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Design, synthesis and anti-ToCV activity of novel 4(3H)quinazolinone derivatives bearing dithioacetal moiety

Guangcheng Zu, Xiuhai Gan, Dandan Xie, Huanyu Yang, Awei Zhang, Shaoyuan Li, Deyu Hu, and Baoan Song

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2	quinazolinone derivatives bearing dithioacetal moiety				
3	Guangcheng Zu, Xiuhai Gan, Dandan Xie, Huanyu Yang, Awei Zhang, Shaoyuan Li,				
4	Deyu Hu*, Baoan Song*				
5	Current address: State Key Laboratory Breeding Base of Green Pesticide and				
6	Agricultural Bioengineering, Key Laboratory of Green Pesticide and Agricultural				
7	Bioengineering, Ministry of Education, Guizhou University, Huaxi District, Guiyang				
8	550025, China.				
9	*Address of the corresponding author				
10	Fax: 0086-851-83622211; E-mail: <u>dyhu@gzu.edu.cn; songbaoan22@yahoo.com</u> .				

ABSTRACT: Tomato chlorosis virus (ToCV) has caused great harm to the 12 production of tomato worldwide. To develop efficient anti-ToCV agents, some novel 13 4(3H)-quinazolinone derivatives containing dithioacetal were designed and 14 synthesized, and their anti-ToCV activities were evaluated by microscale 15 16 thermophoresis (MST) using ToCV coat protein (ToCV-CP) as a new target. The results showed that some compounds had strong binding capacity to ToCV-CP. In 17 particular, compounds C5 and C22 have excellent binding capacity to ToCV-CP, with 18 binding constant values of 0.24 and 0.25 μ M, respectively. Additionally, reduced 19 20 ToCV-CP gene expression levels of 81.05% and 87.59% could be achieved when tomato was treated with compounds C5 and C22, respectively, which were obviously 21 higher than those levels after Ningnanmycin (NNM) treatment (43.88%) and lead 22 23 compound Xiangcaoliusuobingmi (XCLSBM) treatment (63.56%). Therefore, this work indicates that 4(3H)-quinazolinone derivatives containing dithioacetal moiety 24 can be used as novel anti-ToCV agents. 25

26

KEYWORDS: Tomato chlorosis virus, 4(3H)-quinazolinone, dithioacetal, MST, coat
protein, anti-ToCV activity

30 **INTRODUCTION**

Tomato chlorosis virus (ToCV), a plant RNA virus transmitted by whitefly, was first 31 32 discovered in the United States in 1998, and it belongs to the family *Closteroviridae*, genus Crinivirus.^{1,2} ToCV has a wide range of hosts; it not only infects tomato, 33 pepper, potato and other Solanaceae plants but also infects 25 other plant species 34 from eight different families. After being infected with ToCV, the tomato mainly 35 exhibited developmental delay, leaf brittleness, interveinal chlorosis, and limited 36 necrotic flecking.³⁻⁶ In recent years, ToCV has occurred in more than 20 countries 37 around the world and caused enormous economic losses.⁷⁻¹¹ Currently, effective 38 agents to control this plant virus disease are still lacking, resulting in a serious decline 39 in tomato quality and yield. Therefore, the development of efficient anti-ToCV agents 40 41 is urgently needed.

The genome of ToCV consists of two genomic positive-sense RNAs, RNA1 and 42 RNA2, with a total genome length of 16.8 kb,^{12,13} encoding four and nine open 43 reading frames (ORFs)^{14,15}. RNA2 encodes multiple functional proteins, of which, the 44 coat protein (CP) and the minor coat protein (mCP) are mainly involved in virus 45 assembly, coating, and transporting. Coat proteins are essential for the intercellular 46 translocation of this type virus.¹⁶ The loss of CP can prevent the virus from forming a 47 complete viable virion.^{17,18} Therefore, it is an important pathway for development of 48 anti-ToCV agents using ToCV-CP as a target protein.¹⁹ 49

In our previous work, we discovered and reported the anti-plant virus activity of the
dithioacetal structure, which has good antiviral activity against tobacco mosaic virus

