

ORIGINAL ARTICLE

Synthesis and biological evaluation of (*E*)-1-(substituted)-3-phenylprop-2-en-1-ones bearing rhodanines as potent anti-microbial agents

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Abstract

Herein, we report the design, syntheses and *in vitro* anti-microbial activity of two series of rhodanines with chalcone moiety. Anti-microbial tests showed that some of the synthesized compounds exhibited good inhibition (MIC = 1–8 µg/mL) against multi-drug-resistant Gram-positive organisms, including methicillin resistant and quinolone-resistant *Staphylococcus aureus*, in which the compound **4g** was found to be the most potent with minimum inhibitory concentration (MIC) value of 1 µg/mL against two methicillin-resistant *S. aureus*.

Keywords

Anti-bacterial, methicillin-resistant
Staphylococcus aureus, rhodanine

History

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Introduction

The severe infections caused by multi-drug-resistant microorganisms such as methicillin-resistant *Staphylococcus aureus* (MRSA) and glycopeptide-resistant *Staphylococci* (GRSA) have substantially increased over the last few years. However, the number of effective anti-bacterial agents has decreased gradually with the appearance of new resistance mechanisms of microorganisms, which is a major concern in medicine, especially for hospitals now^{1,2}.

Rhodanine scaffold is a powerful device for medicinal chemist and have a broad substrate scope for the synthesis of various heterocyclic moieties with wide range of pharmacological activities such as anti-bacterial^{3–5}, anti-fungal⁶, anti-diabetic⁷, anti-tubercular^{8,9}, anti-HIV^{10,11} and anthelmintic agents^{12,13}. Recently, this scaffold showed important therapeutic targets against plasmodium fatty acid synthesis pathway (FAS-II) in the parasites which is distinct from type I fatty acid synthesis pathway (FAS-I) in human¹⁴. Hardej et al.¹⁵ reported some rhodanine derivatives as anti-microbial agents against MRSA strains.

In our previous study, we designed and synthesized three series of rhodanine derivatives bearing chalcone (Figure 1, **I**), 4-(2-oxo-2-phenylethoxy)benzene (Figure 1, **II**) and 5-aryloxy pyrazole moieties (Figure 1, **III**), respectively^{16–23}. Their anti-bacterial tests *in vitro* showed that these compounds all exhibited good

inhibitory activity against the Gram-positive microorganisms especially for the multi-drug-resistant clinical isolates such as MRSA and quinolone-resistant *S. aureus* (QRSA). Especially in the series of **I**, compound **6q** (A moiety was a naphthalene nucleus) showed the strongest activity with a minimum inhibitory concentration (MIC) value of 2 µg/mL against the multi-drug-resistant clinical isolates²³. In the present work, as a part of our ongoing research, a new series of rhodanine derivatives (**4a–i**) were designed using **6q** as the lead compound, in which the modification of **6q** was focused on reserving the naphthalene nucleus, substituting the acetic acid group on the three-position of the rhodanine with different amino acid side chains (including D- or L-phenylalanine, D- or L-tyrosine, D- or L-valine, D- or L-leucine and L-isoleucine) as shown in Figure 2. Moreover, another series of rhodanine derivatives (**5a–i**) were also been designed, in which the naphthalene nucleus was replaced by a biphenyl group in order to discuss the structure–activity relationship better. Both *R*- and *S*-configuration were involved in the two series of compounds so as to investigate the contribution of configuration on their anti-bacterial activity (Figure 2). Thus, two series of rhodanine derivatives, including 17 new compounds were synthesized and screened for their anti-bacterial activities.

The target compounds (**4a–i** and **5a–i**) were synthesized according to the route described in Scheme 1. The intermediates **1** and **2** were prepared by reacting acetophenones with terephthalaldehyde in the ethanol–NaOH/H₂O condition²⁴. Compounds **4a–i** and **5a–i** were obtained in good yields via a Knoevenagel condensation of compounds **1** (or **2**) and *N*-substituted rhodanines using the reported procedure²⁵. The newly synthesized compounds were characterized by IR, ¹H-NMR and mass spectra. Taking compound **4b** as an example, the mass spectroscopy of **4b** displayed an M + H signal at *m/z* 550, which was corresponding

