Tetrahedron xxx (2013) 1-12

Contents lists available at SciVerse ScienceDirect

Tetrahedron

journal homepage: www.elsevier.com/locate/tet

2-Alkylidene-4-oxothiazolidine S-oxides: synthesis and stereochemistry

Zdravko Džambaski^{a, b}, Rade Marković^{a, c, †}, Erich Kleinpeter^{b, *}, Marija Baranac-Stojanović^{a, c, *}

^a Center for Chemistry ICTM, University of Belgrade, P.O. Box 473, 11000 Belgrade, Serbia ^b Chemisches Institut der Universität Potsdam, Karl-Liebknecht Str. 24-25, D-14476 Potsdam (Golm), Germany ^c Faculty of Chemistry, University of Belgrade, Studentski trg 16, P.O. Box 158, 11000 Belgrade, Serbia

ARTICLE INFO

Article history: Received 23 March 2013 Received in revised form 9 May 2013 Accepted 20 May 2013 Available online xxx

Keywords: 2-Alkylidene-4-oxothiazolidine Sulfoxide Diastereoselectivity Density functional calculations CH…O hydrogen bonds

ABSTRACT

A series of 5-unsubstituted and 5-substituted 2-alkylidene-4-oxothiazolidine-S-oxides were synthesized by the sulfur-oxidation with *m*-CPBA. The stereochemistry of 5-substituted sulfoxides was determined by means of NMR spectroscopy and DFT theoretical calculations. It was found that the thermodynamically less stable *anti*-isomer was initially formed in the course of the oxidation, but it underwent epimerization to the mixture enriched in the more stable *syn*-isomer, during the work-up process. The higher stability of *syn*-isomers is ascribed to the stronger hyperconjugative $\sigma_{C-H} \rightarrow \sigma^*_{S-O}$ interaction versus the weaker $\sigma_{C-C} \rightarrow \sigma^*_{S-O}$ delocalization in their *anti*-counterparts and to the existence of intramolecular 1,5-CH···O hydrogen bonds.

© 2013 Elsevier Ltd. All rights reserved.

Tetrahedror

1. Introduction

The 4-oxothiazolidine core is an important structural motif of biologically active compounds.¹ A wide range of pharmacological activities, such as antibacterial,² antiviral,³ anti-inflammatory,⁴ antifungal,⁵ analgesic,⁶ or anticancer⁷ have been reported. A subclass constitutes 4-oxothiazolidine derivatives possessing an exocyclic double bond at the C-2 position of the ring, with some of them registered as nontoxic, active substances in treatment of neurological diseases (Ralitoline—an antiepileptic),⁸ liver diseases (Piprozoline—a choleretic), or for treatment of hypertension (Etozolin—a diuretic) (Fig. 1).

The sulfoxide functional group is common in pharmaceutically important compounds and many sulfoxides are known to have high biological activity. Sulfur-oxidation of 4-oxothiazolidine ring may result in more potent compounds.⁹ Having this in mind, we investigated the oxidation of a series of 2-alkylidene-4-oxothiazolidines **1** into sulfoxides and analyzed the origin of diastereoselectivity observed in the case of 5-substituted derivatives.

* Corresponding authors. Tel.: +38111 3336740; fax: +38111 2636061 (M.B.-S.); tel.: +49 331 977 5210/5211; fax: +49 331 977 5064/5057 (E.K.); e-mail addresses: ekleinp@

0040-4020/\$ – see front matter @ 2013 Elsevier Ltd. All rights reserved. http://dx.doi.org/10.1016/j.tet.2013.05.087 In addition, sulfoxides are important synthetic intermediates and can be employed for further functionalization of thiazolidines **1**.



2. Results and discussion

The starting 4-oxothiazolidines **4** were prepared by the basecatalyzed reaction of α -mercapto esters **2** and α -substituted nitriles **3**, as already described¹⁰ (Scheme 1). N-Alkylation was achieved with MeI, BnBr, Br(CH₂)₃Br or BrCH₂CO₂Et in moderate to high yields (53–97%), under mild reaction conditions (Table 1 and Scheme 1). After an initial screening of different agents, *meta*chloroperbenzoic acid (*m*-CPBA) was chosen for the oxidation step (Table 2). When used in reagent/substrate 1.5–2/1 molar ratio, sulfone formation was minimized¹¹ and sulfoxides **5** were isolated in good yields (65–95%, Table 1). All the products **1**, **4** and **5** were obtained exclusively as *Z* isomers, concerning the configuration around the C=C double bond.¹² 5-Substituted substrates **1j–r** were oxidized with moderate to good diastereoselectivity, as evidenced



uni-potsdam.de (E. Kleinpeter), mbaranac@chem.bg.ac.rs (M. Baranac-Stojanović). † 1946–2012.

Z. Džambaski et al. / Tetrahedron xxx (2013) 1-12



Ralitoline

Piprozolin

Etozolin





Scheme 1. Reagents and conditions: (i) K_2CO_3 cat., EtOH, reflux; (ii) Mel, BnBr, Br(CH₂)₃Br, or BrCH₂CO₂Et (1.1–1.5 equiv), K_2CO_3 (1 equiv), DMF, rt; (iii) *m*-CPBA (1.5–2 equiv), CH₂Cl₂, 0 °C.

Table 1	
Yields ^a of N-alkylated products 1, sulfoxides 5, experimental and calculated	$^{\rm o}$ syn/anti ratio for 5-substituted derivatives ${\bf 5}$ and their relative free energy

R	\mathbb{R}^1	EWG	1 (%)	5 (%)	Ratio <i>syn/anti</i> exp (calcd)	Relative free energy (kcal/mol) <i>syn/anti</i>
Н	Me	CONHPh	1a (97)	5a (75)	_	_
Н	Me	CONH(CH ₂) ₂ Ph	1b (95)	5b (85)	_	_
Н	Me	CO ₂ Et	1c (73)	5c (80)	_	_
Н	Me	COPh	1d (95)	5d (74)	_	_
Н	Bn	CONHPh	1e (64)	5e (72)	_	_
Н	Bn	CO ₂ Et	1f (82)	5f (95)	_	_
Н	Bn	COPh	1g (89)	5g (69)	_	_
Н	(CH ₂) ₃ Br	CO ₂ Et	1h (94)	5h (84)	_	_
Н	CH ₂ CO ₂ Et	CO ₂ Et	1i (93)	5i (84)	_	_
Me	Me	CONHPh	1j (89)	5j (71)	85/15 (87/13)	0/1.15
Me	Me	CONH(CH ₂) ₂ Ph	1k (87)	5k (65)	86/14 (81/19)	0/0.87
Me	Me	CO ₂ Et	1l (79)	5l (93)	86/14 (85/15)	0/1.01
Me	Me	COPh	1m (53)	5m (73)	85/15 (87/13)	0/1.13
Me	Bn	CONHPh	1n (66)	5n (76)	87/13 (93/7)	0/1.58
Me	Bn	CO ₂ Et	1o (92)	50 (85)	86/14 (91/9)	0/1.41
Me	(CH ₂) ₃ Br	CO ₂ Et	1p (72)	5p (85)	87/13 (83/17)	0/0.93
CH ₂ CO ₂ Et	Bn	CONHPh	1q (77)	5q (79)	79/21 ^c (94/6)	0/1.60
CH ₂ CO ₂ Et	Bn	COPh	1r (93)	5r (71)	77/23 ^c (90/10)	0/1.27

^a Yields of isolated products.

^b At the B3LYP/6-31 G^* level, based on the relative free energy of diastereomers.

^c Not completely equilibrated.



^a Yields of isolated products.

from the NMR spectroscopic data of isolated products where two sets of signals appeared, one prevailing. On the basis of ¹H NMR data, diastereomeric ratio was calculated to range from \sim 79/21 for **5q,r** to \sim 87/13 for **5j**–**p**. It was necessary to identify whether the *syn* or *anti* diastereomer was the major product.