(TMV), potato virus Y (PVY), cucumber mosaic virus (CMV) and ToCV.¹⁹⁻²¹ In 52 addition, we established an efficient and convenient screening method for anti-ToCV 53 54 agents using ToCV-CP as a target protein and some glycoside-containing dithioacetal derivatives were synthesized with good antiviral activity against ToCV.¹⁹ As an 55 important class of nitrogen-containing heterocyclic compounds, 4(3*H*)-56 quinazolinones are widely found in natural products with a range of significant 57 biological activities. Our group found that some 4(3H)-quinazolinone derivatives 58 showed good antiviral activity against CMV and TMV.²²⁻²⁴ However, the biological 59 60 activity of this class of compounds against ToCV has not been reported. In this work, a series of 4(3H)-quinazolinone derivatives containing dithioacetal (Figure 1) were 61 designed, synthesized, and evaluated for their anti-ToCV activities in vitro and in 62 63 vivo. The findings will provide guidance for the design, synthesis and development of anti-ToCV agents. 64

65

Figure 1

66 MATERIALS AND METHODS

67 Chemicals. Analytically pure reagents used in the experiments did not require68 further drying or purification.

69 Instruments. ¹H NMR and ¹³C NMR spectra of the compounds were obtained

vising a Bruker DPX 400 MHz (Bruker, Germany) and a JEOL-ECX500 MHz (JEOL,

Tokyo, Japan) in CDCl₃ or DMSO- d_6 solution. HRMS was performed with a Thermo

72 Scientific Q Exactive (Thermo Scientific, USA). The melting points of the compounds

73 were measured using WRX-4 equipment.

74	General Procedure for the Synthesis of the Intermediates A1-A3 and B1-B9.				
75	Intermediates A1-A3 were synthesized according to the methods reported in the				
76	literature. ²⁵ Intermediates A1-A3 (2.35 mmol), substituted p-hydroxybenzaldehyde				
77	(2.35 mmol) and CH ₃ CN (20 mL) were added to a 50 mL three-necked flask, with				
78	K ₂ CO ₃ (2.82 mmol) as an acid-binding agent. The mixing system was stirred at room				
79	temperature for 10 minutes, and KI (0.23 mmol) was then added. The mixture was				
80	allowed to react under reflux for 8 to 12h. After the reaction completed (as monitored				
81	by TLC), the solvent was removed under vacuum, the residue was then diluted with				
82	water, dichloromethane (50 mL \times 3) was added for extraction, and the two layers were				
83	separated. The organic layer was dried and concentrated to obtain the crude products,				
84	which were recrystallized with anhydrous ethanol to obtain the intermediates B1-B9.				
85	Figure 2				
86	General Procedure for Preparation of the Title Compounds C1-C27. The				
87	intermediates B1-B9 (1.34mmol) were added to a round bottom flask containing				
88	substituted mercaptan (2.68 mmol) and NaHSO ₄ ·SiO ₂ (1.34 mmol) in CH ₂ Cl ₂ solvent				
89	(10 mL). Upon reaction completion, the solvent was removed under vacuum, the				

(10 mL). Upon reaction completion, the solvent was removed under vacuum, the
residue was dissolved with water, dichloromethane (30 mL×3) was added for
extraction, and the two layers were separated. The collected dichloromethane layer
was evaporated, and then purified by flash chromatography with ethyl acetate
/petroleum ether (1:3, v/v) to obtain the title Compounds C1-C27 (Figure 2).
Purification of Tomato Chlorotic Virus Coat Protein (ToCV-CP). The

⁹⁴ Furnication of Foliato Chlorotic virus Coat Frotein (FoCV-CF). The ⁹⁵ ToCV-CP was cloned, expressed and purified according to the method described in 96 the literature.¹⁹

97 Evaluation of Anti-ToCV Activity *in vitro*. Binding of compounds with 98 ToCV-CP was analyzed through microscale thermophoresis (MST) to obtain Kd 99 values by methods described in the literature.^{26,27} Meanwhile, NNM, the lead 100 compound XCLSBM and Ribavirin (RIB) were used as positive controls. All tests 101 were duplicated three times.