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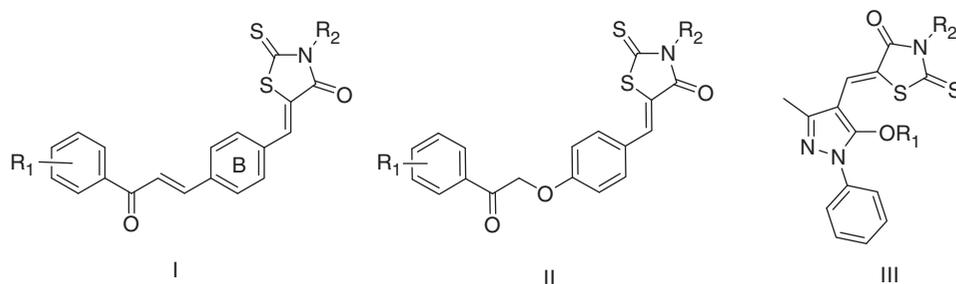
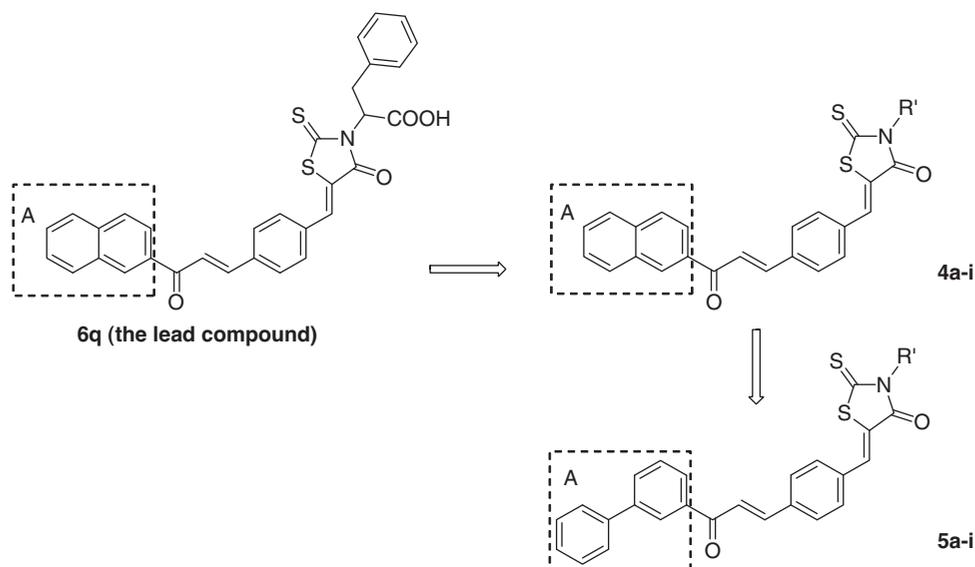
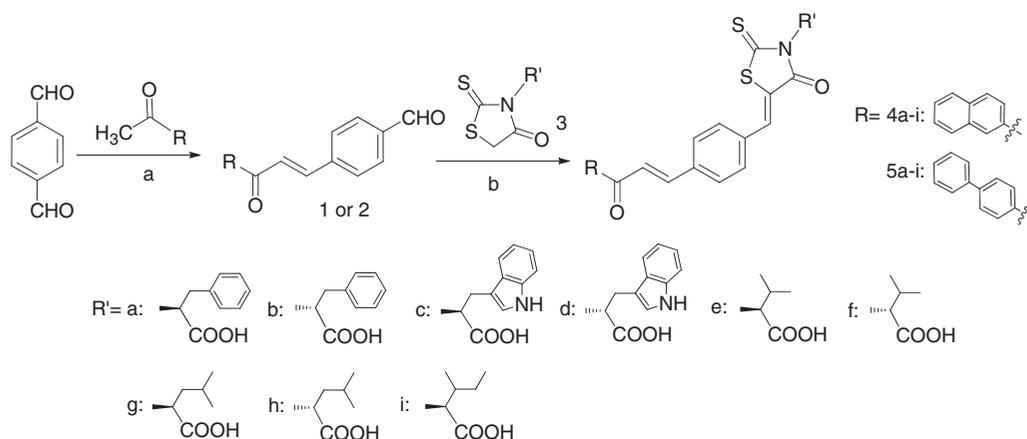
Figure 1. The structures of compounds **I**, **II** and **III**.

Figure 2. Lead compound and structure-based design of the target compounds.

Scheme 1. Synthetic scheme for the synthesis of compounds **4a-i** and **5a-i**.

to its molecular weight of 549. In the $^1\text{H-NMR}$ spectrum of compound **4b**, in addition to the aromatic protons of benzene ring protons ($\delta = 7.15\text{--}8.97$ ppm), a sharp singlet due to vinylic hydrogen was observed at 8.08 ppm, and a broad singlet due to methyne hydrogen linked to nitrogen-atoms was observed at 5.90 ppm. Meanwhile, it has been reported that the (*Z*- or (*E*)-geometry was readily identified by $^1\text{H-NMR}$, as the vinylic proton is more de-shielded in the (*Z*)-isomer than the (*E*)-isomer. In (*Z*)-form, vinylic proton appeared at 7.21 ppm due to the magnetic anisotropy effects of the carbonyl group on the vinylic proton, while in (*E*)-form the resonance should be around 6.50 ppm^{26,27}. In our $^1\text{H-NMR}$ spectra, only a set of signals around 8.08 ppm was appeared, which confirmed our products

were only in (*Z*)-configuration as thermodynamically favored structures.

The physicochemical properties of them are presented in the ‘‘Chemistry’’ section. Their anti-bacterial activities were all evaluated by a serial dilution method to obtain the MIC with different strains, including multi-drug-resistant clinical isolates.

Experimental protocols

Chemistry

Melting points were determined in open capillary tubes and were uncorrected. Reaction courses were monitored by TLC on silica

gel-precoated F254 Merck plates. Developed plates were examined with ultra violet lamps (254 nm). IR spectra were recorded (in KBr) on a FTIR1730. ¹H NMR spectra were measured on a Bruker AV-300 spectrometer using tetramethylsilane as the internal standard. Mass spectra were measured on a matrix assisted laser desorption ionization-time of flight/time of flight (MALDI-TOF/TOF) mass spectrometer (Bruker Daltonik, Bremen, Germany). Specific optical rotation was measured on a Digital automatic polariscope JASCO P-1020 (JASCO, Tokyo, Japan). The major chemicals were purchased from Sigma-Aldrich (St. Louis, MO) and Fluka Companies (Milwaukee, MI).

The general synthesis route of compound 1 and 2

To a stirred solution of terephthalaldehyde (12 mmol) in 15 mL ethanol and 15 mL 5% NaOH, the solution of 1-(naphthalen-2-yl)ethanone or 1-([1,1'-biphenyl]-4-yl)ethanone (10 mmol) in 15 mL ethanol was added drop-wise about 30 min in an ice-bath. The mixture was stirred at room temperature for 8 h and the pH value of the mixture was adjusted to 3 using dilute HCl. After adding 25 mL water into the mixture, the resulting precipitate was filtered, washed with water and recrystallized with ethanol to get the yellow solid.

