SYNTHESIS OF THE ABC RING FRAGMENT OF BREVISIN, A NEW DINOFLAGELLATE POLYCYCLIC ETHER

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Abstract – A polycyclic ether, brevisin was isolated from the red-tide dinoflagellate *Karenia brevis*. Its unique skeletal structure consists of two separate tricyclic ether assemblies connected by a methylene bridge. The ABC ring fragment of brevisin was synthesized *via* Suzuki–Miyaura cross coupling toward a total synthesis of brevisin.

A polycyclic ether, brevisin (1)¹ was isolated from the red tide dinoflagellate *Karenia brevis* which simultaneously produces a range of ladder-frame polyethers, including brevetoxins,²⁻⁴ brevenal and its acetal,⁵ and a cyclic ether amide, brevisamide.⁶ The structure of **1** is characterized by two tricyclic ether ring assemblies linked by a methylene group. One of the assemblies contains a conjugated aldehyde side chain which has been found in brevenal and brevisamide. The interrupted polycyclic ether skeleton of the molecule is quite unique. Common structural features for representative marine polycyclic ethers such as the brevetoxins, ciguatoxins⁷ and yessotoxins⁸ are contiguous fused ether ring assemblies containing middle size (from 7 to 9-membered) ether rings at the center of their polycyclic ether skeletons.⁹

This paper is dedicated to Dr. Akira Suzuki on the occasion of his 80th birthday.

Interestingly, even though the skeletal structure of **1** is divided into two ether ring assemblies by the methylene, **1** inhibited the binding of 42-dihydrobrevetoxin B (PbTx-3) to the VSSC.¹ Stereochemical information of **1** is essential in order to elucidate the relationship between its unique structure and biological activity. However, the relative stereochemistry of the ABC and DEF ring assemblies could be only determined separately due to the shortage of natural brevisin which hampered determination of the stereochemical correlation between the ether ring assemblies and the absolute configuration by NMR methods. Therefore we have begun synthesis of **1** to determine the complete stereostructure of this unusual polyether. Including our group, numerous efforts have been directed toward the synthesis of marine polycyclic ethers using various approaches.¹⁰ Our synthetic strategy based on Suzuki–Miyaura cross coupling¹¹ has led to convergent synthesis of ether ring skeletons of polycyclic ethers¹² and the recent successful synthesis of the related metabolite brevisamide.¹³ Herein we report synthesis of the ABC-ring fragment **2** leading toward the total synthesis of **1** *via* Suzuki–Miyaura cross coupling.



Figure 1. Structure of brevisin (1) and its probable stereostructure of the ABC and DEF ring assemblies.



Scheme 1. Retrosynthetic analysis of 2.¹⁴

Based on the structural similarity of the dienal side chain and the 6-membered ether ring in 1, brevenal, and brevisin, we synthesized the ABC-ring fragment with an 8*S* configuration. Our synthetic strategy is summarized in Scheme 1. The A-ring *exo*-olefin **3** was prepared from 3-benzyloxypropan-1-ol (**5**) and the C-ring ketene acetal phosphate **4** was prepared from 2-deoxy-D-ribose by reported procedures.¹⁵ The A-ring *exo*-olefin **3** and the C-ring ketene acetal phosphate **4** could be linked by a Suzuki–Miyaura cross coupling and subsequent B-ring construction by mixed thioacetalization/methylation to yield the ABC-ring fragment **2**.

The synthesis of the A-ring *exo*-olefin **3** began with 3-benzyloxypropan-1-ol (**5**), which was oxidized with TEMPO and PhI(OAc)₂ to generate aldehyde **6**.¹⁶ Optically active homoallylic alcohol **7** was streoselectively prepared from **6** by the following procedures. Reagent-controlled enantioselective crotylation of aldehyde using (*Z*)-2-butene and (+)- β -methoxydiisopinocampheylborane¹⁷ gave the desired alcohol **7** in 56% yield (90% ee). Determination of the absolute configuration of alcohol **7** was confirmed by the modified Mosher method.¹⁸ The alcohol was esterified with acryloyl chloride and *i*-Pr₂NEt (DIPEA) to furnish acrylate **8** in 92% yield. Treatment of **8** with the second-generation Grubbs' catalyst (Grubbs' II)¹⁹ resulted in successful closure of α , β -unsaturated six-membered lactone **9** in 94% yield, and was subsequently oxidized to diol **10** with OsO₄ and NMO in 88% yield. After protection of the diol with TESOTf to give lactone **11**, the stereostructure of this key intermediate was assigned by NOE correlations and ³*J*_{H,H} coupling constants. NOE correlations between the C-9 methyl and H-11, and the C-9 methyl and H-10 suggested an axial orientation of the C-9 methyl and H-11, and an equatorial orientation of H-10. An NOE correlation and small coupling constant (*J*=3 Hz) between H-9 and H-8 confirmed an axial direction of H-8. The A-ring *exo*-olefin **3** was generated by a Petasis–Tebbe reaction with Cp₂TiMe₂ at 65 °C in 92% yield (Scheme 2).²⁰



Scheme 2. Reagents and conditions (a) TEMPO, PhI(OAc)₂, CH₂Cl₂, 94%; (b) (+)-(*Z*)-crotyldiisopinocampheylborane, Et₂O, THF, -78 °C; then 3 M aq NaOH, H₂O₂, reflux, 56%, 90% ee; (c) acryloyl chloride, DIPEA, CH₂Cl₂, 0 °C, 92%; (d) Grubbs' II, CH₂Cl₂, 94%; (e) OsO₄, NMO, *t*BuOH, H₂O, 0 °C, 88%; (f) TESOTf, 2,6-lutidine, CH₂Cl₂, 0 °C, 87%; (g) Cp₂TiMe₂, toluene, THF, 65 °C, 92%.

