Nucleobases

A Purine-like Nickel(II) Base Pair for DNA**

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Nucleic acids rely on complementary functionalized purine and pyrimidine heterocycles to encode genetic information as G·C and A·T(U) base pairs. Several approaches have been developed to expand the number of available base pairs beyond the two natural pairs, including the use of nonstandard hydrogen-bond donor/acceptor patterns,^[1] van der Waals and hydrophobic interactions,^[2] and metal coordination.^[3] Herein we report the realization of a naturally inspired metallo base pair with a purine core whose design derives from minimal modification of adenine. The resulting base pair (Pur^p·Ni·Pur^p, Figure 1) is found to have the following novel features and properties: 1) greater stability than a G·C base pair, 2) a surprising dimensional resemblance to natural purine–pyrimidine base pairs, and 3) the potential to serve in an ion-activated switch.



Figure 1. Left: The 6-(2'-pyridyl)-purine (Pur^p) metallo base pair. Right: A representation of a hypothetical helix composed of purely $Pur^{p} \cdot Ni^{2+} \cdot Pur^{p}$ base pairs.

Formally, Pur^{P} is derived from adenine by replacing the 6amino group with a pyridyl group. This functional-group interchange places two Lewis basic donor atoms (the purine N1 and pyridine N1' atoms) in an optimal 1,4-relationship for coordinating metal ions. Scheme 1 summarizes the synthesis of Pur^P. The key transformation involved a modified Negishi coupling of pyridyl zinc bromide with chloropurine deoxyriboside $\mathbf{1}^{[4]}$ to provide pyridylpurine deoxyriboside **2**. We

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Scheme 1. Synthesis of 2'-deoxyribosyl-N9-[6-(2'-pyridyl)-purine] phosphoramidite **3.** Tol = 4-toluoyl, THF = tetrahydrofuran, DMT = 4,4'-dimethoxytriphenylmethyl, Pyr = pyridine, OCE = cyanoethyl, DIPEA = N, N-diisopropylethylamine.

initially used the conditions reported for the preparation of ribosyl and acyclic analogues of $2^{[5]}$ but found that an improved yield was possible by switching to a [PdCl₂(PPh₃)₂]catalyst.^[6] Pyridylpurine nucleoside **2** was transformed in three steps into phosphoramidite **3**. DNA containing Pur^P was prepared by using **3** and standard phosphite triester methodology on an ABI394 synthesizer. Complementary dodecamer DNA strands were prepared, each bearing a single Pur^P residue: 5'-d-CTTTCTPur^PTCCCT (**4**) and 5'-d-AGGGAPur^PAGAAAG (**5**). These oligomers were purified by PAGE, and their identities were confirmed by MALDI mass spectrometry.

To assess the viability of Pur^P as the organic component of a metallo base pair, UV-monitored thermal denaturation of complementary dodecamers 4 and 5 bearing single Pur^P residues was performed in the presence of the divalent ions noted in Table 1. Representative denaturation profiles are shown in Figure 2. In all, nine divalent metal ions were screened for their abilities to coordinate to the Pur^P·Pur^P site contained in the 4/5 duplex by measuring the $T_{\rm m}$ value; the $T_{\rm m}$ value of a metal-free control with the same duplex (Table 1, entry 11) was also recorded. As is apparent from the table, only four of the seven divalent ions gave $T_{\rm m}$ values for 4/5 that differed significantly from the metal-free control: Ni²⁺, Co²⁺, Cu²⁺, Zn²⁺, and Ag⁺. Of these five metals, Ni²⁺ is the most stabilizing. Pur^P·Co²⁺·Pur^P leads to a duplex $T_{\rm m}$ value roughly in between those of the 6/10 and 7/11 duplexes bearing T·A and C·G pairs, respectively. More significantly, the Pur^P·Ni²⁺· Pur^P base pair is more stabilizing to a double helix than a C·G base pair by a margin of 6 °C under the conditions reported in Table 1, with 5 μ M NiCl₂. Importantly, the Pur^P·Pur^P 4/5 duplex is highly destabilized in the absence of Ni²⁺ or other **Table 1:** DNA-duplex melting temperatures, T_m , in the presence and absence of divalent ions.^[a]