Having a sulfoxide in hand, it is often a challenge to determine its configuration. Apart from an X-ray analysis requiring welldefined single crystals, general and reliable methods for configurational assignments are lacking. An assessment of stereochemistry directly from NMR spectroscopic data is not possible since coupling constants or NOE contacts with oxygen are inaccessible. Some relations between chemical shifts of neighbouring hydrogen and carbon atoms and the configuration of the sulfoxide group have been observed,¹³ but are applicable only to specific cases. Although magnetic anisotropy of the sulfoxide group has been predicted to be acetylene-like,¹⁴ it is weak due to the single bond character of the SO bond.¹⁵ Unfortunately, in the synthesized compounds 5j-rthe ¹H NMR chemical shift difference of C5–H and C5–CH₃

Please cite this article in press as: Džambaski, Z.; et al., Tetrahedron (2013), http://dx.doi.org/10.1016/j.tet.2013.05.087

2

between the two diastereomers is too small (0.1–0.3 ppm, Table 3) to be of use for structure determination. In addition, all attempts to obtain a single crystal suitable for an X-ray analysis resulted in a mixture of isomers. So, we turned our attention to computational methods, the use of which is now widespread among experimental chemists to solve various stereochemical problems.¹⁶ Using quantum chemical calculations it is possible to accurately predict NMR chemical shifts,¹⁷ which then allows one to distinguish between stereoisomers by comparing calculated and experimental values.

documented γ -gauche effect. It has been suggested by Dračínský et al.^{16a,b} to employ ¹³C NMR chemical shift changes ($\Delta\delta$) induced by sulfur-oxidation as a means to differentiate between stereoisomeric sulfoxides in which the sulfoxide group is contained in various fiveand six-membered rings. The corresponding experimental and theoretically calculated values for the synthesized sulfoxides **5j**–**r** are listed in Table 3 and they proved useful in the present case, as well. These differences obviously arise from the above mentioned steric and electric field effects and go in the following direction:

Table 3

Selected experimental^a and calculated^{b 1}H NMR and ¹³C NMR chemical shifts, and the ¹³C NMR chemical shifts changes ($\Delta\delta$) induced by the oxidation

Compound		¹ H NMR (ppr	¹ H NMR (ppm) ¹³ C NMR (ppm)				$\Delta \delta^{c}$ (ppm)					
		Exp CDCl ₃ /DMSO-d ₆		C5	C5		C5–CH ₃		C5		C5−CH ₃	
		С5—Н	C5−CH ₃	Exp	Calcd	Exp	Calcd	Exp	Calcd	Exp	Calcd	
5j	syn	-/3.89	-/1.36	52.5	57.1	6.6	8.6	12.7	12.5	-12.3	-11.4	
	anti	-/3.65	-/1.40	59.5	63.0	11.7	14.4	19.7	18.3	-7.2	-5.5	
5k	syn	3.31/3.82	1.57/1.33	53.2	56.9	6.8	8.6	12.7	12.3	-12.2	-11.6	
	anti	3.50/3.56	1.49/1.35	59.6	65.5	12.4	13.6	19.2	20.9	-6.6	-6.6	
51	syn	3.42/3.95	1.63/1.36	53.3	57.4	6.7	8.4	12.8	12.3	-12.3	-11.6	
	anti	3.63/3.74	1.56/1.44	59.9	63.5	12.3	14.6	19.3	18.4	-6.7	-5.4	
5m	syn	3.44/3.98	1.65/1.38	53.1	57.0	6.9	8.6	12.9	12.4	-11.6	-11.1	
	anti	3.67/3.78	1.59/1.48	59.7	62.9	12.4	14.6	19.5	18.3	-6.2	-5.1	
5n	syn	-/4.11	-/1.42	52.6	58.0	6.6	8.7	13.0	13.7	-12.5	-11.8	
	anti	-/3.80	-/1.46	59.2	63.1	11.7	13.3	19.6	18.8	-7.4	-7.2	
50	syn	3.48/4.16	1.67/1.42	53.5	58.4	6.7	8.5	13.2	13.7	-12.6	-12.0	
	anti	3.71/3.90	1.58/1.48	59.6	63.5	12.4	13.2	19.2	18.7	-7.0	-7.3	
5p	syn	3.39/-	1.62/-	53.5	57.1	6.6	8.4	13.1	12.4	-12.4	-11.6	
-	anti	3.63/-	1.54/-	59.6	63.2	12.2	14.4	19.2	18.5	-6.8	-5.6	
		C5-H	C5-CH ₂	C5		C5-CH ₂		C5		$C5-CH_2$		
5q	syn	-/4.47	-/2.94, 3.09	53.8	59.5	27.4	27.5	12.8	13.6	-9.4	-12.8	
-	anti	-/3.93	-/3.23, 3.54	61.6	66.6	31.8	33.5	20.6	20.7	-4.9	-6.8	
5r	syn	4.01/4.48	3.23, 3.27/2.95; 3.10	54.6	59.4	27.3	27.6	13.3	13.3	-9.9	-12.5	
	anti	3.75/3.97	3.41, 3.44/-	62.1	66.6	32.3	32.8	20.8	20.8	-5.0	-7.3	

^a Data are from DMSO-*d*₆ solution for **5***j*, **5***n* and **5***q*, and from CDCl₃ solution for all others.

 $^{\rm b}\,$ At the B3LYP/6-31G* theory level, in the gas phase, relative to TMS.

^c Difference between chemical shift values of sulfoxides **5** and thiazolidines **1**.

For this purpose the structures **5j**–**r** were optimized in the gas phase at the B3LYP/6-31G* level.¹⁸ NMR chemical shifts were calculated at the same level using the GIAO method¹⁹ and were plotted against the experimental values (Fig. 2).²⁰ An excellent correlation gave evidence for reliable calculated structures and allowed the stereochemistry determination: the major isomer proved to be *syn*. The most diagnostic chemical shift values are those for carbon atoms at the α - (C5) and β -positions (C5–CH₃), which differ by ~6 ppm between the two isomers (Table 3). A diamagnetic shift observed for *syn*-isomers arises from steric and electric field effects, in the case of the C5–CH₃ being the well





paramagnetic shift for the α -carbon is higher for *anti*-isomers, whereas diamagnetic shift for the β -carbon (CH₃/CH₂ group) is larger for *syn*-isomers. Interestingly, the C5–CH₃ and C5–H signals in the proton NMR spectra exchange their relative position upon shifting from the non-polar solvent to the polar one. Thus, the C5–CH₃ (C5–H) protons of *syn*-isomers resonate at lower (higher) field with respect to *anti*-isomers in chloroform solution, but at higher (lower) field in DMSO solution (Table 3, Fig. S1 in the Supplementary data).

Almost no change in the ¹³C NMR chemical shifts of the C2 atoms experimentally observed after the sulfur-oxidation (Table 4) results from the two opposing effects: paramagnetic shift due to the -I effect of the sulfoxide group and diamagnetic shift coming from the decrease in the push–pull effect of the C=C double bond. This latter effect is the reason for the observed paramagnetic movement of the C2' carbon resonances. The overall result is the decrease in the ¹³C NMR chemical shift difference between the two carbons of the double bond, $\Delta \delta_C =_C$, in the oxidized products (Table 4). Namely, all thiazolidine derivatives **1a-r** and **5a-r** belong to the class of the so-called push-pull alkenes, possessing one or two electron-donating substituents at one end of the double bond and one or two electron-accepting groups at the other. Electronic interactions between the donor and acceptor groups via the C=C double bond reduces its π -bond order and strongly influences its dynamic behaviour and chemical reactivity.

Among several parameters, such as the restricted rotation about the partial C=C double bond,²¹ bond length²² and the occupation quotient π^*/π ,^{21d,23} this push–pull effect can also be quantified by the ¹³C NMR chemical shift difference between the two carbons of the double bond, which increase with increasing push–pull

4

ARTICLE IN PRESS

Z. Džambaski et al. / Tetrahedron xxx (2013) 1-12

Table 4 Experimental^a ¹³C NMR chemical shifts for the two carbons of the C=C double bond and their difference ($\Delta\delta_{c}=_{c}$)

	¹³ C NMR (pp	m)				¹³ C NMR (pp	m)	
	C2	C2′	$\Delta \delta_{C} = C$			C2	C2′	$\Delta \delta_{C} = C$
1a	155.3	93.7	61.6	5a		156.5	102.4	54.1
1b	152.9	93.6	59.3	5b		155.2	102.8	52.4
1c	158.6	90.7	67.9	5c		158.4	100.2	58.2
1d	161.8	95.7	66.1	5d		159.2	102.7	56.5
1e	154.5	94.4	60.1	5e		155.7	102.8	52.9
1f	158.6	90.3	68.3	5f		159.4	98.93	60.5
1g	160.6	97.0	63.6	5g		157.7	104.3	53.4
1h	157.6	90.6	67.0	5h		157.5	100.5	57.0
1i	157.3	90.7	66.6	5i		157.2	100.7	56.5
1j	153.5	93.5	60.0	5j	syn	154.7	103.2	51.5
					anti	154.7	103.8	50.8
1k	153.6	92.3	61.4	5k	syn	153.5	103.8	49.6
					anti	153.5	104.1	49.4
11	157.3	90.2	67.1	51	syn	157.1	100.6	56.5
					anti	157.1	100.9	56.2
1m	160.4	95.3	65.1	5m	syn	157.8	103.1	54.7
					anti	158.0	103.3	54.7
1n	152.5	94.1	58.4	5n	syn	153.8	103.8	50.0
					anti	153.8	104.6	49.2
10	156.2	91.4	64.8	50	syn	155.9	101.7	54.2
					anti	156.0	102.1	53.9
1p	156.3	90.1	66.2	5p	syn	156.1	100.8	55.3
					anti	156.1	101.1	55.0
1q	153.2	94.3	58.9	5q	syn	154.2	103.9	50.3
					anti	154.8	102.8	52.0
1r	159.3	97.0	62.3	5r	syn	156.4	105.4	51.0
					anti	157.6	103.3	54.3

^a Data are from DMSO-d₆ solution for 1a/5a, 1b/5b, 1e/5e, 1f/5f, 1j/5j, 1n/5n and 1q/5q, and from CDCl₃ solution for all others.

character.^{21b,23a,24} The experimental chemical shifts for the C2 and C2' atoms and their differences, $\Delta \delta_{C} = C$, for compounds **1a**–**r** and **5a**–**r** are given in Table 4. The observed decrease in the push–pull activity results from the substitution of a good electron donor, the sulfide, by a poor donor, the sulfoxide. The stabilization energies of the sulfur lone pair conjugative interaction with the π^* -antibonding orbital of the C=C double bond drastically decrease upon oxidation. For example, this energy, obtained from the second order perturbation theory analysis of Fock matrix in the NBO basis,²⁵ for 1c amounts 24.4 kcal/mol, but only 0.7 kcal/mol for 5c. Though significantly reduced, the sulfur lone pair $n \rightarrow \pi^*_{C} =_{C}$ donation in sulfoxides still exists and it has already been shown that the sulfoxide group can act as an electron donor.²⁶ However, its net effect on the double bond of compounds **5** is electron attraction: the π electron density is attracted into the σ^* S–O orbital, as evidenced from the corresponding stabilization energy of 2.6 kcal/mol for **5c**. The reduction in the sulfur lone pair donation to the C=C double bond is mainly caused by the presence of the electronegative oxygen at the sulfur atom, but also by stereoelectronic factors: while one of the sulfur's lone pairs in compounds **1** is p-like and is in plane with the π^* orbital of the double bond, which allows a significant $n_S \rightarrow \pi^*_C =_C$ delocalization, this is not the case for sulfoxides with the sp³ hybridization of the sulfur (Fig. 3). The difference in the $\Delta \delta_C = C$ values between the syn- and anti-isomers is, however, too small and not regular to be of use for stereochemistry determination.

