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# Synthesis and identification in bacterial lipopolysaccharides of 5,7-diacetamido-3,5,7,9-tetradeoxy-D-glycero-D-galacto- and -D-glycero-D-talo-non-2-ulosonic acids

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## Abstract

5,7-Diacetamido-3,5,7,9-tetradeoxy-D-glycero-D-galacto- and -D-glycero-D-talo-non-2-ulosonic acids were synthesized by condensation of 2,4-diacetamido-2,4,6-trideoxy-D-mannose with oxalacetic acid. Comparison of the <sup>1</sup>H and <sup>13</sup>C NMR data and the specific optical rotation values of these monosaccharides and the corresponding L-glycero-D-galacto and L-glycero-D-talo isomers synthesized earlier [Tsvetkov, Y. E.; Shashkov, A. S.; Knirel, Y. A.; Backinowsky, L. V.; Zähringer, U. *Mendeleev Commun.* **2000**, 90–92] with data of the natural compounds enabled the identification in bacterial lipopolysaccharides of derivatives of 5,7-diamino-3,5,7,9-tetradeoxy-D-glycero-D-galacto-non-2-ulosonic (legionaminic) acid and epimers of legionaminic acid at C-4 and C-8. © 2001 Elsevier Science Ltd. All rights reserved.

**Keywords:** 5,7-Diamino-3,5,7,9-tetradeoxynon-2-ulosonic acid; Synthesis; Legionaminic acid, identification of; Lipopolysaccharide components; *Legionella pneumophila*

*N*-Acyl and *O*-acetyl derivatives of 5,7-diamino-3,5,7,9-tetradeoxynon-2-ulosonic acids have been reported as components of the lipopolysaccharides (LPS) of gram-negative bacteria, which play a role in serological specificity and endow the bacterial surface with peculiar physicochemical properties.<sup>1–3</sup> The first monosaccharide of this class discovered in the mid-1980s in the LPS of *Pseudomonas aeruginosa* and *Shigella boydii* was identified as the

L-glycero-L-manno isomer (pseudaminic acid).<sup>4,5</sup> The D-glycero-L-galacto configuration that was ascribed to the second isomer found first in *P. aeruginosa*,<sup>6</sup> was later revised to the L-glycero-D-galacto configuration.<sup>7,8</sup> A homopolymer of a derivative of a 5,7-diamino-3,5,7,9-tetradeoxynon-2-ulosonic acid, called legionaminic acid, was described as the O-chain of the LPS in *Legionella pneumophila* serogroup 1<sup>3,9</sup> and *Pseudomonas fluorescens* ATCC 49271,<sup>10,11</sup> and it was suggested that legionaminic acid has the same L-glycero-D-galacto configuration.<sup>7</sup> While the configurations of the higher sugars in these and some

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other LPS<sup>12–14</sup> (Table 1) were ascribed by NMR spectroscopy without their isolation as monosaccharides, two 5,7-diacetamido-3,5,7,9-tetradeoxynon-2-ulosonic acids were released by mild acid hydrolysis from the LPS of *Vibrio alginolyticus*<sup>15</sup> and *L. pneumophila*,<sup>16</sup> and their configurations were tentatively assigned to be L-glycero-D-galacto and L-glycero-D-talo, respectively.

Aiming at unambiguous identification of these higher sugars, including determination of their absolute configuration, we have synthesized, for the first time, 5,7-diacetamido-3,5,7,9-tetradeoxy-L-glycero-D-galacto-**(1)** and -L-glycero-D-talo-non-2-ulosonic acids (**2**) from 2,4-diacetamido-2,4,6-trideoxy-L-gulose.<sup>17</sup> In this paper, we report the first synthesis of the corresponding D-glycero-D-galacto (**3**) and D-glycero-D-talo (**4**) isomers, which enabled determination of the configurations of the natural monosaccharides, legionaminic acid (Leg) and two of its epimers, 4-epi- and 8-epi-legionaminic acid (4eLeg and 8eLeg, respectively).

