ORIGINAL ARTICLE

Synthesis of 2-azaspiro[3.3]heptane-derived amino acids: ornitine and GABA analogues

Dmytro S. Radchenko · Oleksandr O. Grygorenko · Igor V. Komarov

Received: 18 November 2009/Accepted: 24 December 2009/Published online: 27 January 2010 © Springer-Verlag 2010

Abstract Synthesis of 6-amino-2-azaspiro[3.3]heptane-6-carboxylic acid and 2-azaspiro[3.3]heptane-6-carboxylic acid was performed. Both four-membered rings in the spirocyclic scaffold were constructed by subsequent ring closure of corresponding 1,3-bis-electrophiles at 1,1-*C*- or 1,1-*N*-bis-nucleophiles. The two novel amino acids were added to the family of the sterically constrained amino acids for the use in chemistry, biochemistry, and drug design.

Keywords Tailor-made amino acids · Spiro azetidines · Ornitine analogues · GABA analogues · Molecular rigidity

Introduction

The amino acids which possess sterically constrained molecular framework occupy a distinctive class of organic compounds which arouse much interest of chemists and biologists in the last decade. A number of tailor-made sterically constrained or rigid amino acids were synthesized to date (for recent reviews see Cativiela and Ordóñez 2009; Trabocchi et al. 2008; Komarov et al. 2004). The main driving force of the activity in this area is the quest for new drugs. In general, due to pre-organization of functional groups, the sterically constrained compounds can in

D. S. Radchenko · O. O. Grygorenko · I. V. Komarov Enamine Ltd., Aleksandra Matrosova Street, 23, Kyiv 01103, Ukraine

D. S. Radchenko · O. O. Grygorenko (⊠) · I. V. Komarov Department of Chemistry,
Kyiv National Taras Shevchenko University,
Volodymyrska Street, 64, Kyiv 01033, Ukraine
e-mail: gregor@univ.kiev.ua principle be much more efficient and selective ligands for various biological targets, thus displaying pronounced biological activity (Mann 2008). Recent studies showed that this view of the ligand-target interaction is too simplified (Martin 2007); nevertheless, molecular rigidity is widely regarded as one of the most important property of approved drugs (Feher and Schmidt 2003). Sterically constrained amino acids are especially popular for the design of peptidomimetic drugs. The principles of such design were formulated long ago (Shemyakin et al. 1969; Hruby et al. 1990; Cowell et al. 2004) and have been intensively used since then. As the result, a number of the approved drugs or clinical candidates contain residues of sterically constrained amino acids. It is interesting to note that many natural amino acids are sterically constrained; the Nature has been practicing the concept of rigidification of the biologically important molecules for millions of years.

Natural and synthetic rigid amino acids are either sterically congested or cyclic (polycyclic). This is illustrated in Fig. 1 by three amino acids which occur in Nature: 2-aminoisobutyric acid (1), 2-azetidinecarboxylic acid (2), and 2,4-methanoproline (3). Rigid spirocyclic scaffolds have great potential in providing wide variation of spatial disposition of the functional groups. However, spirocyclic aminoacids are very rare, both among natural and tailormade amino acids. There are only a few reports in the literature on spirocyclic analogues of natural amino acids (Weatherhead et al. 2009; Radchenko et al. 2008; Pellicciari et al. 2002; de Meijere et al. 1999; Tamm et al. 1999).

In this paper, we report syntheses of compounds **4** and **5**—2-azaspiro[3.3]heptane-derived amino acids, spirocyclic rigidified analogues of two natural amino acids, ornitine and γ -aminobutyric acid. To the best of our knowledge, spirocyclic analogues of these biologically important amino acids are not described in the literature to date. The



Fig. 1 Natural sterically constrained amino acids $(1\!-\!3)$ and compounds with 2-azaspiro[3.3]heptane scaffold $(4\!-\!6)$

2-azaspiro[3.3]heptane scaffold has recently captured attention of synthetic and medicinal chemists—compound **6** was suggested as an entry to structural surrogates of 4-substituted piperidines, building blocks for drug design (Meyers et al. 2009). It should also be noted that compounds **4** and **5** are non-chiral; the former being an example of a non-chiral analogue of the chiral natural amino acid, ornitine (for some other examples of non-chiral bicyclic amino acids see Avenoza et al. 2001; Rammeloo et al. 2002; Radchenko et al. 2009).

