

## Pulse radiolysis studies of intermolecular charge transfers involving tryptophan and three-electron-bonded intermediates derived from methionine

Haiying Fu · Mingzhang Lin · Yusa Muroya ·  
Yosuke Katsumura

Received: 26 February 2011 / Accepted: 23 May 2011 / Published online: 14 June 2011  
© Springer Science+Business Media B.V. 2011

**Abstract** The oxidation processes of the radiation-generated, three-electron-bonded intermediates  $\text{AcMet}_2 [\text{S}\cdot\cdot\text{S}]^+$  and  $\text{AcMet} [\text{S}\cdot\cdot\text{Br}]$  were investigated by pulse radiolysis via their reactions with tryptophan (TrpH). These intermediates were derived from *N*-acetyl-methionine amide (*N*-AcMetNH<sub>2</sub>) and *N*-acetyl-methionine methyl ester (*N*-AcMetOMe). The bimolecular rate constant  $k$  of the reaction between each intermediate and L-tryptophan (TrpH) was measured. For *N*-AcMetNH<sub>2</sub>,  $k$  for the reaction of  $\text{AcMet}_2 [\text{S}\cdot\cdot\text{S}]^+$  with TrpH were  $3.4 \times 10^8$  and  $2.2 \times 10^8 \text{ dm}^3 \text{ mol}^{-1} \text{ s}^{-1}$  at pH = 1 and 4.5, respectively. For *N*-AcMetOMe,  $k$  for the reaction of  $\text{AcMet}_2 [\text{S}\cdot\cdot\text{S}]^+$  with TrpH were  $4.0 \times 10^8$  and  $2.8 \times 10^8 \text{ dm}^3 \text{ mol}^{-1} \text{ s}^{-1}$  at pH 1 and 4.5, respectively. The rate constants for the intermolecular transformation of  $\text{Met} [\text{S}\cdot\cdot\text{Br}]$  into  $\text{TrpH}^+$  or Trp were also estimated. For *N*-AcMetNH<sub>2</sub>,  $k$  for the reaction of  $\text{AcMet}_2 [\text{S}\cdot\cdot\text{Br}]$  with TrpH were  $2.6 \times 10^8$  and  $3.3 \times 10^8 \text{ dm}^3 \text{ mol}^{-1} \text{ s}^{-1}$  at pH 1 and 4.5, respectively. Related mechanisms were discussed.

**Keywords** Three-electron-bonded radicals · Methionine derivatives · Tryptophan · Intermolecular electron transfer · Pulse radiolysis

---

H. Fu (✉)  
Laboratory of Radiation Chemistry, Shanghai Institute of Applied Physics,  
Chinese Academy of Sciences, Shanghai 201800, China  
e-mail: fuhaiying@sinap.ac.cn

M. Lin  
Japan Atomic Energy Agency, 2-4 Shirakata Shirane, Tokaimura, Nakagun, Ibaraki 319-1195, Japan

Y. Muroya · Y. Katsumura (✉)  
Nuclear Professional School, School of Engineering, The University of Tokyo,  
2-22 Shirakata Shirane, Tokaimura, Nakagun, Ibaraki 319-1188, Japan  
e-mail: katsu@n.t.u-tokyo.ac.jp



three-electron-bonded species of the types  $\text{Met}_2 [\text{S} \cdot \cdot \text{S}]^+$  or  $\text{Met} [\text{S} \cdot \cdot \text{X}]$  ( $\text{X} = \text{Cl}, \text{Br}, \text{etc.}$ ). Bobrowski and Lubis [13] as well as Prutz et al. [14] investigated the intermolecular electron transfers involved in three-electron-bonded species derived from the oxidation of Met and dipeptides. The dipeptides included  $\text{Met}_2 [\text{S} \cdot \cdot \text{S}]^+$ ,  $\text{Met} [\text{S} \cdot \cdot \text{X}]\text{-Gly}$ , as well as tryptophan (Trp), tyrosine (Tyr), and other amino acids. Other studies demonstrated that intramolecular electron transfers can occur within a (phosphoryl) dipeptide, or within polypeptides such as  $\text{Met-Trp}$  and  $\text{Met-(X)}_n\text{-Tyr}$  ( $\text{X} = \text{Pro}, \text{etc.}, n = 1-5$ ) [12, 15, 16]. The rate of transformation of methionyl into tryptophyl radicals were also shown to depend on peptide concentration. This dependence indicated the occurrence of both inter- and intramolecular processes, which is in contrast with tryptophyltyrosyl peptides. These findings may lead to observable intermolecular charge transfer reactions within the products formed by oxidative attacks on free Met with Trp and Tyr. In the present work, pulse radiolysis was used to demonstrate the oxidizing nature of various intermediates of  $N\text{-AcMetNH}_2$  and  $N\text{-AcMetOMe}$  via the oxidation of Trp in a free aqueous solution. The reaction was examined at different pH levels. Electron transfer processes involving Trp and the sulfur-containing three-electron-bonded intermediates were investigated.

