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J. Org. Chem., Just Accepted Manuscript • Publication Date (Web): 02 Jun 2015

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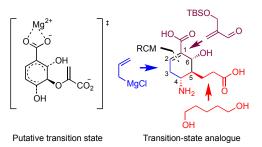
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Stereocontrolled Synthesis of a Potential Transition-State Inhibitor of the Salicylate Synthase MbtI from *Mycobacterium tuberculosis*

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ABSTRACT: Mycobactins are small-molecule iron chelators (siderophores) produced by *Mycobacterium tuberculosis (Mtb)* for iron mobilization. The bifunctional salicylate synthase MbtI catalyzes the first step of mycobactin biosynthesis through the conversion of the primary metabolite chorismate into salicylic acid via isochorismate. We report the design, synthesis and biochemical evaluation of an inhibitor based on the putative transition-state (TS) for the isochorismatase partial reaction of MbtI. The inhibitor mimics the hypothesized charge build-up at C-4 of chorismate in the TS as well as C-O bond-formation at C-6. Another important design element of the inhibitor is replacement of the labile pyruvate side-chain in chorismate with a stable *C*-linked propionate isostere. We developed a stereocontrolled synthesis of the highly functionalized cyclohexene inhibitor that features an asymmetric aldol reaction using a titanium enolate, diastereoselective Grignard addition to a *tert*-butanesulfinyl aldimine, and ring closing olefin metathesis as key steps.

INTRODUCTION

Tuberculosis (TB), the leading cause of infectious disease mortality after HIV-AIDS, is due to members of the Mycobacterium tuberculosis (Mtb) complex that includes seven closely related species.¹ The emergence of multi-drug and extensively-drug resistant strains of *Mtb* has renewed focus on the development of anti-tubercular agents with novel modes of action.² With the exception of pyrazinamide, whose mechanism remains an enigma,³ the antibiotics presently used in TB chemotherapy target a limited set of biochemical pathways in macromolecular and cofactor biosynthesis.⁴ Iron is a required cofactor for more than 40 enzymes in *Mtb*, but is highly restricted in a vertebrate host where the concentration of free iron (Fe^{2+}) is estimated as 10^{-24} M.⁵ In order to obtain iron *Mtb* deploys two complementary strategies: synthesis of the mycobactin siderophores,⁶ which are small-molecule iron chelators that scavenge iron from host tissues and uptake of heme through a specialized heme receptor followed by heme degradation to release the iron.⁷ The relative contribution of each mechanism for iron mobilization *in vivo* is unknown and both may play a role during different stages of infection. Mtb mutants unable to produce mycobactins cannot replicate under iron-restricted conditions and are cleared in vivo, thereby confirming the importance of mycobactin-mediated iron acquisition in Mtb.⁸

The bifunctional salicylate synthase MbtI is responsible for the first committed step in mycobactin biosynthesis through the conversion of chorismate into salicylic acid, which is accomplished in two partial reactions at the same active site.⁹ In the first reaction, MbtI catalyzes the interconversion of chorismate to isochorismate that has been hypothesized to proceed through

a concerted $S_N 2''$ reaction mechanism via transition state **TS1** (Figure 1).^{9c, 10} The isochorismatase activity of MbtI requires Lys205, which nucleophilically activates a water molecule for attack on chorismate at C-6 and Glu252 that polarizes the C-4 hydroxyl-leaving group. In the second reaction, pyruvate is eliminated through an intramolecular [3,3]-sigmatropic rearrangement, formerly a retro-Ene reaction, to afford salicylic acid via bicyclic transition state **TS2** (Figure 1).¹¹

MbtI represents an appealing target for development of inhibitors of mycobactin biosynthesis since it is structurally and biochemically characterized, has no human orthologs, and is conditionally essential under iron-deficient conditions. Following the foundational studies of Abell et al. on inhibitors of chorismate-utilizing enzymes,^{12a-e} Payne and coworkers were the first to disclose inhibitors of MbtI and their most potent compound is the rationally designed substrate mimic 1 (Figure 2A) containing a 2,3-dihydroxybenzoate scaffold.^{12f-g} This compound is a competitive inhibitor and possesses an inhibition constant (K_i) of approximately 10 μ M. In a complementary approach employing high-throughput screening, Vasan and co-workers reported benzimidazole-2-thione **2** as a noncompetitive inhibitor with similar potency to **1** (Figure 2A).¹³ Based on the modest potency of the described MbtI inhibitors, we chose to investigate potential transition-state (TS) inhibitors of MbtI. TS inhibitors exploit the affinity of an enzyme for the TS relative to the Michaelis complex.¹⁴ Capturing even a small percentage of the TS binding energy with an inhibitor, in theory, allows the design of exceptionally potent inhibitors. TS strategies are less successful when the non-catalyzed rate is high since the amount of potential TS

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stabilization will be smaller. In the case of MbtI, the first transition state **TS1** is more desirable to mimic than the second bicyclic transition state **TS2** due to ease of the non-catalyzed sigmatropic rearrangment.^{11, 15}

Inspired by the pioneering work of Kozlowski, Bartlett *et al.*, who reported on the synthesis of **3** as a TS inhibitor for several chorismate-utilizing enzymes,¹⁶ we report the synthesis and characterization of cyclohexene **4** that mimics putative **TS1** through incorporation of an amino group at C-4 to resemble the charged leaving group of **TS1** (Figure 2B). Another key design element is replacement of the sigmatropically labile pyruvate side-chain at C-5 with a stable *C*-linked propionate fragment because MbtI possesses pyruvate lyase activity, unlike the monofunctional chorismate-utilizing enzymes investigated by Kozlowski *et al.*. Interestingly, inhibitor **4** bears a striking resemblance to Oseltamivir (trade name Tamiflu) **5** (Figure 2C), an antiviral drug developed at Gilead that is a TS inhibitor of viral neuraminidase.¹⁷

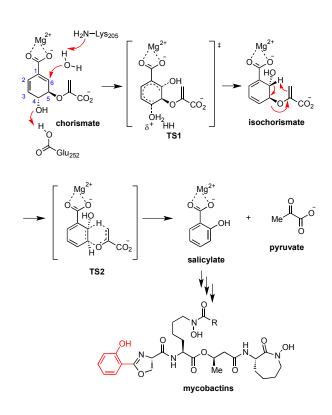


Figure 1. Conversion of chorismate to salicylate via isochorismate catalyzed by MbtI.

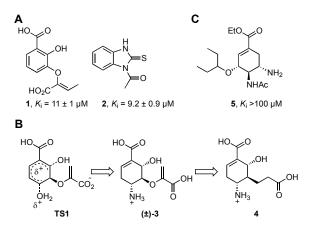


Figure 2. (A) Reported MbtI inhibitors; (B) Rationale for TS inhibitor design of 4; (C) Oseltamivir (Tamiflu) a neuraminidase TS inhibitor. K_i values are with respect to MbtI.

RESULTS AND DISCUSSION

Retrosynthetic Plan. The challenge of the target molecule **4** resides in the construction of three contiguous stereocenters in the highly functionalized cyclohexene scaffold. Our first

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retrosynthetic plan involved disconnection of the C-5 propionate side-chain to thionocarbonate **6** through a radical coupling with *tert*-butyl acrylate. This would allow us to dovetail into the elegant racemic synthesis of **7** reported by Kozlowski and co-workers (Figure 3A).^{16b} We also noted the structural similarity between **4** and Oseltamivir **5** (Figure 2B and 2C), and were inspired by the efficient ring-closing metathesis (RCM) reaction reported by Yao and coworkers¹⁸ for construction of the cyclohexene core of Oseltamivir **5**. Accordingly, as shown in Figure 3B, RCM disconnection of **8** leads to acyclic diene **9** that can be further simplified to syn aldol adduct **10** via an asymmetric Grignard addition employing Ellman's *N*-sulfinamide auxiliary.¹⁹ Intermediate **10** can be readily assembled through syn-aldol reaction between oxazolidinone **11** and aldehyde **12**. An advantage of this second approach is the ability to control stereochemistry on an acyclic precursor using contemporary methods in asymmetric synthesis.