5'-d-CTTTCT**X**TCCCT

3'-d-GAAAGA**Y**AGGGA

Entry	Х·Ү	Duplex	Metal	T _m	$\Delta^{[b]}$
1	Pur [₽] ·Pur [₽]	4/5	NiCl ₂	46.1	+6.0
2	Pur [₽] ∙Pur [₽]	4/5	Ni(NO ₃) ₂	46.6	+6.5
3	Pur [₽] ∙Pur [₽]	4/5	CoCl ₂	38.8	-1.3
4	Pur [₽] ∙Pur [₽]	4/5	CuCl ₂	31.4	-8.7
5	Pur [₽] ∙Pur [₽]	4/5	ZnSO₄	30.8	-9.3
6	Pur [₽] ∙Pur [₽]	4/5	AgNO₃	30.5	-9.6
7	Pur [₽] ∙Pur [₽]	4/5	FeSO₄	28.8	-11.3
8	Pur [₽] ∙Pur [₽]	4/5	MnCl ₂	29.2	-10.9
9	Pur [₽] ∙Pur [₽]	4/5	Eu(NO ₃) ₃	29.1	-11.0
10	Pur [₽] ∙Pur [₽]	4/5	$Pd(NO_3)_2$	27.3	-12.8
11	Pur [₽] ∙Pur [₽]	4/5	_[c]	28.5	-11.6
12	T∙Pur [₽]	6/5	NiCl ₂	27.1	-13.0
13	C∙Pur [₽]	7/5	NiCl ₂	26.6	-13.5
14	A∙Pur [₽]	8/5	NiCl ₂	27.0	-13.1
15	G∙Pur [₽]	9/5	NiCl ₂	29.1	-11.0
16	Τ·Α	6/10	_[c]	36.8	-3.3
17	T·A	6/10	NiCl ₂ ^[d]	37.4	-2.7
18	T·A	6/10	CoCl ₂ ^[d]	36.7	-3.4
19	C∙G	7/11	_[c]	40.2	+0.1
20	C·G	7/11	NiCl ₂ ^[d]	40.1	0.0
21	C∙G	7/11	CoCl ₂ ^[d]	39.9	-0.2

[a] Samples contained 2.5 μm of each DNA strand, 5 μm divalent ion where indicated, 50 mm NaCl, and 10 mm NaH₂PO₄ (pH 7.0). All measurements were performed at least in triplicate. [b] Difference in T_m value relative to that of duplex **7/11** (X·Y=C·G) in the presence of NiCl₂ (entry 20). [c] No divalent metal ions were added. [d] Divalent ion was added in these cases as a control.



Figure 2. Absorbance versus temperature denaturation profiles. Conditions are as reported in Table 1.

divalent ions (Table 1, entry 11): there is a -17.6 °C difference in $T_{\rm m}$ value between the Pur^P·Ni²⁺·Pur^P ($T_{\rm m} = 46.1$ °C) and Pur^P·Pur^P ($T_{\rm m} = 28.5$ °C) base pairs (Table 1, entry 1 versus entry 11). As a control, both the T·A **6/10** and the C·G **7/11** duplexes were denatured in the presence and absence of Ni²⁺ or Co²⁺, and no significant changes were observed in the $T_{\rm m}$ values (Table 1, entries 16–21).

The stability of mismatches between the four natural bases and $\mbox{Pur}^{\mbox{P}}$ was investigated in the presence of \mbox{Ni}^{2+}

(Table 1, entries 12–15). Clearly, none of the natural bases make stable pairs with Pur^P·Ni²⁺. Indeed, the $\Delta T_{\rm m}$ values of the mismatched pairs relative to the Pur^P·Ni²⁺·Pur^P base pair (not the Δ values listed in Table 1 which are relative to the C·G pair) range from -17 to -19.5 °C. By contrast, T·G and C·A mismatches of the parent natural duplex under the same conditions show $\Delta T_{\rm m}$ values of -7.4 and -18.5 °C under the same conditions.^[7] Thus, all four Pur^P·Ni²⁺ mismatches with natural bases are much less stable than the Pur^P·Ni²⁺·Pur^P match. A further point to note is that the instabilities seen for all four Pur^P·Ni²⁺ mismatches with natural bases rival the most severe natural nucleobase mismatches such as C·A.