## Experimental

### Materials

$N$ -acetyl- $L$ -methionine amide ( $N\text{-AcMetNH}_2$ ),  $N$ -acetyl- $L$ -methionine methyl ester ( $N\text{-AcMetOMe}$ ), and  $L$ -tryptophan (TrpH) were obtained from Sigma and were used without further purification. All other compounds, which were of the highest commercially available purity, were from Wako Co., Japan. The pH values of amino acid solutions were adjusted by adding appropriate amounts of  $\text{HClO}_4$ ,  $\text{NaOH}$ , or  $\text{NaH}_2\text{PO}_4$ .

### Apparatus

The pulse radiolysis experiments were performed at the University of Tokyo using an electron linear accelerator (energy = 35 MeV; duration = 20 ns) coupled with an absorption spectroscopic detection system. Absorbed doses were determined by a KSCN dosimeter, which involved taking  $G \times \varepsilon_{(\text{SCN})_2\bullet-} = 5.1 \times 10^4 \text{ m}^2/\text{J}$  [17] for  $\text{N}_2\text{O}$ -saturated  $1 \times 10^{-2} \text{ mol dm}^{-3}$  potassium thiocyanate solutions. The analyzing light came from a 150 W xenon lamp (Hungarian Tungfram), which was pulsed for 3 ms to increase the intensity. Optical absorption was determined with a Bausch-Lomb monochromator, a Hamamatsu R 928 photomultiplier, and a Tektronix 7834 oscilloscope. The oscilloscope monitored the light passing through the irradiation quartz cell with an optical path of 20 mm. The signals were digitized using an Iwatsu 8123 digital oscilloscope and were stored in a private microcomputer for further analysis. The microcomputer software converted the data into give optical densities as well as first and second order kinetic parameters. Further details on the

pulse radiolysis equipment are given elsewhere [18]. For measuring absorption spectra, a higher dose (ca. 35 Gy/pulse) was used. For determining rate constants, a lower dose (10–13 Gy/pulse) was applied.

## Methods

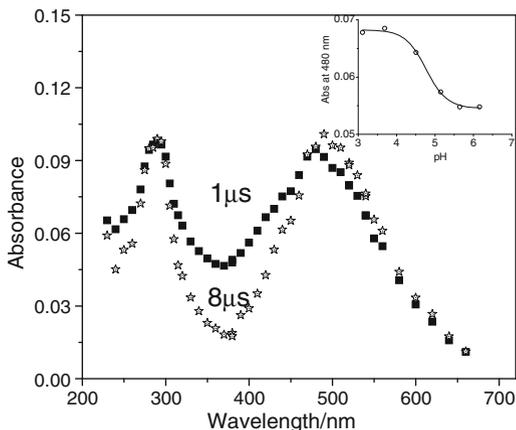
All solutions were prepared with triple-distilled water at room temperature. Before irradiation, the solutions (pH 1) were saturated in high purity Ar for about 30 min to remove oxygen, and in high purity  $N_2O$  (pH > 3) to convert  $e_{aq}^-$  into hydroxyl (OH) radicals. Hydrogen atoms, which cannot easily be converted into OH radicals at pH < 3 (even in the presence of  $N_2O$ ) reacted with Met at a rate constant  $k$  of  $\sim 10^9 \text{ dm}^3 \text{ mol}^{-1} \text{ s}^{-1}$  [19]. However, their reaction products did not interfere with the optical signals observable in our system because earlier optical measurements showed only weak absorption below 300 nm ( $\epsilon_{280} = 200 \text{ dm}^3 \text{ mol}^{-1} \text{ cm}^{-1}$  [20] in acidic Met solutions).

