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Original article

Discovery of novel type II c-Met inhibitors based on BMS-777607

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1. Introduction

c-Met is a prototype member of a subfamily of heterodimeric receptor tyrosine kinases (RTKs). The c-Met pathway is frequently deregulated in a wide variety of human cancers and plays critical roles in cancer formation, progression, and dissemination, as well as the resistance to approved therapies [1–4]. Therefore, the inhibition of increased c-Met signaling may have a significant impact on the treatment of human cancers in which the c-Met pathway is aberrantly activated. In fact, recent clinical trials of c-Met pathwaytargeted agents have yielded convincing evidence to support the potential utility of this class of agents in the treatment of various human cancers [5-7]. To date, the development of small molecular c-Met kinase inhibitors has made remarkable progress, resulting in more than ten candidates reaching clinical trials [8]. These known small molecular inhibitors have been categorized into two types based on their binding mode in the DFG motif (aspartate-phenylalanine-glycine) of the c-Met activation loop. Type I inhibitors bind the DFG-in conformation with a U-shaped geometry and are defined as ATP-competitive inhibitors of the activated kinase. In contrast to type I inhibitors, those that bind the inactivated DFG-out

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ABSTRACT

Twenty-two new analogs based on the structure of BMS-777607 were designed, synthesized, and evaluated to determine their biological activities. Compounds bearing a cyclic sulfonamide or α -chlor-opiperidone scaffold exhibited good activity, which may provide a new basis for further structural optimization. Quinoline-containing analogs exhibited better results than did their counterparts with an aminopyrimidine, aminopyridine, or pyrrolopyridine unit. Two analogs, **22d** and **22e**, stood out as the most potent c-Met inhibitors with IC₅₀s of 0.9 and 1.7 nM, respectively. These two compounds were more potent than BMS-777607 in enzymatic inhibition and cell proliferation studies.

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conformation are defined as type II inhibitors. This type of compounds binds to the same area occupied by the type I inhibitors but also exploits hydrogen bonding and hydrophobic interactions with the allosteric site [9,10].

As disclosed recently, certain mutations near the active site of c-Met may cause resistance of type I inhibitors. In contrast, type II inhibitors are postulated to be more effective against these mutations because their binding interactions extend beyond the entrance to c-Met's active site [11–13]. Initiated by Kirin Brewery Company in 2003 [14], numerous type II inhibitors with different structures have been reported in the past ten years, and some of these are currently in clinical trials or pre-clinical development (Fig. 1) [9]. A good example of these type II inhibitors is BMS-777067, which inhibits the kinase activity of c-Met (IC₅₀ = 3.9 nM), as well as that of Axl (IC₅₀ = 1.1 nM), Ron (IC₅₀ = 1.8 nM), and Tyro3 (IC₅₀ = 4.3 nM) [15]. This compound is now in phase 2 trial because of its excellent *in vivo* efficacy and favorable pharmacokinetic and preclinical safety profiles.

As displayed in Fig 1, most of the type II inhibitors may be disconnected into four units according to their structures and subunit functions. Moiety A is a phenyl ring or *para*-substituted (usually fluorine) phenyl ring. As disclosed by the crystal structure of the c-Met kinase domain in complex with BMS-777607 [15], this aromatic ring occupies a deep hydrophobic pocket defined by three residues (F1134, L1195, and F1200). The main chain of moiety B is usually constituted by five atoms (i.e., six chemical







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Fig. 1. Some representative type II c-Met inhibitors and their structural characteristics.

bonds, as summarized recently by Gong et al. [16,17]), bearing at least one amide bond, with or without a ring on the side chain. In the compound BMS-777607, the carbonyl group in this moiety forms a hydrogen bond with residue D1222. The structure of moiety C is comparatively conserved, bearing a phenyl ring π -stacked with residue F1223 of the DFG motif. In contrast, the structure of moiety D is alterable, and fused pyridine derivates (substituted quinoline, pyrrolopyridine, and thienopyridine) and a simple substituted pyridine are tolerant. The role of the pyridine nitrogen is an anchor to the hinge region through its interaction with M1160 [15].

Based on the unique structure of BMS-777607, we designed and prepared two series of new analogs in this study (Fig. 2). In series 1, moieties D and C in the parent compound were fixed, whereas the pyridone fragment in moiety B was replaced with sulfonamide (cyclic or linear) or substituted piperidone. In addition, several minor modifications were performed on the phenyl ring of moiety A. In the second series, moieties A and C were fixed, and moiety D was replaced with other aromatic rings. Moiety B also underwent several modifications in this series.

2. Chemistry

Substituted *N*-phenyl sulfamoyl acetamides 7a-f were prepared according to the sequence outlined in Scheme 1. Commercially available ethyl 2-(chlorosulfonyl)acetate 1 and substituted anilines 2a-c were coupled in the presence of triethylamine to yield the corresponding linear sulfonamides 3a-c, respectively. The sixmembered cyclic sulfamoyl acetamide esters 3d-f were obtained through the treatment of 3a-c with 1-bromo-3-chloropropane. The hydrolysis of 3a-f gave 4a-f, and the coupling of these compounds with amine 5 [15] under standard conditions yielded the key intermediates **6a–f**. Finally, a Hoffman rearrangement resulted in **7a–f** at high yields.

For the preparation of isomeric sulfamoyl acetamides (11a-c)and isomeric six-membered sulfamoyl acetamides (11d-f) derivates, chloroacetyl chloride was first coupled with substituted anilines (2a-c) to give compounds 8a-c, which were then converted to sulfochlorides 9a-c in the presence of sodium sulfite followed by phosphorous pentachloride. The coupling of amine 5 with sulfochlorides 9a-c produced sulfamoyl acetamide analogs 10a-c, respectively. The treatment of 10a-c with 1,3bromocholoropropane in the presence of potassium carbonate provided the cyclic sulfamoyl acetamide analogs 10d-f. Finally, a Hoffman rearrangement delivered aminopyridines 11a-f from the amide precursors 10a-f, and these reactions were mediated by PhI(OAc)₂ (Scheme 2).

The synthesis of piperidone analogs is displayed in Scheme 3. The coupling of 2-piperidone acid **16a** with amine **5** smoothly produced the corresponding 2-piperidone analog **17a**. The treatment of **17a** with PhI(OAc)₂ gave the desired aminopyridine **18a**. In contrast, the coupling of α -bromo-2-piperidone acid **16b** with amine **5** in the presence of EDC·HCl (2.5 eq) and DMAP afforded the halo-exchanged α -chloro product **17b**. A similar procedure as that described above gave the final product **18b**.

For the preparation of the series 2 compounds, two building blocks (**19**, **20**) were prepared through known procedures [15,18,19]. Carboxylic acid **19a** was activated with SOCl₂ and then coupled with aromatic amine **20a**. The removal of the protecting group resulted in the desired analog **21a** at good yield. Compounds **21b** and **21c** were obtained using similar procedure without the deprotection step. For compounds **22a–e**, the substitution on the pyridone motif was transformed after the coupling of the building blocks **19b** and **20a–c** (Scheme 4).



Fig. 2. Design of two series of novel c-Met inhibitors.

3. Biology

3.1. c-Met kinase assay

The effects of the indicated compound on the activities of c-Met kinases were determined using enzyme-linked immunosorbent assays (ELISAs) with purified recombinant proteins. Briefly, 96-well plates were pre-coated with 20 μ g/mL poly (Glu,Tyr)_{4:1} (Sigma) as the substrate. A 50- μ L aliquot of 10 μ mol/L ATP solution diluted in kinase reaction buffer (50 mmol/L HEPES [pH 7.4], 50 mmol/L MgCl₂, 0.5 mmol/L MnCl₂, 0.2 mmol/L Na₃VO₄, and 1 mmol/L DTT) was added to each well, and 1 μ L of various concentrations of Yhhu3813 diluted in 1% DMSO (v/v) (Sigma) were then added to each reaction well. DMSO (1%, v/v) was used as the negative control. The kinase reaction was initiated by the addition of purified c-Met tyrosine kinase proteins diluted in 49 μ L of kinase reaction buffer. After

incubation for 60 min at 37 °C, the plate was washed three times with phosphate-buffered saline (PBS) containing 0.1% Tween 20 (T-PBS). Anti-phosphotyrosine (PY99) antibody (100 µL; 1:500, diluted in 5 mg/mL BSA T-PBS) was then added. After a 30-min incubation at 37 °C, the plate was washed three times, and 100 μ L of horseradish peroxidase-conjugated goat anti-mouse IgG (1:2000, diluted in 5 mg/mL BSA T-PBS) was added. The plate was then incubated at 37 °C for 30 min and washed three times. A 100-µL aliquot of a solution containing 0.03% H₂O₂ and 2 mg/mL o-phenylenediamine in 0.1 mol/L citrate buffer (pH 5.5) was added. The reaction was terminated by the addition of 50 µL of 2 mol/L H₂SO₄ when the color changed, and the plate was analyzed using a multi-well spectrophotometer (SpectraMAX 190, Molecular Devices) at 490 nm. The inhibition rate (%) was calculated using the following equation: $[1 - (A490/A490 \text{ control})] \times 100\%$. The IC₅₀ values were calculated from the inhibition curves obtained from two separate experiments.



Scheme 1. Reagents and conditions: a) Et₃N, toluene; b) 1-bromo-3-chloropropane, K₂CO₃, DMF; c) NaOH, EtOH; d) EDC·HCl, DMAP; e) Phl(OAC)₂.



Scheme 2. Reagents and conditions: a) Et₃N, CH₂Cl₂; b) Na₂SO₃, EtOH, H₂O; c)PCl₅; d) DIPEA, THF; e) 1-bromo-3-chloropropane, K₂CO₃, DMF; f) Phl(OAc)₂.

3.2. Western blot analysis

MKN45 cells were treated with an increasing dose of the indicated compound for 2 h at 37 °C and then lysed in $1 \times$ SDS sample buffer. The cell lysates were subsequently resolved by 10% SDS-PAGE and transferred to nitrocellulose membranes. The membranes were probed with the appropriate primary antibodies and then with horseradish peroxidase-conjugated anti-rabbit or anti-mouse IgG. The immunoreactive proteins were detected using an enhanced chemiluminescence detection reagent (Thermo Fisher).

3.3. Cell proliferation assay

Cells were seeded in 96-well tissue culture plates. The following day, the cells were exposed to various concentrations of the compounds and further cultured for 72 h. The cell proliferation was then determined using the sulforhodamine B (SRB, Sigma) or the thiazolyl blue tetrazolium bromide (MTT, Sigma) assay. The IC₅₀ values were calculated through fitting of the concentration—response curve using the four-parameter method.

4. Results and discussion

As illustrated in Table 1, most of the designed compounds belonging to series 1 (7a-f, 11a-f, 18a-b) showed moderate to

good inhibition of the c-Met enzyme. The compounds (7a-c) bearing a linear sulfonamide motif in the right site of moiety B (near moiety A) showed IC₅₀s ranging from 290 to 500 nM, whereas the presence of sulfonamide in a six-member ring (7d-7f) resulted in a 1.7–5.3-fold enhancement of the activity (7d vs. 7a, 7e vs. 7b, and **7f** vs. **7c**). The replacement of the amide bond near moiety C with linear sulfonamide (**11a–c**) slightly decreased the enzyme inhibition. However, the presence of cyclic sulfonamide at this position (11d-f) caused a loss of activity. The analog bearing saturated pyridone (i.e., piperidone) (18a) showed weaker activity compared to that bearing cyclic sulfonamide (7d-f). However, the substitution of the α -position of piperidone with chlorine (18b) resulted in a 5-fold enhancement in the activity (18b vs. 18a). The investigation of different groups (H, Me, or F) at the para-site of the phenyl ring in moiety A indicated that *para*-fluoride gave slightly better results (7c, 7f, and 11c). Although all of the compounds (7af, 11a-f, and 18a-b) belonging to series 1 that did not contain a pyridone moiety showed weaker activities compared to the positive control BMS-777607, the novel designed cyclic sulfonamide (7f) and α -chloropiperidone (18b) scaffolds lead to the development other new analogs with improved activity.