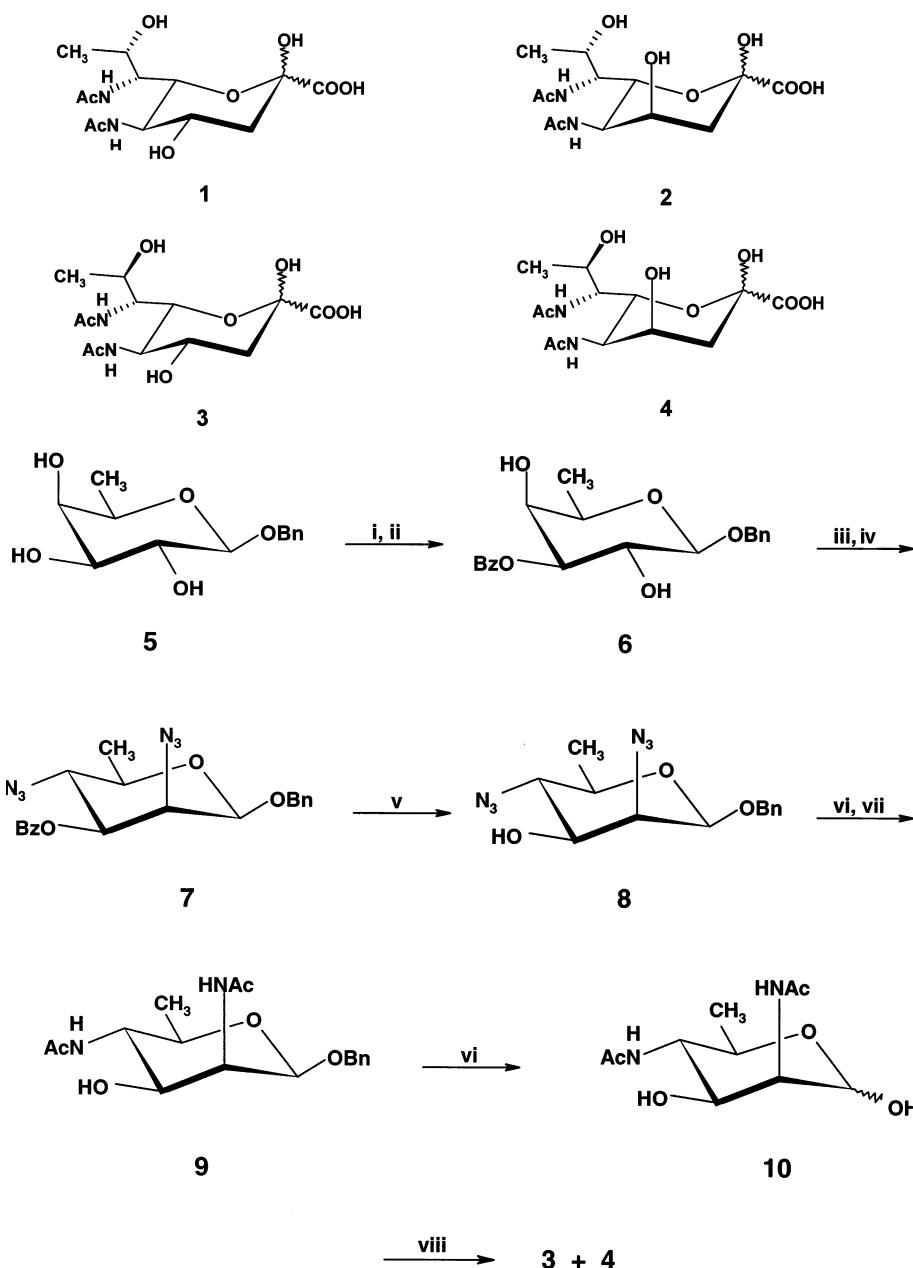
Synthesis was performed starting from benzyl β-D-fucopyranoside **5**,<sup>18</sup> which afforded 3-benzoate **6** on Bu<sub>2</sub>SnO-mediated selective benzoylation (Scheme 1). Conversion of **6** into the corresponding 2,4-ditriflate, followed by

bis-azidation with Bu<sub>4</sub>NN<sub>3</sub>, resulted in diazide **7** having the manno configuration. After debenzoylation (**7** → **8**), reduction of the azido groups was performed by hydrogenation over Pd(OH)<sub>2</sub>–C without affecting the benzyl group<sup>19</sup> and followed by N-acetylation (**8** → **9**). Removal of the benzyl aglycon by hydrogenolysis gave 2,4-diacetamido-2,4,6-trideoxy-D-mannose (**10**) in 38% overall yield from **5**. All new compounds were fully characterized by NMR spectroscopy and elemental analysis.