Materials and methods

General

Solvents were purified according to the standard procedures. (3,3-dimethoxycyclobutane-1,1-diyl)dimethanol 10 (Radchenko et al. 2008) and 2-phenyl-5,5-di(bromomethyl)-1,3-dioxane 16 (Allinger and Tushaus 1965) were prepared using the procedures reported in the literature. All other starting materials were purchased from Acros, Merck, and Fluka. Melting points were measured on MPA100 OptiMelt automated melting point system. Analytical TLC was performed using Polychrom SI F254 plates. Column chromatography was performed using Kieselgel Merck 60 (230-400 mesh) as the stationary phase. ¹H, ¹³C NMR, and all 2D NMR spectra were recorded on a Bruker 170 Avance 500 spectrometer (at 499.9 MHz for protons, 124.9 MHz for carbon-13) and Varian Unity Plus 400 spectrometer (at 400.4 MHz for protons, 100.7 MHz for carbon-13). Chemical shifts are reported in ppm downfield from TMS (¹H, ¹³C) as an internal standard. IR spectra were obtained on Nicolet Nexus 470 spectrometer. v_{max} (cm⁻¹) values in IR spectra are given for the main absorption bands. HPLC– MS analyzes were done on an Agilent 1100 LCMSD SL instrument [chemical ionization (CI)] and Agilent 5890 Series II 5972 GCMS instrument [electron impact ionization (EI)]. Elemental analyzes were performed on Elementar Vario MICRO Cube CHNS/O analyzer.

Methanesulfonic acid 1-methanesulfonyloxymethyl-3,3-dimethoxycyclobutylmethyl ester **11**

To a solution of 10 (4.81 g, 27.4 mmol) in chloroform (50 mL), triethylamine (11.46 mL, 82.2 mmol) was added. The resulting solution was cooled to -30° C, and mesyl chloride (5.09 mL, 65.8 mmol) was added dropwise. After the addition was complete, the reaction mixture was heated to ambient temperature, stirred for 1 h and washed with water (1 \times 25 mL), 10% aqueous citric acid (1 \times 25 mL) and brine $(1 \times 25 \text{ mL})$. Drying of solution with sodium sulfate and evaporation at reduced pressure led to 11 (9.10 g, 100%): mp 96°C (EtOH). MS (m/z, CI) 333 (MH⁺). Anal. calcd for $C_{10}H_{20}O_8S_2 C$ 36.14, H 6.06, S 19.29. Found C 36.19, H 6.31, S 19.04. IR (KBr, cm⁻¹): 1,355 ($v_{as}(SO_2)$), 1,174 ($v_s(SO_2)$). ¹H NMR (DMSO- d^6) δ 4.20 (s, 4H, CH₂OMs), 3.20 (s. 6H), 3.05 (s, 6H), 2.04 (s, 4H, C(CH₂)₂C); ¹³C NMR (DMSO- d^6) δ 98.7 (C(OMe)₂), 72.0 (CH₂OMs), 48.4, 37.1, 36.2, 32.6.

6,6-Dimethoxy-2-(toluene-4-sulfonyl)-2-azaspiro[3.3] heptane **12**

A solution of 11 (9.1 g, 27.4 mmol), potassium carbonate (18.9 g, 137 mmol) and tosyl amide (4.91 g, 28.7 mmol) in DMSO (50 mL) was heated at 85°C for 30 h. After cooling, water (100 mL) was added, and the product was extracted with dichloromethane $(3 \times 50 \text{ mL})$. The combined organic phases were washed with 10% aqueous citric acid (50 mL) and brine (3 \times 50 mL). Drying of the solution with sodium sulfate and evaporation in vacuo led to 12 (5.03 g, 59%) pure enough for the next step. Mp 114°C (EtOH). MS (m/z, CI) 312 (MH⁺), 280. IR (KBr, cm⁻¹): 1,339 ($v_{as}(SO_2)$), 1,280, 1,157 ($v_s(SO_2)$), 1,107, 1,042. ¹H NMR (DMSO- d^6) δ 7.67 (d, J = 8.0 Hz, 2H), 7.48 (d, J = 8.0 Hz, 2H), 3.64 (s, 4H, C(CH₂)₂N), 2.93 (s, 6H, C(OCH₃)₂), 2.42 (s, 3H, C₆H₅CH₃), 2.02 (s, 4H, C(CH₂)₂C); ¹³C NMR (DMSO- d^6) δ 144.4, 131.2, 130.4, 128.7, 99.2 (C(OMe)₂), 62.2, 48.6, 42.2, 28.8, 21.6.