Evaluation of Anti-ToCV Activity in vivo. We used tomato samples (Shouguang, 102 Shandong) infected with ToCV to start in vivo experiments. All of the samples were 103 104 confirmed to be infected with ToCV by polymerase chain reaction (PCR). The PCR primer sequences used are shown in Table 1. Anti-ToCV activity was then assessed in 105 *vivo* according to the previously described method.¹⁹ The primers used in quantitative 106 107 real-time PCR are shown in Table 1. A MiniBEST Plasmid Purification Kit 4.0 (Takara) was used to extract the total RNA of samples, and a Goldenstar RT6 cDNA 108 Synthesis Kit (Tsingke) was used when the extracted total RNA was reverse 109 110 transcribed into cDNA. The quantitative real-time PCR was performed using the Fast qPCR mix SYBR Green kit (Tsingke). 111

112 **RESULTS AND DISCUSSION**

113 **Chemistry.** Figure 2 shows the synthetic routes of 4(3H)-quinazolinone derivatives 114 bearing dithioacetal groups. Methanol was used as a solvent, sodium was added, 115 substituted anthranilic acid and chloroacetonitrile as raw materials, and the mixture 116 was stirred at room temperature for 4-6 h to obtain intermediates **A1-A3**. The second 117 step of the reaction used acetonitrile as a solvent, K₂CO₃ as a catalyst, intermediates 118 A1-A3 and substituted 4-hydroxybenzaldehyde under reflux for 8-12 h to obtain 119 intermediates **B1-B9**. Finally, with dichloromethane as a solvent and NaHSO₄·SiO₂ as 120 a catalyst, intermediates **B1-B9** and substituted mercaptan were reacted at room 121 temperature to obtain the target compounds C1-C27. The chemical structures of these 122 compounds were identified by HRMS, and NMR (Supporting Information).

123

Figure 3

Anti-ToCV Activity of Compounds in vitro. The results of MST experiments are 124 shown in Table 2, which indicates that the target compounds of this series have good 125 126 binding affinity to ToCV-CP, with most of the compounds binding at the micromolar level. In particular, compounds C5 and C22 showed excellent binding capacity to 127 ToCV-CP, with Kd values of 0.24 and 0.25 μ M, which are significantly better than 128 129 that of RIB (15.62 μ M) and slightly superior to those of NNM (0.44 μ M) and lead compound XCLSBM (0.36 μ M). Compounds C9, C10, C12, and C19 showed good 130 binding capacity to ToCV-CP, with Kd values of 0.52, 1.63, 1.66, and 1.02 μ M, 131 132 respectively, which are preferred over that of RIB, but second to those of NNM and lead compound XCLSBM (Figure 3). 133

Structure-activity relationships (SARs). The in vitro binding affinities of ToCV-CP with target compounds showed that when R_2 was a chlorine atom (electron-withdrawing group), these compounds had the worst anti-ToCV activity; for example, C10 > C13 > C16 (H > 2-OCH₃ > 2-Cl); C11 > C14 > C17 (H > 2-OCH₃ > 2-Cl); C12 > C15 > C18 (H > 2-OCH₃ > 2-Cl); C20 > C23 > C26 (H > 2-OCH₃ > 2-Cl); C5 > C2> C8 (2-OCH₃ > H > 2-Cl); C22 > C19 > C25 (2-OCH₃ > H > 2-Cl); however, this rule is not followed by all compounds. For example, C9 > C3 > C6(2-Cl > H > 2-OCH₃) and C27 > C21 > C24 (2-Cl > H> 2-OCH₃). The changes of the substituents of R₁ have a slight effect on the anti-ToCV activity, as fluorine, chlorine and bromine atoms are all electron-withdrawing groups; for example, C19 > C10 >C1 (6-Br > 6-Cl > 6-F), but C20 > C2 > C11 (6-Br > 6-F > 6-Cl).