Synthesis of rhodanine-3-acids (3)

To a solution of appropriate amino acid (30.3 mmol) and sodium hydroxide (30.3 mmol) in 25 mL of water, carbon disulfide (30.3 mmol) was added, and the resulting mixture was stirred vigorously overnight. An aqueous solution of sodium chloroacetate (30.3 mmol) was added to the mixture and stirring was continued at 23 °C for 3 h. Then, the reaction mixture was acidified with dilute HCl until pH 1.0 and refluxed overnight. The reaction mixture was neutralized with saturated NaHCO₃ solution. The resultant solution was acidified again with dilute HCl. The cyclized product was extracted with ethyl acetate, dried over anhydrous sodium sulfate, evaporated under vacuum, and the residue was purified by column chromatography (dichloromethane/methanol = 95:05) to afford a brown liquid.

General procedure for the preparation of compounds 4a–i

To a suspension of **6** (2 mmol) in dry ethanol (10 mL), (*E*)-4-(3-(naphthalen-2-yl)-3-oxoprop-1-en-1-yl)benzaldehyde **2** (2.1 mmol), catalytic amounts of piperidine (0.1 mmol) and glacial acetic acid (0.1 mmol) were added. The resulting mixture was stirred and refluxed overnight. After cooling, the solvent was evaporated *in vacuo*, dried, and purified by silica gel column chromatography (dichloromethane/methanol = 100:1). The yield, melting point and spectral data of each compound are given below.

(*S*)-2-((*Z*)-5-(4-((*E*)-3-(naphthalen-2-yl)-3-oxoprop-1-en-1-yl)benzylidene)-4-oxo-2-thioxothiazolidin-3-yl)-3-phenylpropanoic acid (**4a**)

Yellow solid; yield 62%; m.p. 218–219 °C. $[\alpha]_{\text{D}}^{20}$: –202.3 (*c* = 0.54, acetone). IR (KBr) cm^{-1} : 3449 (OH), 1687 (C=O). ¹H NMR (DMSO-*d*₆, 300 MHz, ppm): δ 3.55 (d, *J* = 5.7 Hz, 2H, CH₂), 5.76 (br.s, 1H, NCH), 7.82 (d, 1H, *J* = 15.7 Hz, CH=CH), 8.09 (s, 1H, C=CH), 8.23 (d, 1H, *J* = 15.7 Hz, CH=CH), 7.16–8.29 (m, 16H, Ar-H), 13.44 (s, 1H, COOH). MS *m/z* 572 ([M + Na]⁺). Electrospray ionization high resolution mass spectrometry (ESI-HRMS) calcd for C₃₂H₂₃NNaO₄S²⁺ ([M + Na]⁺): 572.0961; found: 572.0979.

(*R*)-2-((*Z*)-5-(4-((*E*)-3-(naphthalen-2-yl)-3-oxoprop-1-en-1-yl)benzylidene)-4-oxo-2-thioxothiazolidin-3-yl)-3-phenylpropanoic acid (**4b**)

Yellow solid; yield 15%; m.p. 204–206 °C. $[\alpha]_{\text{D}}^{20}$: 211.9 (*c* = 0.62, acetone). IR (KBr) cm^{-1} : 3424 (OH), 1709 (C=O). ¹H NMR (DMSO-*d*₆, 300 MHz, ppm): δ 3.52 (d, *J* = 5.5 Hz, 2H, CH₂), 5.90 (br.s, 1H, NCH), 7.82 (d, 1H, *J* = 15.7 Hz, CH=CH), 8.08 (s, 1H, C=CH), 8.25 (d, 1H, *J* = 15.7 Hz, CH=CH), 7.15–8.97 (m, 16H, Ar-H), 13.44 (s, 1H, COOH). MS *m/z* 550 (M + H). ESI-HRMS calcd for C₃₂H₂₃NNaO₄S²⁺ ([M + Na]⁺): 572.0961; found: 572.0960.

(*S*)-3-(1*H*-indol-3-yl)-2-((*Z*)-5-(4-((*E*)-3-(naphthalen-2-yl)-3-oxoprop-1-en-1-yl)benzylidene)-4-oxo-2-thioxothiazolidin-3-yl)propanoic acid (**4c**)

Yellow solid; yield 23%; m.p. 130–132 °C. $[\alpha]_{\text{D}}^{20}$: –237.9 (*c* = 0.30, acetone). IR (KBr) cm^{-1} : 3386 (OH), 2889 (NH), 1711 (C=O). ¹H NMR (DMSO-*d*₆, 300 MHz, ppm): δ 3.57–3.75 (m, 2H, CH₂), 5.90 (br.s, 1H, N-CH), 7.82 (d, 1H, *J* = 15.5 Hz, CH=CH), 8.07 (s, 1H, C=CH), 8.24 (d, 1H, *J* = 15.5 Hz, CH=CH), 7.07–8.96 (m, 16H, Ar-H), 10.81 (s, 1H, NH), 13.21 (s, 1H, COOH). MS *m/z* 589 (M + H). ESI-HRMS calcd for C₃₄H₂₄N₂NaO₄S²⁺ ([M + Na]⁺): 611.1070; found: 611.1073.