Most of the compounds belonging to series 2(21a-c, 22a-e) exhibited good to excellent inhibition against c-Met kinase. Of these, three compounds (22c-e) showed better activity than the positive control. If moiety B was fixed as un-substituted pyridone (21a-c), the analogs bearing pyrimidine (21a) or pyrrolopyridine



Scheme 3. Reagents and conditions: a) Cul, K₃PO₄, DMF, 90%; b) ¹BuLi, isobutyl chloroformate, 82%; c) LiOH, 87%; d) Br₂, Et₂O, 92%; e) EDC·HCl, DMAP, 76% for **17a**, 68% for **17b**; f) Phl(OAc)₂, 72% for **18a**, 76% for **18b**.



Scheme 4. Reagents and conditions: a) SOCl₂, DIPEA, THF; b) TFA; c) NaH, EtOH/THF; d) NaH, MeOH/THF.

(21b) showed better results ($IC_{50} = 20-39$ nM) than their counterpart containing a quinoline scaffold (21c). However, the weak activity of 21c may be caused by its poor solubility. A compound bearing the same scaffold (22d) displayed much better activity than its counterparts (22a, 22b), i.e., its activity was as low as 0.9 nM, which is 4-fold stronger than that of BMS-777607. In general, quinoline moiety-containing compounds gave better results (22d vs. 22a, 22b, and BMS-777607). The investigation of compounds with a quinoline unit in moiety D revealed that the substitution of pyridone in moiety B with iodide (22c) or methoxylation (22e) at the 4-position gave good results.

We then evaluated the inhibitory effect of increasing concentrations (0.1, 1, and 10 μ M) of the promising compounds on proliferation of BaF3/TPR-Met cells, which stably express a constitutively active c-Met due to a chromosomal rearrangement. As shown in Fig. 3, five compounds (**21b**, **22b**, **22c**, **22d**, and **22e**) displayed significant inhibitory activity against cell proliferation (>80%) at a concentration of 10 μ M, and three compounds (**22c**, **22d**, and **22e**) showed strong activity at a concentration of 1 μ M. In addition, compounds **22d** and **22e** continued to show significant inhibitory effects even at a concentration of 0.1 μ M, which indicates that these two compounds are more potent than BMS-777607. The IC₅₀ values of **22d** and **22e** with respect to the cell proliferation of the c-Metconstitutively activated MKN45 and BaF3/TPR-Met cells are shown in Table 2. Indeed, these two compounds showed approximately 5–9-fold higher potency than BMS-777607.

Based on the data above, **22d** and **22e** were proven to be potent c-Met inhibitors through both c-Met enzymatic and cell proliferation assays. To further determine whether the c-Met kinase inhibition of these two compounds in a cell-free system can be recapitulated *in vitro*, the cellular c-Met-targeting signal pathway of **22d** in c-Met constitutively activated MKN45 cancer cells was investigated. The results showed that **22d** inhibited c-Met phosphorylation in MKN45 cells in a dose-dependent manner, with complete abolishment at 0.1 μ M (Fig. 4). In addition, Erk1/2 and AKT, the key downstream molecules of c-Met that play important roles in c-Met functioning, were also significantly inhibited as a result of **22d** treatment (Fig. 4). These data support the finding that **22d** inhibits c-Met signaling and, in turn, suppresses c-Met-dependent cell proliferation.

5. Conclusion

In summary, two series of analogs based on BMS-777607 were designed, synthesized, and evaluated to determine their effect on c-Met inhibition. Modifications at moiety B revealed that the novel designed cyclic sulfonamide and α -chloropiperidone scaffolds may provide a new basis for further optimization. The quinoline moiety-containing analogs gave better result than their counterparts bearing aminopyrimidine, aminopyridine, or pyrrolopyridine. Two analogs, namely compounds **22d** and **22e**, stood out as the most potent c-Met inhibitors with IC₅₀s of 0.9 and 1.7 nM, respectively. These two compounds exhibited approximately 5–9-fold higher potency than did BMS-777607 with respect to the inhibition of cell proliferation. Further studies on the structural optimization of these derivatives are currently underway in our laboratory.

6. Experimental protocols

6.1. Chemistry

All chemical reagents were used as supplied unless indicated. Solvents used in organic reactions were distilled under an inert atmosphere. Unless otherwise noted, all reactions were carried out at room temperature and were performed under a positive pressure of argon. Flash column chromatography was performed on silica gel (200–300 mesh, Qingdao, China). Analytical thin layer chromatography (TLC) was performed on glass plates pre-coated with a 0.25 mm thickness of silica gel. ¹H NMR and ¹³C NMR spectra were taken on a Jeol JNM-ECP 600 or a Bruker Avance III 400 spectrometer at rt. Chemical shifts of the ¹H NMR spectra are R₁ c-Met IC₅₀ (nM)

 290.5 ± 18.6

 $_{3}$ CH₃ 494.7 \pm 145.4

 $\textbf{373.2} \pm \textbf{28.5}$

 $\textbf{168.7} \pm \textbf{26.4}$

 $CH_{3} \ 165.5 \pm 30.1$

 $\textbf{70.2} \pm \textbf{3.1}$

 100.8 ± 15.7

 \sim CH₃ 1044 \pm 108.9

 $H \qquad H \qquad H \qquad H \qquad N \qquad S \qquad H \qquad N \qquad S \qquad F \qquad 432.9 \pm 27.0$

Η

Η



Cmpd

7a

7b

7c

7d

7e

7f

11a

11b

c-Met enzymatic activity of the designed compounds.

Moiety D

Cl

 H_2N

CI

 H_2N

Cl

 H_2N

С

 H_2N

Cl

 H_2N

Cl

 H_2N

Cl

 H_2N

Cl

 H_2N

Cl

 H_2N

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Moiety B

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S O Table 1 (continued)

Cmpd	Moiety D	Moiety B	R ₁	c-Met IC ₅₀ (nM)
11d	CI H ₂ N N	N S H	∕s ^s H	Not detected
11e	CI H ₂ N N		⊂ _S S CH <u>a</u>	3 Not detected
11f			F 5 ⁵	Not detected
18a		N N N N	F	427.0 ± 6.1
18b			F	81.0 ± 7.6
21a		N N N N	۶	39.0 ± 0.2
21b		N N N N N N N N N N N N N N N N N N N	۶ کړ	20.0 ± 1.0
21c			۶	29.7% @ 10 μM
22a			F کړ	26.9 ± 3.9
22b			۶	49.6 ± 3.0
22c			F ک ^ر (contin	0.7 ± 0.1 ued on next page)

11c

Table 1 (continued)



The IC₅₀ values are shown as the means \pm SD (nM) from two separate experiments.

expressed in ppm relative to the solvent residual signal 7.26 in CDCl₃ or to tetramethylsilane ($\delta = 0.00$). Chemical shifts of the ¹³C NMR spectra are expressed in ppm relative to the solvent signal 77.00 in CDCl₃ or to tetramethylsilane ($\delta = 0.00$) unless otherwise noted. Electrospray (ESI) mass spectra were recorded on a Global Q-TOF mass spectrometer.

6.2. General methods for preparation of linear sulfonamide 3a-c

Sulfonyl chloride **1** (33 mmol) in THF (10 mL) was added dropwise to a solution of aromatic amine **2** (33 mmol) and triethylamine (4.4 mL, 35 mmol) in 100 mL THF at 0 °C. Upon completion of the addition, the reaction mixture was stirred at room temperature for 1 h before concentrated in vacuo. The residue was dissolved in EtOAc (100 mL), washed with saturated brine (3 \times 20 mL), dried over Na₂SO₄, and concentrated in vacuo. The crude product was purified by flash chromatography on silica gel to give compounds **3a–c**.

6.2.1. Ethyl 2-(N-phenylsulfamoyl)acetate (3a)

Yellow oil; yield 65%; ¹H NMR (CDCl₃, 600 MHz) δ 7.31–7.38 (m, 4H, ArH), 7.25–7.22 (m, 1H, ArH), 7.13 (s, 1H, NH), 4.26 (q, 2H, CH₂O), 3.94 (s, 2H, COCH₂SO₂), 1.30 (t, 3H, *J* = 7.1 Hz, OCH₂CH₃).

6.2.2. Ethyl 2-(N-p-tolylsulfamoyl)acetate (3b)

Yellow solid; yield 72%; ¹H NMR (CDCl₃, 600 MHz) δ 7.21–7.20 (m, 2H, ArH), 7.16 (d, 2H, J = 8.4 Hz, ArH), 6.96 (s, 1H, NH), 4.26 (q, 2H, CH₂O), 3.91 (s, 2H, COCH₂SO₂), 2.33 (s, 3H, CH₃), 1.31 (t, 3H, J = 7.1 Hz, OCH₂CH₃).



Fig. 3. Effect of the promising compounds on BF3/TPR-Met cell proliferation.

6.2.3. Ethyl 2-(N-4'-fulorophenylsulfamoyl)acetate (3c)

Yellow solid; yield 65%; ¹H NMR (CDCl₃, 600 MHz) δ 7.33–7.31 (m, 2H, ArH), 7.09 (s, 1H, NH), 7.07–7.04 (m, 2H, ArH), 4.26 (q, 2H, CH₂O), 3.91 (s, 2H, COCH₂SO₂), 1.31 (t, 3H, J = 7.2 Hz, OCH₂CH₃).

6.3. General methods for preparation of cyclic sulfonamide **3d**-f

A solution of 1-bromo-3-chloropropane (600 μ L, 930 mg, 6 mmol) in DMF (60 mL) was added during 1.5 h to a mixture of sulfonamide **3a–c** (5 mmol) and potassium carbonate (2.07 g, 15 mmol) in DMF (60 mL) at 60 °C. This reaction mixture was stirred until the end of the reaction (checked by TLC). The reaction mixture was then diluted with water (150 mL), acidified with conc. HCl to pH = 1, and extracted with dichloromethane (3 × 50 mL). The organic phase was washed with 2% HCl (3 × 10 mL), dried over sodium sulfate, and evaporated to dryness. The product was recrystallized from a mixture of diethyl ether and hexane.

6.3.1. Ethyl 2-phenyl-1,2-thiazinane-1,1-dioxide-6-carboxylate (3d)

Yellow solid; yield: 75%; ¹H NMR (600 MHz, CDCl₃) δ 7.41–7.30 (m, 4H, ArH), 7.30–7.24 (m, 1H, ArH), 4.35–4.19 (m, 2H, CH₂O), 4.04 (dd, 1H, *J* = 9.9, 4.1 Hz, CH), 4.02–3.92 (m, 1H, CHH), 3.77–3.67 (m, 1H, CHH), 2.65–2.54 (m, 1H, CHH), 2.54–2.46 (m, 1H, CHH), 2.15–2.06 (m, 1H, CHH), 1.98–1.88 (m, 1H, CHH), 1.29 (t, 3H, *J* = 7.1 Hz, CH₂CH₃).