The known lactone 12 was prepared from 2-deoxy-D-ribose in seven steps.¹⁵ Enolization of this lactone with KHMDS in the presence of (PhO)₂P(O)Cl generated the C-ring ketene acetal phosphate 4 in 96% yield. Stereoselective hydroboration of the A-ring exo-olefin 3 with 9-BBN-H produced an alkylborane, which was reacted in situ with the C-ring ketene acetal phosphate 4 in the presence of aqueous Cs₂CO₃ and a catalytic amount of PdCl₂(dppf), to give rise to the cross coupling product 13 in 66% yield from 4. diastereomer could not be detected because of its trace amount. Stereoselective А hydroboration-oxidation of the coupling product 13 furnished alcohol 14 in 77% yield. An NOE correlation between H-14 and the C-19 methyl unambiguously indicated the stereochemistry of 14 as shown in Scheme 3. Oxidation of the resultant hydroxy group with TPAP/NMO led to ketone 15 in 93% yield. Treatment of 15 with EtSH and Zn(OTf)₂ led to removal of the TES group and formation of mixed thioacetal, and following acetylation yielded mixed thioacetals 16 (29%) and 17 (48%) for the two steps. Introduction of the C-15 methyl group in the triacetylated thioacetal **16** was achieved in a one pot manner with *m*CPBA and then AlMe₃ giving rise to triether ring assembly **2** in 98% yield (Scheme 3).^{21,22} The stereochemistry of 2 was confirmed by NOE correlations H-11/the C-15 methyl and H-12/H-8 and H-14.



Scheme 3. Reagents and conditions (a) KHMDS, (PhO) $_2$ P(O)Cl, HMPA, THF, -78 °C, 96%; (b) 3, 9-BBN-H, THF, 50 °C; then 4, 3 M aq Cs₂CO₃, PdCl₂ (dppf), DMF, 50 °C, 66%; (c) BH₃•SMe₂, 0 °C to rt; then 3 M aq NaOH, H₂O₂, 0 °C to rt, 77%; (d) TPAP, NMO, MS4A, CH₂Cl₂, 93%; (e) Zn(OTf) ₂, EtSH, THF, rt; (f) Ac₂O, pyridine, 29% for 16, 48% for 17 (two steps); (g) *m*CPBA, CH₂Cl₂, 0 °C; then AlMe₃, 0 °C, 98%.

Although the ¹H NMR chemical shifts from H-8 to H-18 of the ABC-ring fragment **2** are slightly different from those of the corresponding domain of tetraacetylated brevisin probably due to the absence of a dienal structure and the DEF ring assembly, the coupling constants of **2** agreed well with those of **1**. In summary, the ABC-ring fragment **2** of brevisin was synthesized in a stereocontrolled manner from commercially available materials, 3-benzyloxypropan-1-ol and 2-deoxy-D-ribose. Olefin metathesis was highly effective for construction of the A-ring. The α , β -unsaturated lactone **9** was constructed in only two steps from homoallylic alcohol **7**. Assembly of the A-ring *exo*-olefin **3** and the C-ring ketene acetal phosphate **4** was accomplished by an effective Suzuki–Miyaura coupling reaction. Further study toward the total synthesis of brevisin is now in progress.

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- Physicochemical data of 2: [α]_D²⁹ –15 (*c* 0.13, CHCl₃); ¹H-NMR (500 MHz, CD₃OD) δ 7.36 (5H, m, Ph), 5.16 (1H, br dd, J=2.1, 5.4 Hz), 4.92 (1H, m), 4.53 (1H, d, J=12.2 Hz), 4.47 (1H, d, J=12.2 Hz), 3.95 (1H, m), 3.94 (1H, d, J=11.4 Hz), 3.89 (1H, d, J=11.4 Hz), 3.65 (1H, dd, J=5.1, 12.2 Hz), 3.57 (1H, m), 3.55 (1H, m), 3.46 (1H, dd, J=2.9, 11.5 Hz), 3.46 (1H, ddd, J=3.8, 13.5, 13.5 Hz), 2.16 (3H, s), 2.08 (3H, s), 2.07 (3H, s), 1.89 (1H, m), 1.87 (1H, m), 1.85 (2H, m), 1.73 (1H, ddd, J=4.2, 9.2, 9.2 Hz), 1.69 (1H, dd, J=4.6, 10.9 Hz), 1.58 (1H, m), 1.51 (1H, m), 1.49 (1H, ddd, J=13, 13, 13 Hz), 1.22 (3H, s), 1.19 (3H, s), 1.01 (3H, d, J=7.6 Hz); ¹³C-NMR (100 MHz, CD₃OD) δ 172.2, 172.1, 171.6, 139.9, 129.3, 129.0, 128.7, 80.3, 79.0, 75.9, 74.9, 73.9, 73.3, 73.1, 72.9, 69.5, 68.7, 67.7, 39.3, 36.4, 35.0, 33.2, 30.8, 24.4, 21.2, 20.8, 18.5, 15.7, 11.3, HRMS (FAB), calcd for C₃₁H₄₄O₁₀Na 599.2827 (M+Na⁺), found 599.2823.