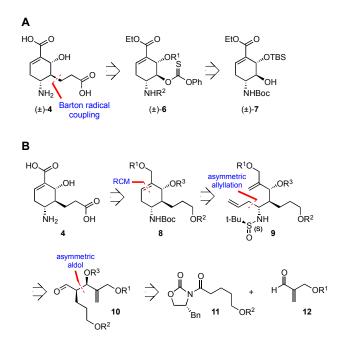
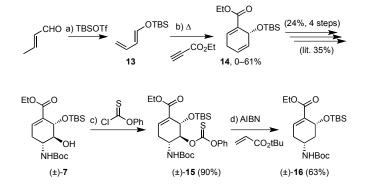


Figure 3. Retrosynthetic analysis.

Radical coupling strategy. Silvloxybutadiene 13 was prepared from crotonaldehyde in 78% yield through modification of the reported procedure^{16b} using TBSOTf. The key Diels-Alder reaction between 13 and ethyl propiolate afforded cyclohexadiene in variable yields from 0–61% (from more than a dozen attempts) with the aromatized benzoate derivative as a major byproduct even though the reaction was rigorously degassed and antioxidants were employed. Notwithstanding the inconsistent yields several grams of 14 were secured and elaborated to cyclohexene derivative (\pm) -7 as described^{16b} in four steps with an overall yield of 24% (average of 70% per step). Conversion to the thionocarbonate (\pm) -6 mediated by N-methylimidazole (NMI) proceeded in 90% yield at room temperature, whereas 4-dimethylaminopyridine (DMAP), a more conventional acylation catalyst resulted in only low conversion (<10%).²⁰ Several conditions were explored to effect a radical coupling with tert-butyl acrylate and other radical traps²¹: however, despite considerable effort, only the deoxygenated product (\pm) -16 was isolated in yields up to 63% yield (84% brsm) likely due to the sterically demanding environment at C-5.

Scheme 1. Attempted introduction of propionate side-chain^a

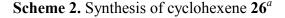


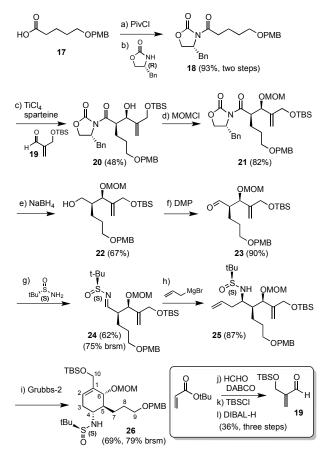
^{*a*}Reagents and conditions: (a) TBSOTf, Et₃N, CH₂Cl₂, -20 °C \rightarrow rt, (b) 2,6-di-*tert*-butyl-4methylphenol (BHT) or methylene blue (0.05 equiv), PhCH₃, 65–80 °C, 6–8 d; (c) N-

methylimidazole (3.5 equiv), CH₂Cl₂, rt, 24 h; (d) representative conditions: (Me₃Si)₃Si (1.05 equiv), *tert*-butyl acrylate (10 equiv), AIBN (0.8 equiv), PhCH₃, reflux, 2 h.

RCM approach. As a result of the capricious yield of the cyclohexadiene 14 coupled with inability to advance intermediate 15, we next focused attention on the complementary RCM route as described below since the propionate side-chain is installed early in the synthesis. Synthesis began with acylation of Evans' (R)-phenylalanine-derived oxazolidinone with the known carboxylic acid 17^{22} under mixed anhydride conditions to afford *N*-acyloxazolidinone 18 in 93% yield. The required aldehyde 19 coupling partner was prepared with a single purification step through Baylis-Hillman reaction²³ of *tert*-butyl acrylate and formaldehyde in aqueous followed by tetrahydrofuran (THF) *tert*-butyldimethylsilyl (TBS) protection and diisobutylaluminum hydride (DIBAL-H) reduction (Scheme 2, bottom insert). Initially, asymmetric boron-mediated aldol reaction²⁴ of **18** with aldehyde **19** using dibutylboron triflate as a Lewis acid in Et₂O provided the syn aldol adduct **20** in high diastereoselectivity (dr > 20:1) but only 36% yield. As an alternative, the titanium enolate of 18 was investigated according to methodology developed by Crimmins and coworkers.²⁵ Treatment of **18** with TiCl₄ and 2.5 equiv of (-)-sparteine in CH₂Cl₂ at -78 °C followed by the addition of aldehyde 19 afforded 20 (dr > 20:1) in an improved 48% yield. Protection as a methoxymethyl (MOM) ether employing CH₃OCH₂Cl and catalytic tetrabutylammonium iodide (TBAI) furnished **21**. The chiral auxiliary was then reductively removed with sodium borohydride²⁶ and Dess-Martin oxidation²⁷ of the resultant alcohol 22 provided aldehyde 23. Condensation with Ellman's (S)-(-)-tertbutanesulfinamide auxiliary yielded tert-butanesulfinyl aldimine 24. As first reported by Xu and

co-workers the combination of pyridinium *p*-toluenesulfonate (PPTS) and anhydrous CuSO₄ dramatically promoted imine formation, which was necessary for the sterically congested aldehyde 23.²⁸ The stereocontrolled installation of the allyl group in 25 was achieved in 87% yield and excellent diastereoselectivity (dr > 20:1) by the addition of allyl magnesium bromide to 24 in CH₂Cl₂ at -78 °C.^{19, 29} Ring-closing metathesis of 25 employing the Grubbs second-generation catalyst³⁰ gave cyclohexene derivative 26 in 95% yield.



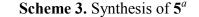


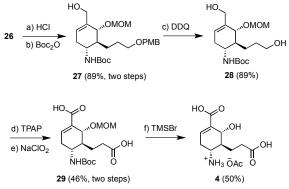
^{*a*}Reagents and conditions: (a) PivCl, Et₃N, THF, 0 °C, 6 h; (b) (*R*)-4-(benzyl)oxazolidin-2-one, *n*-BuLi, -78 °C, 5 min, then added to the mixed anhydride, -78 °C for 30 min, 0 °C for 30 min; (c) TiCl₄, (-)-sparteine, CH₂Cl₂, -78 °C, 1.5 h, then **19** was added, -40 °C for 3 h, -20 °C overnight; (d) CH₃OCH₂Cl, *i*-Pr₂NEt, TBAI, toluene, 90 °C, 7 h; (e) NaBH₄, THF/H₂O (3:1), 0 °C to rt, overnight; (f) Dess-Martin periodinane, NaHCO₃, CH₂Cl₂, rt, 30 min; (g) (*S*)-(–)-2-methyl-2propanesulfinamide, anhydrous CuSO₄, PPTS, CH₂Cl₂, rt, 24 h; (h) allylmagnesium bromide,

CH₂Cl₂, -20 °C, 2.5 h; (i) 14 mol% Grubbs second-generation catalyst, CH₂Cl₂, reflux, 14 h, then DMSO was added, rt, 16 h; (j) HCHO, DABCO, THF–H₂O (1:1), reflux, 16 h; (k) TBSCl, imidazole, CH₂Cl₂, rt, 16 h; (l) DIBAL-H, CH₂Cl₂, -78 °C, 1 h.