## Results and discussion

### Met<sub>2</sub> [S : S]<sup>+</sup> as an oxidant

The transient absorption spectra formed by an  $\bullet OH$ -induced oxidation in a model system containing  $2 \times 10^{-3} \text{ mol dm}^{-3}$  *N*-AcMetNH<sub>2</sub> at pH 1 are shown in Fig. 1. The UV band at  $\lambda$  ca. 290 nm is generally characteristic of a-thioalkyl radicals ( $\cdots\bullet CH-S\cdots$ ). This band resulted from the deprotonation of the  $R_2S^{\bullet+}$ -type radical cation or from the direct H-atom abstraction by  $\bullet OH$  radicals. The broad absorption spectrum with  $\lambda$  max = 480 nm is assigned to the three-electron-bonded dimer intermediate Met<sub>2</sub> [S : S]<sup>+</sup> [20, 21]. This band was formed via OH addition to the sulfur atom in the initial step (Reaction 1):

**Fig. 1** Transient absorption spectra of an aqueous solution containing  $2 \times 10^{-3} \text{ mol dm}^{-3}$  *N*-AcMetNH<sub>2</sub> saturated with  $N_2O$  at pH 1. Dose = 34 Gy/pulse. Inset The absorbance at 480 nm at different pH values



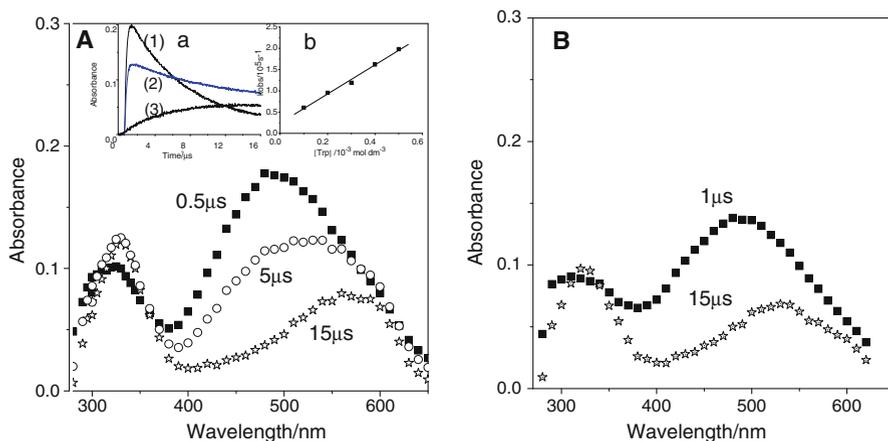


Under the experimental conditions, the OH adducts underwent the elimination of  $\bullet\text{OH}$  with the assistance of external protons (Reaction 2). A subsequent intermolecular equilibration occurred with a second unattacked Met molecule (Reaction 3). The transient absorption at 480 nm was varied by pH values, as shown in Fig. 1 (inset).  $\text{p}K_a$  was determined to be  $4.5 \pm 0.2$ .



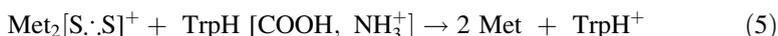
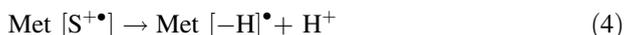
The transient absorption spectra formed by an OH-induced oxidation in a model system containing  $2 \times 10^{-3} \text{ mol dm}^{-3}$  *N*-AcMetNH<sub>2</sub> and  $2 \times 10^{-4} \text{ mol dm}^{-3}$  Trp at pH 1 was shown in Fig. 2A. The concentration of AcMet was much higher than that of TrpH, and the rate constant of Met was  $k(\text{OH} + \text{Met}) = 2.3 \times 10^{10} \text{ dm}^3 \text{ mol}^{-1} \text{ s}^{-1}$  [11] and  $k(\text{OH} + \text{TrpH}) = 1.25 \times 10^{10} \text{ dm}^3 \text{ mol}^{-1} \text{ s}^{-1}$  [22]. Hence, more than 90% of the OH radicals were expected to react with Met. In our experiments, the dimer radical cation formation was completed within 1  $\mu\text{s}$  pulse. About 15  $\mu\text{s}$  after the electron pulse, a new transient species with characteristic absorptions at 325 and 570 nm [see spectrum (O)] was fully developed. This species was the tryptophyl radical cation ( $\text{TrpH}^+$ ) [23], and its development was indicated by the identical growing and decaying processes observed at either 320 or 570 nm.