The free energies, predicted at the same theory level employed for the structure optimizations, favour *syn*-isomers by 0.9–1.6 kcal/ mol (Table 1). Experimentally observed and computed *syn/anti*



ratios are in good agreement and they point to a thermodynamic control of the oxidation reactions. This was corroborated when the oxidation of **11** was monitored by NMR spectroscopy in CDCl₃, at 0 °C: anti-51 was initially formed as the major product, as a result of steric approach control (attack of the oxidizing agent from the less hindered side) (Fig. S2 in the Supplementary data). The initial syn/ anti ratio of 28/72 did not change during the reaction time. However, due to the facile isomerization of sulfoxides having an α hydrogen,^{13b,27} the anti-51 underwent epimerization to the equilibrium mixture (syn/anti 86/14, Table 1) during the work-up process. In order to learn why syn-isomers are more stable than their anti-counterparts donor-acceptor interactions and their stabilization energies have been examined. The main difference in the hyperconjugative interactions between the two isomers comes from the $\sigma_{C5-H} \rightarrow \sigma^*_{S-O}$ existing in the *syn*-isomers (E2 values: 1.42-1.6 kcal/mol; dihedral H-C5-S-O angles: from -161.2 to -167.7°), but replaced by the weaker $\sigma_{C5-C5'} \!\rightarrow \! \sigma^*{}_{S-O}$ interaction in the anti-isomers (E2 values: 0.63-0.79 kcal/mol; dihedral H₃C-C5-S-O angles: from -109.5 to -158.2°) (Fig. 4 and Table 5).



 $R = CH_3, CH_2CO_2Et$

Fig. 4. Hyperconjugative interactions in syn- and anti-sulfoxides 5.

Table 5 Calculated dihedral angles and energies $(E2)^a$ of $\sigma_{C-H} \rightarrow \sigma^*_{S-O}$ and $\sigma_{C-C} \rightarrow \sigma^*_{S-O}$ interactions for *syn*- and *anti*-**5**, respectively

Compound	syn-isomer		anti-isomer		
	$\tau_{H-C5-S-O}$ (°)	σ _{C-H} →σ [*] _{S-O} (kcal/mol)	$\overline{\tau_{C-C5-S-O}}(^{\circ})$	$\sigma_{C-C} \rightarrow \sigma^*_{S-O}$ (kcal/mol)	
5j	-161.6	1.56	-145.1	0.67	
5k	-161.2	1.55	-109.5	_	
51	-161.3	1.56	-141.4	0.63	
5m	-161.4	1.55	-143.6	0.65	
5n	-167.7	1.55	-158.2	0.79	
50	-167.4	1.60	-157.5	0.79	
5p	-162.2	1.57	-144.1	0.67	
5q	-163.4	1.44	-119.4	_	
5r	-162.7	1.42	-113.3	_	

^a Obtained from the second order perturbation theory analysis of Fock matrix in the NBO basis.

In addition, the *syn*-isomers are also preferred by the electrostatic effects, the so-called CH···O hydrogen bonds²⁸ occurring between the positively charged methyl/methylene hydrogens and the negatively charged sulfoxide oxygen atom (Fig. 5; optimized structures of *syn*-**51** and *syn*-**5q** are shown). The calculated partial charge of the hydrogen atom of *syn*-**51** (0.20e; $1e=1.602 \times 10^{-19}$ C) that interacts with the oxygen (-0.60e) is larger than the charges

3. Conclusion

A series of 5-unsubstituted and 5-substituted 2-alkylidene-4oxothiazolidine-*S*-oxides **5** were synthesized by the sulfur-oxidation of the parent sulfides **1** with *m*-CPBA. The products were obtained in good yields, 65–95%, and with moderate to good diastereoselectivity in the case of 5-substituted derivatives (de 54–74%). The stereochemistry of 5-substituted products was elucidated by means of NMR spectroscopy and theoretical calculations. While the stereochemistry of the oxidation step is controlled by the reagent (*m*-CPBA) approach from the less hindered side, the final stereochemical outcome is governed by the relative stability of the isomers and is the result of the facile epimerization at the α -carbon. The greater stability of *syn*-isomers arises from intramolecular 1,5-CH…O hydrogen bonding stabilization and stronger $\sigma_{C-H} \rightarrow \sigma^*_{S-O}$ hyperconjugative interaction versus the weaker $\sigma_{C-C} \rightarrow \sigma^*_{S-O}$ overlap in the *anti*-isomers.

4. Computational details

All calculations were done using Gaussian 03 program package.³⁰ Geometries were optimized at the B3LYP/6-31G* level of theory,¹⁸ followed by frequency calculations to verify the nature of stationary points and to obtain thermochemical parameters. Magnetic



Fig. 5. Optimized structures (B3LYP/6-31G*) of syn-5l and syn-5q.

of the other hydrogen atoms (0.16 and 0.18e) and the H···O distance of 2.56 Å is shorter than the expected van der Waals separation of 2.72 Å. In the case of syn-5q the H…O distance is even shorter and amounts 2.47 Å. Here, another methylene hydrogen is involved in hydrogen bonding with the carbonyl oxygen atom and the calculated partial charges of the two H atoms are very similar (0.224 and 0.215e, Fig. 5). Now, the above mentioned puzzling reversal of position of the C5–CH₃ signals in the ¹H NMR spectra of syn- and anti-isomers in the two solvents of different polarity (Table 3) can be rationalized. Intramolecularly hydrogen bonded CH₃ in syn-isomers resonates at lower field in the chloroform solution with respect to the CH₃ in anti-isomers. In DMSO solution the intermolecular hydrogen bonds formed between the CH₃ and the solvent molecules become stronger with the anti-isomers, resulting in paramagnetic shift of their signals. The downfield movement of proton chemical shift of hydrogen bonded CH protons has already been observed.^{28d,29} In the same manner, the ¹H NMR chemical shift position reversal observed for the C5-H signals can be accounted for by the formation of a hydrogen bond between the C5–H and sulfoxide oxygen in the anti-isomers. This hydrogen bond is weaker than that of the CH₃ group (the CH…O distance for anti-51 amounts 2.64 Å). Substituents attached at the N3 and C2['] positions do not affect the relative stability of isomers and diastereoselectivity of the reaction.

shieldings were computed at the same level using the GIAO method.¹⁹ NMR chemical shifts were calculated relative to TMS. Solvent effects on chemical shift values were modelled as IEF-PCM.³¹

5. Experimental

5.1. General

Melting points were determined on a Stuart SMP10 apparatus. The IR spectra were recorded on a Perkin-Elmer FT-IR 1725X spectrophotometer and are reported as wave numbers (cm⁻¹). The NMR spectra were recorded on a Varian Gemini 2000 spectrometer (¹H at 200 MHz, ¹³C at 50.3 MHz) and on a Bruker Ultrashield Advance III (¹H at 500.26 MHz, ¹³C at 125.79 MHz) in DMSO-d₆ or CDCl₃. Chemical shifts are given in parts per million downfield from TMS as an internal standard. ¹³C NMR resonance assignments were aided by the use of the DEPT technique to determine the number of attached hydrogens. Elemental analyses were performed at the microanalysis laboratory at the Centre for Chemistry ICTM. HRMS was carried out on 6210 Time-of-Flight LC/MS (G1969A, Agilent Technologies) coupled with 1200 Series HPLC system (Agilent Technologies). Thin-layer chromatography (TLC) was carried out on Kieselgel G nach Stahl and spots were visualized by iodine or by 50% H₂SO₄. Column chromatography was carried out on SiO₂ (silica

6

Z. Džambaski et al. / Tetrahedron xxx (2013) 1–12

gel 60 Å, 12–26, ICN Biomedicals). All solvents were distilled before use. DMF was distilled over CaH₂.