Within the double helix, three possible geometries could be envisioned for divalent metal ion complexation by Pur_2^P : square planar, tetrahedral, and D_2^d (a geometry intermediate between square planar and tetrahedral). In the absence of a geometric preference by the metal ion, the most productive geometry for a metallo base pair in forming the double helix is expected to be square planar because this will maximize favorable nearest-neighbor stacking interactions. Low-energy square-planar geometries should be accessible for Ni²⁺, Co²⁺, Cu²⁺, Ag⁺, and Pd²⁺ ions. The first two of these five ions appreciably stabilize the Pur_2^P -bearing helix, whereas the latter three do not. As a result, it may be concluded that geometry alone is an insufficient predictor of metal-ion affinity for Pur^P . A circular dichroism spectrum of duplex 4/5 in the presence of Ni²⁺ is consistent with a B-DNA structure.

To assess the viability of a square-planar geometry for Pur^P·Ni²⁺·Pur^P, an ab initio geometry optimization was performed on the complex by using Gaussian 98^[8] at the B3LYP/ 6-31G*(CHN)/SDD(Ni) level of theory. Figure 3 (left panel) shows the square-planar geometry found to be a (local) minimum on the energy surface. Interestingly, this structure bears an N9-N9' (purine numbering) Pur^P-Pur^P, distance of 9.54 Å, which nearly replicates the N9–N1 purine–pyrimidine distance of 9.05 Å that occurs in natural B-DNA helices for both G·C and A·T base pairs. This suggests $Pur^{P} \cdot Ni^{2+} \cdot Pur^{P}$ is a good dimensional mimic of natural base pairs despite the fact that the metallo base pair incorporates two purine-like components. Superposition of ab initio optimized Pur^P·Ni²⁺·Pur^P and A·T base-pair structures (Figure 3, right panel) further supports this idea and shows that Pur^P coordination of Ni²⁺ is attended by rotation of the Pur^P bases towards the major groove, with a corresponding shortening of the distance between interstrand N9 atoms.

The $Pur^{P} \cdot Ni^{2+} \cdot Pur^{P}$ structure in Figure 3 results from head-to-head dimerization (N1,N1'-Pur^P · Ni^{2+} · N1,N1'-Pur^P). An alternative head-to-tail dimerization mode of Pur^{P} (N1,N1'-Pur^P · Ni^{2+} · N7,N1'-Pur^P) was also investigated computationally. The geometry of this latter complex was found to be highly nonplanar due to ligand encroachment resulting from the 1,5-relationship of the nitrogen atoms (N7,N1') presented by the "tail"-oriented Pur^P. (In contrast, the opposing "head"-oriented Pur^P bears the optimal 1,4-relationship of nitrogen atoms, as found in bipyridine.) Therefore, the head-to-tail dimer is predicted to be less compatible with a helix than the head-to-head dimer.

Metallo base pairs could become functional elements of oligonucleotides that activate or suppress enzymatic activity (for example, transcription or translation) in the presence or absence of a metal ion. For a proof of principle, we have incorporated three consecutive Pur^P residues into a helix to attain "on" and "off" states (in the presence and absence of Ni²⁺, respectively) that are sufficiently insulated from one another to be effectively binary (0 or 1). The following tetradecamer DNA strands were prepared: 5'-d-CTTTCTPur^pPur^pPur^pTCCCT (12) and 5'-d-AGGGAPur^p- $Pur^{P}Pur^{P}AGAAAG$ (13). Gratifyingly, the T_{m} values for the 12/13 duplex under the conditions reported in Table 1 were 64.3 °C in the presence of 10.0 μM NiCl₂ (1.3 equiv per Pur^p residue) and 20.6 °C in the absence of Ni²⁺. Thus, the $T_{\rm m}$ values between the "on" (Ni²⁺ present) and "off" (Ni²⁺ absent) states of this system are separated by 43.7°C, a difference sufficient to produce binary behavior at 37 °C (that is, at this temperature, in the presence of Ni²⁺ the helix is present and in the absence of Ni²⁺ the helix is absent).