The characteristic absorption bands of the protonated tryptophyl radical cation at 325 and 570 nm were fully developed 15  $\mu\text{s}$  after the pulse, even though the 325 nm



**Fig. 2** Transient absorption spectra of  $2 \times 10^{-3} \text{ mol dm}^{-3}$  *N*-AcMetNH<sub>2</sub> and  $2 \times 10^{-4} \text{ mol dm}^{-3}$  Trp aqueous solutions saturated with Ar at **A** pH 1 and **B** pH 4.5. Dose = 34 Gy/pulse. *Inset a* The transient absorption curves observed at (1) 480, (2) 570 nm, and (3) obtained by subtracting (1) from (2) at 570 nm. *Inset b* The plot of observed  $k$  against TrpH concentration for the decay of the  $\text{Met}_2 [\text{S}:\text{S}]^+$  radical cation recorded at 480 nm

absorption was also discernible in the spectrum at 0.5  $\mu\text{s}$ . The formation of  $\text{TrpH}^{+\bullet}$  at this time was only seen below 400 nm, where it absorbed more strongly than  $\text{Met}_2 [\text{S} \cdot \cdot \text{S}]^+$ . These results proved the occurrence of a fast OH-induced intermolecular charge transfer ( $\text{Met}_2 [\text{S} \cdot \cdot \text{S}]^+ \rightarrow \text{TrpH}$ ). The markedly enhanced decay at 480 nm absorption in the presence of Trp also confirmed the oxidation of Trp by a sulfur-centered radical cation. The decay at 480 nm, which occurred exponentially in the absence of TrpH, was found to be faster with increased Trp concentration ( $0.5\text{--}2 \times 10^{-4} \text{ mol dm}^{-3}$ ). A simplified mechanism of the  $\text{Met}_2 [\text{S} \cdot \cdot \text{S}]^+$  decay is summarized as follows:

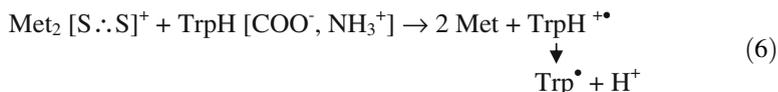


By subtracting the contribution of  $\text{Met}_2 [\text{S} \cdot \cdot \text{S}]^+$  from  $\text{TrpH}^+$  at 570 nm, the dimer intermediate  $\text{Met}_2 [\text{S} \cdot \cdot \text{S}]^+$  was confirmed to decay at the same rate as the rate constant of the tryptophyl cation radical formation [Fig. 2A, inset a (3)]. The experimental rate constant was obtained by following the decay of  $\text{Met}_2 [\text{S} \cdot \cdot \text{S}]^+$  at 480 nm. From Fig. 2A, inset b, a rate constant was determined to be  $3.4 \times 10^8 \text{ dm}^3 \text{ mol}^{-1} \text{ s}^{-1}$ , which is basically consistent with Bobrowski's results [24]. Reaction (5) may only represent an overall reaction if the more reactive species is the molecular radical cation  $\text{AcMet} [\text{S}^{+\bullet}]$  in equilibrium with the dimer. The data presented in the present article did not allow us to distinguish between the two oxidizing species because all data have been recorded at the same Met derivative concentrations. The rate constant  $k_5$  may be exclusively attributed to the reaction of three-electron-bonded species if the rate constants remained unaffected when the Met derivative concentrations were varied.

To compare the oxidation rate constants of both forms of Trp (e.g., zwitterionic  $\text{TrpH} [\text{COO}^-, \text{NH}_3^+]$  and fully protonated  $\text{TrpH} [\text{COOH}, \text{NH}_3^+]$ ),  $\text{Met}_2 [\text{S} \cdot \cdot \text{S}]^+$  was designed to be generated at a pH higher than the  $\text{p}K_a$  of the Trp carboxyl group.

Upon an increased system pH of  $4.5 \pm 0.2$ , the characteristic absorption of  $\text{TrpH}^{+\bullet}$  at 580 nm shifted to a lower wavelength (530 nm; Fig. 2). This shift resulted from the contribution of the neutral radical  $\text{Trp}^{\bullet}$ , which was transferred from  $\text{TrpH}^{+\bullet}$  via deprotonation at  $\text{p}K_a = 4.3$  [25].

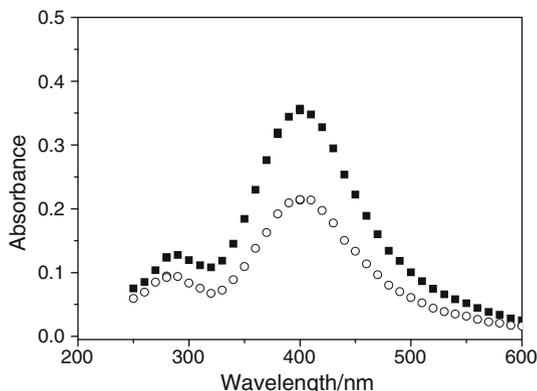
A similar kinetic picture of  $\text{AcMetNH}_2$  at pH 1 was obtained at pH 4.5. The bimolecular rate constant for Reaction (6) was determined by the kinetic analysis of the  $\text{Met}_2 [\text{S} \cdot \cdot \text{S}]^+$  decay at 480 nm ( $k_6 = 2.2 \times 10^8 \text{ dm}^3 \text{ mol}^{-1} \text{ s}^{-1}$ ).