5.2. General procedure for the preparation of *N*-methyl derivatives 1a–d and 1j–m

Methyl iodide (1.1–1.5 mmol) was added to a stirred mixture of 4-oxothiazolidine **4** (1 mmol) and K₂CO₃ (1 mmol) in DMF (2–5 mL) and stirring was continued at rt until the disappearance of the starting material (TLC). The products were precipitated by pouring a threefold quantity of water onto the reaction mixture, filtrated and air-dried. In the case of **1k** and **1m** the precipitate was not suitable for filtration and the aqueous phase was extracted with ethyl acetate (**1k**) or CH₂Cl₂ (**1m**) and the organic layer was washed with water (3×8 mL), brine solution (1×8 mL) and dried over Na₂SO₄. The solvent was removed under reduced pressure and the crude product was purified by column chromatography (gradient toluene/ethyl acetate).

5.2.1. (Z)-(3-Methyl-4-oxothiazolidin-2-ylidene)-N-phenylethanamide (1a). Compound 1a was obtained from 4 (1.76 g; 7.50 mmol; R=H, EWG=CONHPh), K₂CO₃ (1.04 g; 7.50 mmol), CH₃I (1.60 g; 11.25 mmol; 1.5 equiv) in DMF (20 mL) according to the general procedure (reaction time 2 h) as a white solid (1.80 g; 97%); mp 205–208 °C; *R_f*=0.32 (toluene/ethyl acetate 3/2); IR (ATR): *v*=3544, 3457, 3296, 2964, 1712, 1649, 1594, 1560, 1540, 1489, 1441, 1344, 1307, 1195, 1118, 795, 754, 690 cm⁻¹; ¹H NMR (DMSO- d_{6} , 200 MHz): δ 3.09 (s, 3H, CH₃), 3.78 (s, 2H, CH₂S), 5.79 (s, 1H, =CH), 6.70 (t, *J*=7.3 Hz, 1H, p-Ph), 7.28 (m, 2H, m-Ph), 7.59 (d, J=7.8 Hz, 2H, o-Ph), 9.90 (s, 1H, NH_{amide}), 11.53 (broad s, 1H, NH_{lactam}); ¹³C NMR (DMSO-d₆. 50.3 MHz): δ 29.9 (CH₃), 31.4 (CH₂S), 93.7 (=CH), 118.8 (o-Ph), 122.9 (p-Ph), 129.0 (m-Ph), 140.0 (C1-Ph), 155.3 (C=), 165.3 (CO_{amide}), 172.5 (CO_{lactam}); HRMS: calcd for C₁₂H₁₃N₂O₂S [M+H]⁺ 249.0692, found 249.0696; Anal. Calcd for C12H12N2O2S: C, 58.05; H, 4.87; N, 11.28; S, 12.91; found: C, 58.01; H, 4.92; N, 10.98; S, 12.63.

5.2.2. (Z)-(3-Methyl-4-oxothiazolidin-2-ylidene)-N-(2-phenylethyl) ethanamide (1b). Compound 1b was obtained from 4 (1.31 g; 5.00 mmol; R=H, EWG=CONH(CH₂)₂Ph), K₂CO₃ (0.69 g; 5.00 mmol), CH₃I (0.85 g; 6.00 mmol; 1.2 equiv) in DMF (15 mL) according to the general procedure (reaction time 1.5 h) as a white solid (1.31 g; 95%); mp 177–179 °C; *R*_f=0.52 (toluene/acetone 7/3); IR (ATR): *v*=3260, 3066, 2968, 2934, 1712, 1629, 1569, 1550, 1451, 1338, 1286, 1216, 1121, 863, 803, 752, 701 cm⁻¹; ¹H NMR (DMSO- d_6 , 200 MHz): δ 2.72 (t, J=7.3 Hz, 2H, CH₂Ph), 3.02 (s, 3H, CH₃), 3.28–3.38 (m, 2H, NCH₂), 3.72 (s, 2H, CH₂S), 5.59 (s, 1H, =CH), 7.16-7.34 (m, 5H, Ph), 7.84 (t, J=5.6 Hz, 1H, NH_{amide}); ¹³C NMR (DMSO-*d*₆, 50.3 MHz): δ 29.7 (CH₃), 31.3 (CH₂S), 35.6 (CH₂Ph), 40.3 (NCH₂), 93.6 (=CH), 126.3 (p-Ph), 128.6 (o-Ph), 128.8 (m-Ph), 139.8 (C1-Ph), 152.9 (C=), 166.5 (CO_{a-} mide), 172.4 (CO_{lactam}); ¹H NMR (CDCl₃, 200 MHz): δ 2.86 (t, *J*=6.8 Hz, 2H, CH₂Ph), 3.11 (s, 3H, CH₃), 3.55–3.66 (m, 2H, NCH₂), 3.66 (s, 2H, CH₂S), 5.31 (s, 1H, =CH), 5.42 (broad t, 1H, NH_{amide}), 7.19-7.34 (m, 5H, Ph); ¹³C NMR (CDCl₃, 50.3 MHz): δ 29.8 (CH₃), 31.8 (CH₂S), 35.8 (CH₂Ph), 40.5 (NCH₂), 92.7 (=CH), 126.5 (*p*-Ph), 128.6 (*o*-Ph), 128.8 (*m*-Ph), 139.0 (C1–Ph), 155.2 (C=), 166.8 (CO_{amide}), 172.4 (CO_{lactam}); HRMS: calcd for C₁₄H₁₇N₂O₂S [M+H]⁺ 277.1005, found 277.1005; Anal. Calcd for C₁₄H₁₆N₂O₂S: C, 60.85; H, 5.84; N, 10.14; S, 11.60; found: C, 61.15; H, 6.06; N, 10.13; S, 11.33.

5.2.3. (*Z*)-*Ethyl* (3-*methyl*-4-*oxothiazolidin*-2-*ylidene*)*ethanoate* (**1c**). Compound **1c** was obtained from **4** (1.17 g; 6.25 mmol; R=H, EWG=CO₂Et), K₂CO₃ (0.86 g; 6.25 mmol), CH₃I (0.98 g; 6.88 mmol; 1.1 equiv) in DMF (15 mL) according to the general procedure (reaction time 1.5 h) as a white solid (0.94 g; 73%); mp 99–100 °C; R_{f} =0.57 (toluene/ethyl acetate 7/3); IR (KBr): ν =2982, 2942, 1708, 1676, 1562, 1427, 1366, 1340, 1275, 1180, 1118, 805 cm⁻¹; ¹H NMR

(DMSO-*d*₆, 200 MHz): δ 1.20 (t, *J*=7.2 Hz, 3H, CH₃CH₂), 3.08 (s, 3H, NCH₃), 3.85 (s, 2H, CH₂S), 4.10 (q, *J*=7.2 Hz, 2H, CH₂O), 5.57 (s, 1H, =CH); ¹³C NMR (DMSO-*d*₆, 50.3 MHz): δ 14.5 (CH₃CH₂), 29.9 (NCH₃), 31.6 (CH₂S), 59.4 (CH₂O), 89.6 (=CH), 159.7 (C=), 167.2 (CO_{ester}), 172.7 (CO_{lactam}); ¹H NMR (CDCl₃, 200 MHz): δ 1.30 (t, *J*=7.2 Hz, 3H, CH₃CH₂), 3.17 (s, 3H, NCH₃), 3.72 (s, 2H, CH₂S), 4.22 (q, *J*=7.2 Hz, 2H, CH₂O), 5.49 (s, 1H, =CH); ¹³C NMR (CDCl₃, 50.3 MHz): δ 14.3 (CH₃CH₂), 29.9 (NCH₃), 31.8 (CH₂S), 60.0 (CH₂O), 90.7 (=CH), 158.6 (C=), 167.6 (CO_{ester}), 172.3 (CO_{lactam}); HRMS: calcd for C₈H₁₂NO₃S [M+H]⁺ 202.0532, found 202.0525; Anal. Calcd for C₈H₁₁NO₃S: C, 47.75; H, 5.51; N, 6.96; S, 15.93; found: C, 47.71; H, 5.40; N, 7.04; S, 16.12.

5.2.4. (Z)-(3-Methyl-4-oxothiazolidin-2-ylidene)-1-phenylethanone (1d). Compound 1d was obtained from 4 (1.64 g; 7.50 mmol; R=H, EWG=COPh), K₂CO₃ (1.04 g; 7.50 mmol), CH₃I (1.17 g; 8.25 mmol; 1.1 equiv) in DMF (15 mL) according to the general procedure (reaction time 1.5 h) as a pale yellow solid (1.67 g; 95%); mp 173–174 °C; *R_f*=0.39 (toluene/ethyl acetate 4/1); IR (ATR): *v*=3062, 3032, 2974, 2921, 1724, 1614, 1573, 1513, 1419, 1345, 1266, 1206, 1111, 1053, 909, 888, 745, 697 cm⁻¹; ¹H NMR (DMSO-*d*₆, 200 MHz): δ 3.25 (s, 3H, CH₃), 3.85 (s, 2H, CH₂S), 6.92 (s, 1H, =CH), 7.47-7.63 (m, 3H, *m*- and *p*-Ph), 8.01–8.06 (m, 2H, *o*-Ph); ¹³C NMR (DMSO-*d*₆, 50.3 MHz): δ 30.3 (CH₃), 31.5 (CH₂S), 95.5 (=CH), 127.7 (o-Ph), 128.8 (m-Ph), 132.4 (p-Ph), 138.4 (C1-Ph), 162.68 (C=), 172.2 (CO_{lactam}), 187.5 (CO_{ketone}); ¹H NMR (CDCl₃, 200 MHz): δ 3.30 (s, 3H, CH₃), 3.70 (s, 2H, CH₂S), 6.68 (s, 1H, =CH), 7.42-7.58 (m, 3H, m- and p-Ph), 7.92–7.97 (m, 2H, o-Ph); ¹³C NMR (CDCl₃, 50.3 MHz): δ 30.2 (CH₃), 31.7 (CH₂S), 95.7 (=CH), 127.5 (o-Ph), 128.5 (m-Ph), 132.2 (p-Ph), 138.3 (C1–Ph), 161.8 (C=), 172.7 (CO_{lactam}), 188.6 (CO_{ketone}); HRMS: calcd for C₁₂H₁₂NO₂S [M+H]⁺ 234.0583, found 234.0572; Anal. Calcd for C₁₂H₁₁NO₂S: C, 61.78; H, 4.75; N, 6.00; S, 13.74; found: C, 61.44; H, 4.75; N, 5.94; S, 14.01.

