR (CDCl₃): δ (ppm) 1.48-1.68 (large, 6H), 2.56 (s, 3H), 3.45-3.55 (m, 2H), 4.3 (d, 1H, J=10Hz), 4.78 (m, 1H), 4.88 (d, 1H, J=10Hz), 8.52 (s, 1H, H₈ purine), 8.75 (s, 1H, H₂ purine).
¹³C NMR (CDCl₃): δ (ppm) 151.95 (C2), 151.83 (C6),145.87 (C4), 144.15 (C8), 151.8 (C6), 98.95, 97.42, 69.07, 62.5, 29.9, 24.7, 21.4, 18.7.

6-Chloro-9-(5-nitro-2-phenyl-1,3-dioxanne-5-yl)purine:

Trans isomer 7a:

Oil.

IR (CDCl₃), v (cm⁻¹): 1123, 1220, 1311, 1410, 1484, 1565 (NO₂), 1590, 2887, 3134.

M.S.(IE): m/z: 316-314 (M⁺ -HNO₂), 270, 209, 157-155 (6-chloropurine), 105, 77.

¹ H NMR (CDCl₃,): δ (ppm) 4.93 (d, 2H, J= 13Hz, H ax.), 5.43 (d, 2H, J= 13Hz, H eq.), 5.83 (s, 1H, O-C<u>H</u>-O), 7.26 (br s, 5H, H ar), 8.63 (s, 1H, H₈ purine), 8.76 (s, 1H, H₂ purine).

¹³ C NMR (CDCl₃): δ (ppm) 152.47, 152.15, 151.6, 143.9, 135.1, 132.15, 130, 128.5, 126.05, 103.15, 69.45.

Anal. Calcd. for C₁₅ H₁₂ Cl N₅ O₄: C (49.81), H (3.34). Found: C (49.48), H (3.42).

Cis isomer 7b:

Oil.

IR (CDCl₃), v (cm⁻¹): 1134, 1233, 1296, 1388, 1441, 1569 (NO₂), 1595, 2874, 3107.

M.S.(EI): m/z: 316-314 (M⁺ -HNO₂), 270, 209, 157-155 (6-chloropurine), 105, 77.

¹ H NMR (CDCl₃): δ (ppm) 4.75 (d, 2H, J= 13Hz, H ax.), 5.8 (d, 2H, J= 13Hz, H eq.), 5.8 (s, 1H, O-C<u>H</u>-O), 7.45 (br s, 5H, H ar), 8.33 (s, 1H, H₈ purine), 8.8 (s, 1H, H₂ purine).

¹³ C NMR (CDCl₃): δ (ppm) 153, 151.7, 141.3, 135.3, 131.65, 130, 128.8, 126.35, 101.7, 89.85, 68.2.

2-Amino-6-chloro-9-(2-nitro-propane-2-yl)purine 8

m.p.:163°C.

IR (CDCl₃), v (cm⁻¹): 1275, 1567, 1620, 2975, 3425.

M.S. (IC): m/z 259-257 (MH+), 212-210 (MH+-HNO2), 172-170 (2-amino-6-chloropurine+H)+.

¹ H NMR (DMSO d6): δ (ppm) 2.36 (s, 6H), 7.05 (large s, 2H, NH₂), 8.5 (s, 1H, H₈ purine).

¹³ C NMR (DMSO d₆): δ (ppm) 159.5, 153.7, 150.15, 140.6, 123.75, 96.5, 24.85.

Anal. Calcd. for C₈H₉N₆O₂: C: 36.99; H: 3.47. Found: C: 37.06; H: 3.56.

2-Amino-6-chloro-9-[2-nitro-1-(2-oxy-tétrahydropyranyl)-propane-2-yl]purine 9

Oil.

1st diastereoisomer:

IR (CDCl3): v (cm⁻¹) 1020, 1060, 1110, 1120, 1160, 1280, 1320, 1490, 1560, 1600, 2925, 3000-3500, 3400, 3500.

SM (IC): m/z 359-357 (MH⁺), 314-312 (MH⁺- HNO₂), 172-170 (2-amino 6-chloropurine), 85 (THP)⁺

57.

UV (CH₃OH): λ max (nm) 254.

