

The first synthesis of the anhydrophytosphingosine pachastrissamine (jaspine B) from Garner's aldehyde[☆]

N. Sudhakar,^a A. Ravi Kumar,^a A. Prabhakar,^b B. Jagadeesh^b and B. Venkateswara Rao^{a,*}

^aOrganic Chemistry Division III, Indian Institute of Chemical Technology, Hyderabad 500007, India

^bNuclear Magnetic Resonance Division, Indian Institute of Chemical Technology, Hyderabad 500007, India

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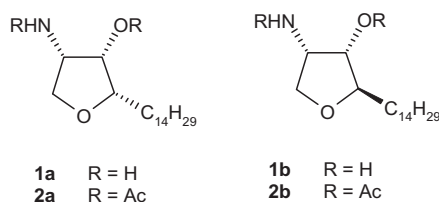
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Abstract—The synthesis of the natural anhydrophytosphingosine pachastrissamine (jaspine B) **1a** from Garner's aldehyde is described.

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Pachastrissamine **1a**, the first natural anhydrophytosphingosine derivative was recently isolated from a marine sponge, *Pachastrissa* sp. (family calthropellidae) by Higa and co-workers¹ and found to possess cytotoxicity at a level of IC₅₀ 0.01 µg/mL against P388, A549, HT29 and Mell 28 cell lines. Later Debitus and co-workers² isolated the same compound from a marine sponge, genus *jaspis*, which is a main source of many cytotoxic compounds such as jaspamides,³ jaspisamides,⁴ isomalabaricane,⁵ toyocamycin and 5-methoxy carbonyltubercidine,⁶ and named as **1a**.

Jaspamides possess interesting biological activities such as antiproliferative (cytotoxic and antimicrobial), anthelmintic, insecticidal and ichthyotoxic activities.⁷



Jaspine B **1a** displayed marked cytotoxicity (IC₅₀ = 0.24 µM) against the A549 human lung carcinoma cell line using the ATP lite assay. Jaspine B **1a** proved to be the most potent compound yet isolated from the *jaspis* genus on this cell line, cf. pectenotoxin II (IC₅₀ > 10 µM),⁸ bengamide Y (IC₅₀ = 12.8 µM),⁹ bengamide Z (IC₅₀ = 10.5 µM).¹⁰ The important biological activity and the novel structural features of **1a** prompted us to undertake its total synthesis.

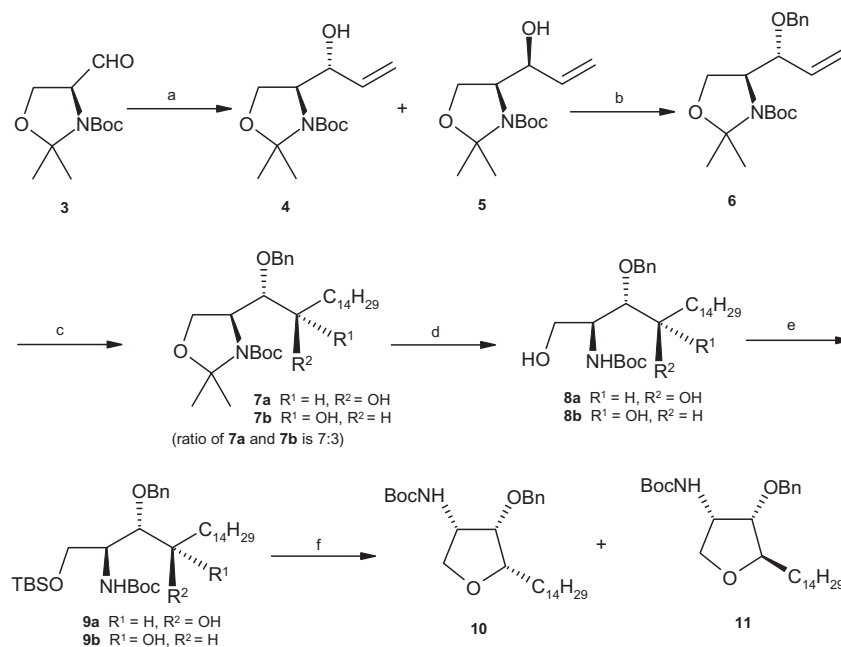
The convergent total synthesis of **1a** started from the Garner's aldehyde.¹¹ Stereoselective addition of vinyl-magnesium bromide to **3** afforded a 6:1 mixture of allylic alcohols **4** and **5**, which were separated by column chromatography.¹² The alcohol functionality of compound **4** was protected as its *O*-benzyl ether to give **6**. Oxidative degradation of the alkene functionality with ozone afforded the aldehyde, which then underwent Grignard addition using C₁₄H₂₉MgBr in THF to give compounds **7a** and **7b** as an inseparable diastereomeric mixture (ratio 7:3 by ¹H NMR), which was carried further as such (Scheme 1).