2-(Toluene-4-sulfonyl)-2-azaspiro[3.3] heptan-6-one **13**

Compound **12** (3 g, 9.64 mmol) was mixed with 3 N HCl (100 mL) and stirred vigorously for 12 h. The solid formed was filtered and dried on air to yield **13** (2.41 g, 94%): mp

180–182°C (EtOH); MS (*m*/*z*) 266 (MH⁺). Anal. calcd for C₁₃H₁₅NO₃S C 58.85, H 5.70, N 5.28, S 12.08. Found C 58.99, H 5.45, N 5.03, S 11.87. IR (KBr, cm⁻¹): 1,785 (*v*(C=O)), 1,338 (*v*_{as}(SO₂)), 1,158 (*v*_s(SO₂)). ¹H NMR (DMSO-*d*⁶) δ 7.69 (d, J = 7.5 Hz, 2H), 7.49 (d, J = 7.5 Hz, 2H), 3.85 (s, 4H, C(CH₂)₂N), 3.07 (s, 4H, C(CH₂)₂C), 2.42 (s, 3H, C₆H₅CH₃); ¹³C NMR (DMSO-*d*⁶) δ 205.8, 144.5, 131.5, 130.4, 128.7, 61.5, 57.0, 27.8, 21.6.

2-(Toluene-4-sulfonyl)-2,7,9-triazadispiro[3.1.4.1]undecane-8,10-dione **14**

Ketone 13 (0.8 g, 3.02 mmol), potassium cyanide (0.3 g, 4.61 mmol) and ammonium carbonate (0.87 g, 9.05 mmol) were dissolved in a mixture of EtOH (8 mL) and water (3.3 mL). The mixture was stirred at 50°C for 2 days and acidified with 1 N HCl to pH = 1. The precipitate formed was filtered and dried to yield 0.62 g (61%) of 14 as a white solid: mp 241°C (EtOH); MS (m/z, CI) 336 (MH⁺), 311. Anal. calcd for C₁₅H₁₇N₃O₄S C 53.72, H 5.11, N 12.53, S 9.56. Found C 53.93, H 5.11, N 12.72, S 9.69. IR (KBr, cm^{-1}): 1,758 and 1,710 (v(C=O)), 1,402, 1,340 $(v_{as}(SO_2)), 1,161 (v_s(SO_2)).$ ¹H NMR (DMSO- d^6) δ 10.56 (s, 1H, NH), 8.1 (s, 1H, NH), 7.69 (d, J = 6.5 Hz, 2H), 7.48 (d, J = 6.5 Hz, 2H), 3.76 (br s, 2H), 3.66 (br s, 2H), 2.41 (s, 3H), 2.24 (m, 4H); ¹³C NMR (DMSO-d⁶) δ 178.6, 156.4, 144.5, 131.3, 130.4, 128.7, 63.0, 61.1, 57.6, 42.2, 30.2, 21.6.

6-amino-2-azaspiro[3.3]heptane-6-carboxylic acid 4

Compound 14 (200 mg, 0.57 mmol) was added to solid sodium amalgam (3.9 g) in methanol (15 mL). The reaction mixture was heated at reflux for 6 h. The solution was decanted from the liquid amalgam, and the residue was washed with methanol (10 mL). The solvent was evaporated in vacuo and the residue was dissolved in DME (4 mL). Boc₂O (0.68 g, 3.1 mmol), NEt₃ (0.25 mL, 1.79 mmol) and DMAP (2 mg, 0.016 mmol) were added in succession. Reaction mixture was stirred overnight at ambient temperature and concentrated in vacuo. The residue was taken up in CH₂Cl₂ (15 mL), washed with 1 N HCl (2 \times 5 mL), saturated aqueous Na₂CO₃ (1 \times 5 mL) and brine $(1 \times 5 \text{ mL})$, dried over Na₂SO₄, filtered and concentrated to yield 15 as a yellowish oil, 0.254 g (93%, >90% purity by ¹H NMR), which was used in the next step without further purification.