¹ H NMR (CDCl₃): δ (ppm) 1.33-1.76 (large, 6H), 2.43 (s, 3H), 3.6 (m, 1H), 3.83 (m, 1H),

4.48 (d, 1H, J=10Hz), 4.6 (d, 1H, J=10Hz and s, 1H), 5.26 (broad s, 2H, NH₂), 8.21 (s, 1H, H₈ purine). ¹³ C NMR (CDCl₃): δ (ppm) 158.95 (C2), 144.35 (C4), 140.95 (C8), 124.9 (C5), 100.2, 97.4, 69.85, 63.3, 30.15, 25.05, 21.45, 19.30.

Anal. Calcd. for C13H17CIN6O4: C: 43.51; H: 4.74. Found: C: 43.38; H: 4.65.

2nd diastereoisomer:

SM (IC): m/z 359-357 (MH⁺), 314-312 (MH⁺- HNO₂), 172-170 (2-amino 6-chloropurine), 85 (THP)⁺,

57.

UV (CH₃OH): λ max (nm) 251.

¹ H NMR (CDCl₃): δ (ppm) 1.33-1.75 (large, 6H), 2.43 (s, 3H), 3.5 (m, 2H), 4.23 (d, 1H, J=10Hz), 4.8 (d, 1H, J=10Hz and s, 1H), 5.26 (broad s, 1H, NH₂), 8.1 (s, 1H, H₈ purine).

¹³ C NMR (CDCl₃): δ (ppm) 159.1 (C2), 154.1 (C6), 151.95 (C4), 140.50 (C8), 125.25 (C5), 98.9, 97.2, 69.1, 62.35, 30, 24.9, 21.35, 18.7.

Synthesis of 6-isopropoxy-9-(5-nitro-2-phenyl-1,3-dioxanne-5-yl)purine 10:

Sodium hydride (50 mg, 2 mmol washed twice by pentane), and CH_2Cl_2 (10 ml) introduced in a pyrex flask caped with a rubber septum are cooled in an ice bath and stirred for 10 min. under argon. Isopropyl alcohol (154 µl, 2 mmol) is slowly added by syringe and the mixture is kept at room temperature for 15 min. The purine derivative **7a** (723 mg, 2 mmol) dissolved in CH_2Cl_2 (10 ml) is then introduced by syringe and stirring is maintained at room temperature for 1 hour. After classical work-up, the crude product is purified by silica gel colum chromatography ($CH_2Cl_2/MeOH : 95/5$) to give **8**, white solid (700 mg, 91%) slowly recrytallized for several days in pure MeOH to yield the monocrystal required for X-ray crystallography.

m.p.: 180°C.

IR (CDCl₃), v (cm⁻¹): 1113, 1155, 1250, 1315, 1470, 1566 (NO₂), 1600, 1610, 2915, 3050, 3075. M.S.(EI): m/z : 385 (M⁺), 339 (M⁺ –NO₂), 297, 234, 191.

¹ H NMR (CDCl₃): δ (ppm) 1.5 (d, 6H, J= 6Hz, (C<u>H</u>₃)₂CHO), 4.9 (d, 2H, J= 13Hz, H ax.), 5.46 (d, 2H, J= 13Hz, H eq.), 5.68 (m, 1H, J= 6Hz, pur-O-C<u>H</u>), 5.82 (s, 1H, O-C<u>H</u>-O), 7.36 (br s, 5H, H ar), 8.42 (s, 1H, H₈ purine), 8.5 (s, 1H, H₂ purine).

Anal. Calcd. for C₁₈ H₁₉ N₅ O₅: C (56.10), H (4.94). Found: C (56.08), H (4.73).

Crystal data: $C_{18}H_{19}N_5O_5$, M= monoclinic, space group C2/c, a= 21.411(9), b= 12.577(6), c= 15.032(7) Å, β = 112.88(7)°, U= 3729 Å³. Dc= 0.8*0.4*0.4 mm³. A Philips automatic diffractometer, graphite monochromator, λ = 0.7107 Å, ω = 0.025°. s⁻¹, larg. 1.4°, 3 reflexions /3hours. 3764 reflexions were recorded, of which 2856 unique reflexions were observed with I>3.0 σ (I).

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