5.2.5. (*Z*)-(3,5-*Dimethyl*-4-oxothiazolidin-2-ylidene)-*N*-phenylethanamide (**1***j*). Compound **1***j* was obtained from **4** (248 mg; 1.00 mmol; R=Me, EWG=CONHPh), K₂CO₃ (138 mg; 1.00 mmol), CH₃I (213 mg; 1.50 mmol; 1.5 equiv) in DMF (5 mL) according to the general procedure (reaction time 2.5 h) as a white solid (0.234 g; 89%); mp 168–170 °C; *R*_f=0.30 (toluene/ethyl acetate 7/3); IR (ATR): *v*=3341, 1697, 1658, 1567, 1492, 1440, 1337, 1302, 1175, 1130, 1054, 761, 692 cm⁻¹; ¹H NMR (DMSO-d₆, 200 MHz): δ 1.46 (d, *J*=7.3 Hz, 3H, CH₃CH), 3.11 (s, 3H, NCH₃), 4.02 (q, *J*=7.3 Hz, 1H, CHS), 5.80 (s, 1H, =CH), 7.00 (t, *J*=7.3 Hz, 1H, *p*-Ph), 7.28 (m, 2H, *m*-Ph), 7.59 (d, *J*=7.2 Hz, 2H, o-Ph), 9.88 (s, 1H, NH_{amide}); ¹³C NMR (DMSO*d*₆, 50,3 MHz): δ 18.9 (CH₃CH), 29.6 (NCH₃), 39.8 (CHS), 93.5 (=CH), 118.7 (o-Ph), 122.8 (*p*-Ph), 128.9 (*m*-Ph), 139.9 (C1–Ph), 153.5 (C=), 165.1 (CO_{amide}), 175.2 (CO_{lactam}); HRMS: calcd for C₁₃H₁₅N₂O₂S [M+H]⁺ 263.0849, found 263.0855.

5.2.6. (*Z*)-(3,5-Dimethyl-4-oxothiazolidin-2-ylidene)-N-(2-phenylethyl) ethanamide (1k). Compound 1k was obtained from 4 (275 mg; 1.00 mmol; R=Me, EWG=CONH(CH₂)₂Ph), K₂CO₃ (138 mg; 1.00 mmol), CH₃I (170 mg; 1.20 mmol; 1.2 equiv) in DMF (4 mL) according to the general procedure (reaction time 1.5 h). Column chromatography (eluent: gradient toluene/ethyl acetate 100/0 to 60/ 40) gave pure **1k** as a colourless oil (234 mg; 87%). Crystallization from ethanol/water mixture 2/1 (v/v) gave a white solid, mp 87–89 °C; R_{f} =0.33 (toluene/ethyl acetate 3/2); IR (ATR): ν =3313, 3058, 2979, 2933, 1712, 1644, 1575, 1285, 1202, 1132, 1054, 794, 736, 702 $\rm cm^{-1};\,{}^1H$ NMR (DMSO-d₆, 200 MHz): δ 1.42 (d, J=7.2 Hz, 3H, CH₃CH), 2.72 (t, J=7.3 Hz, 2H, CH₂Ph), 3.03 (s, 3H, NCH₃), 3.28-3.38 (m, 2H, NCH₂), 3.94 (q, J=7.2 Hz, 1H, CHS), 5.59 (s, 1H, =CH), 7.17-7.33 (m, 5H, Ph), 7.82 (t, *J*=5.6 Hz, 1H, NH); ¹³C NMR (DMSO-*d*₆, 50.3 MHz): δ 19.1 (*C*H₃CH), 29.8 (NCH₃), 35.6 (CH₂Ph), 39.6 (NCH₂), 40.2 (CHS), 93.4 (=CH), 126.3 (p-Ph), 128.6 (o-Ph), 128.8 (m-Ph), 139.8 (C1-Ph), 151.2 (C=), 166.4 (CO_{amide}), 175.2 (CO_{lactam}); ¹H NMR (CDCl₃, 200 MHz): δ 1.58 (d,

J=7.1 Hz, 3H, *CH*₃CH), 2.85 (t, *J*=7.0 Hz, 2H, CH₂Ph), 3.10 (s, 3H, NCH₃), 3.53–3.63 (m, 2H, NCH₂), 3.83 (q, *J*=7.1 Hz, 1H, CHS), 5.32 (s, 1H, =CH), 5.58 (broad t, 1H, NH), 7.18–7.36 (m, 5H, Ph); ¹³C NMR (CDCl₃, 50,3 MHz): δ 19.0 (CH₃CH), 29.9 (NCH₃), 35.8 (CH₂Ph), 40.2 (NCH₂), 40.5 (CHS), 92.3 (=CH), 126.4 (*p*-Ph), 128.6 (*o*-Ph), 128.7 (*m*-Ph), 138.9 (C1–Ph), 153.6 (C=), 166.8 (CO_{amide}), 175.5 (CO_{lactam}); HRMS: calcd for C₁₅H₁₉N₂O₂S [M+H]⁺ 291.1162, found 291.1167.

5.2.7. (*Z*)-*Ethyl* (3,5-*dimethyl*-4-*oxothiazolidin*-2-*ylidene*)*ethanoate* (**1**). Compound **1** was obtained from **4** (R=Me, EWG=CO₂Et) as already described.³²

5.2.8. (Z)-(3,5-Dimethyl-4-oxothiazolidin-2-ylidene)-1-phenylethanone (1m). Compound 1m was obtained from 4 (350 mg; 1.50 mmol; R=H, EWG=COPh), K₂CO₃ (207 mg; 1.50 mmol), CH₃I (245 mg; 1.725 mmol; 1.15 equiv) in DMF (6 mL) according to the general procedure (reaction time 1.5 h). Column chromatography (eluent: gradient toluene/ethyl acetate 100/0 to 80/20) gave pure **1m** as a white solid (196 mg; 53%); mp 130–132 °C; R_f =0.52 (toluene/ethyl acetate 4/1); IR (ATR): ν =3394, 3061, 2980, 2938, 1711, 1624, 1516, 1347, 1224, 1052, 756, 704 $\rm cm^{-1};\,{}^{1}H$ NMR (DMSO-*d*₆, 200 MHz): δ 1.49 (d, *J*=7.2 Hz, 3H, CH₃CH), 3.27 (s, 3H, NCH₃), 4.08 (q, J=7.2 Hz, 1H, CHS), 6.93 (s, 1H, =CH), 7.47-7.63 (m, 3H, *m*- and *p*-Ph), 8.01–8.06 (m, 2H, o-Ph); ¹³C NMR (DMSO-*d*₆, 50.3 MHz): δ 18.4 (CH₃CH), 30.5 (NCH₃), 39.8 (CHS), 95.4 (=CH), 127.7 (o-Ph), 128.8 (m-Ph), 132.4 (p-Ph), 138.4 (C1-Ph), 160.9 (C=), 176.1 (CO_{lactam}), 187.6 (CO_{ketone}); ¹H NMR (CDCl₃, 200 MHz): δ 1.63 (d, *J*=7.3 Hz, 3H, CH₃CH), 3.31 (s, 3H, NCH₃), 3.88 (q, J=7.3 Hz, 1H, CHS), 6.67 (s, 1H, =CH), 7.42–7.57 (m, 3H, m- and p-Ph), 7.92–7.98 (m, 2H, o-Ph); ¹³C NMR (CDCl₃, 50.3 MHz): δ 18.6 (CH₃CH), 30.3 (NCH₃), 40.2 (CHS), 95.3 (=CH), 127.5 (o-Ph), 128.5 (m-Ph), 132.2 (p-Ph), 138.4 (C1-Ph), 160.4 (C=), 176.0 (CO_{lactam}), 188.6 (CO_{ketone}); HRMS: calcd for C₁₃H₁₄NO₂S [M+H]⁺ 248.0740, found 248.0746.