Deprotection of the acetonide group in compound **7** was achieved using 80% AcOH to give compounds **8a** and **8b**, which were converted into the *O*-silyl ethers **9a,b**. The hydroxyl groups in compounds **9a,b** were converted into mesylates, which were treated with TBAF to give the five membered cyclic compounds **10** and **11** in a 7:3 ratio, which were easily separable by column chromatography. The confirmation of structures and assignments for the tetrahydrofurans **10** and **11** was achieved by detailed 1D and 2D NMR studies including DQF-COSY and NOESY experiments. For **10** strong NOE

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*Corresponding author. Tel.: +91 040 27160123x2614; fax: +91 040 27160512; e-mail: venky@iict.res.in



Scheme 1. Reagents and conditions: (a) vinylmagnesium bromide, THF, 0°C–rt, 12 h, (b) BnBr, THF, NaH (60% w/w), 0°C–rt, 12 h, 92%, (c) (i) O₃, DCM, –78°C, 1 h, (CH₃)₂S, (ii) C₁₄H₂₉MgBr, THF, 12 h, rt (83% for two steps), (d) 80% AcOH, 0°C–rt, 12 h, 91%, (e) TBDMS-Cl, imidazole, dry DCM, DMAP, 0°C–rt, 12 h, 86%, (f) (i) MsCl, Et₃N, DCM, 0°C–rt, 2 h, (ii) TBAF, THF, rt, 2 h, 88% (overall yield 28.3% for compound **10** and 12.1% for compound **11**), (7:3 diastereoisomeric mixture for **10** and **11**).

cross peaks were observed between NH–H1', H2–H4 and H5–H6, whereas for **11** the NOE cross peaks between NH–H1', H2–H5, H3–H5 and H4–H6 confirmed the structures as shown in Figure 1.

The *O*-benzyl ether in compound **10** was cleaved under Na/liq. NH₃ conditions to afford alcohol **12**. Compound **12** on treatment with 50% TFA–DCM gave the target molecule pachastrissamine (jaspine B) **1a** as its TFA salt (**Scheme 2**).

The diacetate derivative of pachastrissamine **2a** was synthesized from compound **1a** using Et₃N and Ac₂O. The ¹H NMR of the TFA salt of **1a** [its optical rotation [α]_D²⁵ +17.1 (*c* 0.4, EtOH)], {lit.¹ [α]_D²⁵ +18 (*c* 0.1, EtOH)} and

electrospray analysis: *m/z* 300 (M⁺+1–CF₃COOH) and for the diacetate derivative were identical with the reported values.^{1,2}

Compound **11** was also converted into the C-2 epimer of **1a** as its TFA salt **1b** and the di-acetate derivative **2b** was prepared using the same strategy (**Scheme 3**). The spectral data of diacetate **2b** is matched with the C-12 side chain derivative, which was synthesized earlier.¹³ Compound **1b** is also reported as synthetic derivative of phytostrissamine.¹⁴

In conclusion we have developed a strategy for the synthesis of pachastrissamine (jaspine B) **1a** and its diacetate derivative **2a** from Garner's aldehyde. The C-2 epimer of pachastrissamine (jaspine B) was also synthesized.

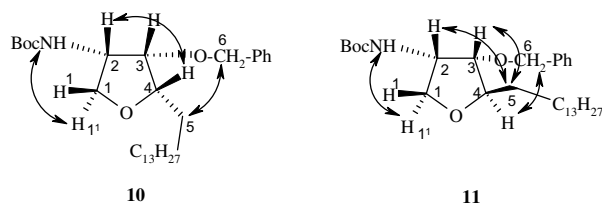
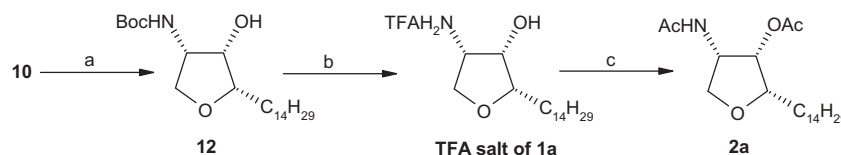


Figure 1. NOEs observed in compounds **10** and **11**.

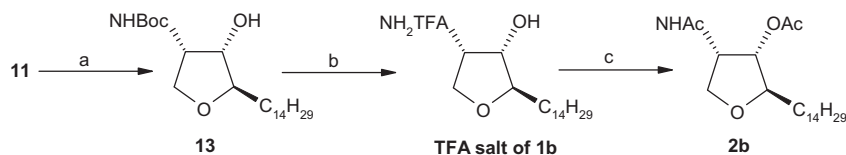


Scheme 2. Reagents and conditions: (a) Na/liq. NH₃, THF, –78°C, 30 min, 96%, (b) 50% TFA–DCM, rt, 6 h, 87%, (c) Et₃N, Ac₂O, DCM, 0°C–rt, 4 h, 94% (overall yield for **2a** is 22.3%).