6.3.2. Ethyl 2-(p-tolyl)-1,2-thiazinane-1,1-dioxide-6-carboxylate (**3e**)

Yellow solid; yield 74%; ¹H NMR (600 MHz, CDCl₃) δ 7.24–7.18 (m, 2H, ArH), 7.17–7.13 (m, 2H, ArH), 4.41–4.17 (m, 2H, CH₂O), 4.03 (dd, 1H, *J* = 10.1, 4.1 Hz, COCHSO₂), 3.99–3.87 (m, 1H, CHH), 3.75–3.59 (m, 1H, CHH), 2.67–2.53 (m, 1H, CHH), 2.53–2.40 (m, 1H, CHH), 2.33 (s, 3H, CH₃), 2.16–2.01 (m, 1H, CHH), 2.00–1.86 (m, 1H, CHH), 1.29 (t, 3H, *J* = 7.1 Hz, CH₂CH₃).

6.3.3. Ethyl 2-(4-fluorophenyl)-1,2-thiazinane-1,1-dioxide-6-carboxylate (**3f**)

Yellow solid; yield 77%; ¹H NMR (600 MHz, CDCl₃) δ 7.35–7.27 (m, 2H, ArH), 7.08–6.99 (m, 2H, ArH), 4.35–4.17 (m, 2H, CH₂O), 4.04 (dd, 1H, *J* = 9.5, 4.2 Hz, COCHSO₂), 3.94–3.86 (m, 1H, CH*H*), 3.71–3.62 (m, 1H, CH*H*), 2.62–2.54 (m, 1H, CH*H*), 2.54–2.47 (m, 1H, CH*H*), 2.20–2.04 (m, 1H, CH*H*), 1.98–1.83 (m, 1H, CH*H*).

6.4. General methods for preparation of amide **6a**-**f**

Ethyl ester **3** (3 mmol) was treated with KOH (672 mg, 12 mmol) in a mixed solvent of methanol (5 mL) and water (5 mL) for 3 h. After most of the solvent was evaporated, the solution was acidified to pH 1 with 1 M HCl and extracted with EtOAc (3×20 mL). The organic extracts were combined and washed with brine (2×5 mL). Evaporation of the solvent gave the corresponding acid, which was used directly in the next step.

EDC·HCl (1.2 g, 6.25 mmol) was added to a suspension of the carboxylic acid and the amine [15] (**5**, 705 mg, 2.5 mmol) in THF (25 mL) at 0 °C followed by DMAP (30 mg, 0.25 mmol). The reaction mixture was warmed to room temperature and stirred overnight. After diluted with EtOAc (150 mL), the whole mixture was washed with 1 M HCl (3 × 10 mL), 5% NaHCO₃ (3 × 10 mL), and brine (3 × 10 mL), dried over Na₂SO₄, and concentrated in vacuo. The residue was purified by column chromatography to give corresponding amides **6a**–**f**.

Table 2Effects of 22d and 22e on cell proliferation (nM).

Cmpd	BaF3/TPR-Met	MKN45
22d 22e BMS777607	$19.5 \pm 8.8 \\ 36.3 \pm 1.2 \\ 188.4 \pm 9.3$	$\begin{array}{c} 39.7 \pm 9.1 \\ 57.7 \pm 6.6 \\ 285.8 \pm 22.7 \end{array}$

The IC ₅₀ valu	es are shown a	s the means \pm SD	(nM)	from two se	parate exp	periments
			· · ·			



Fig. 4. 22d suppresses c-Met phosphorylation and downstreams signaling in MKN45 cells.

6.4.1. 3-Chloro-4-(2-fluoro-4-(2-(N-phenylsulfamoyl)acetamido) phenoxy)picolinamide (**6a**)

Gray solid; yield 55%; ¹H NMR (DMSO- d_6 , 600 MHz) δ 10.74 (s, 1H, NH), 10.13 (s, 1H, NH), 8.34 (d, 1H, J = 5.5 Hz, ArH), 8.08 (s, 1H, NH), 7.82 (dd, 1H, J = 12.8, 2.3 Hz, ArH), 7.78 (s, 1H, NH), 7.45–7.34 (m, 4H, ArH), 7.30 (s, 1H, NH), 7.29 (d, 1H, J = 7.4 Hz, ArH), 7.15–7.12 (m, 1H, ArH), 6.83 (d, 1H, J = 5.0 Hz), 4.18 (s, 1H, COCH₂SO₂); ¹³C NMR (DMSO- d_6 , 150 MHz) δ 167.1, 162.9, 161.0, 160.5, 154.8, 152.7, 149.4, 138.2, 135.9, 129.8, 124.8, 124.4, 121.2, 119.7, 116.9, 116.7, 111.3, 108.8, 58.0.

6.4.2. 3-Chloro-4-(2-fluoro-4-(2-(N-(p-tolyl)sulfamoyl)acetamido) phenoxy)picolinamide (**6b**)

White solid; yield 57%; ¹H NMR (DMSO- d_6 , 600 MHz) δ 10.71 (s, 1H, NH), 9.95 (s, 1H, NH), 8.33 (d, 1H, J = 6.1 Hz, ArH), 8.07 (s, 1H, NH), 7.83 (dd, 1H, J = 12.8, 1.9 Hz, ArH), 7.78 (s, 1H, NH), 7.44 (t-like, 1H, J = 9.0, 8.7 Hz, ArH), 7.38 (d, 1H, J = 8.7 Hz, ArH), 7.17 (m, 4H, ArH), 6.83 (d, 1H, J = 5.0 Hz), 4.11 (s, 1H, COCH₂SO₂), 2.26 (s, 1H, CH₃); ¹³C NMR (DMSO- d_6 , 150 MHz) δ 167.1, 161.1, 160.5, 154.8, 154.3, 152.7, 149.4, 138.4, 135.9, 135.5, 134.3, 130.1, 124.4, 121.9, 116.9, 116.7, 111.3, 108.8, 57.7, 20.9.

6.4.3. 3-Chloro-4-(2-fluoro-4-(2-(N-(4-fluorophenyl)sulfamoyl) acetamido)phenoxy)picolinamide (**6c**)

White solid; yield 48%; ¹H NMR (DMSO-*d*₆, 600 MHz) δ 10.73 (s, 1H, NH), 10.11 (s, 1H, NH), 8.33 (d, 1H, *J* = 5.5 Hz, ArH), 8.08 (s, 1H, NH), 7.83 (dd, 1H, *J* = 12.8, 1.9 Hz, ArH), 7.78 (s, 1H, NH), 7.45–7.36 (m, 2H, ArH), 7.33–7.31 (m, 2H, ArH), 7.23–7.19 (m, 2H, ArH), 6.83 (d, 1H, *J* = 5.5 Hz), 4.13 (s, 1H, COCH₂SO₂); ¹³C NMR (DMSO-*d*₆, 150 MHz) δ 167.1, 161.1, 160.5, 159.1, 154.8, 149.4, 138.3, 135.9, 134.3, 124.4, 116.9, 116.7, 116.5, 116.4, 111.3, 108.8, 57.8.

6.4.4. N-(4-((2-carbamoyl-3-chloropyridin-4-yl)oxy)-3fluorophenyl)-2-phenyl-1,2-thiazinane-6-carboxamide 1,1-dioxide

(*6d*) White solid; yield 43%; ¹H NMR (600 MHz, Acetone- d_6) δ 9.73 (s, 1H, NH), 8.34 (d, J = 5.5 Hz, 1H, NH), 7.94 (dt, 1H, J = 8.5, 1.0 Hz, ArH), 7.72 (dt, 1H, J = 8.3, 1.0 Hz, ArH), 7.58–7.53 (m, 1H, ArH), 7.49 (dt, 1H, J = 8.9, 1.7 Hz, ArH), 7.47–7.36 (m, 5H, ArH), 6.99 (s, 1H, NH), 6.91 (d, 1H, *J* = 5.5 Hz, ArH), 4.30 (dd, 1H, *J* = 10.9, 3.7 Hz, COCHSO₂), 4.06–3.94 (m, 1H, CH*H*), 3.82–3.71 (m, 1H, CH*H*), 2.77–2.62 (m, 1H, CH*H*), 2.54–2.42 (m, 1H, CH*H*), 2.17–2.05 (m, 2H, CH₂); ¹³C NMR (150 MHz, Acetone-*d*₆) δ 162.9, 162.0, 153.7, 152.6, 148.9, 142.3, 142.2, 129.9, 129.8, 127.9, 127.8, 127.7, 127.6, 124.5, 117.2, 112.2, 100.8, 66.0, 61.3, 60.5, 54.1, 36.2, 28.1, 24.0, 23.8, 14.4.

6.4.5. N-(4-((2-carbamoyl-3-chloropyridin-4-yl)oxy)-3-

fluorophenyl)-2-(p-tolyl)-1,2-thiazinane-6-carboxamide 1,1-dioxide (6e)

White solid; yield 55%; ¹H NMR (600 MHz, Acetone- d_6) δ 9.73 (s, 1H, NH), 8.33 (d, 1H, J = 5.6 Hz, ArH), 7.94 (dt, 2H, J = 8.4, 0.9 Hz, ArH), 7.72 (dt, 2H, J = 8.4, 1.0 Hz, ArH), 7.55 (ddd, 1H, J = 8.2, 6.9, 0.9 Hz, ArH), 7.51–7.47 (m, 1H, ArH), 7.43 (ddd, 1H, J = 8.4, 6.9, 1.0 Hz, ArH), 7.37 (t, 1H, J = 8.8 Hz, ArH), 7.30–7.26 (m, 2H, ArH), 7.24–7.18 (m, 2H, ArH), 6.99 (s, 1H, NH), 6.91 (dd, 1H, J = 5.5, 1.1 Hz, ArH), 4.28 (dd, 1H, J = 10.9, 3.7 Hz, COCHSO₂), 4.03–3.95 (m, 1H, CHH), 3.70 (m, 1H, CHH), 2.73–2.62 (m, 1H, CHH), 2.53–2.44 (m, 1H, CHH), 2.33 (s, 3H, CH₃), 2.12–2.06 (m, 2H, CH₂); ¹³C NMR (150 MHz, Acetone- d_6) δ 163.0, 162.9, 155.3, 153.7, 152.6, 148.9, 138.2, 137.1, 129.9, 124.5, 117.0, 116.5, 112.2, 65.9, 65.9, 54.3, 36.1, 28.2, 23.7, 14.4.

6.4.6. N-(4-((2-carbamoyl-3-chloropyridin-4-yl)oxy)-3fluorophenyl)-2-(4-fluorophenyl)-1,2-thiazinane-6-carboxamide

1,1-dioxide (**6***f*) White solid; yield 49%; ¹H NMR (600 MHz, Acetone- d_6) δ 9.73 (s, 1H, NH), 8.34 (d, 1H, J = 5.5 Hz, ArH), 7.98–7.89 (m, 1H, ArH), 7.65 (s, 1H, ArH), 7.53–7.47 (m, 1H, ArH), 7.47–7.43 (m, 2H, ArH), 7.37 (t, 1H, J = 8.9 Hz, ArH), 7.22–7.15 (m, 2H, ArH), 6.94 (s, 1H, NH), 6.91 (dd, 1H, J = 5.5, 1.1 Hz, ArH), 4.32 (dd, 1H, J = 10.6, 3.7 Hz, COCHSO₂), 4.02–3.96 (m, 1H, CHH), 3.77–3.72 (m, 1H, CHH), 2.73–2.61 (m, 1H, CHH), 2.54–2.43 (m, 1H, CHH), 2.17–2.07 (m, 2H, CH₂); ¹³C NMR (150 MHz, Acetone- d_6) δ 162.2, 161.2, 151.8, 148.1, 138.8, 137.0, 129.6, 129.5, 127.0, 126.8, 123.7, 116.3, 111.4, 65.1, 59.7, 53.4, 27.4, 23.1, 20.1.