The remaining synthetic operations involve oxidation of the primary alcohols at C-9 and C-10, removal of the *tert*-butanesulfinamide auxiliary, and deprotection of the MOM group at C-6 of cyclohexene 26. We initially sought to adjust the oxidation level at C-9 and C-10 and maintain the *tert*-butanesulfinamide as a nitrogen protecting group. However we discovered the tert-butanesulfinamide moiety is readily oxidized under mild oxidation conditions to a sulfonamide,¹⁹ which undergoes facile elimination due to its axial conformation combined with the enhanced acidity of the vicinal allylic proton. Thus, the sulfinamide auxiliary was first removed along with the TBS group by treatment with 4 M HCl in dioxane–MeOH. The resultant amino alcohol was Boc-protected to afford 27 in 89% yield over two steps. Subsequent PMB deprotection with 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (DDQ) gave diol 28. Direct oxidation to the dicarboxylic acid 29 was very low yielding, so we employed a two-stage procedure with tetrapropylammonium perruthenate (TPAP)³¹ to provide an intermediate dialdehyde that was converted to dicarboxylic acid 29 through Pinnick-Lindgren oxidation.³² Deprotection of **29** with trimethylsilyl bromide³³ in CH₂Cl₂ at -78 °C afforded enantiopure target compound 4 as the acetate salt that was purified by silica gel chromatography employing a quaternary solvent system consisting of CHCl₃-MeOH-H₂O-AcOH.







^{*a*}Reagents and conditions: (a) 4 M HCl in 1,4-dioxane, MeOH, 0 °C, 1 h; (b) Boc₂O, Et₃N, 1,4-dioxane/H₂O (2:1), rt, 1 h; (c) DDQ, CH₂Cl₂–H₂O (20:1), rt, 2 h; (d) tetrapropylammonium perruthenate, NMO, 4 Å molecular sieves, CH₂Cl₂, rt, 15 min; (e) NaClO₂, 2-methyl-2-butene, *t*-BuOH–THF–H₂O (5:1:1), rt, 2 h; (f) TMSBr, CH₂Cl₂, -78 °C, 1 h.

The relative configurations of three contiguous stereocenters in **5** were confirmed by the 2D NOESY studies of cyclohexene **28**. As shown in Figure 4, NOE correlation of H-4 with H-7 and H-8 indicates an *anti* relationship between H-4 and H-5. Due to NOE correlation of H-6 with H-7 and H-8 as well as the correlation of H-5 with the CH_2 of the MOM group at C-6, we conclude that H-5 and H-6 also have an *anti* relationship.

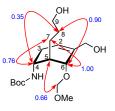


Figure 4. Key NOE correlations of **28**. The integral of the H-6/H-9 cross-peak was used as the internal calibrant (its integral was set to 1.00).

Biological Evaluation. The putative transition-state inhibitor **4** was evaluated for enzyme inhibition against recombinant MbtI under initial-velocity conditions as described,¹³ but showed less than 10% inhibition at 100 μ M. The modest potency of **4** clearly indicates it is a poor TS

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mimic. To rationalize the observed activity, we docked **4** into MbtI using the recently reported co-crystal structure of MbtI with a chorismate analog.³⁴ Introduction of a CH₂ moiety in place of the C-5 oxygen atom of chorismate, led to loss of key hydrogen bond with Arg405 while the protonated C-4 amino group made a potentially repulsive electrostatic interaction with Arg405 (Figure S1).

CONCLUSION

We designed and synthesized an inhibitor based on the hypothetical transition state of the isochorismate partial reaction catalyzed by MbtI wherein the C-4 hydroxyl group of chorismate is protonated by Glu252 resulting in bond cleavage and concomitant C-O bond formation at C-6 due to nucleophilic activation of a water molecule by Lys205. MbtI is a bifunctional enzyme and also catalyzes pyruvate elimination via an intramolecular [3,3]-signatropic reaction. In order to prevent this potential reaction from occurring in our inhibitor, the pyruvate side-chain was replaced with a stable propionate isostere. Two complementary synthetic routes were explored to the target inhibitor 4. The initial route capitalized on the beautiful chemistry developed by Bartlett and Kozlowski for the preparation of a cyclohexene intermediate (\pm) -7. Frustrated by our inability to install the propionate side-chain through a radical-mediated process and the fickle yield in the key Diels-Alder reaction, we undertook a novel synthetic route to enantiopure 4. This second approach featured an asymmetric aldol reaction of a titanium enolate, a diastereoselective Grignard addition to a *tert*-butylsulfinyl aldimine, and ring closing olefin metathesis as key steps. Enzyme inhibition studies revealed 4 is a poor TS mimic and potentially suggests an alternate TS that does not involve charge build-up at C-4 and/or a more prominent role of the C-5 oxygen atom in chorismate for binding. These studies provide a synthetic and mechanistic foundation for future efforts to develop TS-based inhibitors of MbtI.

EXPERIMENTAL SECTION

General Method. All reactions were carried out under a dry Ar atmosphere using oven-dried glassware and magnetic stirring. The solvents were dried before use as follows: THF and Et₂O were heated at reflux over sodium benzophenone ketyl; toluene was heated at reflux over sodium; CH₂Cl₂ was dried over CaH₂. Anhydrous N,N-diisopropylethylamine, triethylamine were used directly as purchased. Commercially available reagents were used without further purification unless otherwise noted. Aluminum TLC sheets (silica gel 60 F₂₅₄) of 0.2-mm thickness were used to monitor the reactions. The spots were visualized with short wavelength UV light or by charring after spraying with a solution prepared from one of the following solutions: phosphomolybdic acid (5.0 g) in 95% EtOH (100 mL); p-anisaldehyde solution (2.5 mL of panisaldehyde, 2 mL of AcOH, and 3.5 mL of conc. H₂SO₄ in 100 mL of 95% EtOH); or ninhydrin solution (0.3g ninhydrin in 100 ml of *n*-butanol; add 3 ml AcOH). Flash chromatography was carried out with silica gel 60 (230-400 ASTM mesh). NMR spectra were obtained on a 400-MHz or 600-MHz spectrometer. Proton chemical data are reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, br = broad), coupling constant, and integration. Chemical shifts were referenced on residual solvent peaks: CDCl₃ (δ = 7.26 ppm for ¹H NMR and 77.00 ppm for ¹³C NMR), CD₂Cl₂ (δ = 5.32 ppm

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for ¹H NMR and 53.84 ppm for ¹³C NMR), CD₃OD ($\delta = 3.31$ ppm for ¹H NMR and 49.00 ppm for ¹³C NMR), D₂O ($\delta = 4.79$ ppm for ¹H NMR). Optical rotations were measured at rt in a 1.0-dm cell. High-resolution mass spectrometry was performed on an linear trap quadrupole (LTQ) mass spectrometer with electrospray ionization and a resolution of 60,000 (at m/z=400).