Condensation of **10** with oxalacetic acid in the presence of sodium tetraborate at pH 10.5<sup>20</sup> gave a mixture of **3** and **4**. The individual compounds **3**, [α]<sub>D</sub> + 27.2° (c 1, water), and **4**, [α]<sub>D</sub> – 12.5° (c 1, water), were isolated in 7 and 10% yield, respectively, by anion-exchange chromatography on Dowex 1 × 8 (aq 0.3 M formic acid) followed by reversed-phase C<sub>18</sub> HPLC (aq 0.05% CF<sub>3</sub>CO<sub>2</sub>H). The <sup>1</sup>H and <sup>13</sup>C NMR spectroscopic data (Table 2) showed that **3** and **4** are mixtures of α and β anomers in the ratio 1:18 and 1:5.4, respectively, and proved their configurations. Particularly, the J<sub>3a,4</sub>, J<sub>4,5</sub>, and J<sub>5,6</sub> coupling constant values clearly indicated that the substituents at C-4,5,6 in **3** and C-5,6 in **4** are equatorial, whereas the hydroxy group at C-4 in **4** is axial.

Table 1  
Some naturally occurring isomers of 5,7-diamino-3,5,7,9-tetradeoxynon-2-ulosonic acid

Source (lipopolysaccharide of)	Configuration		
	Originally ascribed	Previously revised to <sup>7,8</sup>	Revised to/confirmed in this work
<i>Legionaminic acid (Leg)</i>			
<i>Legionella pneumophila</i>	D-glycero-L-galacto <sup>9</sup>	L-glycero-D-galacto	D-glycero-D-galacto
<i>Pseudomonas fluorescens</i>	D-glycero-L-galacto <sup>10,11</sup>	L-glycero-D-galacto	D-glycero-D-galacto
<i>Vibrio salmonicida</i>	L-glycero-D-galacto <sup>8</sup>		D-glycero-D-galacto
<i>Acinetobacter baumannii</i>	L-glycero-D-galacto <sup>12</sup>		D-glycero-D-galacto
<i>Vibrio alginolyticus</i>	D-glycero-L-galacto <sup>15</sup>	L-glycero-D-galacto	D-glycero-D-galacto
<i>8-Epilegionaminic acid (8eLeg)</i>			
<i>Pseudomonas aeruginosa</i>	D-glycero-L-galacto <sup>6</sup>	L-glycero-D-galacto	L-glycero-D-galacto
<i>Salmonella arizonae</i>	D-glycero-L-galacto <sup>13</sup>	L-glycero-D-galacto	L-glycero-D-galacto
<i>Yersinia ruckerii</i>	D-glycero-L-galacto <sup>14</sup>	L-glycero-D-galacto	L-glycero-D-galacto
<i>4-Epilegionaminic acid (4eLeg)</i>			
<i>Legionella pneumophila</i>	L-glycero-D-talo <sup>16</sup>		D-glycero-D-talo



Scheme 1. **i**, Bu<sub>2</sub>SnO, benzene, reflux; **ii**, BzCl, rt; **iii**, Tf<sub>2</sub>O, pyridine, CH<sub>2</sub>Cl<sub>2</sub>, 0 °C; **iv**, Bu<sub>4</sub>NN<sub>3</sub>, toluene, 100 °C; **v**, MeONa, MeOH, rt; **vi**, H<sub>2</sub>, Pd(OH)<sub>2</sub>-C, MeOH, rt; **vii**, Ac<sub>2</sub>O, MeOH, rt; **viii**, oxalacetic acid, Na<sub>2</sub>B<sub>4</sub>O<sub>7</sub>, pH 10.5, rt.

Comparison of the <sup>1</sup>H and <sup>13</sup>C NMR data of synthetic and natural compounds (Table 2) suggested that 5,7-diacetamido-3,5,7,9-tetra-deoxynon-2-ulosonic acids from LPS of *V. alginolyticus* (Leg5Ac7Ac)<sup>15</sup> and *L. pneumophila* (4eLeg5Ac7Ac)<sup>16</sup> have the same relative configurations as **3** and **4** and differ from those of **1** and **2**, respectively. In addition to the *J*<sub>H,H</sub> coupling constant values, this conclusion was based on the <sup>13</sup>C NMR chemical shifts for C-6 and C-8, which are indicative of

the configuration at the side chain. The specific optical rotation values of the natural compounds, [α]<sub>D</sub> + 25.2° (*c* 0.5, water) for Leg5Ac7Ac<sup>15</sup> and [α]<sub>D</sub> − 3.9° (*c* 0.2, water) for 4eLeg5Ac7Ac (this work), compared with the data for **3** and **4**, respectively (see above), indicated that they have the same absolute configurations as well. The identity of the monosaccharide from *L. pneumophila* was confirmed by GLC-MS of the acetylated (+)-2-butyl ester (+)-2-butyl glycosides.