Compound **15** (0.25 g) was dissolved in DME (5 mL), and 1 N NaOH (5 mL) was added. The mixture was stirred overnight at ambient temperature. The resulting solution was partitioned between Et_2O (5 mL) and water (10 mL), the aqueous layer was washed with Et_2O (1 × 5 mL), acidified with 3 N HCl to adjust pH = 1 and stirred for 2 h. Water was evaporated in vacuo, the residue was purified by ion-exchange chromatography using Amberlite IR-120(plus) ion-exchange resin (25 g) as the stationary phase and 5% aqueous ammonia as an eluent yielding the amino acid **4** (45 mg, 54% from **15**): mp > 250°C, dec. (EtOH–H₂O). Anal. calcd for C₇H₁₂N₂O₂ C 53.83, H 7.74, N 17.94. Found C 53.99, H 7.95, N 18.04 IR (KBr, cm⁻¹): 3,479 and 3,430 (ν (NH₂)), 1,633 (ν _{as}(COO⁻)), 1,590 (δ (NH₂⁺), 1,397 (ν _s(COO⁻)). ¹H NMR (D₂O) δ 4.22 (s, 2H) 4.06 (s, 2H), 2.73 (d, J = 14.5 Hz, 2H), 2.53 (d, J = 14.5 Hz, 2H); ¹³C NMR (D₂O) δ 176.9, 58.3, 56.1, 53.7, 40.8, 34.3.

Diisopropyl 7-phenyl-6,8-dioxaspiro[3.5]nonane-2,2-dicarboxylate **17**

To a suspension of NaH (60% in mineral oil, 67 g, 1.67 mol) in dry DMF (700 mL), diisopropyl malonate (286 g, 1.52 mmol) was added dropwise (the temperature being maintained below 70°C) followed by 2-phenyl-5,5-di(bromomethyl)-1,3-dioxane 16 (266 g, 0.76 mol). The resulting reaction mixture was heated at 140°C for 15 h. After cooling, saturated NH₄Cl solution (1.5 L) was added, and the product was extracted with hexanes $(3 \times 500 \text{ mL})$. The combined organic extracts were dried over sodium sulfate and concentrated in vacuo. A crystalline product formed was separated from a liquid residue by filtration, washed with hexane $(2 \times 100 \text{ mL})$ and dried to yield pure 17 (195 g, 72%) as a white solid: mp 90°C (Hexanes); MS (m/z, EI): 375 (M⁺–H), 333, 317, 291. Anal. calcd. for $C_{12}H_{14}Br_2O_2$ C 41.17, H 4.03, Br 45.65. Found C 41.32, H 3.89, Br 45.58. IR (KBr, cm⁻¹): 1,740, 1,720 (v(C=O)). ¹H NMR (CDCl₃) δ 7.47 (m, 2H), 7.36 (m, 3H), 5.41 (s, 1H, PhCH), 5.07 (m, J = 6.5 Hz, 2H, CH(CH₃)₂), 4.15 (d, J = 11 Hz, 2H), 3.75 (d, J = 11 Hz, 2H), 2.76 (s, 2H), 2.19 (s, 2H), 1.25(d, 6.5 Hz); 13 C NMR (CDCl₃) δ 171.3, 138.1, 129.0, 128.3, 126.1, 101.4, 75.4, 69.1, 47.3, 37.5, 31.6, 31.3, 21.6.

Diisopropyl 3,3-bis(hydroxymethyl)cyclobutane-1,1dicarboxylate **18**

To a solution of **17** (10 g, 26.6 mmol) in MeOH (100 mL), 10% Pd/C (2 g) was added, and the resulting suspension was hydrogenated at 5 atm and ambient temperature upon stirring for 48 h. Filtration of the catalyst and removal of the solvent in vacuo yielded **18** (7.4 g, 96%) as a colorless oil which was used in the next step without further purification. MS (*m*/*z*, CI) 289 (MH⁺). ¹H NMR (CDCl₃) δ 5.04 (m, *J* = 6.0 Hz, 2H, CH(CH₃)₂), 3.70 (br. s, 4H, CH₂OH), 2.66 (br. s, 2H, CH₂OH), 2.37 (s, 4H, C(CH₂)₂C), 1.22 (d, *J* = 6.0 Hz, 12H, CH(CH₃)₂); ¹³C NMR (CDCl₃) δ 171.6 (COO*i*Pr), 69.2, 68.5, 47.2 (C(COO*i*Pr)₂), 37.9 (C(CH₂OH)₂), 32.7 (C(CH₂)₂C), 21.5 (CH(CH₃)₂). Diisopropyl 3,3-bis(((methylsulfonyl)oxy) methyl)cyclobutane-1,1-dicarboxylate **19**