$\text{Met}_2 [\text{S} \cdot \cdot \text{Br}]$  as an oxidant

An oxidized sulfur center can be stabilized not only as a  $\text{S} \cdot \cdot \text{S}^+$  radical cation complex, but also as a  $\text{S} \cdot \cdot \text{X}$  three-electron-bonded to a halogen atom ( $\text{X} = \text{Cl}$  or  $\text{Br}$ ). After the pulse radiolysis of  $\text{N}_2\text{O}$ -saturated aqueous solutions containing

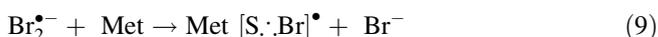
**Fig. 3** Transient absorption spectra of  $2 \times 10^{-3}$  mol dm $^{-3}$  *N*-AcMetNH $_2$  and  $5 \times 10^{-2}$  mol dm $^{-3}$  KBr in N $_2$ O-saturated aqueous solutions at pH 4.3 recorded 1 (filled square) and 8 (open circle)  $\mu$ s after the pulse



$2 \times 10^{-3}$  mol dm $^{-3}$  *N*-AcMetNH $_2$  and 0.05 mol dm $^{-3}$  KBr at pH 4.3, a transient species with a maximum absorption at 400 nm was obtained (Fig. 3). This absorption was assigned to the three-electron-bonded intermediate AcMet [S $\cdot$ :Br] $^{\bullet}$ , consistent with the results of Hiller et al. [26]. A simple reaction mechanism could be described as follows:



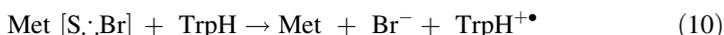
Subsequently, the quantitative reaction of Br $_2^{\bullet-}$  with Met derivatives could be described as:



The rate constants of the formation of AcMet [S $\cdot$ :Br] $^{\bullet}$  were determined as  $k_9 = 6.8 \times 10^8$  mol dm $^{-3}$  s $^{-1}$  at pH 2.  $k_9$  was deduced from a series of the observed decay rates of the radical anion Br $_2^{\bullet-}$  at different *N*-AcMetNH $_2$  concentrations. At pH 10,  $k$  was  $6.4 \times 10^8$  mol dm $^{-3}$  s $^{-1}$  for *N*-AcMetNH $_2$  and was  $7.5 \times 10^8$  mol dm $^{-3}$  s $^{-1}$  for *N*-AcMetOMe.

Subsequently, the oxidation of Trp by Met [S $\cdot$ :Br] was investigated in N $_2$ O-saturated solutions containing  $5 \times 10^{-2}$  mol dm $^{-3}$  KBr,  $2 \times 10^{-3}$  mol dm $^{-3}$  AcMetNH $_2$ , and up to  $2 \times 10^{-4}$  mol dm $^{-3}$  Trp at pH 1.

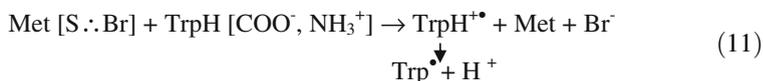
Given that  $k(\text{Br}_2^{\bullet-} + \text{Met}) > k(\text{Br}_2^{\bullet-} + \text{TrpH})$  [26] and the Trp to Met concentration ratio was  $\leq 0.1$ , the direct oxidation of TrpH by Br $_2^{\bullet-}$  was almost negligible. Therefore, any formation process observed after the electron pulse at longer wavelengths (e.g., 580 nm) and at wavelengths identical with the initial decay of Met [S $\cdot$ :Br] (e.g., 400 nm) can be attributed to the formation of the TrpH $^{\bullet+}$  radical cation. This formation was via the direct oxidation of the Trp molecule by Met [S $\cdot$ :Br] (Reaction 10):



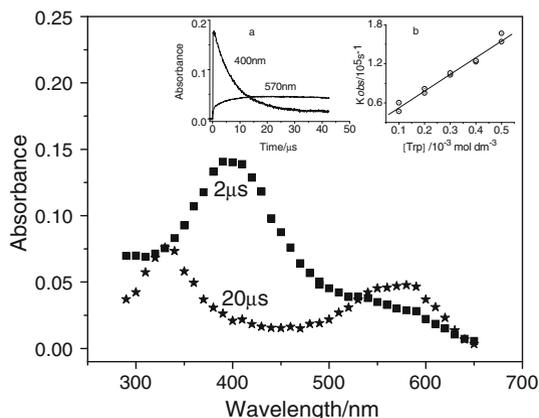
The absorption spectra measured 2 and 20  $\mu$ s after the pulse radiolysis of  $2 \times 10^{-3}$  mol dm $^{-3}$  *N*-AcMetNH $_2$ ,  $2 \times 10^{-4}$  mol dm $^{-3}$  Trp, and  $5 \times 10^{-2}$  mol dm $^{-3}$  KBr in N $_2$ O-saturated aqueous solutions at pH 1 was shown in Fig. 4.