(*R*)-3-(1*H*-indol-3-yl)-2-((*Z*)-5-(4-((*E*)-3-(naphthalen-2-yl)-3-oxoprop-1-en-1-yl)benzylidene)-4-oxo-2-thioxothiazolidin-3-yl)propanoic acid (**4d**)

Yellow solid; yield 17%; m.p. 120–122 °C. $[\alpha]_{\text{D}}^{20}$: 227.6 (*c* = 0.63, CH₃CO₂C₂H₅). IR (KBr) cm^{-1} : 3406 (OH), 2909 (NH), 1713 (C=O). ¹H NMR (DMSO-*d*₆, 300 MHz, ppm): δ 3.55–3.78 (m, 2H, CH₂), 5.89 (br.s, 1H, N-CH), 7.84 (d, 1H, *J* = 15.6 Hz, CH=CH), 8.09 (s, 1H, C=CH), 8.27 (d, 1H, *J* = 15.6 Hz, CH=CH), 6.89–8.99 (m, 16H, Ar-H), 10.83 (s, 1H, NH), 13.37 (s, 1H, COOH). MS *m/z* 589 (M + H). ESI-HRMS calcd for C₃₄H₂₄N₂NaO₄S²⁺ ([M + H]⁺): 589.1250; found: 589.1244.

(*S*)-3-methyl-2-((*Z*)-5-(4-((*E*)-3-(naphthalen-2-yl)-3-oxoprop-1-en-1-yl)benzylidene)-4-oxo-2-thioxothiazolidin-3-yl)butanoic acid (**4e**)

Yellow solid; yield 28%; m.p. 206–208 °C. $[\alpha]_{\text{D}}^{20}$: 45.2 (*c* = 0.59, CHCl₃). IR (KBr) cm^{-1} : 3406 (OH), 1713 (C=O). ¹H NMR (DMSO-*d*₆, 300 MHz, ppm): δ 0.75 (d, 3H, *J* = 6.8 Hz, CH-CH₃), 1.20 (d, 3H, *J* = 6.5 Hz, CH-CH₃), 2.69–2.75 (m, 1H, CH(CH₃)₂), 5.19 (br.s, 1H, N-CH), 7.84 (d, 1H, *J* = 15.6 Hz, CH=CH), 7.92 (s, 1H, C=CH), 8.26 (d, 1H, *J* = 15.6 Hz, CH=CH), 7.63–8.97 (m, 11H, Ar-H), 13.22 (s, 1H, COOH). MS *m/z* 502 (M + H). ESI-HRMS calcd for C₂₈H₂₃NNaO₄S²⁺ ([M + Na]⁺): 524.0961; found: 524.0947.

(*R*)-3-methyl-2-((*Z*)-5-(4-((*E*)-3-(naphthalen-2-yl)-3-oxoprop-1-en-1-yl)benzylidene)-4-oxo-2-thioxothiazolidin-3-yl)butanoic acid (**4f**)

Yellow solid; yield 15%; m.p. 136–139 °C. $[\alpha]_{\text{D}}^{20}$: 45.2 (*c* = 0.59, CHCl₃). IR (KBr) cm^{-1} : 3437 (OH), 1714 (C=O). ¹H NMR (DMSO-*d*₆, 300 MHz, ppm): δ 0.77 (d, 3H, *J* = 6.5 Hz, CH-CH₃), 1.21 (d, 3H, *J* = 6.1 Hz, CH-CH₃), 2.70–2.77 (m, 1H, CH(CH₃)₂), 5.18 (br.s, 1H, N-CH), 7.86 (d, 1H, *J* = 15.7 Hz, CH=CH), 7.94 (s, 1H, C=CH), 8.28 (d, 1H, *J* = 15.7 Hz, CH=CH), 7.35–8.99 (m, 11H, Ar-H), 13.18 (s, 1H, COOH). MS *m/z* 502 (M + H). ESI-HRMS calcd for C₂₈H₂₃NNaO₄S²⁺ ([M + Na]⁺): 524.0961; found: 524.0966.

(*S*)-4-methyl-2-((*Z*)-5-(4-((*E*)-3-(naphthalen-2-yl)-3-oxoprop-1-en-1-yl)benzylidene)-4-oxo-2-thioxothiazolidin-3-yl)pentanoic acid (**4g**)

Yellow solid; yield 31%; m.p. 150–152 °C. $[\alpha]_D^{20}$: -5.1 ($c = 0.58$, CHCl₃). IR (KBr) cm⁻¹: 3260 (OH), 1721 (C=O). ¹H NMR (DMSO-*d*₆, 300 MHz, ppm): δ 0.87 (d, 3H, $J = 6.8$ Hz, CHCH₃), 0.92 (d, 3H, $J = 6.3$ Hz, CHCH₃), 1.47–1.53 (m, 1H, CH(CH₃)₂), 2.02–2.23 (m, 2H, CH₂), 5.61 (br.s, 1H, NCH), 7.84 (d, 1H, $J = 15.7$ Hz, CH=CH), 8.13 (s, 1H, C=CH), 8.27 (d, 1H, $J = 15.7$ Hz, CH=CH), 7.63–8.98 (m 11H, Ar-H), 13.37 (s, 1H, COOH). MS m/z 516 (M+H). ESI-HRMS calcd for C₂₉H₂₅NNaO₄S₂⁺ ([M+Na]⁺): 538.1117; found: 538.1130.

(*R*)-4-methyl-2-((*Z*)-5-(4-((*E*)-3-(naphthalen-2-yl)-3-oxoprop-1-en-1-yl)benzylidene)-4-oxo-2-thioxothiazolidin-3-yl)pentanoic acid (**4h**)

Yellow solid; yield 16%; m.p. 198–200 °C. $[\alpha]_D^{20}$: 6.0 ($c = 0.63$, CHCl₃). IR (KBr) cm⁻¹: 3441 (OH), 1721 (C=O). ¹H NMR (DMSO-*d*₆, 300 MHz, ppm): δ 0.86 (d, 3H, $J = 6.8$ Hz, CHCH₃), 0.91 (d, 3H, $J = 6.3$ Hz, CHCH₃), 1.46–1.52 (m, 1H, CH(CH₃)₂), 2.01–2.20 (m, 2H, CH₂), 5.59 (br.s, 1H, NCH), 7.83 (d, 1H, $J = 15.6$ Hz, CH=CH), 8.12 (s, 1H, C=CH), 8.25 (d, 1H, $J = 15.6$ Hz, CH=CH), 7.64–8.97 (m, 11H, Ar-H), 13.37 (s, 1H, COOH). MS m/z 516 (M+H). ESI-HRMS calcd for C₂₉H₂₅NNaO₄S₂⁺ ([M+Na]⁺): 538.1117; found: 538.1118.