145

Figure 4

Anti-ToCV Activity of Compounds C5 and C22 in vivo. After confirming 146 tomato infection of ToCV by polymerase chain reaction (PCR), the PCR reaction 147 148 produced an expected site amplicon (774 bp), as shown in Figure 4. The side branches of tomato infected with ToCV were cut off and soaked it in tap water for 5 days, and 149 then situated in a 50 μ g/mL compound solution for 7 days. The result showed that the 150 151 expression levels of the ToCV-CP gene in different compound treatment groups showed large differences. As shown in Figure 5, 81.05% and 87.59% reductions in 152 ToCV-CP gene expression levels could be achieved in the groups treated with 153 154 compounds C5 and C22 in the tomato sample, respectively, which were obviously superior to the water (CK) treatment, the NNM treatment (43.88%) and the lead 155 compound XCLSBM treatment (63.56%) counterparts. The results of in vivo 156 experiments showed that the expression level of ToCV-CP in tomato can be 157 significantly reduced by compounds C5 and C22, which have certain controlling 158 effects on tomato chlorotic virus. 159

160

Figure 5

161 In summary, twenty-seven novel 4(3H)-quinazolinone derivatives bearing

162	dithioacetal moiety were designed and synthesized. The results showed that most of
163	the compounds had good binding capacity to ToCV-CP. In particular, compounds C5
164	and C22 had excellent Kd to ToCV-CP, with values of 0.24 and 0.25 μ M, respectively.
165	The <i>in vivo</i> experiments further indicate that the ToCV-CP gene expression levels are
166	reduced when tomatoes are treated with compounds C5 and C22. Our present work
167	indicated that novel 4(3H)-quinazolinone derivatives containing dithioacetal moiety
168	can be used as novel anti-ToCV agents to control this plant virus disease.

170 ASSOCIATED CONTENT

171 Supporting information

- 172 Characterization data, ¹H and ¹³C NMR spectra, and HRMS for the title compounds
- 173 C1–C27 are shown in the Supplementary Information. The data of the interactions
- between C1–C27 and ToCV-CP are also listed in the Supporting Information.

175 AUTHOR INFORMATION

176 Corresponding Authors

- *E-mail for Baoan Song: songbaoan22@yahoo.com. *Phone: 86-851-88292170.
- 178 Fax: 86-851-83622211.
- *E-mail: dyhu@gzu.edu.cn. Tel.: 851-8829-2170. Fax: 86-8518829-2170.
- 180 **ORCID**
- 181 Baoan Song: 0000-0002-4237-6167
- 182 Deyu Hu: 0000-0001-7843-371X
- 183 Guangcheng Zu: 0000-0003-4677-2957
- 184 Xiuhai Gan: 0000-0002-4070-0824
- 185 Dandan Xie: 0000-0002-2531-1038
- 186 Huanyu Yang: 0000-0001-5842-9166
- 187 Awei Zhang: 0000-0003-0479-9705
- 188 Shaoyuan Li: 0000-0001-6280-9129

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- 193 The authors declare no competing financial interest.

ABBREVIATIONS

- ¹H NMR, ¹H nuclear magnetic resonance; ¹³C NMR, ¹³C nuclear magnetic resonance;
- 196 HRMS, High-resolution mass spectrometry; NNM, Ningnanmycin; RIB, Ribavirin;
- 197 XCLSBM, Xiangcaoliusuobingmi; ToCV, tomato chlorosis virus; CP, coat protein;
- 198 MST, microscale thermophoresis; ToCV-CP, Tomato chlorosis virus coat protein.

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299 Figure Information

- **Figure 1**. Design of the title compounds.
- **Figure 2**. Synthesis route of the title compounds **C1-C27**.
- **Figure 3**. Microscale thermophoresis (MST) results of the title compounds.
- **Figure 4**. Results of samples tested *in vivo* through polymerase chain reaction (PCR).
- **Figure 5**. Effects of compounds on the inhibition of ToCV-CP levels in tomato *in*

vivo.