6.5. General methods for preparation of compounds 7a-f

To amide **6** (0.2 mmol) in ethyl acetate (2 mL), acetonitrile (2 mL), and water (1 mL) at 0 °C was added iodobenzene diacetate (82 mg, 0.26 mmol). After stirring at room temperature for 2 h, saturated NaHCO₃ (3 mL) was added, followed by 30 mL of ethyl acetate. The mixture was filtered, and the filtrate was washed with brine (3 × 5 mL), dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by flash chromatography on silica gel to give compounds **7a–f**.

6.5.1. N-(4-((2-amino-3-chloropyridin-4-yl)oxy)-3-fluorophenyl)-2-(N-phenylsulfamoyl)acetamide (**7a**)

White solid; yield 67%; ¹H NMR (600 MHz, Methanol-*d*₄) δ 7.73 (dd, 1H, *J* = 12.6, 2.5 Hz, ArH), 7.69 (d, 1H, *J* = 5.9 Hz, ArH), 7.39–7.23 (m, 5H, ArH), 7.22–7.07 (m, 2H, ArH), 5.96 (dd, 1H, *J* = 5.9, 0.9 Hz, ArH); ¹³C NMR (150 MHz, Methanol-*d*₄) δ 162.5, 162.1, 156.0, 154.4, 147.6, 147.5, 138.5, 138.4, 130.3, 126.2, 124.4, 122.9, 117.4, 110.2, 110.1, 102.1; MS (ESI pos ion) *m*/*z*: calcd for C₁₉H₁₆ClFN₄O₄S, 450.1; found, 450.9 (M+H).

6.5.2. N-(4-((2-amino-3-chloropyridin-4-yl)oxy)-3-fluorophenyl)-2-(N-(p-tolyl)sulfamoyl)acetamide (**7b**)

White solid; yield 71%; ¹H NMR (600 MHz, Acetone- d_6) δ 9.84 (s, 1H, NH), 8.76 (s, 1H, NH), 7.85 (d, 1H, J = 12.8 Hz, ArH), 7.80 (d, 1H, J = 5.8 Hz, ArH), 7.39 (d, 2H, J = 8.8 Hz, ArH), 7.34 (d, 2H, J = 8.23 Hz, ArH), 7.29 (t, 1H, J = 8.8 Hz, ArH), 7.19 (d, 3H, J = 8.0 Hz, ArH), 6.03 (d, 1H, J = 5.5 Hz, ArH), 5.88 (s, 1H, NH), 5.87 (s, 1H, NH), 4.12 (s, 2H, COCH₂SO₂), 2.31 (s, 3H, CH₃); ¹³C NMR (150 MHz, Acetone- d_6) δ 161.7, 161.1, 158.4, 155.5, 153.8, 148.1, 135.9, 135.8, 130.6, 124.3,

123.4, 116.8, 116.7, 109.4, 109.3, 101.8, 57.3, 20.8; MS (ESI pos ion) *m*/*z*: calcd for C₂₀H₁₈ClFN₄O₄S, 464.1; found, 465.1 (M+H).

6.5.3. N-(4-((2-amino-3-chloropyridin-4-yl)oxy)-3-fluorophenyl)-2-(N-(4-fluorophenyl)sulfamoyl)acetamide (7c)

White solid; yield 73%; ¹H NMR (600 MHz, Methanol- d_4) δ 7.75 (dd, 1H, J = 12.6, 2.5 Hz, ArH), 7.69 (d, 1H, J = 5.9 Hz, ArH), 7.41–7.34 (m, 2H, ArH), 7.31–7.24 (m, 1H, ArH), 7.18 (t, 1H, J = 8.8 Hz, ArH), 7.09–7.02 (m, 2H, ArH), 5.96 (dd, 1H, J = 5.9, 1.0 Hz, ArH); ¹³C NMR (150 MHz, Methanol- d_4) δ 162.7, 162.5, 158.7, 156.0, 154.4, 147.4, 138.5, 134.7, 125.9, 125.8, 124.5, 117.4, 116.9, 116.8, 110.2, 102.1; MS (ESI pos ion) m/z: calcd for C₁₉H₁₅ClF₂N₄O₄S, 468.0; found, 469.0 (M+H).

6.5.4. N-(4-((2-amino-3-chloropyridin-4-yl)oxy)-3-fluorophenyl)-2-phenyl-1,2-thiazinane-6-carboxamide 1,1-dioxide (**7d**)

White solid; yield 72%; ¹H NMR (600 MHz, CDCl₃) δ 9.09 (s, 1H, NH), 7.87–7.54 (m, 2H, ArH), 7.33–7.22 (m, 5H, ArH), 7.20 (s, 1H, NH), 7.16–7.11 (m, 1H, ArH), 7.03 (t, 1H, *J* = 8.6 Hz, ArH), 5.94 (dd, 1H, *J* = 6.0, 1.0 Hz, ArH), 5.48 (s, 2H, NH₂), 4.09 (dd, 1H, *J* = 9.8, 4.0 Hz, COCHSO₂), 3.96–3.85 (m, 1H, CHH), 3.75–3.65 (m, 1H, CHH), 2.70–2.59 (m, 1H, CHH), 2.59–2.50 (m, 1H, CHH), 2.12–2.01 (m, 1H, CHH), 2.00–1.86 (m, 1H, CHH); ¹³C NMR (150 MHz, CDCl₃) δ 162.0, 161.7, 155.1, 154.6, 153.0, 142.0, 129.5, 128.0, 127.3, 123.3, 116.5, 116.4, 109.8, 109.6, 103.2, 101.8, 64.7, 53.8, 27.4, 23.4; MS (ESI pos ion) *m/z*: calcd for C₂₂H₂₀ClFN₄O₄S, 490.1; found, 490.9 (M+H).

6.5.5. N-(4-((2-amino-3-chloropyridin-4-yl)oxy)-3-fluorophenyl)-2-(p-tolyl)-1,2-thiazinane-6-carboxamide 1,1-dioxide (**7e**)

White solid; yield 67%; ¹H NMR (600 MHz, CDCl₃) δ 9.14 (s, 1H, NH), 7.72 (d, 1H, *J* = 6.3 Hz, ArH), 7.70 (dd, 2H, *J* = 12.0, 2.5 Hz, ArH), 7.24–7.19 (m, 4H, ArH), 7.17 (d, 2H, *J* = 8.3 Hz, ArH), 7.09 (t, 1H, *J* = 8.6 Hz, ArH), 6.01 (dd, 1H, *J* = 6.3, 1.0 Hz, ArH), 5.54 (s, 2H, NH₂), 4.16–4.10 (m, 1H, COCHSO₂), 3.97–3.88 (m, 1H, CHH), 3.78–3.67 (m, 1H, CHH), 2.75–2.65 (m, 1H, CHH), 2.65–2.55 (m, 1H, CHH), 2.33 (s, 3H, CH₃), 2.13–2.06 (m, 1H, CHH), 2.03–1.94 (m, 1H, CHH); ¹³C NMR (150 MHz, CDCl₃) δ 161.6, 161.3, 155.8, 154.8, 153.1, 144.1, 138.3, 137.6, 137.1, 136.5, 130.1, 127.1, 123.3, 116.5, 116.4, 112.7, 109.8, 109.7, 102.9, 101.9, 64.5, 53.8, 27.3, 23.4, 21.1; MS (ESI pos ion) *m/z*: calcd for C₂₃H₂₂ClFN₄O₄S, 504.1; found, 504.9 (M+H).

6.5.6. N-(4-((2-amino-3-chloropyridin-4-yl)oxy)-3-fluorophenyl)-2-(4-fluorophenyl)-1,2-thiazinane-6-carboxamide 1,1-dioxide (**7f**)

White solid; yield 77%; ¹H NMR (600 MHz, CDCl₃) δ 9.12 (s, 1H, NH), 7.73 (d, 1H, *J* = 6.0 Hz, ArH), 7.70 (dd, 1H, *J* = 11.9, 2.5 Hz, ArH), 7.34–7.29 (m, 2H, ArH), 7.21–7.17 (m, 1H, ArH), 7.09 (t, 1H, *J* = 8.6 Hz, ArH), 7.07–7.03 (m, 2H, ArH), 6.00 (dd, 1H, *J* = 6.0, 1.0 Hz, ArH), 5.41 (s, 2H, NH₂), 4.15 (dd, 1H, *J* = 9.6, 4.0 Hz, COCHSO₂), 3.94–3.86 (m, 1H, CHH), 3.74–3.68 (m, 1H, CHH), 2.75–2.65 (m, 1H, CHH), 2.65–2.56 (m, 1H, CHH), 2.20–2.08 (m, 1H, CHH), 2.02–1.90 (m, 1H, CHH); ¹³C NMR (150 MHz, CDCl₃) δ 162.8, 161.3, 161.2, 161.0, 156.0, 154.8, 153.1, 144.8, 136.3, 135.9, 129.2, 123.3, 116.4, 116.3, 116.2, 116.1, 109.6, 102.8, 101.9, 64.5, 60.5, 53.9, 27.3, 23.3, 14.2; MS (ESI pos ion) *m/z*: calcd for C₂₃H₂₂ClF₂N₄O₄S, 508.1; found, 508.9 (M+H).

6.6. General methods for preparation of compounds **8a**–*c*

Triethylamine (14 mL) was added to a solution of the aromatic amine (**2**, 50 mmol) in dry DCM (100 mL) at 0 °C. The mixture was stirred at room temperature for another 5 h after the addition of 2-chloroacetyl chloride (5.65 mL) at 0 °C. Ethyl acetate (150 mL) was added; the whole organic layer was washed with 1 M HCl (3×30 mL), saturated NaHCO₃ (2×20 mL), and brine (2×20 mL). The organic solvent was dried over Na₂SO₄ and then concentrated

in vacuo. The crude product was purified by recrystallization from ethyl acetate and hexane.

6.6.1. 2-Chloro-N-phenylacetamide (8a)

Gray solid; yield 89%; ¹H NMR (600 MHz, CDCl₃) δ 8.30−8.22 (m, 1H, ArH), 7.57−7.50 (m, 2H, ArH), 7.36 (dd, 2H, *J* = 8.5, 7.4 Hz, ArH), 7.21−7.14 (m, 1H, ArH), 4.18 (s, 2H, CH₂).

6.6.2. 2-Chloro-N-(p-tolyl)acetamide (8b)

Gray solid; yield 93%; ¹H NMR (600 MHz, CDCl₃) δ 8.22 (s, 1H, NH), 7.43–7.40 (m, 1H, ArH), 7.17–7.14 (m, 1H, ArH), 4.17 (s, 1H, CH₂), 2.33 (s, 3H, CH₃).

6.6.3. 2-Chloro-N-(4-fluorophenyl)acetamide (8c)

Gray solid; yield 95%; ¹H NMR (600 MHz, CDCl₃) δ 8.23 (s, 1H, NH), 7.53–7.49 (m, 2H, ArH), 7.07–7.03 (m, 2H, ArH), 4.19 (s, 1H, CH₂).

6.7. General methods for preparation of amide 10a-c

Sulfonyl chloride **9** (0.6 mmol) was diluted with dry THF (2 mL) and added to a solution of amine **5** (141 mg, 0.5 mmol) and DIPEA (174 μ L, 1 mmol) in dry THF (10 mL). The mixture was stirred for another 1 h before concentrated in vacuo. The residue was redissolved in ethyl acetate (50 mL) and washed with brine (3 × 10 mL). After dried over Na₂SO₄ and concentrated in vacuo, the residue was purified by column chromatography to give corresponding amide **10a**–c.