1. Spectral data

1.1. TFA salt of **1b**

[α]_D²⁵ +14.46 (*c* 1.5, EtOH); (¹H NMR, 300 MHz, CD₃OD): δ 0.89 (t, 3H, *J* = 7.3 Hz), 1.22–1.39 (m,



Scheme 3. Reagents and conditions: (a) Na/liq. NH_3 , THF, -78°C , 30 min, 96%, (b) 50% TFA–DCM, rt, 6 h, 89%, (c) Et_3N , Ac_2O , DCM, 0°C –rt, 4 h, 94% (overall yield for **2b** is 9.8%).

22H), 1.41–1.67 (m, 4H), 3.64–3.75 (m, 3H), 3.98–4.04 (m, 1H), 4.10–4.17 (m, 1H); ^{13}C NMR (CD_3OD , 125 MHz): δ 85.20, 74.33, 69.36, 53.65, 34.02, 32.95, 30.67, 30.65, 30.59, 30.55, 30.35, 26.76, 23.61, 14.34; electrospray analysis: m/z 300 ($\text{M}^+ + 1 - \text{CF}_3\text{COOH}$).

1.2. Compound 10

$[\alpha]_D^{25} + 1.08$ (c 1.1, CHCl_3); ^1H NMR (500 MHz, CDCl_3): δ 0.88 (t, 3H, $J = 6.85\text{ Hz}$), 1.26 (br s, 24H), 1.44 (br s, 9H, $\text{CH}_3\text{-Boc}$), 1.62 (m, 2H, H-5), 3.61 (dd, 1H, $J_{1,1'} = 8.3\text{ Hz}$, $J_{1,2} = 7.2\text{ Hz}$, H-1'), 3.80 (dt, 1H, $J_{3,4} = 4.5\text{ Hz}$, $J_{4,\text{CH}_2} = 6.4\text{ Hz}$, H-4), 3.91 (dd, 1H, $J_{1,1'} = 8.3\text{ Hz}$, $J_{1,2} = 7.7\text{ Hz}$, H-1), 3.95 (dd, 1H, $J_{3,4} = 4.5\text{ Hz}$, $J_{2,3} = 6.0\text{ Hz}$, H-3), 4.35 (dddd, $J_{1,2} = 6.7\text{ Hz}$, $J_{1,2} = 7.4\text{ Hz}$, $J_{2,3} = 6.2\text{ Hz}$, $J_{\text{NH},2} = 7.2\text{ Hz}$, H-2), 4.58 (s, 2H, H-6), 5.0 (d, 1H, $J = 8.3\text{ Hz}$, NH), 7.30–7.40 (m, 5H, phenyl); ^{13}C NMR (75 MHz, CDCl_3): δ 155.55, 137.79, 128.47, 127.91, 81.90, 79.56, 79.14, 74.24, 70.56, 53.21, 31.89, 29.63, 29.56, 29.30, 28.37, 26.33, 22.63, 14.01; FABMS: m/z 490 ($\text{M}^+ + 1$).

1.3. Compound 11

$[\alpha]_D^{25} + 5.1$ (c 1, CHCl_3); ^1H NMR (500 MHz, CDCl_3): δ 0.88 (t, 3H, $J = 6.9\text{ Hz}$), 1.26 (br s, 24H), 1.45 (br s, 9H, $\text{CH}_3\text{-Boc}$), 1.47 (m, 2H, H-5), 3.57 (dd, 1H, $J_{1,1'} = 8.7\text{ Hz}$, $J_{1,2} = 7.4\text{ Hz}$, H-1'), 3.66 (dd, 1H, $J_{3,4} = 4.3\text{ Hz}$, $J_{2,3} = 6.2\text{ Hz}$, H-3), 3.83 (dt, 1H, $J_{3,4} = 4.3\text{ Hz}$, $J_{4,\text{CH}_2} = 6.3\text{ Hz}$, H-4), 4.10 (dd, 1H, $J_{1,1'} = 8.7\text{ Hz}$, $J_{1,2} = 6.7\text{ Hz}$, H-1), 4.22 (dddd, $J_{1,2} = 6.7\text{ Hz}$, $J_{1,2} = 7.4\text{ Hz}$, $J_{2,3} = 6.2\text{ Hz}$, $J_{\text{NH},2} = 7.2\text{ Hz}$, H-2), 4.53 (s, 2H, H-6), 5.10 (d, 1H, $J = 7.2\text{ Hz}$, NH), 7.28–7.40 (m, 5H, phenyl); ^{13}C NMR (75 MHz, CDCl_3): δ 155.64, 137.51, 128.48, 127.99, 127.90, 82.50, 81.58, 79.58, 72.24, 71.12, 51.54, 33.92, 31.90, 29.64, 29.54, 29.32, 28.37, 25.72, 22.65, 14.05; FABMS: m/z 490 ($\text{M}^+ + 1$).

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