To a solution of 18 (6.96 g, 24 mmol) in dichloromethane (100 mL), triethylamine (10 mL, 72 mmol) was added. The resulting mixture was cooled to -30° C, and mesyl chloride (4.49 mL, 58 mmol) was added dropwise. After the addition was complete, the reaction mixture was heated to ambient temperature and stirred for 1 h, then washed with water $(1 \times 50 \text{ mL})$, 10% aqueous citric acid $(1 \times 50 \text{ mL})$ and brine $(1 \times 50 \text{ mL})$. The organic phase was dried over sodium sulfate and evaporated at reduced pressure to give dimesylate 19 (10.7 g, 100%) as a white solid. Mp 87°C (EtOH). MS (m/z, CI) 445 (MH⁺). Anal. calcd for C₁₆H₂₈O₁₀S₂ C 43.23, H 6.35, S 14.43. Found C 43.07, H 6.18, S 14.51. IR (KBr, cm^{-1}): 1,720 $(v(C=O)), 1,365 (v_{as}(SO_2)), 1,274, 1,176 (v_s(SO_2)).$ ¹H NMR (CDCl₃) δ 5.07 (sept, J = 6.0 Hz, 2H, CH(CH₃)₂), 4.28 (s, 4H, CH₂OSO₂CH₃), 3.06 (s, 6H, CH₂OSO₂CH₃), 2.51 (s, 4H, C(CH₂)₂C), 1.25 (d, J = 6.0 Hz, 12H, CH(CH₃)₂); ¹³C NMR (CDCl₃) δ : 170.6 (COO*i*Pr), 70.5, 69.7, 37.4, 35.7, 32.2, 21.5 (CH(CH₃)₂).

Diisopropyl 2-((4-methylphenyl)sulfonyl)-2azaspiro[3.3]heptane-6,6-dicarboxylate **20**

A solution of 19 (9.17 g, 20.6 mmol), potassium carbonate (14.2 g, 103 mmol), and tosylamide (3.7 g, 21.7 mmol) in DMSO (50 mL) was heated at 85°C for 12 h. After cooling, water (100 mL) was added, and the product was extracted with EtOAc (3 \times 50 mL). The combined organic phases were washed with 10% aqueous citric acid (50 mL) and brine $(3 \times 50 \text{ mL})$, dried over sodium sulfate and evaporated in vacuo to yield 20 (8.3 g, 81%) pure enough for the next step: mp 70°C (EtOH); MS (m/z, CI): 424 (MH^+) , 382, 340. IR (KBr, cm⁻¹): 1,744, 1,722 (v(C=O)), 1,341 ($v_{as}(SO_2)$), 1,279, 1,159 ($v_s(SO_2)$), 1,102, 1,080. ¹H NMR (CDCl₃) δ 7.70 (d, J = 7.5 Hz, 2H), 7.35 (d, J = 7.5 Hz, 2H), 4.99 (sept, J = 6.0 Hz, 2H, CH(CH₃)₂), 3.74 (s, 4H, C(CH₂)₂N), 2.61 (s, 2H, CH₂), 2.49 (s, 3H, $C_6H_4CH_3$), 2.45 (s, 2H, CH₂), 1.19 (d, J = 6 Hz, 12H, CH(CH₃)₂); ¹³C NMR (CDCl₃) δ 170.6, 144.1, 131.6, 129.7, 128.4, 69.2, 62.1, 48.3, 41.0, 39.0, 32.5, 21.6, 21.5.

2-((4-methylphenyl)sulfonyl)-2-azaspiro[3.3]heptane-6,6-dicarboxylic acid **21**

To a solution of **20** (0.824 g, 1.95 mmol) in ethanol (5 mL), a solution of sodium hydroxide (0.26 g, 6.5 mmol) in ethanol (5 mL) was added. The resulting mixture was refluxed for 2 h, then cooled, filtered and washed with ethanol (2 \times 2 mL) to give dipotassium salt of **21**. It was dissolved in water (5 mL), and the solution was acidified