(*E*)-1-(*tert*-Butyldimethylsilyloxy)-1,3-butadiene (13). To a solution of freshly distilled crotonaldehyde (5.00 mL, 60.3 mmol, 1.00 equiv) in CH₂Cl₂ (70 mL) at -10 °C was added Et₃N (12.0 mL, 36.6 mmol, 1.40 equiv) and the reaction stirred 5 min. Next, TBSOTf (15.0 mL, 65.3 mmol, 1.10 equiv) was slowly added and the reaction warmed to 23 °C over 2 h, then stirred an additional 16 h at 23 °C. The reaction was partitioned between CH₂Cl₂ (75 mL) and saturated aqueous NaHCO₃ (75 mL), dried (MgSO₄) and concentrated under reduced pressure. The title compound was purified by distillation over a Vigreux column (bp 60–62°C/11 mm Hg) to afford the title compound (8.70 g, 78%) as a colorless liquid. The analytical data (¹H, ¹³C NMR and HRMS) matched the reported data for this compounds prepared by an alternate procedure [reported yield 61% using crotonaldehyde (1.0 equiv) TBSCl (1.2 equiv), Et₃N (1.2 equiv), ZnCl₂ (0.013 equiv), hydroquinone (0.02 equiv), benzene, 80 °C, 24 h].^{16b}

(±)-(4*R*,5*S*,6*S*) Ethyl 4-(*tert*-butoxycarbonylamino)-6-(*tert*-butyldimethylsilyloxy)-5-(phenoxythiocarbonyloxy)cyclohex-1-enecarboxylate (15). To a solution of alcohol (±)- 7^{16b} (76.0 mg, 0.180 mmol, 1.00 equiv) in CH₂Cl₂ (5 mL) at 23 °C was added phenyl thionochloroformate (50.0 µL, 0.36 mmol, 2.00 equiv) followed by *N*-methylimidazole (50.0 µL, 0.65 mmol, 3.60 equiv). The reaction mixture was stirred for 16 h at 23 °C then quenched with H₂O (30 mL) and extracted with CH₂Cl₂ (3 × 30 mL). The combined organic extracts were washed with saturated aqueous NaCl (30 mL), dried (Na₂SO₄) and concentrated *in vacuo*. Purification by flash chromatography (5:1 hexane–EtOAc) afforded the title compound (89 mg, 90%) as white solid: ¹H NMR (400 MHz, CDCl₃) δ 0.16 (s, 3H), 0.30 (s, 3H), 0.90 (s, 9H), 1.33 (t, J = 7.1 Hz, 3H), 1.42 (s, 9H), 2.44–2.69 (m, 2H), 4.12–4.25 (m, 1H), 4.25–4.38 (m, 2H), 4.83 (s, 1H), 5.49–5.57 (m, 1H), 6.24 (d, J = 9.1 Hz, 1H), 7.04–7.11 (m, 3H), 7.27–7.32 (m, 1H), 7.37–7.44 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ –5.1, –4.5, 14.3, 18.0, 25.8, 28.3, 29.6, 43.9, 60.9, 63.7, 79.0, 79.5, 121.7, 126.7, 129.55, 129.64, 138.8, 153.3, 154.9, 165.9, 193.6; HRMS (ESI+) calcd for C₂₇H₄₁NNaO₇SSi⁺[M + Na]⁺ 574.2265, found 574.2266 (error 0.2 ppm).

(±)-(4R.6R) Ethyl 4-(tert-butoxycarbonylamino)-6-(tert-butyldimethylsilyloxy)cyclohex-1-enecarboxylate (16). To a solution of ester (\pm) -15 (44.0 mg, 0.080 mmol, 1.00 equiv) in refluxing toluene (4 mL) was added dropwise a solution of (Me₃Si)₃SiH (40 µL, 0.12 mmol, 1.55 equiv), AIBN (7 mg, 0.04 mmol, 0.50 equiv) and tert-butyl acrylate (20 µL, 0.12 mmol, 1.55 equiv) in toluene (2 mL). The reaction mixture was heated at reflux for 2 h. After cooling to room temperature, the solvent was removed *in vacuo*. Purification by flash chromatography (5:1 hexane–EtOAc) afforded the title compound (20 mg, 63%) as white solid: ¹H NMR (400 MHz, $CDCl_3$) $\delta 0.10$ (s, 3H), 0.18 (s, 3H), 0.88 (s, 9H), 1.31 (t, J = 7.1 Hz, 3H), 1.40 (s, 9H), 1.73 (dt, J= 14.3, 3.8 Hz, 1H), 2.01–2.09 (m, 1H), 2.27–2.39 (m, 1H), 2.55 (dd, J = 20.0, 5.1 Hz, 1H), 4.07 (q, J = 4.4 Hz, 1H), 4.10-4.20 (m, 1H), 4.21-4.32 (m, 1H), 4.75-4.81 (m, 1H), 6.55 (d, J = 8.1) ^{13}C 6.91-6.98 NMR (100)CDCl₃) Hz. 1H). 1H): MHz. (m.

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 δ -5.0, -4.8, 14.3, 18.0, 25.8, 28.4, 32.6, 33.9, 42.0, 60.6, 62.7, 78.6, 132.0, 138.8, 155.3, 166.3; HRMS (ESI+) calcd for C₂₀H₃₇NNaO₅Si⁺ [M + Na]⁺ 422.2333, found 422.2337 (error 0.9 ppm).

(R)-3-[5-(4-Methoxybenzyloxy)pentanoyl]-4-benzyloxazolidin-2-one (18). To a solution of acid 17²² (10.66 g, 44.7 mmol, 1.15 equiv) in THF (500 mL) at -78 °C was added Et₃N (10.8 mL, 77.2 mmol, 2.00 equiv) followed by pivalovl chloride (6.00 mL, 49.2 mmol, 1.15 equiv) and the reaction was warmed to 0 °C. The resulting thick white precipitate was stirred for 6 h at 0 °C and then re-cooled to -78 °C. In a separate flask, n-BuLi (2.5 M in hexane, 17.0 mL, 42.5 mmol, 1.10 equiv) was added by syringe over 5 min to a solution of (R)-4-benzyl-2-oxazolidinone (6.84 g, 38.6 mmol, 1.00 equiv) in THF (100 mL) at -78 °C. The oxazolidinone solution was transferred by cannula to the flask containing the mixed anhydride. The mixture was stirred for 30 min at -78 °C and 30 min at 0 °C, then guenched with saturated aqueous NH₄CI (200 mL). THF was removed under reduced pressure and the aqueous layer was extracted with CH_2Cl_2 (3 × 200 mL). The combined organic layers were washed with 10% aqueous NaOH solution, dried (MgSO₄), and concentrated. The crude residue was purified by flash chromatography $(3:1\rightarrow2:1\rightarrow3:2 \text{ hexanes-EtOAc})$ to afford the title compound (14.2 g, 93%) as a colorless oil: $[\alpha]_{p}^{23}$ -37.8 (c 3.20, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 1.67–1.75 (m, 2H), 1.76–1.85 (m, 2H), 2.75 (dd, J = 13.4, 9.6 Hz, 1H), 2.89–3.05 (m, 2H), 3.29 (dd, J = 13.3, 3.2 Hz, 1H), 3.51 (t, J = 6.2 Hz, 2H), 3.80 (s, 3H), 4.14–4.19 (m, 2H), 4.45 (s, 2H), 4.66 (dddd, J = 13.3, 10.1, 7.1, 3.5Hz, 1H), 6.87–6.92 (m, 2H), 7.19–7.23 (m, 2H), 7.25–7.37 (m, 5H); ¹³C NMR (100 MHz, CDCl₃) δ 21.0, 29.0, 35.1, 37.8, 55.0, 55.2, 66.1, 69.6, 72.5, 113.7, 127.2, 128.9, 129.2, 129.3, 130.6,

135.3, 153.4, 159.0, 173.0; HRMS (ESI+) calcd for $C_{23}H_{27}NNaO_5^+$ [M + Na]⁺ 420.1781, found 420.1786 (error 1.2 ppm).