6.7.1. 3-Chloro-4-(2-fluoro-4-(2-oxo-2-(phenylamino) ethylsulfonamido)phenoxy)picolinamide (**10a**)

Yellow solid; yield 65%; ¹H NMR (600 MHz, DMSO- d_6) δ 10.49 (s, 1H, NH), 10.36 (s, 1H, NH), 8.34 (d, 1H, J = 5.6 Hz, ArH), 8.06 (d, 1H, J = 2.3 Hz, ArH), 7.76 (s, 1H, NH), 7.58–7.54 (m, 2H, ArH), 7.42 (t, 1H, J = 9.0 Hz, ArH), 7.36–7.29 (m, 3H, ArH), 7.19 (dd, 1H, J = 8.5, 2.5 Hz, ArH), 7.10 (t, 1H, J = 7.3 Hz, ArH), 6.79 (d, 1H, J = 5.6 Hz, ArH), 4.30 (s, 2H, CH₂); ¹³C NMR (150 MHz, DMSO- d_6) δ 167.6, 166.5, 160.0, 159.8, 154.2, 154.0, 152.4, 148.7, 138.3, 137.3, 128.9, 124.0, 119.2, 117.2, 116.4, 110.6, 109.1, 109.0, 57.7.

6.7.2. 3-Chloro-4-(2-fluoro-4-(2-oxo-2-(p-tolylamino) ethylsulfonamido)phenoxy)picolinamide (**10b**)

Yellow solid; yield 59%; ¹H NMR (600 MHz, DMSO- d_6) δ 10.47 (s, 1H, NH), 10.28 (s, 1H, NH), 8.33 (d, J = 5.6 Hz, 1H, ArH), 8.07 (s, 1H, NH), 7.76 (s, 1H, NH), 7.47–7.39 (m, 3H, ArH), 7.32 (dd, 1H, J = 12.2, 2.5 Hz, ArH), 7.21–7.16 (m, 1H, ArH), 7.13 (d, J = 8.3 Hz, 2H, ArH), 6.79 (d, 1H, J = 5.6 Hz, ArH), 4.27 (s, 2H, SO₂CH₂CO), 2.25 (s, 3H, CH₃); ¹³C NMR (150 MHz, DMSO- d_6) δ 166.6, 159.9, 159.6, 154.2, 154.0, 152.4, 149.4, 148.7, 137.3, 135.9, 133.1, 129.3, 124.0, 120.4, 119.3, 117.2, 116.4, 110.7, 109.1, 109.0, 105.9, 57.6, 20.5.

6.7.3. 3-Chloro-4-(2-fluoro-4-(2-((4-fluorophenyl)amino)-2oxoethylsulfonamido)phenoxy)picolinamide (**10c**)

Yellow solid; yield 61%; ¹H NMR (600 MHz, DMSO- d_6) δ 10.50 (s, 1H, NH), 10.44 (s, 1H, NH), 8.34 (d, 1H, J = 5.6 Hz, ArH), 8.06 (s, 1H, NH), 7.78–7.74 (m, 1H, ArH), 7.61–7.56 (m, 2H, ArH), 7.42 (t, 1H, J = 9.0 Hz, ArH), 7.31 (dd, 1H, J = 12.2, 2.5 Hz, ArH), 7.21–7.15 (m, 3H, ArH), 6.79 (d, 1H, J = 5.5 Hz, ArH), 4.28 (s, 2H, SO₂CH₂CO); ¹³C NMR (150 MHz, DMSO- d_6) δ 166.5, 166.4, 159.9, 159.8, 159.2, 157.6, 157.3, 154.2, 154.0, 152.4, 148.7, 135.7, 134.7, 124.0, 121.3, 121.1, 117.2, 116.4, 115.6, 115.4, 110.6, 109.1, 108.9, 57.6.

6.8. General methods for preparation of cyclic sulfonamide 10d-f

Similar to the preparation method of **3d**-f.

6.8.1. 2-(4-((2-Carbamoyl-3-chloropyridin-4-yl)oxy)-3-

fluorophenyl)-N-phenyl-1,2-thiazinane-6-carboxamide 1,1-dioxide (10d)

From **10a**; white solid; yield 52%; ¹H NMR (600 MHz, Acetoned₆) δ 9.39 (s, 1H, NH), 8.38 (d, 1H, J = 5.3 Hz, ArH), 7.69–7.65 (m, 2H, ArH), 7.48 (dd, 1H, J = 11.8, 2.5 Hz, ArH), 7.44 (t, 1H, J = 8.8 Hz, ArH), 7.40–7.37 (m, 1H, ArH), 7.35–7.31 (m, 2H, ArH), 7.13–7.09 (m, 1H, ArH), 6.95 (s, 1H, NH), 6.89 (dd, 1H, J = 5.4, 1.1 Hz, ArH), 4.39 (dd, 1H, J = 10.3, 3.9 Hz, CH), 4.09–4.02 (m, 1H, CHH), 3.92–3.86 (m, 1H, CHH), 2.73–2.62 (m, 1H, CHH), 2.54–2.46 (m, 1H, CHH), 2.23–2.13 (m, 1H, CHH), 2.13–2.07 (m, 1H, CHH); ¹³C NMR (150 MHz, Acetoned₆) δ 162.2, 161.6, 154.9, 153.3, 152.7, 148.9, 140.9, 139.9, 139.4, 129.6, 125.0, 124.7, 124.5, 123.8, 120.4, 120.3, 116.8, 116.6, 112.6, 66.0, 54.1, 28.3, 23.4.

6.8.2. 2-(4-((2-Carbamoyl-3-chloropyridin-4-yl)oxy)-3fluorophenyl)-N-(p-tolyl)-1,2-thiazinane-6-carboxamide 1,1dioxide (**10e**)

From **10b**; white solid; yield 59%; ¹H NMR (600 MHz, Acetoned₆) δ 9.29 (s, 1H, NH), 8.37 (d, 1H, J = 5.4 Hz, ArH), 7.65 (s, 1H, NH), 7.55–7.52 (m, 2H, ArH), 7.47 (dd, 1H, J = 11.8, 2.6 Hz, ArH), 7.43 (t, 1H, J = 8.8 Hz, ArH), 7.39–7.36 (m, 1H, ArH), 7.16–7.12 (m, 2H, ArH), 6.96 (s, 1H, NH), 6.89 (dd, 1H, J = 5.5, 1.1 Hz, ArH), 4.36 (dd, 1H, J = 10.2, 3.8 Hz, CH), 4.08–4.00 (m, 1H, CHH), 3.92–3.85 (m, 1H, CHH), 2.72–2.63 (m, 1H, CHH), 2.52–2.46 (m, 1H, CHH), 2.28 (s, 3H, CH₃), 2.20–2.13 (m, 1H, CHH), 2.12–2.06 (m, 1H, CHH); ¹³C NMR (150 MHz, Acetone- d_6) δ 161.2, 160.8, 152.5, 152.0, 148.2, 140.2, 139.2, 136.1, 133.6, 129.2, 123.8, 123.1, 119.7, 119.6, 115.9, 111.8, 104.3, 100.0, 65.2, 53.3, 27.5, 22.6, 20.0.

6.8.3. 2-(4-((2-Carbamoyl-3-chloropyridin-4-yl)oxy)-3fluorophenyl)-N-(4-fluorophenyl)-1,2-thiazinane-6-carboxamide 1,1-dioxide (**10**f)

From **10c**; white solid; yield 54%; ¹H NMR (600 MHz, DMSO- d_6) δ 10.40 (s, 1H, NH), 8.39 (d, 1H, J = 5.5 Hz, ArH), 8.06–8.04 (m, 1H, ArH), 7.95 (s, 1H, NH), 7.76 (s, 1H, NH), 7.64–7.60 (m, 2H, ArH), 7.56 (dd, 1H, J = 11.8, 2.6 Hz, ArH), 7.47 (t, 1H, J = 9.0 Hz, ArH), 7.35 (ddd, 1H, J = 5.5, 0.9 Hz, ArH), 4.33 (dd, 1H, J = 10.7, 3.8 Hz, CH), 3.99–3.91 (m, 1H, CHH), 3.83–3.76 (m, 1H, CHH), 2.55–2.47 (m, 1H, CHH), 2.40–2.33 (m, 1H, CHH), 2.01–1.93 (m, 2H, CHH); ¹³C NMR (150 MHz, DMSO- d_6) δ 167.0, 162.8, 161.9, 160.1, 159.7, 158.1, 154.0, 152.4, 149.4, 140.1, 139.0, 135.3, 124.3, 123.6, 121.8, 121.7, 117.3, 116.2, 116.1, 115.9, 111.7, 65.3, 56.3, 53.6, 36.3, 31.3, 30.1, 27.8, 22.6.

6.9. General method for preparation of **11a**-**f**

Similar to the preparation of **7a**–**f**.

6.9.1. 2-(N-(4-((2-amino-3-chloropyridin-4-yl)oxy)-3-fluorophenyl)sulfamoyl)-N-phenylacetamide (**11a**)

From **10a**; white solid; yield 72%; ¹H NMR (600 MHz, Acetone*d*₆) δ 9.58 (s, 1H, NH), 9.19 (s, 1H, NH), 7.79 (d, *J* = 5.7 Hz, 1H, ArH), 7.64–7.60 (m, 2H, ArH), 7.45 (dd, 1H, *J* = 12.0, 2.5 Hz, ArH), 7.37– 7.29 (m, 4H, ArH), 7.12 (tt, 1H, *J* = 7.4, 1.1 Hz, ArH), 6.02 (dd, 1H, *J* = 5.8, 1.0 Hz, ArH), 5.86 (s, 2H, NH₂), 4.25 (s, 2H, SO₂CH₂CO); ¹³C NMR (150 MHz, Acetone-*d*₆) δ 161.0, 160.9, 158.4, 155.7, 154.0, 148.3, 148.1, 139.3, 139.0, 138.9, 137.5, 137.3, 129.7, 125.1, 124.5, 120.3, 119.2, 119.1, 111.5, 111.3, 101.9, 57.8; MS (ESI pos ion) *m/z*: calcd for C₁₉H₁₆ClFN₄O₄S, 450.1; found, 450.9 (M+H).

6.9.2. 2-(N-(4-((2-amino-3-chloropyridin-4-yl)oxy)-3-fluorophenyl)sulfamoyl)-N-(p-tolyl)acetamide (**11b**)

From **10b**; white solid; yield 62%; ¹H NMR (600 MHz, Acetone*d*₆) δ 9.54 (s, 1H, NH), 7.78 (d, 1H, *J* = 5.7 Hz, ArH), 7.50 (dd, 2H, *J* = 8.6, 2.0 Hz, ArH), 7.44 (dd, 1H, *J* = 12.1, 2.5 Hz, ArH), 7.36–7.32 (m, 1H, ArH), 7.30 (t, 1H, *J* = 8.6 Hz, ArH), 7.14 (d, 2H, *J* = 8.3 Hz, ArH), 6.02 (dd, 1H, *J* = 5.7, 1.0 Hz, ArH), 5.87 (s, 2H, NH₂), 4.23 (s, 2H, SO₂CH₂CO), 2.28 (s, 3H, CH₃); ¹³C NMR (150 MHz, Acetone-*d*₆) δ 160.9, 160.8, 158.3, 155.6, 154.0, 148.0, 147.7, 138.9, 138.8, 137.4, 137.3, 136.8, 134.5, 130.0, 124.5, 120.3, 120.2, 119.1, 111.4, 111.3, 101.8, 57.6, 20.8; MS (ESI pos ion) *m*/*z*: calcd for C₂₀H₁₈ClFN₄O₄S, 464.1; found, 465.1 (M+H).