Table 2  
500 MHz  $^1\text{H}$  and 125 MHz  $^{13}\text{C}$  NMR data of synthetic compounds **1–4** and natural compounds Leg5Ac7Ac and 4eLeg5Ac7Ac for solutions in  $\text{D}_2\text{O}$  at 30 °C ( $\delta$ , ppm, related to internal acetone,  $\delta_{\text{H}}$  2.225,  $\delta_{\text{C}}$  31.45;  $J_{\text{H,H}}$ , Hz)

Compound	$\text{H-3eq}$ ( $J_{\text{3eq},\text{3ax}}$ )	$\text{H-3ax}$ ( $J_{\text{3ax},4}$ )	$\text{H-4}$ ( $J_{\text{3eq},4}$ )	$\text{H-5}$ ( $J_{4,5}$ )	$\text{H-6}$ ( $J_{5,6}$ )	$\text{H-7}$ ( $J_{6,7}$ )	$\text{H-8}$ ( $J_{7,8}$ )	$\text{H-9}$ ( $J_{8,9}$ )	$\text{CH}_3\text{CON}$
<i><sup>1</sup>H NMR</i>									
<b>1<math>\alpha</math></b>	2.67 (12.9)	1.69 (12)	3.81 (4.6)	3.65 (10.2)	4.82 (10)	3.90	3.97	1.17 (6.3)	
<b>1<math>\beta</math></b> <sup>17</sup>	2.30 (13.1)	1.85 (12.2)	3.92 (4.8)	3.70 (10.2)	4.14 (10.3)	3.93 (2.0)	3.89 (6.4)	1.16 (6.2)	1.96, 2.00
<b>2<math>\alpha</math></b> <sup>17</sup>	2.65 (14.4)	1.94 (2.7)	4.08 (3.6)	3.84 (2.7)	4.47 (10.5)	3.89	4.08	1.28 (6.3)	1.96, 2.04
<b>2<math>\beta</math></b> <sup>17</sup>	2.18 (14.9)	2.13 (3.3)	4.11 (2.9)	3.89 (2.8)	4.48 (10.6)	3.95 (1.3)	3.96	1.21 (5.7)	1.97, 2.01
<b>3<math>\alpha</math></b>	2.73 (12.9)	1.71 (11.9)	3.82 (4.7)	3.68	3.93 (10.3)	3.84	3.94	1.16	1.97, 2.00
<b>3<math>\beta</math></b>	2.31 (13.1)	1.87 (11.7)	3.98 (4.8)	3.72 (10.3)	4.31 (10.5)	3.91 (1.9)	3.85 (8.9)	1.16 (6.2)	1.99, 2.00
<b>4<math>\alpha</math></b>	2.69 (14.4)	1.95 (3.5)	4.10 (3.0)	3.86 (2.9)	4.55 (10.8)	3.88 (2.3)	4.00 (8.6)	1.20 (6.4)	1.96, 2.00
<b>4<math>\beta</math></b>	2.19 (14.9)	2.14 (3.4)	4.13 (3.0)	3.90 (2.9)	4.63 (10.8)	3.92	3.92	1.18 (5.5)	1.98, 1.99
Leg5Ac7Ac $\beta$ <sup>a</sup>	2.20 (13)	1.80 (11.5)	3.93 (5)	3.70 (10.3)	4.22 (10.4)	3.84 (1.8)	3.84	1.14 (6.3)	
4eLeg5Ac7Ac $\alpha$ <sup>b</sup>	2.67 (14.5)	1.95 (3.1)	4.09 (2.6)	3.86 (2.6)	4.54 (11)	3.86 (2.2)	4.00 (8.5)	1.20 (6.6)	1.96, 2.01
4eLeg5Ac7Ac $\beta$ <sup>b</sup>	2.17 (14.9)	2.13 (3.1)	4.12 (2.6)	3.90 (2.6)	4.62 (11)	3.91 (<2)	3.91	1.18 (5.3)	1.99, 2.00
<i><sup>13</sup>C NMR</i>									
	C-1	C-2	C-3	C-4	C-5	C-6	C-7	C-8	C-9
<b>1<math>\alpha</math></b>	174.3	96.5	41.2	68.9	53.7	75.2	54.4	69.4	19.8
<b>1<math>\beta</math></b> <sup>17</sup>	40.4	68.3	54.1	72.9	54.4	69.3	69.3	19.8	23.1; 23.4
<b>2<math>\alpha</math></b> <sup>17</sup>	40.0	66.6	50.1	72.6	54.9	69.7	69.7	19.9	23.0; 23.1
<b>2<math>\beta</math></b> <sup>17</sup>	37.7	66.9	50.0	68.5	54.9	69.2	69.2	19.9	174.6 <sup>c</sup> ; 175.2
<b>3<math>\alpha</math></b>	41.4	69.2	53.4	73.2	54.4	68.0	68.0	20.4	23.0; 23.4
<b>3<math>\beta</math></b>	174.4 <sup>c</sup>	96.6	40.3	68.4	53.9	70.9	54.4	67.5	20.4
<b>4<math>\alpha</math></b>	40.2	66.9	49.7	70.3	55.0	68.2	68.2	20.4	
<b>4<math>\beta</math></b>	37.6	67.1	49.7	66.5	54.7	67.2	67.2	20.4	23.0; 23.1
Leg5Ac7Ac $\beta$ <sup>a</sup>	175.1	97.7	40.9	67.9	54.1	70.8	54.5	68.3	20.4
4eLeg5Ac7Ac $\alpha$ <sup>b</sup>	96.5	37.6	66.9	49.8	70.2	54.9	68.1	20.4	22.9; 23.1
4eLeg5Ac7Ac $\beta$ <sup>b</sup>	174.9 <sup>c</sup>		67.2	49.7	66.5	54.6	67.4	20.4	174.6 <sup>c</sup> ; 174.9 <sup>c</sup>