524		Tuble 1. Thinkis used in this study				
	Primer	Sequence (5'-3')	Purpose			
ToCP F ToCP R		ATGGAGAACAGTGCCGTTGC				
		TTAGCAACCAGTTATCGATGC	ToCV identification ^a			
	qToCP-F2	TAGATGATGGCGTAGATGAC				
	qToCP-R 2	CTAGTGGAGTGTACCTTCAAT	ToCV qPCR ^b			
	qActin-F	TGCCATTCTCCGTCTTGACT	Tomoto noforma on och			
	qActin-R	TGCAGTCTCGAGTTCCTGTT	Tomato reference genes ^o			
325	^a Primer seque	ences were reported by Segev et al. ¹⁰				
326	 ^a Primer sequences were reported by Segev <i>et al.</i>¹⁰ ^b Primer sequences were provided by Zhou Tao of China Agricultural Universit Friendship 					
327	Friendship.					
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Table 1. Primers used in this study

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342 Table 2. Compound structures and their binding constants (Kd) with the ToCV coat

343 protein

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	٥	O NH		
Compound	IX.		6	V d voluo (uM)
Compound	S. S.		Ku value (μ IVI)	
			R ₃	
	R ₁	R ₂	R ₃	
C1	6-F	Н	CH ₂ CH ₃	3.49 ± 2.27
C2	6-F	Н	$CH_2CH_2CH_3$	5.67±2.92
C3	6-F	Н	$CH(CH_3)_2$	3.03+1.42
C4	6-F	2-OCH ₃	CH ₂ CH ₃	15.44±5.99
C5	6-F	2-OCH ₃	$CH_2CH_2CH_3$	0.24 ± 0.07
C6	6-F	2-OCH ₃	$CH(CH_3)_2$	33.95±20.84
C7	6-F	2-Cl	CH ₂ CH ₃	11.56±7.00
C8	6-F	2-Cl	$CH_2CH_2CH_3$	59.65±23.49
С9	6-F	2-Cl	$CH(CH_3)_2$	0.52 ± 0.09
C10	6-Cl	Н	CH ₂ CH ₃	1.63±0.73
C11	6-Cl	Н	CH ₂ CH ₂ CH ₃	9.98+3.08
C12	6-Cl	Н	$CH(CH_3)_2$	1.66±0.54
C13	6-Cl	2-OCH ₃	CH ₂ CH ₃	10.43±6.81
C14	6-Cl	2-OCH ₃	CH ₂ CH ₂ CH ₃	67.60±30.87
C15	6-Cl	2-OCH ₃	$CH(CH_3)_2$	28.74±10.69
C16	6-Cl	2-Cl	CH ₂ CH ₃	15.22±3.64
C17	6-Cl	2-Cl	CH ₂ CH ₂ CH ₃	73.11±52.75
C18	6-Cl	2-Cl	$CH(CH_3)_2$	42.91±29.93
C19	6-Br	Н	CH ₂ CH ₃	1.02+0.39
C20	6-Br	Н	CH ₂ CH ₂ CH ₃	4.82±3.13
C21	6-Br	Н	$CH(CH_3)_2$	25.50±8.79
C22	6-Br	2-OCH ₃	CH ₂ CH ₃	0.25±0.06
C23	6-Br	2-OCH ₃	CH ₂ CH ₂ CH ₃	42.69±22.76
C24	6-Br	2-OCH ₃	$CH(CH_3)_2$	48.65±32.42
C25	6-Br	2-Cl	CH ₂ CH ₃	9.64±8.79
C26	6-Br	2-Cl	CH ₂ CH ₂ CH ₃	44.61±24.51
C27	6-Br	2-Cl	CH(CH ₃) ₂	10.71±4.15
ningnanmycin ^a			/-	0.44±0.26
Ribavirin ^a				15.62±7.15
XCLSBM ^b				0 36±0 24

³⁴⁴ ^{*a*}Commercially available agrichemicals were used as controls.

 b Lead compound was reported by Zhang *et al.*²⁰





Figure 1. Design of the title compounds.

379x82mm (300 x 300 DPI)



Figure 2. Synthesis route of the title compounds C1-C27.

197x113mm (300 x 300 DPI)



Figure 3. Microscale thermophoresis (MST) results of the title compounds.

279x165mm (200 x 200 DPI)



Figure 4. Results of samples tested in vivo through polymerase chain reaction (PCR).

61x83mm (96 x 96 DPI)



Figure 5. Effects of compounds on the inhibition of ToCV-CP levels in tomato in vivo.

192x122mm (150 x 150 DPI)



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