6.9.3. 2-(N-(4-((2-amino-3-chloropyridin-4-yl)oxy)-3-fluorophenyl)sulfamoyl)-N-(4-fluorophenyl)acetamide (**11c**)

From **10c**; white solid; yield 68%; ¹H NMR (600 MHz, Acetone*d*₆) δ 9.62 (s, 1H, NH), 9.04 (s, 1H, NH), 7.79 (d, 1H, *J* = 5.5 Hz, ArH), 7.68–7.62 (m, 2H, ArH), 7.44 (dd, 1H, *J* = 12.0, 2.4 Hz, ArH), 7.36– 7.29 (m, 2H, ArH), 7.14–7.08 (m, 2H, ArH), 6.02 (dd, 1H, *J* = 5.6, 0.9 Hz, ArH), 5.85 (s, 2H, NH₂), 4.24 (s, 2H, CH₂); ¹³C NMR (150 MHz, Acetone-*d*₆) δ 160.9, 160.8, 158.2, 155.5, 153.9, 147.9, 147.7, 138.8, 138.7, 137.3, 137.2, 135.5, 124.4, 122.2, 122.1, 122.0, 119.0, 118.9, 116.1, 115.9, 111.3, 111.1, 102.1, 101.7, 57.6; MS (ESI pos ion) *m/z*: calcd for C₁₉H₁₅ClF₂N₄O₄S, 468.0; found, 469.0 (M+H).

6.9.4. 2-(4-((2-Amino-3-chloropyridin-4-yl)oxy)-3-fluorophenyl)-N-phenyl-1,2-thiazinane-6-carboxamide 1,1-dioxide (**11d**)

From **10d**; white solid; yield 66%; ¹H NMR (600 MHz, CDCl₃) δ 8.79 (s, 1H, NH), 7.75 (d, 1H, *J* = 6.0 Hz, ArH), 7.55–7.47 (m, 2H, ArH), 7.31 (t, 2H, *J* = 7.9 Hz, ArH), 7.25 (dd, 1H, *J* = 11.6 Hz, ArH), 7.20–7.10 (m, 3H, ArH), 6.01 (d, 1H, *J* = 5.9 Hz, ArH), 5.47 (s, 2H, NH₂), 4.15 (dd, 1H, *J* = 9.2, 4.0 Hz, CH), 3.97–3.86 (m, 1H, CHH), 3.85–3.71 (m, 1H, CHH), 2.76–2.67 (m, 1H, CHH), 2.67–2.58 (m, 1H, CHH), 2.23–2.13 (m, 1H, CHH), 2.02–1.91 (m, 1H, CHH); ¹³C NMR (150 MHz, CDCl₃) δ 160.7, 160.5, 156.3, 154.5, 152.9, 145.1, 140.5, 138.4, 138.3, 137.2, 129.1, 125.2, 123.6, 123.5, 123.1, 120.3, 116.6, 116.5, 103.3, 102.3, 100.0, 64.7, 53.7, 29.8, 27.4, 23.2, 22.8; MS (ESI pos ion) *m/z*: calcd for C₂₂H₂₀CIFN₄O₄S, 490.1; found, 490.9 (M+H).

6.9.5. 2-(4-((2-Amino-3-chloropyridin-4-yl)oxy)-3-fluorophenyl)-N-(p-tolyl)-1,2-thiazinane-6-carboxamide 1,1-dioxide (**11e**)

From **10e**; white solid; yield 58%; ¹H NMR (600 MHz, Acetoned₆) δ 9.28 (s, 1H, NH), 7.82 (d, 1H, J = 5.6 Hz, ArH), 7.58–7.48 (m, 2H, ArH), 7.45–7.37 (m, 1H, ArH), 7.32 (d, 2H, J = 5.6 Hz, ArH), 7.13 (d, 2H, J = 8.2 Hz, ArH), 6.04 (d, 2H, J = 5.8 Hz, ArH), 5.91 (d, 1H, J = 8.7 Hz, ArH), 4.34 (dd, 1H, J = 10.3, 3.8 Hz, CH), 4.10–3.97 (m, 1H, CHH), 3.92–3.80 (m, 1H, CHH), 2.72–2.60 (m, 1H, CHH), 2.53–2.43 (m, 1H, CHH), 2.28 (s, 3H, CH₃), 2.18–2.12 (m, 1H, CHH), 2.12–2.06 (m, 1H, CHH); MS (ESI pos ion) m/z: calcd for C₂₃H₂₂ClFN₄O₄S, 504.1; found, 504.9 (M+H).

6.9.6. 2-(4-((2-Amino-3-chloropyridin-4-yl)oxy)-3-fluorophenyl)-N-(4-fluorophenyl)-1,2-thiazinane-6-carboxamide 1,1-dioxide (**11f**)

From **10f**; white solid; yield 68%; ¹H NMR (600 MHz, Acetone*d*₆) δ 9.46 (s, 1H, NH), 7.82 (d, 1H, *J* = 5.7 Hz, ArH), 7.71–7.66 (m, 2H, ArH), 7.45–7.40 (m, 1H, ArH), 7.34–7.29 (m, 2H, ArH), 7.13–7.08 (m, 2H, ArH), 6.04 (dd, 1H, *J* = 5.7, 0.9 Hz, ArH), 5.90 (s, 2H, NH₂), 4.35 (dd, 1H, *J* = 10.4, 3.8 Hz, CH), 4.07–4.00 (m, 1H, CHH), 3.88–3.83 (m, 1H, CHH), 2.71–2.63 (m, 1H, CHH), 2.53–2.45 (m, 1H, CHH), 2.19–2.12 (m, 1H, CHH), 2.08–2.06 (m, 1H, CHH); ¹³C NMR (150 MHz, Acetone-*d*₆) δ 162.2, 160.8, 158.5, 155.1, 148.2, 140.9, 140.8, 140.3, 140.2, 135.7, 124.4, 123.5, 122.4, 122.3, 122.2, 116.7, 116.5, 116.2, 116.1, 102.3, 100.8, 66.0, 54.2, 28.3, 23.5; MS (ESI pos ion) *m/z*: calcd for C₂₃H₂₂ClFN₄O₄S, 504.1; found, 504.9 (M+H).

6.10. Preparation of 1-(4-fluorophenyl)piperidin-2-one (14)

1-Fluoro-4-iodobenzene (2.22 g, 10 mmol) and piperidin-2-one (1.2 g, 12 mmol) were added to 30 mL of dry DMF, followed by the

addition of K₃PO₄ (6.36 g, 30 mmol) and CuI (190 mg, 0.1 mmol). The mixture solution was heated to 100 °C for 12 h, before filtered through celite. After washed with ethyl acetate (3 × 10 mL), the combined organic phase was concentrated and the residue was purified by column go give 1.73 g (90%) of compound **14** as yellow solid. ¹H NMR (600 MHz, CDCl₃) δ 7.20 (dd, 2H, *J* = 8.5, 5.0 Hz, ArH), 7.06 (t, 2H, *J* = 8.4 Hz, ArH), 3.60 (t, 2H, *J* = 5.4 Hz, NCH₂), 2.54 (t, 2H, *J* = 6.2 Hz, COCH₂), 2.01–1.84 (m, 4H, CH₂CH₂); ¹³C NMR (150 MHz, CDCl₃) δ 170.2, 161.9, 160.2, 139.3, 128.0, 127.9, 115.9, 51.9, 32.8, 23.5, 21.5.

6.11. Preparation of isobutyl 1-(4-fluorophenyl)-2-oxopiperidine-3-carboxylate (**15**)

Piperidone (14, 386 mg, 2 mmol) was dissolved in 20 mL of dry THF and cooled to -78 °C. After the addition of ^tBuLi (1.4 mL, 1.6 M in THF, 2.2 mmol) and stirred at this temperature for 4 h, 400 μ L (2 mmol) of isobutyl chlorofomate was added. 10 min later, the reaction was quenched by 2 mL of saturated NH₄Cl. The mixture was diluted with water (20 mL) and extracted with EtOAc $(3 \times 20 \text{ mL})$. The combined organic layer was washed with brine, dried over Na₂SO₄, and concentrated in vacuo. The residue was purified by column to give compound 15 (480 mg, 82%) as yellow wax. ¹H NMR (600 MHz, CDCl₃) δ 7.25–7.20 (m, 2H, ArH), 7.09–7.03 (m, 2H, ArH), 3.99 (dd, 1H, J = 10.6, 6.7 Hz, CH), 3.83 (d, 1H, *J* = 6.7 Hz, CH*H*), 3.70–3.61 (m, 1H, CH*H*), 3.58 (t, 1H, *J* = 6.9 Hz, CH), 2.32-2.24 (m, 1H, CHH), 2.23-2.16 (m, 1H, CHH), 2.12-2.04 (m, 1H, CHH), 2.02–1.87 (m, 2H, CH, CHH), 0.94 (d, 6H, J = 6.6 Hz, CH₃ \times 2); ¹³C NMR (150 MHz, CDCl₃) δ 171.0, 166.3, 162.1, 160.4, 138.8, 127.9, 127.9, 116.2, 116.0, 100.0, 71.5, 51.6, 49.6, 27.8, 25.3, 21.4, 19.1.

6.12. Preparation of 1-(4-fluorophenyl)-2-oxopiperidine-3-carboxylic acid (**16a**)

To a solution of **15** (217 mg, 0.74 mmol) in THF/MeOH/H₂O (1/1/ 1, 3 mL in total) at 0 °C was added LiOH monohydrate (94 mg, 2.2 mmol). The reaction mixture was warmed to room temperature and stirred for 5 h. The solution was acidified to pH 1 with 1 M HCl and extracted with EtOAc (3 × 20 mL). The organic extracts were combined and washed with brine (2 × 5 mL). Evaporation of the solvent gave the corresponding acid **16a** (152 mg, 87%) as white solid. ¹H NMR (600 MHz, DMSO-*d*₆) δ 12.61 (s, 1H, OH), 7.33–7.28 (m, 1H, ArH), 7.25–7.19 (m, 1H, ArH), 3.69–3.55 (m, 2H, NCH₂), 3.43 (dd, 1H, *J* = 8.2, 6.5 Hz, CH), 2.16–2.10 (m, 1H, CHH), 2.08–2.02 (m, 1H, CHH), 1.98–1.91 (m, 1H, CHH), 1.91–1.83 (m, 1H, CHH).

6.13. Preparation of 3-bromo-1-(4-fluorophenyl)-2-oxopiperidine-3-carboxylic acid (**16b**)

To a solution of acid **16a** (220 mg, 0.93 mmol) in Et₂O (5 mL) was added liquid Br₂ (48 μ L, 0.93 mmol) at 0 °C. The reaction mixture was stirred for 2 h, before concentrated in vacuo. The residue was purified by column, giving compound **16b** (265 mg, 91%) as white solid. ¹H NMR (600 MHz, Acetone-*d*₆) δ 13.03 (s, 1H, OH), 7.44–7.40 (m, 2H, ArH), 7.25–7.20 (m, 2H, ArH), 4.04 (td, 1H, *J* = 12.1, 4.6 Hz, NCHH), 3.82 (ddt, 1H, *J* = 13.0, 6.3, 2.4 Hz, NCHH), 2.77–2.69 (m, 1H, CHH), 2.62–2.56 (m, 1H, CHH), 2.53–2.43 (m, 1H, CHH), 2.19–2.12 (m, 1H, CHH); ¹³C NMR (150 MHz, Acetone-*d*₆) δ 166.4, 162.5, 160.9, 140.6, 140.4, 129.0, 128.9, 116.2, 52.1, 32.3, 20.4.

6.14. Preparation of 17a and 17b

The procedure is very similar to the preparation of **6**.