with 2 N HCl to pH = 2. Extraction of the resulting mixture with diethyl ether (3 × 5 mL), drying of the combined organic extracts over magnesium sulfate and evaporation of the solvent afforded dicarboxylic acid **21** (0.614 g, 93%) as a white solid: mp 231°C (EtOH); MS (*m*/*z*): 340 (MH⁺). Anal. calcd for C₁₅H₁₇NO₆S C 53.09, H 5.05, N 4.13, S 9.45. Found C 52.97, H 5.12, N 4.01, S 9.77. IR (KBr, cm⁻¹): 2,860–3,500 (*v*(O–H)), 1,708 (*v*(C=O)), 1,338 (*v*_{as}(SO₂)), 1,160 (*v*_s(SO₂)). ¹H NMR (DMSO-*d*⁶) δ 12.77 (br s, 2H, COOH), 7.68 (d, *J* = 7.5 Hz, 2H), 7.48 (d, *J* = 7.5 Hz, 2H), 3.65 (s, 4H, C(CH₂)₂N), 2.42 (s, 3H, C₆H₄CH₃), 2.31 (s, 4H, C(CH₂)₂C); ¹³C NMR (DMSO-*d*⁶) δ 172.9, 144.4, 131.6, 130.3, 128.6, 62.5, 48.0, 38.9, 32.5, 21.54.

2-[(4-methylphenyl)sulfonyl]-2-azaspiro[3.3]heptane-6-carboxylic acid **22**

Dicarboxylic acid 21 (1.58 g, 4.66 mmol) was dissolved in pyridine (20 mL) and heated at reflux for 8 h. Then most of the solvent was evaporated in vacuo, 2 N HCl was added to adjust pH = 1, and the product was extracted with diethyl ether $(3 \times 20 \text{ mL})$. The combined organic extracts were dried over magnesium sulfate and evaporated in vacuo to give 22 (1.29 g, 94%) as a white solid: mp 136°C (EtOH); MS (m/z, CI) 296 (MH⁺). Anal. calcd for C₁₄H₁₇NO₄S C 56.93, H 5.80, N 4.74, S 10.86. Found C14H17NO4S C 56.76, H 5.83, N 4.59, S 11.00. IR (KBr, cm⁻¹): 2,560– 3.070 (v(O–H)), 1,705 (v(C=O)), 1,343 (v_{as}(SO₂)), 1,158 $(v_{s}(SO_{2}))$. ¹H NMR (DMSO-d⁶) δ 12.10 (br s, 1H, COOH), 7.67 (d, J = 8.0 Hz, 2H), 7.47 (d, J = 8.0 Hz, 2H), 3.67 (s, 2H, C(CH₂)(CH₂)N), 3.59 (s, 2H, C(CH₂)(CH₂)N), 2.78 (m, 1H, CHCOOH), 2.42 (s, 3H, C₆H₄CH₃), 2.05 (m, 4H, CH(CH₂)₂C); ¹³C NMR (DMSO- d^6) δ 176.1, 144.4, 131.4, 130.3, 128.7, 62.7, 62.2, 35.2, 34.3, 32.4, 21.6.

2-azaspiro[3.3]heptane-6-carboxylic acid 5

Carboxylic acid **22** (1.29 g, 4.37 mmol) was dissolved in methanol (100 mL), and solid sodium amalgam (15 g) was added to the solution in one portion. The reaction mixture was refluxed for 8 h and then cooled. The solution was decanted from the liquid amalgam, and the residue was washed with methanol (30 mL). The solvent was evaporated in vacuo, the residue was dissolved in water, and 3 N HCl was added to adjust pH = 1. The resulting aqueous solution was washed with diethyl ether and evaporated. Purification of the residue by ion-exchange chromatography (Amberlite IR-120(plus) ion-exchange resin (50 g), 5% aqueous ammonia as an eluent) yielded amino acid **5** (0.48 g, 77%): mp 297°C, dec. (EtOH–H₂O). Anal. calcd for C₇H₁₁NO₂ C 59.56, H 7.85, N 9.92. Found C 59.65, H 7.88, N 10.01. IR (KBr, cm⁻¹): 3,459 (ν (N–H)), 1,621



Scheme 2 Synthesis of amino acid 4

Scheme 1 Retrosynthetic

 $(v_{as}(COO^{-})), 1,569 (\delta(NH_{2}^{+}), 1,394 (v_{s}(COO^{-}))).$ ¹H NMR (D₂O) δ 4.07 (s, 2H, 1-CH₂), 3.96 (s, 2H, 3-CH₂), 2.80 (quint, J = 8.5 Hz, 1H, CHCOO), 2.39 (t, J = 11.0 Hz, 2H, 5-CHH and 7-CHH), 2.26 (t, J = 11.0 Hz, 2H, 5-CHH and 7-CHH); ¹³C NMR (D₂O) δ 183.9 (COO), 58.0 (1-CH₂), 57.2 (3-CH₂), 36.8, 35.8, 34.6.