5.3. General procedure for the preparation of *N*-benzyl derivatives 1e–g, 1n,o and 1q,r

Benzyl bromide (1.2 mmol) was added to a stirred mixture of 4-oxothiazolidine **4** (1 mmol) and K₂CO₃ (1–1.05 mmol) in DMF (2–10 mL) and stirring was continued at rt until the disappearance of the starting material (TLC). The reaction mixture was neutralized with satd NH₄Cl solution. The aqueous phase was extracted with ethyl acetate ($3\times2-5$ mL) and the organic layer was washed with water ($3\times2-5$ mL), brine solution ($1\times2-5$ mL) and dried over Na₂SO₄. The solvent was removed under reduced pressure and the crude product was purified by column chromatography (gradient toluene/ethyl acetate).

5.3.1. (Z)-(3-Benzyl-4-oxothiazolidin-2-ylidene)-N-phenylethanamide (1e). Compound was 1e obtained from 4 (234.3 mg; 1.00 mmol; R=H, EWG=CONHPh), K₂CO₃ (138.2 mg; 1.00 mmol), benzyl bromide (205.3 mg, 1.20 mmol) in DMF (5 mL) according to the general procedure (reaction time 2.5 h). Column chromatography (eluent: gradient toluene/ethyl acetate 100/0 to 70/30) gave pure **1e** as a white solid (208.9 mg; 64%); mp 216–218 °C; *R*_f=0.64 (toluene/ethyl acetate 3/2); IR: v=3283, 3244, 3142, 1712, 1595, 1547, 1489, 1440, 1317, 1243, 1156, 861, 792, 752, 691 cm⁻¹; ¹H NMR (DMSO-*d*₆, 200 MHz): δ 3.92 (s, 2H, CH₂S), 4.85 (s, 2H, CH₂Ph), 5.78 (s, 1H, ==CH), 6.95–7.56 (m, 10H, 2× Ph), 9.82 (s, 1H, NH); ¹³C NMR (DMSO-*d*₆, 50.3 MHz): δ 31.4 (CH₂S), 46.3 (CH₂Ph), 94.4 (=CH), 119.0, 123.0, 127.0, 127.9, 129.1, 135.2, 139.8, 154.5 (C=), 165.3 (CO_{amide)}, 173.2 (CO_{lactam}); HRMS: calcd for C₁₈H₁₇N₂O₂S [M+H]⁺ 325.1005, found 325.1008; Anal. Calcd for C₁₈H₁₆N₂O₂S: C, 66.64; H, 4.97; N, 8.64; S, 9.88; found: C, 66.25; H, 5.07; N, 8.61; S, 9.72.

5.3.2. (*Z*)-*Ethyl* (3-*benzyl-4-oxothiazolidin-2-ylidene)ethanoate* (**1***f*). Compound **1***f* was obtained from **4** (187.2 mg; 1.00 mmol;

R=H, EWG=CO₂Et), K₂CO₃ (138.2 mg; 1.00 mmol), benzyl bromide (205.3 mg, 1.20 mmol) in DMF (5 mL) according to the general procedure (reaction time 2.5 h). Column chromatography (eluent: gradient toluene/ethyl acetate 100/0 to 85/15) gave pure 1f as a white solid (227.4 mg; 82%); mp 89–91 °C; *R_f*=0.52 (toluene/ethyl acetate 4/1); IR (KBr): v=2981, 2930, 1718, 1681, 1560, 1289, 1159, 1039, 967, 788, 691 cm⁻¹; ¹H NMR (DMSO- d_6 , 200 MHz): δ 1.15 (t, *I*=7.1 Hz, 3H, CH₃), 4.01 (s, 2H, CH₂S), 4.03 (q, *I*=7.1 Hz, 2H, CH₂O), 4.90 (s, 2H, CH₂Ph), 5.47 (s, 1H, =CH), 7.23–7.41 (m, 5H, Ph); ¹³C NMR (DMSO-d₆, 50.3 MHz): δ 14.4 (CH₃), 31.4 (CH₂S), 45.8 (CH₂Ph), 59.5 (CH₂O), 90.3 (=CH), 126.9 (o-Ph), 127.8 (p-Ph), 128.9 (m-Ph), 135.2 (C1–Ph), 158.6 (C=), 167.0 (CO_{ester}), 173.1 (CO_{lactam}); ¹H NMR (CDCl₃, 200 MHz): δ 1.25 (t, *J*=7.3 Hz, 3H, CH₃), 3.81 (s, 2H, CH₂S), 4.15 (q, J=7.3 Hz, 2H, CH₂O), 4.88 (s, 2H, CH₂Ph), 5.47 (s, 1H, =CH), 7.20–7.36 (m, 5H, Ph); 13 C NMR (CDCl₃, 50.3 MHz): δ 14.3 (CH₃), 31.6 (CH₂S), 46.8 (CH₂Ph), 60.0 (CH₂O), 91.8 (=CH), 126.8 (o-Ph), 128.0 (p-Ph), 128.9 (m-Ph), 134.0 (C1-Ph), 157.5 (C=), 167.6 (CO_{es-} ter), 172.6 (CO_{lactam}); HRMS: calcd for C₁₄H₁₆NO₃S [M+H]⁺ 278.0845, found 278.0854; Anal. Calcd for C₁₄H₁₅NO₃S: C, 60.63; H, 5.45; N, 5.05; S, 11.56; found: C, 60.36; H, 5.42; N, 5.04; S, 11.65.

5.3.3. (Z)-(3-Benzyl-4-oxothiazolidin-2-ylidene)-1-phenylethanone (1g). Compound 1g was obtained from 4 (328.9 mg; 1.50 mmol; R=H, EWG=COPh), K₂CO₃ (207.4 mg; 1.50 mmol), benzyl bromide (307.8 g, 1.80 mmol) in DMF (7 mL) according to the general procedure (reaction time 1 h). Column chromatography (eluent: gradient toluene/ethyl acetate 100/0 to 90/10) gave pure 1g as a white solid (412.3 mg; 89%); mp 169–170 °C; R_f=0.51 (toluene/ethyl acetate 4/1): IR: v=3059, 3031, 2968, 2930, 1708, 1633, 1575, 1515, 1348, 1216, 1176, 915, 883, 764, 695 cm⁻¹; ¹H NMR (DMSO- d_{6} , 200 MHz): δ 4.02 (s, 2H, CH₂S), 5.11 (s, 2H, CH₂Ph), 6.90 (s, 1H, =CH), 7.24–7.91 (m, 10H, 2× Ph); 13 C NMR (DMSO- d_6 , 50.3 MHz): δ 31.4 (CH₂S), 46.0 (CH₂Ph), 96.1 (=CH), 127.3, 127.5, 127.8, 128.9, 128.9, 132.4, 135.6, 138.8, 161.7 (C=), 173.7 (CO_{lactam}), 187.4 (CO_{ketone}); ¹H NMR (CDCl₃, 200 MHz): δ 3.81 (s, 2H, CH₂S), 5.01 (s, 2H, CH₂Ph), 6.67 (s, 1H, =CH), 7.26–7.79 (m, 10H, $2 \times$ Ph); ¹³C NMR (CDCl₃, 50.3 MHz): δ 31.6 (CH₂S), 47.2 (CH₂Ph), 97.0 (=CH), 126.9, 127.4, 128.2, 128.5, 129.1, 132.2, 134.2, 138.3, 160.6 (C=), 173.1 (CO_{lactam}), 188.6 (CO_{ketone}); HRMS: calcd for C₁₈H₁₆NO₂S [M+H]⁺ 310.0896, found 310.0890; Anal. Calcd for C18H15NO2S: C, 69.88; H, 4.89; N, 4.53; S, 10.36; found: C, 70.15; H, 4.94; N, 4.40; S, 10.28.

5.3.4. (Z)-(3-Benzyl-5-methyl-4-oxothiazolidin-2-ylidene)-N-phenylethanamide (1n). Compound 1n was obtained from 4 (124.2 mg; 0.50 mmol; R=Me, EWG=CONHPh), K₂CO₃ (69.1 mg; 0.50 mmol), benzyl bromide (102.6 mg, 0.60 mmol) in DMF (5 mL) according to general procedure (reaction time 2 h). Column chromatography (eluent: gradient toluene/ethyl acetate 100/0 to 80/20) gave pure **1n** as a white solid (111.6 mg; 66%); mp 166–167 °C; *R*_f=0.47 (toluene/ethyl acetate 4/1); IR (ATR): v=3291, 3056, 3030, 2976, 2932, 1717, 1644, 1560, 1493, 1440, 1317, 1159, 975, 731, 691 cm⁻¹; ¹H NMR (DMSO-d₆, 200 MHz): δ 1.52 (d, J=7.2 Hz, 3H, CH₃CH), 4.17 (q, J=7.2 Hz, 1H, CHS), 4.86 (s, 2H, CH₂Ph), 5.77 (s, 1H, =CH), 6.98 (t, J=7.3 Hz, 1H, p-Ph), 7.22-7.43 (m, 7H, Ph), 7.53 (d, J=7.8 Hz, 2H, o-Ph), 9.84 (s, 1H, NH_{amide}); ¹³C NMR (DMSO- d_6 , 50.3 MHz): δ 19.1 (CH₃), 39.5 (CHS), 46.2 (CH₂Ph), 94.1 (=CH), 118.8, 122.9, 126.7, 127.7, 128.9, 135.1 (C1–Ph), 139.6 (C1–Ph), 152.6 (C=), 164.9 (CO_{a-} mide), 175.8 (CO_{lactam}); HRMS: calcd for $C_{19}H_{19}N_2O_2S$ [M+H]⁺ 339.1162, found 339.1157.