6.14.1. 3-Chloro-4-(2-fluoro-4-(1-(4-fluorophenyl)-2-

oxopiperidine-3-carboxamido)phenoxy)picolinamide (**17a**) From **16a**; white solid; yield 76%; ¹H NMR (600 MHz, DMSO-*d*₆) δ 10.56 (s, 1H, NH), 8.33 (d, 1H, *J* = 5.5 Hz, ArH), 8.07 (s, 1H, NH), 7.91 (dd, 1H, *J* = 12.9, 2.4 Hz, ArH), 7.77 (s, 1H, NH), 7.44 (dd, 1H, *J* = 8.9, 2.4 Hz, ArH), 7.41 (t, 1H, *J* = 8.8 Hz, ArH), 7.36–7.32 (m, 2H, ArH), 7.26–7.20 (m, 2H, ArH), 6.84 (dd, 1H, *J* = 5.5, 1.1 Hz, ArH), 3.76–3.68 (m, 1H, CH), 3.65–3.57 (m, 2H, CH₂), 2.21–2.13 (m, 2H, CH₂), 2.12– 2.04 (m, 1H, CH*H*), 1.96–1.87 (m, 1H, CH*H*); ¹³C NMR (150 MHz, DMSO-*d*₆) δ 169.3, 166.5, 160.9, 160.0, 159.3, 154.2, 153.8, 152.2, 148.7, 139.4, 138.5, 138.4, 134.8, 134.7, 128.3, 128.2, 123.7, 116.3, 115.9, 115.6, 115.4, 110.6, 107.9, 107.8, 54.9, 51.2, 50.5, 48.6, 24.8, 21.2; MS (ESI pos ion) *m*/*z*: calcd for C₂₄H₁₉ClF₂N₄O₄, 500.1; found, 501.1 (M+H).

6.14.2. 3-Chloro-4-(4-(3-chloro-1-(4-fluorophenyl)-2oxopiperidine-3-carboxamido)-2-fluorophenoxy)picolinamide (**17b**)

From **16b**; white solid; yield 68%; ¹H NMR (600 MHz, CDCl₃) δ 10.11 (s, 1H, NH), 8.24 (d, 1H, *J* = 5.5 Hz, ArH), 7.80 (dd, 1H, *J* = 11.9, 2.5 Hz, ArH), 7.54 (d, 1H, *J* = 3.9 Hz, ArH), 7.33–7.22 (m, 3H, ArH), 7.18–7.11 (m, 3H, NH), 6.68 (dd, 1H, *J* = 5.5, 1.1 Hz, ArH), 6.15 (d, 1H, *J* = 3.4 Hz, ArH), 3.85–3.78 (m, 1H, CHH), 3.73–3.68 (m, 1H, CHH), 2.96–2.87 (m, 1H, CHH), 2.65–2.56 (m, 1H, CHH), 2.45–2.34 (m, 1H, CHH), 2.17–2.07 (m, 1H, CHH); ¹³C NMR (150 MHz, CDCl₃) δ 166.8, 166.0, 164.9, 162.6, 161.8, 160.9, 154.7, 153.0, 148.3, 147.0, 137.9, 136.9, 136.8, 136.7, 127.9, 127.8, 123.4, 121.2, 116.7, 116.6, 116.5, 111.7, 109.8, 109.6, 64.4, 52.6, 33.8, 19.4; MS (ESI pos ion) *m/z*: calcd for C₂₄H₁₈Cl₂F₂N₄O₄, 534.1; found, 535.1 (M+H), 537.1 (M+H+2).

6.15. Preparation of 18a and 18b

The procedure is very similar to the preparation of **11**.

6.15.1. N-(4-((2-amino-3-chloropyridin-4-yl)oxy)-3-fluorophenyl)-1-(4-fluorophenyl)-2-oxopiperidine-3-carboxamide (**18a**)

From **17a**; white solid; yield 72%; ¹H NMR (600 MHz, CDCl₃) δ 10.00 (s, 1H, NH), 7.71 (dd, 1H, *J* = 12.6, 2.5 Hz, ArH), 7.24–7.18 (m, 3H, ArH), 7.17–7.10 (m, 3H, ArH), 7.02 (dt, 1H, *J* = 9.0, 1.8 Hz, ArH), 6.62 (t, 1H, *J* = 8.9 Hz, ArH), 5.01 (s, 2H, NH₂), 3.65 (dq, 2H, *J* = 7.2, 4.3, 3.4 Hz, NCH₂), 3.54 (t, *J* = 6.3 Hz, 1H, CH), 2.59–2.49 (m, 1H, CHH), 2.21–2.15 (m, 1H, CHH), 2.10–2.05 (m, 1H, CHH), 2.04–1.98 (m, 1H, CHH); ¹³C NMR (150 MHz, CDCl₃) 169.4, 165.7, 162.5, 160.9, 155.6, 154.7, 151.1, 146.5, 139.9, 138.5, 134.5, 134.4, 128.3, 128.2, 123.3, 116.9, 116.8, 116.7, 116.6, 116.5, 115.3, 115.2, 109.4, 109.2, 108.9, 52.8, 47.6, 47.5, 29.8, 22.9, 21.8; MS (ESI pos ion) *m/z*: calcd for C₂₃H₁₉ClF₂N₄O₃, 472.1; found, 473.1 (M+H).

6.15.2. N-(4-((2-amino-3-chloropyridin-4-yl)oxy)-3-fluorophenyl)-3-chloro-1-(4-fluorophenyl)-2-oxopiperidine-3-carboxamide (**18b**)

From **17b**; white solid; yield 76%; ¹H NMR (600 MHz, CDCl₃) δ 10.02 (s, 1H, NH), 7.77 (d, 1H, *J* = 5.8 Hz, ArH), 7.74 (dd, 1H, *J* = 12.0, 2.5 Hz, ArH), 7.26–7.23 (m, 2H, ArH), 7.23–7.20 (m, 1H, ArH), 7.16–7.11 (m, 3H, ArH), 5.99 (dd, 1H, *J* = 5.8, 1.0 Hz, ArH), 5.04 (s, 2H, NH₂), 3.84–3.74 (m, 1H, CH*H*), 3.74–3.66 (m, 1H, CH*H*), 2.94–2.82 (m, 1H, CH*H*), 2.63–2.57 (m, 1H, CH*H*), 2.43–2.34 (m, 1H, CH*H*), 2.14–2.09 (m, 1H, CH*H*); ¹³C NMR (150 MHz, CDCl₃) δ 166.9, 164.8, 162.6, 161.0, 160.5, 156.6, 155.0, 153.3, 148.6, 147.8, 146.7, 137.9, 136.2, 127.8, 123.4, 116.7, 116.6, 116.2, 109.6, 109.4, 102.6, 102.1, 91.8, 64.2, 52.7, 33.8, 19.5; MS (ESI pos ion) *m/z*: calcd for C₂₃H₁₈Cl₂F₂N₄O₃, 506.1; found, 507.1 (M+H), 509.1 (M+H+2).

6.16. Preparation of fragments 19a-b and 20a-c

Following the known procedure [15,18,19].

6.17. Preparation of N-(4-((2-aminopyrimidin-4-yl)oxy)-3fluorophenyl)-1-(4-fluorophenyl)-2-oxo-1,2-dihydropyridine-3carboxamide (**21a**)

To the suspension of carboxylic acid 19a (27 mg, 0.11 mmol) in drv toluene (5 mL) was added sulfurvl dichloride (0.5 mL). The mixture was stirred for 3 h before concentrated in vacuo. The residue was redissolved in 5 mL of dry toluene and concentrated again. The resulting yellow solid was then dissolved in 5 mL of dry THF and the solution was cooled to 0 °C. Compound 20a (30 mg, 0.1 mmol) and DIPEA (50.6 µL, 0.29 mmol) was added to the mixture, respectively. After stirred at 0 °C for 15 min and at room temperature for 1 h, the reaction mixture was diluted with water (10 mL) and then extracted with EtOAc (3 \times 20 mL). The combined organic layer was dried over Na₂SO₄ and concentrated in vacuo, giving a yellow solid (38 mg, 67%), which was directly dissolved in TFA (1 mL). The reaction mixture was heated to reflux for 6 h before concentrated in vacuo. The residue was purified by column, giving compound 21a (24 mg, 77%) as pale yellow solid. ¹H NMR (600 MHz, Acetone- d_6) δ 8.70–8.60 (m, 1H, ArH), 8.35 (d, 1H, J = 7.0 Hz, ArH), 8.10 (dd, 1H, J = 12.7, 2.3 Hz, ArH), 8.06 (dd, 1H, *J* = 6.6, 2.2 Hz, ArH), 7.67–7.63 (m, 2H, ArH), 7.48-7.45 (m, 1H, ArH), 7.42-7.38 (m, 2H, ArH), 7.36 (t, 1H, J = 8.6 Hz, ArH); ¹³C NMR (150 MHz, Acetone- d_6) δ 172.2, 163.2, 162.6, 159.3, 155.4, 153.8, 150.2, 150.1, 146.0, 144.6, 139.1, 135.1, 130.2, 130.0, 121.8, 117.1, 116.9, 108.9, 107.8, 107.7, 99.2, 99.1; MS (ESI pos ion) *m*/*z*: calcd for C₂₂H₁₅F₂N₅O₃, 435.1; found, 436.0 (M+H).

6.18. General procedure for the preparation of **21b**, **21c**, **23**, **25** and **22c**

Similar to the preparation of **21a**, but omitting the second step (treatment with TFA).

6.18.1. N-(4-((7H-pyrrolo[2,3-d]pyrimidin-4-yl)oxy)-3fluorophenyl)-1-(4-fluorophenyl)-2-oxo-1,2-dihydropyridine-3carboxamide (**21b**)

From **19a** and **20b**; white solid; yield 34%; ¹H NMR (600 MHz, DMSO- d_6) δ 11.83 (s, 1H, NH), 11.62 (s, 1H, NH), 8.14 (dd, 1H, J = 7.2, 2.3 Hz, ArH), 7.84 (s, 1H, ArH), 7.66 (dd, 1H, J = 6.5, 2.3 Hz, ArH), 7.51 (dd, 1H, J = 12.6, 2.5 Hz, ArH), 7.18–7.13 (m, 2H, ArH), 7.05 (t, 1H, J = 3.0 Hz, ArH), 7.01–6.92 (m, 4H, ArH), 6.27 (t, 1H, J = 6.9 Hz, ArH), 6.14 (dd, 1H, J = 3.6, 1.7 Hz, ArH); ¹³C NMR (150 MHz, DMSO- d_6) δ 163.2, 162.1, 161.6, 161.4, 155.1, 154.1, 153.5, 150.5, 145.5, 144.6, 137.4, 137.3, 136.8, 135.8, 135.7, 129.9, 129.8, 126.1, 125.2, 120.7, 116.7, 116.6, 116.5, 107.5, 104.6, 100.0, 98.4; MS (ESI pos ion) *m*/*z*: calcd for C₂₄H₁₅F₂N₅O₃, 459.1; found, 460.0 (M+H), 482.1 (M+Na).

6.18.2. N-(4-((6,7-dimethoxyquinolin-4-yl)oxy)-3-fluorophenyl)-1-(4-fluorophenyl)-2-oxo-1,2-dihydropyridine-3-carboxamide (**21c**)

From **19a** and **20c**; white solid; yield 78%; ¹H NMR (600 MHz, DMSO- d_6) δ 11.98 (s, 1H, NH), 8.52 (d, 1H, J = 5.4 Hz, ArH), 8.50 (d, 1H, J = 3.0 Hz, ArH), 8.47 (d, 1H, J = 3.0 Hz, ArH), 8.05 (dd, 1H, J = 12.7, 2.5 Hz, ArH), 7.66–7.61 (m, 2H, ArH), 7.58–7.54 (m, 2H, ArH), 7.50–7.41 (m, 4H, ArH), 3.96 (s, 3H, OCH₃), 3.95 (s, 3H, OCH₃); ¹³C NMR (150 MHz, DMSO- d_6) δ 162.9, 160.5, 159.8, 154.3, 153.0, 149.7, 148.2, 144.2, 142.0, 136.2, 135.6, 129.6, 129.4, 128.2, 128.0, 125.5, 124.3, 121.0, 116.2, 116.0, 114.6, 112.2, 108.9, 108.7, 106.9, 102.3, 99.0, 55.8; MS (ESI pos ion) m/z: calcd for C₂₉H₂₁F₂N₃O₅, 529.1; found, 530.0 (M+H).