(2*S*)-3-methyl-2-((*Z*)-5-(4-((*E*)-3-(naphthalen-2-yl)-3-oxoprop-1-en-1-yl)benzylidene)-4-oxo-2-thioxothiazolidin-3-yl)pentanoic acid (**4i**)

Yellow solid; yield 36%; m.p. 160–162 °C. $[\alpha]_D^{20}$: -45.8 ($c = 0.63$, CHCl₃). IR (KBr) cm⁻¹: 2967 (OH), 1717 (C=O). ¹H NMR (DMSO-*d*₆, 300 MHz, ppm): δ 0.79 (t, 3H, $J = 7.2$ Hz, CH₂CH₃), 0.92–0.97 (m, 2H, CH₂CH₃), 1.15 (d, 3H, $J = 6.4$ Hz, CHCH₃), 1.20–1.24 (m, 1H, CHCH₃), 5.23 (br.s, 1H, NCH), 7.83 (d, 1H, $J = 15.8$ Hz, CH=CH), 8.15 (s, 1H, C=CH), 8.26 (d, 1H, $J = 15.8$ Hz, CH=CH), 7.62–8.97 (m, 11H, Ar-H), 12.95 (s, 1H, COOH). MS m/z 538 (M+Na⁺). ESI-HRMS calcd for C₂₉H₂₅NNaO₄S₂⁺ ([M+H]⁺): 516.1298; found: 516.1298.

General procedure for the preparation of compounds 5a–i

To a suspension of **6** (2 mmol) in dry ethanol (10 mL), (*E*)-4-(3-([1,1'-biphenyl]-4-yl)-3-oxoprop-1-en-1-yl)benzaldehyde **3** (2.1 mmol), catalytic amounts of piperidine (0.1 mmol) and glacial acetic acid (0.1 mmol) were added. The resulting mixture was stirred and refluxed overnight. After cooling, the solvent was evaporated *in vacuo*, dried and purified by silica gel column chromatography (dichloromethane/methanol = 100:1). The yield, melting point and spectral data of each compound are given below.

(*S*)-2-((*Z*)-5-(4-((*E*)-3-([1,1'-biphenyl]-4-yl)-3-oxoprop-1-en-1-yl)benzylidene)-4-oxo-2-thioxothiazolidin-3-yl)-3-phenylpropanoic acid (**5a**)

Yellow solid; yield 38%; m.p. 123–125 °C. $[\alpha]_D^{20}$: -206.5 ($c = 0.61$, CH₃CO₂C₂H₅). IR (KBr) cm⁻¹: 3005 (OH), 1711 (C=O). ¹H NMR (DMSO-*d*₆, 300 MHz, ppm): δ 3.54 (s, 2H, $J = 5.6$ Hz, CH₂), 5.91 (br.s, 1H, NCH), 7.16–8.30 (m, 20H, Ar-H), 7.78 (s, 1H, C=CH), 13.53 (s, 1H, COOH). MS m/z 576 (M+H). ESI-HRMS calcd for C₃₄H₂₅NNaO₄S₂⁺ ([M+Na]⁺): 598.1117; found: 598.1126.

(*R*)-2-((*Z*)-5-(4-((*E*)-3-([1,1'-biphenyl]-4-yl)-3-oxoprop-1-en-1-yl)benzylidene)-4-oxo-2-thioxothiazolidin-3-yl)-3-phenylpropanoic acid (**5b**)

Yellow solid; yield 21%; m.p. 98–100 °C. $[\alpha]_D^{20}$: 187.2 ($c = 0.63$, CH₃CO₂C₂H₅). IR (KBr) cm⁻¹: 3028 (OH), 1713 (C=O). ¹H NMR (DMSO-*d*₆, 300 MHz, ppm): δ 3.51 (s, 2H, $J = 5.5$ Hz, CH₂), 5.83 (br.s, 1H, NCH), 7.10–8.21 (m, 20H, Ar-H), 7.77 (s, 1H, C=CH), 13.32 (s, 1H, COOH). MS m/z 576 (M+H). ESI-HRMS calcd for C₃₄H₂₅NNaO₄S₂⁺ ([M+Na]⁺): 598.1117; found: 598.1122.

(*S*)-2-((*Z*)-5-(4-((*E*)-3-([1,1'-biphenyl]-4-yl)-3-oxoprop-1-en-1-yl)benzylidene)-4-oxo-2-thioxothiazolidin-3-yl)-3-(1*H*-indol-3-yl)propanoic acid (**5c**)

Yellow solid; yield 35%; m.p. 108–112 °C. $[\alpha]_D^{20}$: -269.1 ($c = 0.61$, CH₃CO₂C₂H₅). IR (KBr) cm⁻¹: 3398 (OH), 2890 (NH), 1706 (C=O). ¹H NMR (DMSO-*d*₆, 300 MHz, ppm): δ 3.55–3.80 (m, 2H, CH₂), 5.89 (br.s, 1H, NCH), 7.08–8.29 (m, 20H, Ar-H), 7.78 (s, 1H, C=CH), 10.83 (s, 1H, NH), 13.53 (s, 1H, COOH). MS m/z 615 (M+H). ESI-HRMS calcd for C₃₆H₂₆N₂NaO₄S₂⁺ ([M+Na]⁺): 637.1226; found: 637.1246.