Although there is some indication of a weak absorption band at around 320 nm, the spectrum at 2  $\mu\text{s}$  is presumably caused by Met [S $\cdot$ :Br]. However, the spectra after 20  $\mu\text{s}$  were identical to the spectra of the Met solutions in Fig. 2. In agreement with previous assignments, these spectra after 20  $\mu\text{s}$  are attributed to Trp radical cations. Accordingly, Reaction (10) indeed proceeded with  $k_{10} = 2.6 \times 10^8 \text{ dm}^3 \text{ mol}^{-1} \text{ s}^{-1}$ .

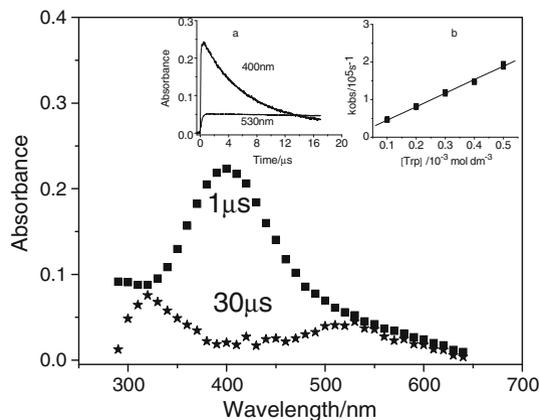
Upon an increased pH of 4.50, the 580 nm absorption of the tryptophyl radical shifted to lower wavelengths (530 nm; Fig. 5). This shift was consistent with the known effect of pH on the transient absorption maxima of tryptophyl species. The rate of the intermolecular oxidation of the Trp zwitterionic form by Met [S $\cdot$ :Br] was practically the same with the fully protonated molecule ( $k_{11} = 3.3 \times 10^8 \text{ dm}^3 \text{ mol}^{-1} \text{ s}^{-1}$ ).



Finally, the rate constants of the intermolecular electron transfers from Trp to the three-electron-bonded intermediates are listed in Table 1. The intermediates were derived from the oxidation of AcMetNH<sub>2</sub> and AcMetOMe by both  $\cdot\text{OH}$  radicals and Br<sub>2</sub> $\cdot^-$  radical anions in aqueous solutions at different pH values. Apparently, intermolecular charge transfers within peptides containing Met and aromatic amino acids such as Trp occurred very rapidly. Similarly efficient intramolecular charge



**Fig. 4** Transient absorption spectra of  $2 \times 10^{-3} \text{ mol dm}^{-3}$  *N*-AcMetNH<sub>2</sub>,  $2 \times 10^{-4} \text{ mol dm}^{-3}$  Trp, and  $5 \times 10^{-2} \text{ mol dm}^{-3}$  KBr in N<sub>2</sub>O-saturated aqueous solutions at pH 1 recorded 2 (filled square) and 20 (star)  $\mu\text{s}$  after the pulse at  $\sim 35 \text{ Gy/pulse}$ . *Inset a* The transient absorption curves for the decay of Met<sub>2</sub> [S $\cdot$ :Br] at 400 nm and the formation of TrpH<sup>+</sup> at 570 nm. *Inset b* The plot of the observed decay rate constants of the transient species Met<sub>2</sub> [S $\cdot$ :Br] (400 nm) against Trp concentrations at pH 1 ( $\sim 12 \text{ Gy/pulse}$ )



**Fig. 5** Transient absorption spectra of  $2 \times 10^{-3} \text{ mol dm}^{-3}$  *N*-AcMetNH<sub>2</sub>,  $2 \times 10^{-4} \text{ mol dm}^{-3}$  Trp, and  $5 \times 10^{-2} \text{ mol dm}^{-3}$  KBr in N<sub>2</sub>O-saturated aqueous solutions at pH 4.4 recorded 1 (filled square) and 30 (star)  $\mu\text{s}$  after the pulse. *Inset a* The transient absorption curves for the decay of Met<sub>2</sub> [S $\cdot$ :Br] at 400 nm and the formation of TrpH<sup>+</sup> at 530 nm. *Inset b* The plot of the observed decay rate constants of transient species Met<sub>2</sub> [S $\cdot$ :Br] (400 nm) against tryptophan concentrations at pH 4.4 ( $\sim 12 \text{ Gy/pulse}$ )