6.18.3. N-(3-fluoro-4-((2-((4-methoxybenzyl)amino)pyrimidin-4yl)oxy)phenyl)-1-(4-fluorophenyl)-4-iodo-2-oxo-1,2dihydropyridine-3-carboxamide (**23**)

From **19b** and **20a**; yellow solid; yield 56%. Used directly for the next step without further structure characterization.

6.18.4. N-(4-((7H-pyrrolo[2,3-d]pyrimidin-4-yl)oxy)-3-fluorophenyl)-1-(4-fluorophenyl)-4-iodo-2-oxo-1,2-dihydropyridine-3-carboxamide (**25**)

From **19b** and **29b**; white solid; yield 46%; Used directly for the next step without further structure characterization.

6.18.5. N-(4-((6,7-dimethoxyquinolin-4-yl)oxy)-3-fluorophenyl)-1-(4-fluorophenyl)-4-iodo-2-oxo-1,2-dihydropyridine-3-carboxamide (**22c**)

From **19b** and **20c**; white solid; yield 76%; ¹H NMR (600 MHz, CDCl₃) δ 11.61 (s, 1H, NH), 8.43 (d, 1H, *J* = 5.5 Hz, ArH), 7.89 (dd, 1H, *J* = 12.1, 2.6 Hz, ArH), 7.57–7.44 (m, 2H, ArH), 7.39–6.97 (m, 8H, ArH), 6.40 (d, 1H, *J* = 5.5 Hz, ArH), 3.99 (s, 3H, OCH₃), 3.98 (s, 3H, OCH₃); ¹³C NMR (150 MHz, CDCl₃) δ 162.1, 161.3, 160.5, 157.3, 156.3, 155.1, 154.0, 153.4, 150.3, 137.8, 135.4, 134.9, 133.0, 128.4, 128.3, 128.2, 125.3, 123.6, 121.9, 120.4, 120.0, 117.1, 117.0, 116.8, 116.5, 115.7, 100.0, 99.7, 56.6, 56.4; MS (ESI pos ion) *m*/*z*: calcd for C₂₉H₂₀F₂IN₃O₅, 655.0; found, 655.8 (M+H).

6.19. 4-Ethoxy-N-(3-fluoro-4-((2-((4-methoxybenzyl)amino) pyrimidin-4-yl)oxy)phenyl)-1-(4-fluorophenyl)-2-oxo-1,2-dihydropyridine-3-carboxamide (**24**)

Sodium hydride (7 mg, 0.16 mmol, 60% dispersion in mineral oil) was added slowly to a solution of ethanol (1 mL) and THF (1 mL) under argon and the resulting mixture was stirred at rt for 5 min. After re-cooled to 0 °C, compound 23 (76 mg, 0.11 mmol) was added to the mixture. The resulting solution was kept stirring at 0 °C for 10 min, and then warmed to rt and stirred for 1 h. The reaction mixture was concentrated in vacuo. The resulting crude solid was suspended in ethyl acetate, washed with saturated aqueous sodium bicarbonate solution, and water. The organic layer was dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by column to give the compound 24 (48 mg, 72%) as white solid. ¹H NMR (600 MHz, CDCl₃) δ 8.09 (s, 1H, NH), 7.86 (d, 1H, *J* = 12.3 Hz, ArH), 7.48 (dd, 1H, *J* = 7.9, 2.0 Hz, ArH), 7.33 (ddt, 2H, J = 6.5, 4.4, 2.0 Hz, ArH), 7.28–7.18 (m, 4H, ArH), 7.16–6.98 (m, 2H, ArH), 6.79 (d, 2H, J = 7.9 Hz, ArH), 6.33 (dd, 1H, J = 7.9, 2.0 Hz, ArH), 6.13 (dd, 1H, J = 5.7, 1.9 Hz, ArH), 5.52 (dd, 1H, J = 4.5, 2.2 Hz, ArH), 4.32 (q, 2H, J = 7.1 Hz, OCH₂), 4.28 (s, 1H, NH), 4.03–4.01 (m, 1H, NCHH), 3.87-3.77 (m, 1H, NCHH), 3.76 (s, 3H, OCH₃), 1.55 (t, 3H, J = 7.0 Hz, CH₃); ¹³C NMR (150 MHz, CDCl₃) δ 170.9, 163.7, 163.5, 162.2, 161.9, 161.7, 158.8, 153.4, 140.5, 128.9, 128.7, 123.7, 116.9, 116.7, 113.9, 109.1, 106.6, 103.4, 100.0, 98.5, 97.4, 66.4, 55.4, 33.3, 14.8; MS (ESI pos ion) m/z: calcd for C₃₂H₂₇F₂N₅O₅, 599.2; found, 600.1 (M+H).

6.20. Preparation of N-(4-((7H-pyrrolo[2,3-d]pyrimidin-4-yl)oxy)-3-fluorophenyl)-4-ethoxy-1-(4-fluorophenyl)-2-oxo-1,2dihydropyridine-3-carboxamide (**22b**)

Similar to the preparation of **24** but using **25** as starting material. White solid; yield 46%; ¹H NMR (600 MHz, DMSO-*d*₆) δ 12.31 (s, 1H, NH), 10.56 (s, 1H, NH), 8.30 (s, 1H, ArH), 7.97–7.79 (m, 2H, ArH), 7.63–7.27 (m, 7H, ArH), 6.60 (d, 1H, *J* = 3.5 Hz, ArH), 6.53 (d, 1H, *J* = 7.9 Hz, ArH), 4.26 (q, 2H, *J* = 6.9 Hz, OCH₂CH₃), 1.31 (t, 3H, *J* = 7.0 Hz, OCH₂CH₃); ¹³C NMR (150 MHz, DMSO-*d*₆) δ 165.0, 162.9, 161.5, 161.1, 160.5, 153.9, 150.5, 141.3, 136.9, 129.7, 129.6, 125.0, 116.5, 116.3, 111.9, 104.5, 98.3, 96.5, 65.6, 31.2, 15.2; MS (ESI pos ion) *m*/*z*: calcd for C₂₆H₁₉F₂N₅O₄, 503.1; found, 503.9 (M+H), 526.1 (M+Na).

6.21. Preparation of N-(4-((6,7-dimethoxyquinolin-4-yl)oxy)-3-fluorophenyl)-4-ethoxy-1-(4-fluorophenyl)-2-oxo-1,2-dihydropyridine-3-carboxamide (**22d**)

Similar to the preparation of **24** but using **22c** as starting material. White solid; yield 71%; ¹H NMR (400 MHz, CDCl₃) δ 11.66 (s, 1H, NH), 8.48 (d, 1H, J = 5.3 Hz, ArH), 7.97 (dd, 1H, J = 12.6, 2.4 Hz, ArH), 7.59 (s, 1H, ArH), 7.53 (d, 1H, J = 7.8 Hz, ArH), 7.42 (s, 1H, ArH), 7.39–7.33 (m, 3H, ArH), 7.26–7.22 (m, 2H, ArH), 7.16 (t, 1H, J = 8.7 Hz, ArH), 6.42 (dd, 1H, J = 5.2, 0.8 Hz, ArH), 6.38 (d, 1H, J = 7.9 Hz, ArH), 4.37 (q, 2H, J = 7.0 Hz, OCH₂CH₃), 4.06 (s, 3H, OCH₃), 4.05 (s, 3H, OCH₃), 1.59 (t, 3H, J = 7.0 Hz, OCH₂CH₃); ¹³C NMR (100 MHz, CDCl₃) δ 171.0, 164.0, 163.8, 162.1, 160.4, 155.7, 152.9, 149.6, 149.0, 146.9, 140.6, 137.8, 137.7, 136.0, 128.8, 128.7, 123.5, 117.0, 116.8, 116.6, 115.7, 107.9, 106.5, 102.4, 99.7, 97.5, 66.5, 56.3, 14.8; MS (ESI pos ion) m/z: calcd for C₃₁H₂₅F₂N₃O₆, 573.2; found, 574.1 (M+H).

6.22. Preparation of N-(4-((6,7-dimethoxyquinolin-4-yl)oxy)-3-fluorophenyl)-1-(4-fluorophenyl)-4-methoxy-2-oxo-1,2-dihydropyridine-3-carboxamide (**22e**)

Similar to the preparation of **24** but using **22c** as starting material; ethanol was replaced with methanol. White solid; yield 71%; ¹H NMR (400 MHz, CDCl₃) δ 11.75 (s, 1H, NH), 8.49 (d, 1H, *J* = 5.3 Hz, ArH), 8.40 (dd, 1H, *J* = 12.6, 2.4 Hz, ArH), 7.59 (s, 1H, ArH), 7.57 (d, 1H, *J* = 7.8 Hz, ArH), 7.42 (s, 1H, ArH), 7.39–7.33 (m, 3H, ArH), 7.29–7.27 (m, 1H, ArH), 7.26–7.23 (m, 1H, ArH), 7.16 (t, 1H, *J* = 8.7 Hz, ArH), 6.44–6.41 (m, 2H, ArH), 4.13 (s, 3H, OCH₃), 4.06 (s, 3H, OCH₃); ¹³C NMR (100 MHz, CDCl₃) δ 171.8, 163.7, 162.1, 160.4, 155.7, 153.2, 152.9, 149.6, 149.1, 146.9, 140.9, 137.8, 137.7, 136.5, 135.9, 128.8, 128.7, 123.5, 117.1, 116.8, 116.5, 115.7, 107.9, 106.3, 102.4, 99.6, 96.8, 57.5, 56.31, 56.26; MS (ESI pos ion) *m/z*: calcd for C₃₀H₂₃F₂N₃O₆, 559.2; found, 559.8 (M+H).

6.23. Preparation of N-(4-((2-aminopyrimidin-4-yl)oxy)-3fluorophenyl)-4-ethoxy-1-(4-fluorophenyl)-2-oxo-1,2dihydropyridine-3-carboxamide (**22a**)

Compound **24** (40 mg, 0.07 mmol) was dissolved in TFA and heated to reflux for 6 h. The solvent was removed before the residue was purified by column, giving title compound (23 mg, 77%) as pale yellow solid. ¹H NMR (600 MHz, DMSO- d_6) δ 10.57 (s, 1H, NH), 8.26 (d, 1H, *J* = 6.3 Hz, ArH), 7.87–7.80 (m, 4H, ArH + NH₂), 7.47–7.43 (m, 2H, ArH), 7.40 (dd, 1H, *J* = 8.9, 2.4 Hz, ArH), 7.39–7.31 (m, 3H, ArH), 4.25 (q, 2H, *J* = 7.0 Hz, OCH₂), 1.29 (t, 3H, *J* = 7.0 Hz, CH₃); ¹³C NMR (150 MHz, DMSO- d_6) δ 170.2, 164.5, 162.7, 160.7, 160.0, 154.1, 152.5, 140.9, 138.7, 138.6, 136.4, 129.2, 129.1, 124.2, 116.0, 115.9, 115.4, 111.5, 97.1, 96.1, 65.1, 14.7; MS (ESI pos ion) *m*/*z*: calcd for C₂₄H₁₉F₂N₅O₄, 479.1; found, 480.1 (M+H).

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Appendix A. Supplementary data

Supplementary data related to this article can be found at http://dx.doi.org/10.1016/j.ejmech.2014.04.056.

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