(*R*)-2-((*Z*)-5-(4-((*E*)-3-([1,1'-biphenyl]-4-yl)-3-oxoprop-1-en-1-yl)benzylidene)-4-oxo-2-thioxothiazolidin-3-yl)-3-(1*H*-indol-3-yl)propanoic acid (**5d**)

Yellow solid; yield 19%; m.p. 126–128 °C. $[\alpha]_D^{20}$: 271.1 ($c = 0.60$, CHCl₃). IR (KBr) cm⁻¹: 3410 (OH), 2897 (NH), 1709 (C=O). ¹H NMR (DMSO-*d*₆, 300 MHz, ppm): δ 3.55–3.76 (m, 2H, CH₂), 5.90 (br.s, 1H, NCH), 7.49–8.07 (m, 20H, Ar-H), 7.77 (s, 1H, C=CH), 10.83 (s, 1H, NH), 13.47 (s, 1H, COOH). MS m/z 615 (M+H). ESI-HRMS calcd for C₃₆H₂₆N₂NaO₄S₂⁺ ([M+Na]⁺): 637.1226; found: 637.1236.

(*S*)-2-((*Z*)-5-(4-((*E*)-3-([1,1'-biphenyl]-4-yl)-3-oxoprop-1-en-1-yl)benzylidene)-4-oxo-2-thioxothiazolidin-3-yl)-3-methylbutanoic acid (**5e**)

Yellow solid; yield 26%; m.p. 120–123 °C. $[\alpha]_D^{20}$: -31.4 ($c = 0.57$, CHCl₃). IR (KBr) cm⁻¹: 2942 (OH), 1719 (C=O). ¹H NMR (DMSO-*d*₆, 300 MHz, ppm): δ 0.75 (d, 3H, $J = 6.8$ Hz, CHCH₃), 1.19 (d, 3H, $J = 6.4$ Hz, CHCH₃), 2.70–2.76 (m, 1H, CH(CH₃)₂), 5.17 (br.s, 1H, NCH), 7.49–8.29 (m, 15H, Ar-H), 7.77 (s, 1H, C=CH), 13.14 (s, 1H, COOH). MS m/z 528 (M+H). ESI-HRMS calcd for C₃₀H₂₅NNaO₄S₂⁺ ([M+H]⁺): 528.1298; found: 528.1310.

(*R*)-2-((*Z*)-5-(4-((*E*)-3-([1,1'-biphenyl]-4-yl)-3-oxoprop-1-en-1-yl)benzylidene)-4-oxo-2-thioxothiazolidin-3-yl)-3-methylbutanoic acid (**5f**)

Yellow solid; yield 20%; m.p. 89–92 °C. $[\alpha]_D^{20}$: 28.5 ($c = 0.38$, CHCl₃). IR (KBr) cm⁻¹: 2959 (OH), 1721 (C=O). ¹H NMR (DMSO-*d*₆, 300 MHz, ppm): δ 0.90 (d, 3H, $J = 7.3$ Hz, CHCH₃), 1.19 (d, 3H, $J = 6.5$ Hz, CHCH₃), 2.69–2.74 (m, 1H, CH(CH₃)₂), 5.17 (br.s, 1H, NCH), 7.43–8.29 (m, 15H, Ar-H), 7.77 (s, 1H, C=CH), 13.03 (s, 1H, COOH). MS m/z 528 (M+H). ESI-HRMS calcd for C₃₀H₂₅NNaO₄S₂⁺ ([M+H]⁺): 528.1298; found: 528.1294.

(*S*)-2-((*Z*)-5-(4-((*E*)-3-([1,1'-biphenyl]-4-yl)-3-oxoprop-1-en-1-yl)benzylidene)-4-oxo-2-thioxothiazolidin-3-yl)-4-methylpentanoic acid (**5g**)

Yellow solid; yield 32%; m.p. 110–112 °C. $[\alpha]_D^{20}$: -16.9 ($c = 0.59$, CH₃CO₂C₂H₅). IR (KBr) cm⁻¹: 3028 (OH), 1713 (C=O). ¹H NMR (DMSO-*d*₆, 300 MHz, ppm): δ 0.87 (d, 3H, $J = 6.5$ Hz,

CHCH₃), 0.92 (d, 3H, *J* = 6.5 Hz, CHCH₃), 1.47–1.53 (m, 1H, CH(CH₃)₂), 2.02–2.21 (m, 2H, CH₂), 5.60 (br.s, 1H, N–CH), 7.42–8.29 (m, 15H, Ar–H), 7.78 (s, 1H, C=CH), 13.37 (s, 1H, COOH). MS *m/z* 542 (M+H). ESI-HRMS calcd for C₃₁H₂₇NNaO₄S₂⁺ ([M+Na]⁺): 564.1274; found: 564.1291.

(*R*)-2-((*Z*)-5-(4-((*E*)-3-([1,1'-biphenyl]-4-yl)-3-oxoprop-1-en-1-yl)benzylidene)-4-oxo-2-thioxothiazolidin-3-yl)-4-methylpentanoic acid (**5h**)

Yellow solid; yield 13%; m.p. 96–99 °C. [α]_D²⁰: 19.3 (*c* = 0.45, CH₃CO₂C₂H₅). IR (KBr) cm⁻¹: 3045 (OH), 1712 (C=O). ¹H NMR (DMSO-*d*₆, 300 MHz, ppm): δ 0.86 (d, 3H, *J* = 6.5 Hz, CHCH₃), 0.89 (d, 3H, *J* = 6.4 Hz, CHCH₃), 1.46–1.52 (m, 1H, CH(CH₃)₂), 2.01–2.23 (m, 2H, CH₂), 5.60 (br.s, 1H, NCH), 7.45–8.59 (m, 15H, Ar–H), 7.76 (s, 1H, C=CH), 13.51 (s, 1H, COOH). MS *m/z* 564 ([M+Na]⁺). ESI-HRMS calcd for C₃₀H₂₅NNaO₄S₂⁺ ([M+Na]⁺): 564.1274; found: 564.1279.