Results and discussion

The construction of the 2-azaspiro[3.3]heptane core in both 4 and 5 was done by consequent closure of the cyclobutane and azetidine rings using bis-alkylation of malonate and tosylamide, respectively. Nevertheless, the origin of the bis-electrophile necessary for the azetidine ring construction was different. Whereas in the case of 4 the corresponding $C(CH_2X)_2$ unit originated from the malonate, in the case of 5 the fragment was already present in the starting molecule. Thus, isomeric dibromides 7 and 8 were suggested to be the appropriate starting compounds for the synthesis of 4 and 5, respectively (Scheme 1).

The syntheses are outlined in the Schemes 2 and 3. To obtain the ornitine analogue 4, the dibromide 7 was transformed to dimesylate 11 analogously to the method reported previously by our group (Radchenko et al. 2008). Reaction of 11 with tosyl amide under mild basic conditions resulted in the construction of 2-azaspiro[3.3]heptane ring system. Deprotection of 12 led to the formation of ketone 13^{1} which underwent Bucherer–Lieb reaction giving the dispirocyclic hydantoine 14. Hydrolysis of 14 by heating with excess of the strong alkali was unsuccessful and led to the formation of many unidentified by-products, probably due to the azetidine ring opening. On other hand, relatively mild condition of the hydrolysis (heating to 60°C with 3.5 equivalents of 10% aqueous NaOH) resulted in no reaction. Therefore, we transformed 14 to the tris-Bocderivative 15; compounds of that type are known to hydrolyze under mild conditions (Wysong et al. 1996). Indeed, two-step hydrolysis of 15 afforded the amino acid 4

¹ While the manuscript was in the preparation, an alternative synthesis of 6-oxo-2-azaspiro[3.3]heptane derivative was reported (Meyers et al. 2009).





which was isolated by ion-exchange chromatography (Scheme 2).

In the synthesis of the GABA analogue 5, the dibromide 16, which is available commercially in bulk quantities, was used to alkylate diisopropyl malonate to form the spiro-1,3-dioxane 17. Deprotection of the diol moiety followed by bis-mesylation set up the stage for the azetidine ring formation. In the next step, 2-azaspiro[3.3]heptane ring system was constructed in a manner analogous to the transformation of 11-12 discussed above. Compound 20 thus obtained underwent the classic reaction sequence in malonate chemistry, hydrolysis followed by decarboxylation, leading to the formation of the carboxylic acid 22. Pyridine was used as the solvent for the decarboxylation, as in this case the decarboxylation conditions were mild. Finally, detosylation of 22 yielded the amino acid 5 which was isolated by ion-exchange chromatography (Scheme 3).

Conclusions

The use of 2-azaspiro[3.3]heptane scaffold in construction of functionalized sterically constrained amino acids was demonstrated for the first time. The synthesized compounds, 6-amino-2-azaspiro[3.3]heptane-6-carboxylic acid and 2-azaspiro[3.3]heptane-6-carboxylic acid are analogues of the natural compounds (ornitine and GABA, respectively) which play important roles in biological processes. Therefore, they can be advantageously used in medicinal chemistry and drug design.