<sup>a</sup> Obtained by mild acid hydrolysis of LPS of *V. alginolyticus* strain 945-80.<sup>15</sup>

<sup>b</sup> Obtained by mild acid hydrolysis of O-deacylated LPS of *L. pneumophila* serogroup 1, strain Philadelphia 1 (this work).

<sup>c</sup> Assignment within one row could be interchanged.

The data obtained enabled also determination of the configuration of the higher sugars that could not be isolated as monosaccharides. Based on the  $J_{\text{H,H}}$  coupling constant and  $^{13}\text{C}$  NMR chemical shift data, it was concluded that legionaminic acid in the homopolymer O-chain of LPS of *L. pneumophila*<sup>9</sup> has the same relative configuration as **3** (e.g., compare  $\delta_{\text{C-6}}$  73.5 and  $\delta_{\text{C-8}}$  68.2 in Leg5Ac7Ac $\alpha$ <sup>9</sup> with the values  $\delta_{\text{C-6}}$  73.2 and  $\delta_{\text{C-8}}$  68.0 in **3** $\alpha$  but  $\delta_{\text{C-6}}$  75.2 and  $\delta_{\text{C-8}}$  69.4 in **1** $\alpha$ ). It has also the same absolute configuration, as shown by GLC–MS of the acetylated (+)-2-butyl ester (+)-2-butyl glycosides derived from LPS. Therefore, the configuration of Leg in the *L. pneumophila* polysaccharide should be revised from L-glycero-D-galacto<sup>7–9</sup> to D-glycero-D-galacto (Table 1). In contrast, the L-glycero-D-galacto configuration that had been ascribed to the isomer found in LPS of *P. aeruginosa*<sup>6–8</sup>, could be confirmed. Indeed, this sugar is characterized by the values  $\delta_{\text{C-6}}$  74.8 and  $\delta_{\text{C-8}}$  70.1<sup>6</sup> and thus represents 8eLeg5Ac7Ac. Similarly, it was concluded that derivatives of Leg are present in the O-chain polysaccharides of *Vibrio salmonicida* LPS,<sup>8</sup> *Pseudomonas fluorescens*<sup>10,11</sup> and *Acinetobacter baumannii*,<sup>12</sup> whereas derivatives of 8eLeg are components of LPS of *Salmonella arizona*e<sup>13</sup> and *Yersinia ruckerii*<sup>14</sup> (Table 1).

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