(*S*)-2-((*Z*)-5-(4-((*E*)-3-([1,1'-biphenyl]-4-yl)-3-oxoprop-1-en-1-yl)benzylidene)-4-oxo-2-thioxothiazolidin-3-yl)-3-methylpentanoic acid (**5i**)

Yellow solid; yield 25%; m.p. 108–111 °C. [α]_D²⁰: –27.2 (*c* = 0.61, CH₃CO₂C₂H₅). IR (KBr) cm⁻¹: 3426 (OH), 1717 (C=O). ¹H NMR (DMSO-*d*₆, 300 MHz, ppm): δ 0.80 (t, 3H, *J* = 7.0 Hz, CH₂CH₃), 0.94–0.97 (m, 2H, CH₂CH₃), 1.16 (d, 3H, *J* = 6.3 Hz, CHCH₃), 1.27–1.33 (m, 1H, CHCH₃), 5.24 (br.s, 1H, NCH), 7.42–8.30 (m, 15H, Ar–H), 7.78 (s, 1H, C=CH), 13.29 (s, 1H, COOH). MS *m/z* 542 (M+H). ESI-HRMS calcd for C₃₀H₂₅NNaO₄S₂⁺ ([M+Na]⁺): 564.1274; found: 564.1284.

Pharmacology

Evaluation of anti-bacterial activity in vitro

The micro-organisms used in the present study were *S. aureus* (*S. aureus* RN 4220, *S. aureus* KCTC 209 and *S. aureus* KCTC 503) and *Escherichia coli* (*E. coli* 1356). The strains of multi-drug-resistant clinical isolates were MRSA (MRSA CCARM 3167 and MRSA CCARM 3506) and QRSA (QRSA CCARM 3505 and QRSA CCARM 3519). Clinical isolates were collected from various patients hospitalized in several clinics.

Test bacteria were grown to mid-log phase in Mueller-Hinton broth (MHB) and diluted 1000-fold in the same medium. The bacteria of 10⁵ CFU/mL were inoculated into MHB and dispensed at 0.2 mL/well in a 96-well microtiter plate. As positive controls, oxacillin and norfloxacin were used. Test compounds were prepared in DMSO, the final concentration of which did not exceed 0.05%. A twofold serial dilution technique²⁸ was used to obtain final concentrations. The MIC was defined as the concentration of a test compound that completely inhibited bacteria growth during 24 h incubation at 37 °C. Bacteria growth was determined by measuring the absorption at 650 nm using a microtiter enzyme-linked immunosorbent assay (ELISA) reader. All experiments were carried out three times.

Evaluation of cytotoxicity in vitro

Human cervical (Hela) cell monolayers were used as an *in vitro* model of cervicovaginal epithelium for testing the cytotoxicity of the new compounds. Hela cells were grown in Dulbecco-modified Eagle medium (DMEM) supplemented with fetal bovine serum (10%), and antibiotics (penicillin–streptomycin mixture [100 U/mL]). Cells at 80–90% confluence were split by trypsin (0.25% in phosphate buffered saline (PBS); pH 7.4), and the medium was changed at 24 h intervals. The cells were cultured at 37 °C in a 5% CO₂ incubator. The cells were grown to three passages and

approximately 1 × 10⁴ cells were seeded into each well of a 96-well plate and allowed to incubate overnight to allow cells to attach to the substrate. After 24 h, the medium was replaced with DMEM supplemented with 10% fetal bovine serum (FBS) containing various concentrations of test compounds and incubated for 48 h. Then, 10 μl of methyl thiazolyl tetrazolium (MTT) solution (5 mg/mL in PBS) was added to each well. After incubation for 4 h, the medium was removed and the resulting formazan crystals were dissolved with 100 μl DMSO. After shaking 10 min, the optical density was measured at 570 nm using a microtiter ELISA reader. The assay was conducted four times. The IC₅₀ values were defined as the concentrations inhibiting 50% of cell growth.

Results and discussion

Anti-bacterial activity

The anti-microbial assay was carried out using the following bacterial strains *S. aureus* RN 4220, *S. aureus* KCTC 209, *S. aureus* KCTC 503, MRSA CCARM 3167 and 3506, QRSA CCARM 3505 and 3519 and *E. coli* 1356. The *in vitro* anti-bacterial activity was evaluated using a 96-well microtiter plate and a serial dilution method to obtain the MIC. Oxacillin, norfloxacin, gatifloxacin and moxifloxacin were used as positive controls.

Table 1 summarized the results obtained for the MICs of the 18 target compounds (**4a–i** and **5a–i**) against three Gram-positive strains (*S. aureus* RN 4220, *S. aureus* KCTC 209 and *S. aureus* KCTC 503) and one Gram-negative strain (*E. coli* 1356). For *S. aureus* RN 4220, it could be found that most of the compounds (except **4b**, **4c**, **4d**, **4i**, **5c** and **5d**) showed good inhibition with MIC values in the range of 1–16 μg/mL. In particular, compounds **4g** and **4h** (MIC = 1 μg/mL) had a twofold more potent activity than the positive control norfloxacin (MIC = 2 μg/mL), and comparable activity to the positive control oxacillin (MIC = 1 μg/mL). For *S. aureus* KCTC 209, most of the

Table 1. Inhibitory activity (MIC*, μg/mL) of compounds **4a–i** and **5a–i** against bacteria.