2-(*tert*-Butyldimethylsilyloxymethyl)acrolein (19). To a solution of *tert*-butyl 2-(hydroxymethyl)acrylate prepared according to the reported procedure^{23, 35} (10.0 g, 63.2 mmol, 1.00 equiv) in CH₂Cl₂ (500 mL) at 0 °C was added imidazole (11.1 g, 165 mmol, 2.61 equiv) followed by *tert*-butyldimethylsilyl chloride (19.1 g, 126 mmol, 2.00 equiv). The reaction mixture was stirred at 23 °C overnight. After 16 h, the solvent was removed *in vacuo* to provide a colorless oil, which was dissolved in 10:1 hexane–EtOAc (220 mL). The solution was passed through a short pad of silica gel, which was washed with hexane–EtOAc (10:1). The filtrate was concentrated *in vacuo* and dried under high vacuum to afford a colorless oil, which was then used directly in the next step without further purification.

To the solution of the crude *tert*-butyl 2-(*tert*-butyldimethylsilyloxymethyl)acrylate prepared above in CH₂Cl₂ (300 mL) at -78 °C was added diisobutylaluminum hydride (100 mL, 1.0 M in hexane, 100 mmol) dropwise. The reaction mixture was stirred for 1 h at -78 °C, then diluted with Et₂O (300 mL), and allowed to slowly warm to 23 °C. While the reaction mixture slowly warmed to room temperature, the reaction mixture was quenched by the sequential dropwise addition of H₂O (4 mL), 15% aqueous NaOH solution (4 mL), and H₂O (10 mL). After warming to 23 °C, the reaction mixture was vigorously stirred for 30 min, then treated with anhydrous MgSO₄, and stirred for an additional 15 min. The mixture was filtered through a pad of Celite and the resulting filtrate was concentrated *in vacuo* to afford the title compound (6.00 g, 50%,

 two steps) as a colorless oil, whose ¹H and ¹³C NMR agreed with the reported data for **19** prepared by an alternate synthetic route.³⁶

(R)-4-Benzyl-3-[(2R,3R)-4-(tert-butyldimethylsilyloxy)methyl-2-{3-[(4-

methoxybenzyl)oxy|propyl}-3-(methoxymethoxy)pent-4-enoyl]oxazolidin-2-one (21). To a solution of *N*-acyloxazolidinone 18 (3.00 g, 7.50 mmol, 1.00 equiv) in CH₂Cl₂ (100 mL) at - 78 °C was added a 1.0 M solution of TiCl₄ in CH₂Cl₂ (8.50 mL, 8.50 mmol, 1.13 equiv) dropwise and the solution was stirred for 10 min. (–)-Sparteine (4.30 mL, 18.8 mmol, 2.50 equiv) was added dropwise to the mixture and the solution stirred at -78 °C for 1.5 h. Freshly distilled aldehyde 19 (3.00 g, 15.0 mmol, 2.00 equiv) was then added dropwise. The reaction was stirred for 1.5 h at -78 °C, 3 h at -40 °C, and 12 h at -20 °C. The reaction was quenched at -40 °C by addition of half-saturated aqueous NH₄Cl (100 mL) and quickly transferred to a separatory funnel. The organic layer was separated and the aqueous layer extracted with CH₂Cl₂ (2 × 100 mL). The combined organic layers were dried (MgSO₄), filtered and concentrated. Purification of the crude residue by flash chromatography (10:1 \rightarrow 8:1 \rightarrow 7:1 \rightarrow 6:1 PhMe–EtOAc) provided aldol adduct 20 (2.13 g, 48%) as a colorless oil, which was directly carried onto the next step.

Chloromethyl methyl ether (1.69 g, 21.0 mmol, 5.90 equiv) was added to a solution of aldol adduct **20** (2.13 g, 3.56 mmol, 1.0 equiv), *i*-Pr₂NEt (3.66 mL, 21.0 mmol, 5.90 equiv), and Bu₄NI (131 mg, 0.36 mmol, 0.10 equiv) in PhMe (50 mL) and the mixture stirred at 90 °C for 7 h. The reaction mixture was cooled to rt and treated with saturated aqueous NaHCO₃ (30 mL). The layers were separated and the aqueous layer was extracted with EtOAc (3×50 mL). The combined organic extracts were washed with saturated aqueous NaCl (30 mL), dried (Na₂SO₄)

and concentrated. Purification by flash chromatography (4:1 \rightarrow 3:1 hexane–EtOAc) afforded the title compound (1.88 g, 82%) as colorless oil: $[\alpha]_D^{23}$ +13.5 (*c* 1.10, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 0.00 (s, 6H), 0.84 (s, 9H), 1.57–1.67 (m, 2H), 1.82–1.92 (m, 2H), 2.55 (dd, *J* = 13.2, 10.2 Hz, 1H), 3.22 (dd, *J* = 13.2, 3.0 Hz, 1H), 3.27 (s, 3H), 3.37–3.42 (m, 2H), 3.70 (s, 3H), 3.97–4.13 (m, 4H), 4.16–4.23 (m, 1H), 4.26 (d, *J* = 7.9 Hz, 1H), 4.35 (s, 2H), 4.41 (d, *J* = 6.8 Hz, 1H), 4.47 (dddd, *J* = 12.9, 9.8, 6.4, 3.0 Hz, 1H), 4.55 (d, *J* = 6.8 Hz, 1H), 5.09 (s, 1H), 5.29 (s, 1H), 6.76–6.81 (m, 2H), 7.10–7.15 (m, 2H), 7.15-7.27 (m, 5H); ¹³C NMR (100 MHz, CDCl₃) δ - 5.6, 18.2, 25.0, 25.7, 27.0, 37.7, 46.4, 55.0, 55.7, 55.8, 62.1, 65.8, 69.7, 72.3, 77.9, 94.2, 112.8, 113.5, 127.1, 128.8, 129.1, 129.2, 130.5, 135.3, 145.8, 152.9, 158.9, 173.3; HRMS (ESI+) calcd for C₃₅H₅₁NNaO₈Si⁺ [M + Na]⁺ 664.3276, found 664.3283 (error 1.05 ppm).

(2S,3R)-4-{[(tert-Butyldimethylsilyl)oxy]methyl}-2-{3-[(4-methoxybenzyl)oxy]propyl}-3-

(methoxymethoxy)pent-4-en-1-ol (22). Sodium borohydride (450 mg, 11.7 mmol, 4.00 equiv) was added in one portion to a solution of oxazolidinone 21 (1.88 g, 2.93 mmol, 1.00 equiv) in 3:1 THF–H₂O (60 mL) at 0 °C. The reaction mixture was allowed to warm to 23 °C and stirred overnight (~16 h). The reaction was quenched by the slow addition of saturated aqueous NH₄Cl (60 mL) and extracted with diethyl ether (3 × 50 mL). The combined organic extracts were washed with saturated aqueous NaCl, dried (MgSO₄), and concentrated under reduced pressure. Purification by flash chromatography (3:1→2:1→3:2 hexane–EtOAc) afforded the title compound (920 mg, 67%) as a colorless oil: $[\alpha]_D^{23}$ +60.7 (*c* 0.400, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 0.00 (s, 6H), 0.84 (s, 9H), 1.22–1.34 (m, 1H), 1.43–1.59 (m, 2H), 1.59–1.71 (m, 2H), 2.67 (br s, 1H), 3.35 (s, 3H), 3.41 (t, *J* = 6.3 Hz, 2H), 3.58–3.63 (m, 2H), 3.76 (s, 3H), 4.09 (s,

2H), 4.19 (d, J = 6.1 Hz, 1H), 4.39 (s, 2H), 4.48 (d, J = 6.5 Hz, 1H), 4.58 (d, J = 6.5 Hz, 1H), 5.12 (s, 1H), 5.32 (s, 1H), 6.83 (d, J = 8.5 Hz, 2H), 7.22 (d, J = 8.5 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ -5.58, -5.56, 18.2, 22.7, 25.8, 27.3, 43.1, 55.0, 55.8, 62.4, 63.2, 70.2, 72.4, 78.5, 94.6, 112.9, 113.6, 129.0, 130.5, 146.1, 159.0; HRMS (ESI+) calcd for C₂₅H₄₄NaO₆Si⁺ [M + Na]⁺ 491.2799, found 491.2806 (error 1.4 ppm).