5.3.5. (*Z*)-*Ethyl* (3-*benzyl*-5-*methyl*-4-*oxothiazolidin*-2-*ylidene*)*ethanoate* (**10**). Compound **10** was obtained from **4** (150.9 mg; 0.75 mmol; R=Me, EWG=CO₂Et), K₂CO₃ (103.6 mg; 0.75 mmol), benzyl bromide (153.9 mg, 0.90 mmol) in DMF (6 mL) according to the general procedure (reaction time 2.5 h). Column chromatography (eluent: gradient toluene/ethyl acetate 100/0 to 90/10) gave

Z. Džambaski et al. / Tetrahedron xxx (2013) 1–12

pure **1o** as a white solid (200.8 mg; 92%); mp 96–97 °C; *R*_f=0.48 (toluene/ethyl acetate 9/1); IR (KBr): v=3028, 2976, 2936, 1714, 1686, 1568, 1372, 1342, 1290, 1156, 1041, 969, 784, 693 cm⁻¹; ¹H NMR (DMSO-*d*₆, 200 MHz): δ 1.14 (t, *J*=7.0 Hz, 3H, CH₃CH₂), 1.54 (d, J=7.1 Hz, 3H, CH₃CH), 4.03 (q, J=7.0 Hz, 1H, CH₂O), 4.27 (q, J=7.1 Hz, 2H, CHS), 4.91 (s, 2H, CH₂Ph), 5.86 (s, 1H, =CH), 7.21-7.41 (m, 5H, Ph); 13 C NMR (DMSO- d_6 , 50.3 MHz): δ 14.4 (CH₃CH₂), 19.0 (CH₃CH), 40.1 (CHS), 45.9 (CH₂Ph), 59.5 (CH₂O), 90.3 (=CH), 126.8 (o-Ph), 127.8 (p-Ph), 129.0 (m-Ph), 135.2 (C1-Ph), 156.8 (C=), 166.9 (CO_{ester}), 176.1 (CO_{lactam}); ¹H NMR (CDCl₃, 200 MHz): δ 1.24 (t, J=7.2 Hz, 3H, CH₃CH₂), 1.68 (d, J=7.2 Hz, 3H, CH₃CH), 4.00 (q, *I*=7.2 Hz, 1H, CHS), 4.15 (q, *I*=7.2 Hz, 2H, CH₂O), 4.85 (d, *I*=15.8 Hz, 1H, CH_AH_BPh), 4.90 (d, *J*=15.8 Hz, 1H, CH_AH_BPh), 5.44 (s, 1H, =CH), 7.17–7.40 (m, 5H, Ph); ¹³C NMR (CDCl₃, 50.3 MHz): δ 14.3 (CH₃CH₂), 19.3 (CH₃CH), 40.4 (CHS), 46.8 (CH₂Ph), 60.0 (CH₂O), 91.4 (=CH), 126.7 (o-Ph), 127.9 (p-Ph), 128.9 (m-Ph), 134.2 (C1-Ph), 156.2 (C=), 167.6 (CO_{ester}), 175.9 (CO_{lactam}); HRMS: calcd for C₁₅H₁₈NO₃S [M+H]⁺ 292.1002, found 292.1001; Anal. Calcd for C₁₅H₁₇NO₃S: C, 61.83; H, 5.88; N, 4.81; S, 11.01; found: C, 61.50; H, 5.64; N, 4.90; S, 10.83.

5.3.6. (Z)-(3-Benzyl-5-ethoxycarbonylmethyl-4-oxothiazolidin-2ylidene)-N-phenylethanamide (1q). Compound 1q was obtained from 4 (160.2 mg; 0.50 mmol; R=CH₂CO₂Et, EWG=CONHPh), K₂CO₃ (69.1 mg; 0.50 mmol), benzyl bromide (102.6 mg, 0.60 mmol) in DMF (5 mL) according to the general procedure (reaction time 2.5 h). Column chromatography (eluent: gradient toluene/ethyl acetate 100/0 to 75/25) gave pure 1q as a white solid (157.3 mg; 77%); *R_f*=0.36 (toluene/ethyl acetate 4/1); mp 209–211 °C; IR (ATR): v=3307, 3137, 2985, 2927, 1715, 1648, 1598, 1569, 1497, 1444, 1326, 1164, 1030, 765, 697 cm⁻¹; ¹H NMR (DMSOd₆, 200 MHz): δ 1.18 (t, *J*=7.2 Hz, 3H, CH₃), 3.02 (dd, *J*_{AB}=17.4 Hz, J_{AX}=7.2 Hz, 1H, CH_AH_BCO₂Et), 3.14 (dd, J_{AB}=17.4 Hz, J_{BX}=4.6 Hz, 1H, CH_AH_BCO₂Et), 4.11 (q, J=7.2 Hz, 2H, CH₂O), 4.42 (dd, J_{AX}=7.2 Hz, J_{BX} =4.6 Hz, 1H, CH_XS), 4.87 (s, 2H, CH₂Ph), 5.75 (s, 1H, =CH), 6.98 (t, J=7.3 Hz, 1H, p-Ph), 7.22–7.43 (m, 7H, Ph), 7.53 (d, J=7.8 Hz, 2H, o-Ph), 9.82 (s, 1H, NH_{amide}); 13 C NMR (DMSO- d_6 , 50.3 MHz): δ 14.2 (CH₃), 36.8 (CH₂CO₂Et), 40.9 (CHS), 46.5 (CH₂Ph), 60.8 (CH₂O), 94.3 (=CH), 118.8, 122.9, 126.9, 127.7, 128.9, 135.1, 139.7, 153.2 (C=), 165.0 (CO_{amide}), 170.3 (CO_{ester}), 174.2 (CO_{lactam}); HRMS: calcd for C₂₂H₂₂N₂NaO₄S [M+Na]⁺ 433.1192, found 433.1177.

5.3.7. (Z)-(3-Benzyl-5-ethoxycarbonylmethyl-4-oxothiazolidin-2ylidene)-1-phenylethanone (1r). Compound 1r was obtained from 4 (305.4 mg; 1.00 mmol; R=CH₂CO₂Et, EWG=COPh), K₂CO₃ (138.2 mg; 1.00 mmol), benzyl bromide (205.3 mg, 1.20 mmol) in DMF (7 mL) according to the general procedure (reaction time 1 h). Column chromatography (eluent: gradient toluene/ethyl acetate 100/0 to 80/20) gave pure **1r** as a white solid (367.1 mg; 93%); mp 105–106 °C; R_f =0.57 (toluene/ethyl acetate 4/1); IR: ν =3422, 3063, 2990, 1715, 1609, 1573, 1501, 1486, 1371, 1214, 1172, 915, 763, 694 cm⁻¹; ¹H NMR (CDCl₃, 200 MHz): δ 1.26 (t, *J*=7.3 Hz, 3H, CH₃), 3.02 (dd, J_{AB}=14.4 Hz, J_{AX}=8.4 Hz, 1H, CH_AH_BCO₂Et), 3.21 (dd, J_{AB}=14.4 Hz, J_{BX}=4.0 Hz, 1H, CH_AH_BCO₂Et), 4.19 (q, J=7.3 Hz, 2H, CH₂O), 4.26 (dd, J_{AX}=8.4 Hz, J_{BX}=4.0 Hz, 1H, CH_XS), 5.03 (s, 2H, CH₂Ph), 6.66 (s, 1H, =CH), 7.26–7.78 (m, 10H, $2 \times$ Ph); ¹³C NMR (CDCl₃, 50.3 MHz): δ 14.1 (CH₃), 37.2 (CH₂CO₂Et), 41.3 (CHS), 47.4 (CH₂Ph), 61.4 (CH₂O), 97.0 (=CH), 126.9, 127.4, 128.1, 128.5, 129.0, 132.2, 134.2, 138.3, 159.3 (C=), 169.7 (CO_{ester}), 174.7 (CO_{lactam}), 186.6 (CO_{ketone}); HRMS: calcd for $C_{22}H_{22}NO_4S$ [M+H]⁺ 396.1264, found 396.1272; Anal. Calcd for C₂₂H₂₁NO₄S: C, 66.82; H, 5.35; N, 3.54; S, 8.11; found: C, 66.90; H, 5.46; N, 5.37; S, 8.30.