References

- Allinger NL, Tushaus LA (1965) Conformational Analysis. XLIII. Stereochemical studies in the cyclobutane ring system. J Org Chem 65:1945–1951. doi:10.1021/jo01017a057
- Avenoza A, Cativiela C, Busto JH, Fernández-Recio MA, Peregrina JM, Rodrígues F (2001) New synthesis of 7-azabicyclo[2.2.1]heptane-1-carboxylic acid. Tetrahedron 57:545–548. doi:10.1016/S0040-4020(00)01023-1
- Cativiela C, Ordóñez M (2009) Recent progress on the stereoselective synthesis of cyclic quaternary α-amino acids. Tetrahedron Asymmetry 20(1):1–63. doi:10.1016/j.tetasy.2009.01.002
- Cowell SM, Lee YS, Cain JP, Hruby VJ (2004) Exploring Ramachandran and chi space: conformationally constrained amino acids and peptides in the design of bioactive polypeptide ligands. Curr Med Chem 11(21):2785–2798. doi:10.2174/ 0929867043364270
- de Meijere A, Ernst K, Zuck B, Brandl M, Kozhushkov SI, Tamm M, Yufit DS, Howard JAK, Labahn T (1999) Cyclopropyl building blocks for organic synthesis, 53. Convenient syntheses of novel α- and β-amino acids with spiropentyl groups. Eur J Org Chem 1999(11):3105–3115. doi:10.1002/(SICI)1099-0690(199911)1999: 11<3105::AID-EJOC3105>3.0.CO;2-1
- Feher M, Schmidt JM (2003) Property distributions: differences between drugs, natural products, and molecules from combinatorial chemistry. J Chem Inf Comput Sci 43(1):218–227. doi: 10.1021/ci0200467
- Hruby VJ, Al-Obeidi F, Kazmierski W (1990) Emerging approach in the molecular design of receptor-selective peptide ligands: conformational, topographical and dynamic considerations. Biochem J 268:249–262 http://www.biochemj.org/bj/268/0249/ bj2680249_browse.htm
- Komarov IV, Grigorenko AO, Turov AV, Khilya VP (2004) Conformationally rigid cyclic α-amino acids in the design of peptidomimetics, peptide models and biologically active compounds. Russ Chem Rev 73:785–810. doi:10.1070/RC2004v 073n08ABEH000912
- Mann A (2008) Conformational restriction and/or steric hindrance in medicinal chemistry. In: Wermuth CG (ed) Practice of medicinal

chemistry, 3rd edn. Academic Press/Elsevier, Amsterdam, pp 363–379 ISBN: 0123741947

- Martin SF (2007) Preorganization in biological systems: are conformational constraints worth the energy? Pure Appl Chem 79(2):193–200. doi:10.1351/pac200779020193
- Meyers MJ, Muizebelt I, van Wiltenburg J, Brown DL, Thorarensen A (2009) Synthesis of *tert*-butyl 6-oxo-2-azaspiro[3.3]heptane-2carboxylate. Org Lett 11(16):3523–3525. doi:10.1021/ol901325s
- Pellicciari R, Marinozzi M, Camaioni E, del Carmen Nùnez M, Costantino G, Gasparini F, Giorgi G, Macchiarulo A, Subramanian N (2002) Spiro[2.2]pentane as a dissymmetric scaffold for conformationally constrained analogues of glutamic acid: focus on racemic 1-aminospiro[2.2]pentyl-1, 4-dicarboxylic Acids. J Org Chem 67(16):5497–5507. doi:10.1021/jo020138v
- Radchenko DS, Grygorenko OO, Komarov IV (2008) Synthesis of conformationally restricted glutamic acid analogues based on the spiro[3.3]heptane scaffold. Tetrahedron Asymmetry 19:2924– 2930. doi:10.1016/j.tetasy.2008.12.016
- Radchenko DS, Kopylova N, Grygorenko OO, Komarov IV (2009) Conformationally restricted nonchiral pipecolic acid analogues. J Org Chem 74:5541–5544. doi:10.1021/jo900842w
- Rammeloo T, Stevens CV, De Kimpe N (2002) Synthesis of 2,4-methanoproline analogues via an addition-intramolecular

substitution sequence. J Org Chem 67:6509-6513. doi:10.1021/ jo025897s

- Shemyakin MM, Ovchinnikov YA, Ivanov VT (1969) Topochemical investigations on peptide systems. Angew Chem Int Ed 8(7):492–499. doi:10.1002/anie.196904921
- Tamm M, Thutewohl M, Ricker CB, Bes MT, de Meijere A (1999) Cyclopropyl building blocks in organic synthesis, 51. An easy access to 1-azaspiropentane-2-carboxamides—the first derivatives of a new type of amino acids. Eur J Org Chem (9):2017– 2024. doi:10.1002/(SICI)1099-0690(199909)1999:9<2017:: AID-EJOC2017>3.0.CO;2-0
- Trabocchi A, Scarpi D, Guarna A (2008) Structural diversity of bicyclic amino acids. Amino acids 34(1):1–24. doi:10.1007/ s00726-007-0588-y
- Weatherhead RA, Carducci MD, Mash EA (2009) Synthesis of conformationally constrained diaminodicarboxylic acid derivatives. J Org Chem 74:8773–8778. doi:10.1021/jo901892d
- Wysong CL, Yokum TS, Morales GA, Gundry RL, McLaughlin ML, Hammer RP (1996) 4-Aminopiperidine-4-carboxylic acid: a cyclic α,α-disubstituted amino acid for preparation of watersoluble highly helical peptides. J Org Chem 61:7650–7651. doi: 10.1021/jo961594k