**Table 1** The rate constants of the intermolecular electron transfers from Trp to AcMteNH<sub>2</sub> and to AcMetOMe in aqueous solutions

Initiator	$k \text{ (mol dm}^{-3} \text{ s}^{-1}\text{)}$					
	AcMetNH <sub>2</sub>		AcMetOMe		Met <sup>a</sup>	
pH	1	4.3	1	4.5	1	4.3
$\bullet\text{OH}$	$3.42 \times 10^8$	$2.2 \times 10^8$	$4.0 \times 10^8$	$2.8 \times 10^8$	$3.8 \times 10^8$	
$\text{Br}_2^{\bullet-}$	$2.6 \times 10^8$	$3.3 \times 10^8$	$3.2 \times 10^8$	$3.6 \times 10^{8b}$	$4.6 \times 10^8$	$4.6 \times 10^8$

<sup>a</sup> From Bobrowski [24]

<sup>b</sup> pH 5.5

transfers with rate constants in the order of  $10^2\text{--}10^4 \text{ s}^{-1}$  were also found to occur between Trp and Tyr residues in numerous synthetic and natural peptides [27, 28].

## Conclusions

Intermolecular electron transfers from Trp to three-electron-bonded intermediates derived from the oxidation of *N*-AcMetNH<sub>2</sub> and *N*-AcMetOMe by  $\bullet\text{OH}$  and  $\text{Br}_2^{\bullet-}$  radicals were investigated. The reactions took place in N<sub>2</sub>O-saturated aqueous solutions with pH values of 1.0–4.5. A series of rate constants for the intermolecular electron transfers were determined. The results revealed that Trp–Met pairs can form very favorable redox couples in biological systems, which is consistent with earlier reports of more efficient and rapid intramolecular charge transfers in synthetic as well as natural peptides containing Met.

**Acknowledgments** The authors are grateful to the staff of LINAC in the University of Tokyo for their technical assistance and encouragement in pulse radiolysis experiments.