(2S,3R)-4-{[(tert-Butyldimethylsilyl)oxy]methyl}-2-{3-[(4-methoxybenzyl)oxy]propyl}-3-(methoxymethoxy)pent-4-enal (23). To a solution of alcohol 22 (920 mg, 1.96 mmol, 1.00 equiv) in CH₂Cl₂ (20 mL) at 23 °C was added solid NaHCO₃ (1.65 g, 19.6 mmol, 10.0 equiv) followed by Dess-Martin periodinane (1.25 g, 2.94 mmol, 1.50 equiv). The reaction was stirred for 30 min at 23 °C then guenched by 1:1 10% aqueous Na₂S₂O₃-saturated aqueous NaHCO₃ (20 mL). The layers were separated and the aqueous layer was extracted with CH₂Cl₂ (3×20 ml). The combined organic layers were dried (MgSO₄) and concentrated under reduced pressure. The resulting residue was dissolved in a minimum amount of 3:1 hexane-EtOAc and filtered through a short pad of silica gel, which was rinsed with 3:1 hexane–EtOAc (200 mL). The filtrate was concentrated to afford the title compound (821 mg, 90%) as colorless oil: ¹H NMR (400 MHz, $CDCl_3$) δ 0.00 (s, 6H), 0.84 (s, 9H), 1.43–1.54 (m, 1H), 1.57–1.79 (m, 3H), 2.50 (dddd, J = 11.5, 8.9, 5.6, 2.7 Hz, 1H), 3.26 (s, 3H), 3.36 (t, J = 6.0 Hz, 2H), 3.73 (s, 3H), 4.05 (s, 2H), 4.34 (s, 2H), 4.41 (d, J = 6.7 Hz, 1H), 4.44 (d, J = 5.9 Hz, 1H), 4.56 (d, J = 6.8 Hz, 1H), 5.08 (s, 1H), 5.27 (s, 1H), 6.80 (d, J = 8.6 Hz, 2H), 7.19 (d, J = 8.6 Hz, 2H), 9.61 (d, J = 2.2 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ -5.54, -5.52, 18.2, 21.2, 25.8, 27.6, 54.6, 55.1, 55.9, 63.2, 69.7, 72.4, 75.8, 94.1, 113.6, 114.0, 129.1, 130.5, 144.8, 159.0, 203.1; HRMS (ESI+) calcd for $C_{25}H_{42}NaO_6Si^+[M + Na]^+ 489.2643$, found 489.2647 (error 0.8 ppm).

(S,E)-N-[(2S,3R)-4-{[(tert-Butyldimethylsilyl)oxy]methyl}-2-{3-[(4-

methoxybenzyl)oxy]propyl}-3-(methoxymethoxy)pent-4-en-1-ylidene]-tert-butylsulfinamide

(24). To a solution of aldehyde 23 (820 mg, 1.76 mmol, 1.00 equiv) in CH₂Cl₂ (10 mL) was added (S)-(-)-tert-butylsulfinamide (235 mg, 1.94 mmol, 1.10 equiv), anhydrous CuSO₄ (562 mg, 3.52 mmol, 2.00 equiv) and pyridinium *p*-toluenesulfonate (442 mg, 1.76 mmol, 1.00 equiv). The mixture was stirred at 23 °C for 24 h, then filtered through a pad of Celite, and the filter cake was washed with CH₂Cl₂. The combined organic filtrate was washed with saturated aqueous NaCl, dried (MgSO₄), and concentrated. Purification by flash chromatography $(3:1\rightarrow 2:1)$ hexane–EtOAc) afforded the title compound (616 mg, 62%, 75% brsm) as a colorless oil: $[\alpha]_D^{23}$ +165 (c 0.700, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 0.00 (s, 6H), 0.84 (s, 9H), 1.13 (s, 9H), 1.42-1.69 (m, 3H), 1.80-1.91 (m, 1H), 2.73-2.83 (m, 1H), 3.30 (s, 3H), 3.36 (t, J = 6.1 Hz, 2H),3.72 (s, 3H), 4.06 (s, 2H), 4.24 (d, J = 7.1 Hz, 1H), 4.34 (s, 2H), 4.42 (d, J = 6.7 Hz, 1H), 4.58 (d, J = 6.7 Hz, 1H), 5.06 (s, 1H), 5.25 (s, 1H), 6.80 (d, J = 8.3 Hz, 2H), 7.17 (d, J = 8.3 Hz, 2H), 7.87 (d, J = 6.8 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ -5.71, -5.69, 18.0, 22.2, 24.2, 25.7, 27.2, 48.7, 54.9, 55.8, 56.6, 62.3, 69.4, 72.2, 77.8, 93.7, 113.5, 114.1, 128.9, 130.4, 144.6, 158.9, 169.8; HRMS (ESI+) calcd for $C_{29}H_{51}NNaO_6SSi^+$ [M + Na]⁺ 592.3099, found 592.3105 (error 1.0 ppm).

(S)-N-[(4R,5S,6R)-7-{[(tert-Butyldimethylsilyl)oxy]methyl}-5-{3-[(4-

methoxybenzyl)oxy]propyl}-6-(methoxymethoxy)octa-1,7-dien-4-yl]-tert-butylsulfinamide

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(25). To a solution of *tert*-butanesulfinyl imine 24 (600 mg, 1.05 mmol, 1.00 equiv) in CH₂Cl₂ (20 mL) was added allylmagnesium bromide (1.0 M in Et₂O, 3.15 mL, 3.15 mmol, 3.00 equiv) at -78 °C. The reaction was warmed to -20 °C and stirred for 2.5 h at this temperature then quenched with saturated aqueous NH₄Cl (20 mL). The layers were separated and the aqueous layer was extracted with CH_2Cl_2 (3 × 20 ml). The combined organic layers were dried (MgSO₄), filtered, and concentrated. The resulting residue was dissolved in a minimum amount of Et₂O and filtered through a short pad of silica gel, which was rinsed with Et₂O. The filtrate was concentrated to provide the title compound (560 mg, 87%) as colorless oil: $[\alpha]_D^{23}$ +80.1 (*c* 0.600, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 0.02 (s, 6H), 0.87 (s, 9H), 1.16 (s, 9H), 1.37–1.51 (m, 2H), 1.55-1.79 (m, 3H), 2.35-2.45 (m, 1H), 2.49-2.59 (m, 1H), 3.22 (d, J = 8.3 Hz, 1H), 3.28 (s, 3H), 3.35-3.38 (m, 3H), 3.74 (s, 3H), 3.99 (d, J = 15.0 Hz, 1H), 4.05 (d, J = 14.6 Hz, 1H), 4.16(d, J = 7.3 Hz, 1H), 4.36 (s, 2H), 4.39 (d, J = 6.7 Hz, 1H), 4.56 (d, J = 6.7 Hz, 1H), 5.05-5.10 (m, 2000)3H), 5.34 (s, 1H), 5.66–5.80 (m, 1H), 6.81 (d, J = 8.2 Hz, 1H), 7.19 (d, J = 8.2 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ -5.55, -5.52, 18.2, 22.6, 22.8, 25.7, 29.5, 38.5, 42.7, 55.0, 55.9, 56.0, 56.5, 62.1, 70.0, 72.3, 79.0, 93.9, 113.5, 113.9, 118.6, 129.0, 130.5, 134.5, 145.3, 158.9; HRMS (ESI+) calcd for $C_{32}H_{58}NO_6SSi^+$ [M + H]⁺ 612.3749, found 612.3754 (error 0.8 ppm).