In summary, Pur^P leads to a metallo base pair with Ni²⁺ selectivity that is more stable than natural G·C and A·T base pairs. Additionally, Pur^P·Ni²⁺·Pur^P is orthogonal in its pairing properties relative to the four genomic nucleobases: all mismatches are highly destabilizing to the helix in comparison to the parent metallo base pair. Finally, Pur^P·Ni²⁺·Pur^P appears to resemble natural base pairs dimensionally and has the potential to serve as the functional component of an ion-activated switch. Given the resemblance between natural purines and Pur^P it is possible that enzymes, including those



Figure 3. Left: Stereoview of the structure of $Pur^{p} \cdot Ni^{2+} \cdot Pur^{p}$ with optimized geometry obtained with Gaussian 98.^[8] Right: Superposition of optimized $Pur^{p} \cdot Ni^{2+} \cdot Pur^{p}$ and A-T structures. The superposition was guided by $Pur^{p} N9/A N9$, $Pur^{p} N9/T N1$, and the hydrogen atoms attached to these nitrogen atoms.

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involved in DNA replication, might recognize the Pur^P·Ni²⁺· Pur^P base pair. We are actively pursuing this possibility.

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- a) C. Switzer, S. E. Moroney, S. A. Benner, *J. Am. Chem. Soc.* 1989, *111*, 8322–8323; b) J. A. Piccirilli, T. Krauch, S. E. Moroney, S. A. Benner, *Nature* 1990, *343*, 33–47.
- [2] a) B. A. Schweitzer, E. T. Kool, J. Am. Chem. Soc. 1995, 117, 1863–1872; b) D. L. McMinn, A. K. Ogawa, Y. Q. Wu, J. Q. Liu, P. G. Schultz, F. E. Romesberg, J. Am. Chem. Soc. 1999, 121, 11585–11586.
- [3] a) E. Meggers, P. L. Holland, W. B. Tolman, F. E. Romesberg, P. G. Schultz, J. Am. Chem. Soc. 2000, 122, 10714-10715; b) H. Weizman, Y. Tor, J. Am. Chem. Soc. 2001, 123, 3375-3376; c) K. Tanaka, Y. Yamada, M. Shionoya, J. Am. Chem. Soc. 2002, 124, 8802-8803; d) T. Tanaka, A. Tengeiji, T. Kato, N. Toyama, M. Shiro, M. Shionoya, J. Am. Chem. Soc. 2002, 124, 12494-12498; e) N. Zimmerman, E. Meggers, P. G. Schultz, J. Am. Chem. Soc. 2002, 124, 13684-13685; f) K. Tanaka, A. Tengeji, T. Kato, N. Toyama, M. Shionoya, Science 2003, 299, 1212-1213; g) C. Brotschi, C. J. Leumann, Nucleosides Nucleotides Nucleic Acids 2003, 22, 1195-1197.
- [4] Z. Kazimierczuk, H. B. Cottam, G. R. Revankar, R. K. Robins, J. Am. Chem. Soc. 1984, 106, 6379-6382.
- [5] M. Hocek, A. Holy, I. Vortuba, H. Dvoáková, Collect. Czech. Chem. Commun. 2001, 66, 483–499.
- [6] A. Lützen, M. Hapke, Eur. J. Org. Chem. 2002, 2292-2297.
- [7] H. Hashimoto, M. G. Nelson, C. Switzer, J. Am. Chem. Soc. 1993, 115, 7128-7134.
- [8] Gaussian 98 (Revision A.7), M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, V. G. Zakrzewski, J. A. Montgomery, R. E. Stratmann, J. C. Burant, S. Dapprich, J. M. Millam, A. D. Daniels, K. N. Kudin, M. C. Strain, O. Farkas, J. Tomasi, V. Barone, M. Cossi, R. Cammi, B. Mennucci, C. Pomelli, C. Adamo, S. Clifford, J. Ochterski, G. A. Petersson, P. Y. Ayala, Q. Cui, K. Morokuma, D. K. Malick, A. D. Rabuck, K. Raghavachari, J. B. Foresman, J. Cioslowski, J. V. Ortiz, B. B. Stefanov, G. Liu, A. Liashenko, P. Piskorz, I. Komaromi, R. Gomperts, R. L. Martin, D. J. Fox, T. Keith, M. A. Al-Laham, C. Y. Peng, A. Nanayakkara, C. Gonzalez, M. Challacombe, P. M. W. Gill, B. G. Johnson, W. Chen, M. W. Wong, J. L. Andres, M. Head-Gordon, E. S. Replogle, J. A. Pople, Gaussian, Inc., Pittsburgh, PA, **1998**.