## References

1. O.B. Morozova, S.E. Korchak, R.Z. Sagdeev, A.V. Yurkovskaya, Time-resolved chemically induced dynamic nuclear polarization studies of structure and reactivity of methionine radical cations in aqueous solution as a function of pH. *J. Phys. Chem. A* **109**, 10459–10466 (2005)
2. M. Davies, The oxidative environment and protein damage. *Biochim. Biophys. Acta* **1703**, 93–109 (2005)
3. E.R. Stadtman, H.V. Remmen, A. Richardson, N.B. Wehra, R.L. Levine, Methionine oxidation and aging. *Biochim. Biophys. Acta* **1703**, 135–140 (2005)
4. C. Schoneich, D. Pogocki, G.L. Hug, K. Bobrowski, Free radical reactions of methionine in peptides: mechanisms relevant to  $\beta$ -amyloid oxidation and Alzheimer's disease. *J. Am. Chem. Soc.* **125**, 13700–13713 (2003)
5. I. Carmichael, Theoretical studies on some  $S[ \cdot ]N$  three-electron bonded radical cations. *Acta Chem. Scand.* **51**, 567–571 (1997)
6. C. Schoneich, D. Pogocki, P. Wisniowski, G.L. Hug, K. Bobrowski, Intramolecular Sulfur–Oxygen bond formation in radical cations of *N*-acetylmethionine amide. *J. Am. Chem. Soc.* **122**, 10224–10225 (2000)
7. I. Fourre, J. Berges, C. Houee-Levin, Structural and topological studies of methionine radical cations in dipeptides: electron sharing in two-center three-electron bonds. *J. Phys. Chem. A* **114**, 7359–7368 (2010)
8. K. Bobrowski, G.L. Hug, D. Pogocki, B. Marciniak, C. Schoneich, Stabilization of sulfide radical cations through complexation with the peptide bond: mechanisms relevant to oxidation of proteins containing multiple methionine residues. *J. Phys. Chem. B* **111**, 9608–9620 (2007)
9. D. Pogocki, Ch. Schoneich, Redox properties of Met(35) in neurotoxic beta-amyloid peptide. A molecular modeling study. *Chem. Res. Toxicol.* **15**, 408–418 (2002)
10. D.A. Butterfield,  $\beta$ -Amyloid-associated free radical oxidative stress and neurotoxicity: implications for Alzheimer's disease. *Chem. Res. Toxicol.* **10**, 495–506 (1997)
11. K.-O. Hiller, K.D. Asmus, Oxidation of methionine by  $X_2^{\bullet+}$  in aqueous solution and characterization of some  $S \cdot X$  three-electron bonded intermediates, a pulse radiolysis study. *Int. J. Radiat. Biol.* **46**, 583–595 (1981)
12. G.S. Chu, S.D. Yao, Z.C. Zhang, W.F. Wang, M.W. Zhang, Intermolecular electron transfer involving phosphoryl and nonphosphoryl methionine and tryptophan, a pulse radiolysis study. *Radiat. Phys. Chem.* **50**, 385–391 (1997)
13. K. Bobrowski, R. Lubis, Intermolecular charge transfer involving tryptophan, tyrosine and three electron-bonded intermediates derived from methionine. *Int. J. Radiat. Biol.* **50**, 1039–1050 (1986)
14. W.A. Prutz, J. Butler, E.J. Land, A.J. Swallow, The role of sulphur peptide function in free radical transfer: a pulse radiolysis study. *Int. J. Radiat. Biol.* **55**, 539–556 (1989)
15. W.A. Prutz, J. Butler, E.J. Land, Methionyl-tyrosyl radical transitions initiated by  $Br_2^{\bullet+}$  in peptide model systems and ribonuclease A. *Int. J. Radiat. Biol.* **47**, 149–156 (1985)
16. K. Bobrowski, K.L. Wierzchowski, J. Holcman, M. Ciurak, Pulse radiolysis studies of intramolecular electron transfer in model peptides and proteins. *Int. J. Radiat. Biol.* **62**, 507–516 (1992)
17. G.V. Buxton, C.R. Stuart, Re-evaluation of the thiocyanate dosimeter for pulse radiolysis. *J. Chem. Soc. Faraday Trans* **91**, 279–281 (1991)
18. H.Y. Fu, M.Z. Lin, Y. Muroya, K. Hata, Y. Katsumura, Free radical scavenging reactions and antioxidant activity of silybin: Mechanistic aspects and pulse radiolytic studies. *Free Radic. Res.* **43**, 887–897 (2009)
19. L.K. Mee, S.J. Adelstein, Inactivation of ribonuclease S-peptide by the primary aqueous radicals. *Radiat. Res.* **60**, 422–431 (1974)
20. K.-O. Hiller, B. Masloch, M. Gobl, K.-D. Asmus, Mechanism of the OH radical induced oxidation of methionine in aqueous solution. *J. Am. Chem. Soc.* **103**, 2734–2743 (1981)
21. K. Bobrowski, C. Schöneich, J. Holcman, K.D. Asmus, OH radical induced decarboxylation of methionine-containing peptides. Influence of peptide sequence and net charge. *J. Chem. Soc. Perkin Trans.* **2**, 353–362 (1991)

22. R.C. Armstrong, A.J. Swallow, Pulse- and gamma radiolysis of aqueous solutions of tryptophan. *Radiat. Res.* **40**, 563–579 (1969)
23. J.L. Redpath, R. Santus, J. Ovidia, U. Grossweiner, The oxidation of tryptophan by radical cations. *Int. J. Radiat. Biol.* **27**, 201–204 (1975)
24. K. Bobrowski, R. Łubis, Intermolecular charge transfer involving tryptophan, tyrosine and three electron-bonded intermediates derived from methionine. *Int. J. Radiat. Biol.* **50**, 1039–1050 (1986)
25. R.F. Evans, C.A. Ghiron, W.A. Volkert, R.R. Kuntz, R. Santus, M. Bazin, Flash photolysis of *N*-acetyl-L-tryptophanamide; acid-base equilibrium of radical transients. *Chem. Phys. Lett.* **42**, 43–45 (1976)
26. K.-O. Hiller, K.-O. Lier, K.-D. Asmus,  $Tl^{2+}$  and  $Ag^{2+}$  metal-ion induced oxidation of methionine in aqueous solution. A pulse radiolysis study. *Int. J. Radiat. Biol.* **40**, 597–604 (1980)
27. W.A. Prutz, J. Butler, E.J. Land, A.J. Swallow, Direct demonstration of electron transfer between tryptophan and tyrosine in proteins. *Biochem. Biophys. Res. Commun.* **96**, 408–414 (1980)
28. A.G. Griesbeck, J. Neudörfl, A. deKiff, Photoinduced electron-transfer chemistry of the bielectrophoric *N*-phthaloyl derivatives of the amino acids tyrosine, histidine and tryptophan. *Beilstein J. Org. Chem.* **7**, 518–524 (2011)