(S)-N-[(1R,5R,6S)-4-{[(tert-Butyldimethylsilyl)oxy]methyl}-6-{3-[(4-

methoxybenzyl)oxy]propyl}-5-(methoxymethoxy)cyclohex-3-en-1-yl]-tert-butylsulfinamide(26).[1,3-bis-(2,4,6-Trimethylphenyl)-2-imidazolidinylidene]dichloro(phenylmethylene)(tricyclohexylphosphine)ruthenium(13.8 mg,0.016 mmol, 0.14 equiv) was added to a solution of diene 25 (70.0 mg, 0.114 mmol, 1.0 equiv)

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in CH₂Cl₂ (20 mL) and the reaction was heated at reflux for 14 h. The reaction was cooled to 23 °C, then treated with DMSO (1.5 mL),³⁷ and stirred 16 h at 23 °C. The reaction mixture was washed with saturated aqueous NaCl (50 mL), and the organic layer was dried (MgSO₄), filtered and concentrated. Purification by flash chromatography (2:1 \rightarrow 3:2 \rightarrow 1:1 hexane–EtOAc) afforded the title compound (63 mg, 95%) as a light brown oil: ¹H NMR (400 MHz, CDCl₃) δ - 0.01 (s, 3H), 0.00 (s, 3H), 0.85 (s, 9H), 1.11 (s, 9H), 1.16–1.29 (m, 2H), 1.54–1.70 (m, 2H), 1.99–2.07 (m, 1H), 2.23–2.34 (m, 1H), 2.48–2.59 (m, 1H), 3.26–3.46 (m, 6H), 3.73 (s, 3H), 3.92 (s, 1H), 4.02 (d, *J* = 12.8 Hz, 1H), 4.14 (d, *J* = 12.8 Hz, 1H), 4.35 (s, 2H), 4.54 (d, *J* = 6.6 Hz, 1H), 4.60–4.69 (m, 2H), 5.65 (s, 1H), 6.80 (d, *J* = 8.2 Hz, 2H), 7.17 (d, *J* = 8.2 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ -5.6, -5.4, 18.0, 22.5, 25.7, 26.5, 27.5, 29.2, 40.9, 53.5, 55.0, 55.5, 55.6, 64.3, 69.5, 71.6, 72.4, 95.3, 113.5, 121.5, 129.0, 130.3, 135.0, 158.9; HRMS (ESI+) calcd for C₃₀H₅₃NNaO₆SSi⁺ [M + Na]⁺ 606.3255, found 606.3252 (error 0.5 ppm).

tert-Butyl [(1*R*,5*R*,6*S*)-4-(hydroxymethyl)-6-{3-[(4-methoxybenzyl)oxy]propyl}-5-(methoxymethoxy)cyclohex-3-en-1-yl]carbamate (27). To a solution of 26 (55 mg, 0.094 mmol, 1.0 equiv) in MeOH (2.0 mL) was added dropwise 4 M HCl in 1,4-dioxane (190 μ L, 0.75 mmol, 8.0 equiv) at 0 °C. The resulting mixture was stirred at 0 °C for 1 h then quenched with saturated aqueous NaHCO₃ (3 mL). Methanol was removed under reduced pressure and the aqueous layer was treated with CH₂Cl₂ (2 mL) and 6 M aqueous NaOH (2 mL) at 0 °C. After stirring for 10 min at 0 °C, the layers were separated. The aqueous layer was extracted with CH₂Cl₂ (3 × 3 mL) and the combined organic layers were dried (K₂CO₃) and concentrated under

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reduced pressure to afford the crude amino alcohol as a viscous yellow oil, which was used directly in the next step without further purification.

To the crude amino alcohol in 1:2 H₂O-1,4-dioxane (3 mL) at 23 °C was added Et₃N (26 µL, 0.19 mmol, 2.0 equiv) followed by di-tert-butyl dicarbonate (28 mg, 0.13 mmol, 1.4 equiv) in 1,4-dioxane (0.5 mL). The reaction mixture was stirred for 1 h at 23 °C, then partitioned between H₂O (10 mL) and CH₂Cl₂ (10 mL). The aqueous layer was extracted with CH₂Cl₂ (3×3 mL) and the combined organic layers were dried (MgSO₄), filtered, and concentrated. Purification by flash chromatography (1:1 hexane-EtOAc) afforded the title compound (39 mg, 89%) as a colorless oil: ¹H NMR (400 MHz, CDCl₃) δ 1.21–1.31 (m, 2H), 1.42 (s, 9H), 1.63– 1.74 (m, 2H), 1.96–2.11 (m, 2H), 2.19–2.26 (m, 2H), 3.37–3.46 (m, 5H), 3.80 (s, 3H), 3.82–3.89 (m, 1H), 3.96 (s, 1H), 4.09 (d, J = 12.0 Hz, 1H), 4.18 (d, J = 12.7 Hz, 1H), 4.40 (s, 2H), 4.64 (d, J = 6.7 Hz, 1H), 4.74 (d, J = 6.7 Hz, 1H), 5.76 (br s, 1H), 5.93 (d, J = 8.1 Hz, 1H), 6.87 (d, J = 6.7 Hz 8.5 Hz, 2H), 7.23 (d, J = 8.5 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 27.0, 27.68, 27.72, 28.4, 40.0, 46.3, 55.2, 56.1, 65.4, 69.7, 72.5, 74.0, 78.8, 96.2, 113.8, 124.6, 129.2, 130.5, 134.8, 155.4, 159.1; HRMS (ESI+) calcd for $C_{25}H_{39}NNaO_7^+$ [M + Na]⁺ 488.2619, found 488.2621 (error 0.4 ppm).

tert-Butyl

[(1*R*,5*R*,6*S*)-4-(hydroxymethyl)-6-(3-hydroxypropyl)-5-

(methoxymethoxy)cyclohex-3-en-1-yl]carbamate (28). To a solution of 27 (40 mg, 0.086 mmol, 1.0 equiv) in 20:1 CH₂Cl₂–H₂O (10 mL) was added DDQ (29 mg, 0.13 mmol, 1.5 equiv) at 0 °C. The reaction was stirred for 2 h at 23 °C, then partitioned between saturated aqueous NaHCO₃ (10 mL) and CH₂Cl₂ (10 mL). The aqueous layer was extracted with CH₂Cl₂ (2 × 10