Compound	Gram-positive strains			Gram-negative strains
	<i>Staphylococcus aureus</i>			<i>Escherichia coli</i>
	4220	209	503	1356
4a	8	16	>64	>64
4b	>64	>64	>64	>64
4c	>64	>64	>64	>64
4d	>64	>64	>64	>64
4e	4	8	8	>64
4f	4	32	8	>64
4g	1	4	2	>64
4h	1	8	2	>64
4i	>64	>64	>64	>64
5a	16	8	8	>64
5b	8	8	8	>64
5c	>64	>64	>64	>64
5d	>64	>64	>64	>64
5e	8	8	8	>64
5f	16	32	16	>64
5g	8	8	2	>64
5h	16	64	64	>64
5i	8	8	8	>64
Norfloxacin	2	2	2	16
Oxacillin	1	1	1	>64
Gatifloxacin	0.25	2	4	16
Moxifloxacin	0.25	2	2	>64

*The anti-bacterial testing was carried out three times, and the MICs are average of them.

compounds (except **4b**, **4c**, **4d**, **4i**, **5c** and **5d**) displayed poor inhibition with MICs ranging from 4 to 32 µg/mL, while these compounds showed better inhibitory activity against *S. aureus* KCTC 503, whose MICs were mostly in the range of 2–8 µg/mL. Unfortunately, all of the test compounds did not exhibit any inhibition against the Gram-negative strain *E. coli* 1356 (MICs > 64 µg/mL).

The compounds **4a–i** and **5a–i** were also evaluated for their inhibitory activity against MRSA and QRSA. As shown in Table 2, compounds **4a–i** and **5a–i** (except **4b**, **4c**, **4d**, **4i**, **5c** and **5d**) presented the high levels of activity with MIC values of 2–8 µg/mL. For MRSA 3167 and 3506, most compounds presented comparable or much more potent activities than norfloxacin. Among them, compound **4g**, with MIC value of 1 µg/mL, showed eightfold more potent than norfloxacin (MIC = 8 µg/mL) and 64-fold more potent than oxacillin (MIC > 64 µg/mL), twofold more potent than gatifloxacin (MIC = 2 µg/mL), and comparable with moxifloxacin (MIC = 1 µg/mL). For QRSA 3505 and 3519, compounds **4e**, **4g**, **4h** and **5h** also presented higher levels of activity (MIC = 2 µg/mL), which were slightly less active than oxacillin (MIC = 1 µg/mL) but much more potent than norfloxacin (MIC > 64 µg/mL), gatifloxacin (MIC = 8 or 4 µg/mL) and moxifloxacin (MIC = 4 µg/mL).

Most of synthesized compounds (**4a–i** and **5a–i**) exhibit strong anti-bacterial activity, although the mechanism of action is not yet clearly understood. However, in this study, some preliminary remarks on the structure–activity relationship can be drawn from the results of bioactivities. (1) There is no remarkable difference of anti-bacterial activity between **4a–i** and **5a–i**. (2) The activity of *S*-configuration compound is a little better than the *R*-configuration compound. For example, the activity of compound **4g** (*S*-configuration) is higher than that of compound **4h** (*R*-configuration) generally. (3) The MIC values of the compounds derived from different amino acids differed greatly. For example, the compounds bearing tryptophan-derived rhodanines (**4c**, **4d**, **5c** and **5d**) did not show any inhibition against

all of the selected bacteria at 64 µg/mL. While the compounds bearing leucine-derived rhodanines (**4g**, **4h**, **5g** and **5h**) exhibited a strong inhibitory activity with MICs of 1–4 µg/mL against the four drug-resistance bacterial strains. The cytotoxicity of the selected compound (**4g**) was also evaluated. The IC₅₀ value (HeLa cells) of **4g** is 2.41, which is comparable with its MIC value.

Conclusion

Based on our previous work, we synthesized two new series of rhodanine derivatives (**4a–i** and **5a–i**) and evaluated their anti-bacterial activities. Most of the compounds showed good anti-bacterial activities against Gram-positive bacteria as well as multi-drug-resistant strains of clinical isolates. Among them, compound **4g**, with MIC value of 1 µg/mL, showed eightfold more potent than norfloxacin (MIC = 8 and 4 µg/mL) and 64-fold more potent than oxacillin (MIC > 64 µg/mL). Efforts to determine the reason for its anti-bacterial activity are ongoing and will be reported in due course.

Declaration of interest

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Table 2. Inhibitory activity (MIC*, µg/mL) of compounds **4a–i** and **5a–i** against clinical isolates of multidrug-resistant Gram-positive strains.

Compound	Multi-drug-resistant Gram-positive strains			
	MRSA		QRSA	
	3167	3506	3505	3519
4a	2	2	4	4
4b	>64	>64	>64	>64
4c	>64	>64	>64	>64
4d	>64	>64	>64	>64
4e	2	2	2	2
4f	2	4	4	4
4g	1	1	2	2
4h	1	2	2	2
4i	>64	>64	>64	>64
5a	4	8	4	8
5b	2	4	4	4
5c	>64	>64	>64	>64
5d	>64	>64	>64	>64
5e	2	4	4	4
5f	8	8	8	8
5g	2	4	4	4
5h	2	2	2	2
5i	4	4	4	8
Norfloxacin	8	4	>64	>64
Oxacillin	>64	>64	1	1
Gatifloxacin	2	1	8	4
Moxifloxacin	1	1	4	4

*The anti-bacterial testing was carried out three times, and the MICs are average of them.

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