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mL) and the combined organic layers were dried (Na₂SO₄), filtered, and concentrated. Purification by flash chromatography (Et₂O \rightarrow EtOAc) afforded the title compound (23 mg, 89%) as a colorless oil: ¹H NMR (400 MHz, CDCl₃) δ 1.21–1.31 (m, 2H), 1.42 (s, 9H), 1.58–1.72 (m, 2H), 2.04–2.12 (m, 1H), 2.20–2.26 (m, 2H), 2.30–2.40 (m, 2H), 3.43 (s, 3H), 3.55–3.68 (m, 2H), 3.80–3.89 (m, 1H), 3.97–4.01 (m, 1H), 4.07 (d, *J* = 12.4 Hz, 1H), 4.18 (d, *J* = 12.6 Hz, 1H), 4.67 (d, *J* = 6.8 Hz, 1H), 4.77 (d, *J* = 6.8 Hz, 1H), 5.72–5.79 (m, 1H), 5.93 (d, *J* = 7.5 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 26.6, 27.8, 28.4, 30.4, 39.9, 46.4, 56.1, 62.4, 65.1, 73.7, 78.9, 96.2, 124.7, 134.9, 155.5; HRMS (ESI+) calcd for C₁₇H₃₁NNaO₆⁺ [M + Na]⁺ 368.2044, found 368.2046 (error 0.5 ppm).

(4R,5S,6R)-4-tert-(Butylcarbonyl)amino-5-(2-carboxyethyl)-6-

(methoxymethoxy)cyclohex-1-enecarboxylic acid (29). To a solution of diol 28 (23 mg, 0.076 mmol, 1.0 equiv), *N*-methylmorpholine-*N*-oxide (53 mg, 0.45 mmol, 6.0 equiv), and powdered 4 Å molecular sieves (35 mg) in CH_2Cl_2 (1 mL) at 23 °C was added tetrapropylammonium perruthenate (2.0 mg, 0.0056 mmol, 0.075 equiv). The mixture was stirred for 15 min at 23 °C, then filtered through a short pad of silica gel, eluting with EtOAc. The filtrate was concentrated under reduced pressure to afford the crude dialdehyde, which was directly used in the next step without further purification.

To a solution of freshly prepared dialdehyde and 2-methyl-2-butene (0.20 mL, 1.9 mmol, 25 equiv) in 5:1:1 *tert*-butyl alcohol–THF–H₂O (1 mL) at 0 °C was slowly added a solution of sodium chlorite (80% w/w technical grade, 52 mg, 0.46 mmol, 6.0 equiv) and sodium phosphate monobasic monohydrate (63 mg, 0.46 mmol, 6.0 equiv) in H₂O (0.3 mL). The resulting

suspension was stirred at 23 °C for 2 h. The reaction was quenched with saturated aqueous NaHSO₃ (3 mL) at 0 °C and the aqueous layer was extracted with CH₂Cl₂ (3 × 5 mL). The combined organic layers were dried (Na₂SO₄), filtered, and concentrated under reduced pressure. Purification by flash chromatography on silica gel³⁸ (210:25:4 \rightarrow 180:25:4 CHCl₃–MeOH–AcOH) afforded the title compound (13 mg, 46% for two steps) as a colorless oil that was ~90% pure: ¹H NMR (400 MHz, CDCl₃) δ 1.41 (s, 9H), 1.44–1.54 (m, 2H), 2.18–2.25 (m, 1H), 2.39–2.60 (m, 4H), 3.40 (s, 3H), 3.88–3.95 (m, 1H), 4.31–4.34 (m, 1H), 4.72 (d, *J* = 6.9 Hz, 1H), 4.86 (d, *J* = 6.9 Hz, 1H), 6.29 (d, *J* = 8.1 Hz, 1H), 7.14–7.18 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 25.0, 28.4, 29.0, 31.8, 40.1, 45.4, 56.1, 71.8, 79.3, 97.5, 128.0, 142.4, 155.5, 171.1, 178.2; HRMS (ESI–) calcd for C₁₇H₂₆NO₈⁻ [M – H]⁻ 372.1664, found 372.1668 (error 1.1 ppm).

(4*R*,5*S*,6*R*)-4-Amino-5-(2-carboxyethyl)-6-hydroxycyclohex-1-enecarboxylic acid acetate salt (4). To a solution of dicarboxylic acid 29 (13 mg, 0.035 mmol, 1.0 equiv) in CH₂Cl₂ (1 mL) was added TMSBr (40 μ L, 0.30 mmol, 8.6 equiv) at -78 °C. The reaction mixture was stirred for 1 h at -78 ° then concentrated *in vacuo* during which time the flask warmed to 23 °C. The residue was washed with Et₂O, and purified by flash chromatography on silica gel³⁸ (65:25:4:2 \rightarrow 100:56:8:8 CHCl₃–MeOH–H₂O–AcOH) to afford the acetate salt of the title compound (5.0 mg, 50%) as a white solid: [α]_D²³ +6.2 (*c* 0.40, H₂O); ¹H NMR (600 MHz, D₂O) δ 1.57–1.64 (m, 2H), 2.22–2.29 (m, 1H), 2.46 (t, *J* = 7.4 Hz, 2H), 2.56 (d, *J* = 13.5 Hz, 1H), 2.78 (d, *J* = 13.5 Hz, 1H), 3.68–3.74 (m, 1H), 4.55–4.59 (m, 1H), 6.84–6.88 (m, 1H); ¹³C NMR (150 MHz, D₂O) δ 21.1 (AcOH), 24.0, 65.0, 32.7, 40.7, 46.9, 65.1, 131.8*, 134.3, 172.4*, 177.4*

(AcOH), 179.6* (Chemical shifts denoted by * are derived from HMBC); HRMS (ESI–) calcd for $C_{10}H_{14}NO_5^{-}$ [M – H]⁻ 228.0877, found 228.0883 (error 2.6 ppm).

MbtI Assay. Reactions were performed under initial velocity conditions in a total volume of 50 μ L at 37 °C for 30 min and the production of salicylic acid was monitored continuously by following changes in fluorescence at 420 nm with excitation at 305 nm on a microplate reader. Assays were set up in duplicate and contained 0.5 μ M MbtI in reaction buffer (100 mM Tris-HCl, pH 8.0, 1 mM MgCl₂, 50 μ M chorismate, and 0.0025% Igepal CA-630). A three-fold serial dilution of inhibitor in H₂O was added to black 384 well plates coated with a non-binding surface (Greiner). A positive control (H₂O only) and negative control (10 mM EDTA) were also included. The IC₅₀ values were calculated from the Hill equation (eq 1). In this equation the fractional activity (v_i/v_0) versus inhibitor concentration was fit by non-linear regression analysis using GraphPad prism version 6.0 where v_i is the reaction velocity at a given [I] and v_0 is the reaction velocity of the DMSO control, and *h* is the Hill slope.

$$\frac{v_i}{v_0} = \frac{1}{1 + ([I]/IC_{50})^h} \tag{1}$$

ASSOCIATED CONTENT

Supporting Information: copies of ¹H NMR and ¹³C NMR spectra of all new compounds and Figure illustrating a docked pose of **4** in MbtI. This material is available free of charge via the Internet at <u>http://pubs.acs.org</u>.

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Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENT

This work was supported by a grant from the National Institutes of Health Grant (RO1 AI070219 to CCA) and the National Natural Science Foundation of China (21402058 to ZL). We thank Dr. Youlin Xia from NMR center of University of Minnesota for NMR spectroscopic analysis. Mass spectrometric analysis was carried out in the Analytical Biochemistry Shared Resource of the Masonic Cancer Center, University of Minnesota, funded in part by Cancer Center Support Grant CA-77598.

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