5.4. Synthesis of *N*-bromoalkyl derivatives 1h and 1p

Compounds **1h** and **1p** were obtained as already described.³³

5.5. (*Z*)-Ethyl (3-ethoxycarbonylmethyl-4-oxothiazolidin-2-ylidene)ethanoate (1i)

Compound **1i** was obtained from **4** (187.2 mg; 1.00 mmol; R=H, EWG=CO₂Et), K₂CO₃ (145.1 mg; 1.05 mmol), ethyl 2-bromoacetate (200.4 mg, 1.20 mmol, 0.14 mL) in DMF (2 mL) at rt (reaction time 1 h). The reaction mixture was then diluted with CHCl₃ (3 mL) and water (3 mL) was added. The organic layer was separated and the water layer extracted with $CHCl_3$ (3×2 mL). The chloroform extracts were combined, washed with water $(3 \times 3 \text{ mL})$, brine $(1 \times 3 \text{ mL})$ and dried over Na₂SO₄. The solvent was evaporated under reduced pressure. Thus obtained product was stirred with *n*-hexane for 2 h and filtered to give pure 1i as a white solid (111.6 mg; 93%); mp 103–104 °C; *R_f*=0.65 (toluene/ethyl acetate 7/3); IR (ATR): *v*=2985, 1746, 1687, 1571, 1301, 1219, 1167, 1047, 1026, 976, 794 $\rm cm^{-1};\ ^1H$ NMR (CDCl₃, 200 MHz): δ 1.19 (t, J=7.2 Hz, 6H, 2× CH₃), 3.79 (s, 2H, CH₂S), 4.20 (q, J=7.2 Hz, 2H, CH₂O), 4.24 (q, J=7.2 Hz, 2H, CH₂O), 4.40 (s, 2H, NCH₂), 5.33 (s, 1H, =CH); ¹³C NMR (CDCl₃, 50.3 MHz): δ 14.0 (CH₃), 14.3 (CH₃), 31.4 (CH₂S), 44.2 (NCH₂), 60.1 (CH₂O), 62.1 (CH₂O), 90.7 (=CH), 157.3 (C=), 166.0 (CO_{ester}), 167.3 (CO_{ester}), 172.1 (CO_{lactam}); HRMS: calcd for C₁₁H₁₆NO₅S [M+H]⁺ 274.0744, found 274.0735.

5.6. General procedure for the oxidation of thiazolidine derivatives 1

A solution of *m*-CPBA (1.5–2.0 mmol) in CH₂Cl₂ was added dropwise within 5 min to a solution of thiazolidine **1** (1.0 mmol) in CH₂Cl₂ at 0 °C. The mixture was stirred at 0 °C for additional 30–60 min until the disappearance of the starting material (TLC). After the addition of 10% aq Na₂S₂O₃ (5–15 mL), the solution was extracted with CH₂Cl₂ (3×2–5 mL). The combined organic layers were washed with 10% K₂CO₃ (2×2–5 mL), water (2×2–5 mL) and finally with brine (1×2–5 mL). The organic layer was dried over Na₂SO₄ and solvent was removed under reduced pressure. Thus obtained crude product was purified by column chromatography.

5.6.1. (Z)-(3-Methyl-1,4-dioxothiazolidin-2-ylidene)-N-phenylethanamide (5a). Compound 5a was obtained from 1a (300 mg; 1.20 mmol; 18 mL CH₂Cl₂) and *m*-CPBA (428 mg; 2.48 mmol; 24 mL CH₂Cl₂) according to the general procedure (reaction time 30 min). Column chromatography (eluent: gradient toluene/acetone 100/ 0 to 30/70) gave pure 5a as a white solid (248 mg; 75%);mp 228–231 °C (decomposes); *R*_f=0.39 (toluene/acetone 1/1); IR (KBr): *v*=3303, 3263, 3198, 3097, 3054, 2931, 1711, 1677, 1625, 1552, 1493, 1439, 1339, 1108, 1041, 845, 771, 696 cm⁻¹; ¹H NMR (DMSO-*d*₆, 200 MHz): δ 3.11 (s, 3H, CH₃), 3.50 (d, *J*=17.4 Hz, 1H, CH_aH_bS), 4.10 (d, J=17.4 Hz, 1H, CH_aH_bS), 6.04 (s, 1H, =CH), 7.08 (t, J=7.3 Hz, 1H, p-Ph), 7.35 (m, 2H, m-Ph), 7.66 (d, J=7.4 Hz, 2H, o-Ph), 10.35 (s, 1H, NH_{amide}); ¹³C NMR (DMSO-*d*₆, 50.3 MHz): δ 29.3 (CH₃), 52.1 (CH₂S), 102.6 (=CH), 119.3 (o-Ph), 123.8 (p-Ph), 129.2 (m-Ph), 139.2 (C1-Ph), 156.8 (C=), 162.3 (CO_{amide}), 170.1 (CO_{lactam}); HRMS: calcd for C₁₂H₁₃N₂O₃S [M+H]⁺ 265.0641, found 265.0637.

5.6.2. (*Z*)-(3-*Methyl*-1,4-*dioxothiazolidin*-2-*ylidene*)-*N*-(2-*phenylethyl*) *ethanamide* (*5b*). Compound **5b** was obtained from **1b** (156 mg; 0.60 mmol; 15 mL CH₂Cl₂) and *m*-CPBA (214 mg; 0.90 mmol; 20 mL CH₂Cl₂) according to the general procedure (reaction time 30 min). Column chromatography (eluent: gradient toluene/acetone 100/0 to 30/70) gave pure **5b** as a white solid (149 mg; 85%);mp 139–141 °C (decomposes); *R*_f=0.27 (toluene/acetone 1/1); IR (KBr): *v*=3519, 3421, 3239, 3072, 2934, 1724, 1652, 1605, 1286, 1124, 1040, 850, 705 cm⁻¹; ¹H NMR (DMSO-*d*₆, 200 MHz): δ 2.77 (t, *J*=7.3 Hz, 2H, CH₂Ph), 3.03 (s, 3H, CH₃), 3.36–3.46 (m, 3H, NCH₂ and CH_AH_BS), 4.03 (d, *J*=17.4 Hz, 1H, CH_AH_BS), 5.86 (s, 1H, =CH), 7.17–7.35 (m, 5H, Ph), 8.33 (t,

J=5.3 Hz, 1H, NH_{amide}); ¹³C NMR (DMSO-*d*₆, 50.3 MHz): δ 29.2 (CH₃), 35.3 (CH₂Ph), NCH₂ is covered by DMSO, 52.1 (CH₂S), 102.8 (=CH), 126.4 (*p*-Ph), 128.7 (*o*-Ph), 128.9 (*m*-Ph), 139.6 (C1–Ph), 155.2 (C=), 163.6 (CO_{amide}), 170.0 (CO_{lactam}); HRMS: calcd for C₁₄H₁₇N₂O₃S [M+H]⁺ 293.0954, found 293.0960.

5.6.3. (Z)-Ethyl (3-methyl-1,4-dioxothiazolidin-2-ylidene)ethanoate (5c). Compound 5c was obtained from 1c (151 mg; 0.75 mmol; 6 mL CH₂Cl₂) and *m*-CPBA (267 mg; 1.125 mmol; 10 mL CH₂Cl₂) according to the general procedure (reaction time 60 min). Column chromatography (eluent: gradient toluene/acetone 100/0 to 70/30) gave pure **5c** as a pale yellow solid (128 mg; 80%);mp 128–129 °C; *R*_f=0.36 (toluene/acetone 7/3); IR (KBr): *v*=3046, 2924, 1711, 1619, 1276, 1185, 1116, 1040, 847 cm⁻¹; ¹H NMR (DMSO-*d*₆, 200 MHz): δ 1.25 (t, J=7.1 Hz, 3H, CH₃CH₂), 3.09 (s, 3H, NCH₃) 3.54 (d, J=17.4 Hz, 1H, CH_AH_BS), 4.12 (d, J=17.4 Hz, 1H, CH_AH_BS), 4.20 (q, J=7.2 Hz, 2H, CH₂O), 5.87 (s, 1H, =CH); ¹³C NMR (DMSO- d_6 , 50.3 MHz): δ 14.4 (CH₃CH₂), 29.4 (NCH₃), 52.2 (CH₂S), 60.6 (CH₂O), 98.3 (=CH), 160.6 (C=), 164.9 (CO_{ester}), 170.3 (CO_{lactam}); ¹H NMR (CDCl₃, 200 MHz): δ 1.34 (t, J=7.2 Hz, 3H, CH₃CH₂), 3.19 (s, 3H, NCH₃) 3.60 (d, J=17.7 Hz, 1H, CH_AH_BS), 3.86 (d, J=17.7 Hz, 1H, CH_AH_BS), 4.31 (q, J=7.2 Hz, 2H, CH₂O), 5.72 (s, 1H, =CH); ¹³C NMR (CDCl₃, 50.3 MHz): δ 14.0 (CH₃CH₂), 29.4 (NCH₃), 51.5 (CH₂S), 61.3 (CH₂O), 100.2 (=CH), 158.4 (C=), 164.6 (CO_{ester}), 168.6 (CO_{lactam}); HRMS: calcd for C₈H₁₂NO₄S [M+H]⁺ 218.0418, found 218.0486; Anal. Calcd for C₈H₁₁NO₄S: C, 44.23; H, 5.10; N, 6.45; S, 14.76; found: C, 44.33; H, 5.19; N, 6.43; S. 14.48.

5.6.4. (Z)-(3-Methyl-1.4-dioxothiazolidin-2-ylidene)-1-phenylethanone (5d). Compound 5d was obtained from 1d (175 mg; 0.75 mmol; 12 mL CH₂Cl₂) and *m*-CPBA (269 mg; 1.13 mmol; 20 mL CH₂Cl₂) according to the general procedure (reaction time 40 min). Column chromatography (eluent: gradient toluene/acetone 100/0 to 70/30) gave pure 5d as a pale yellow solid (187 mg; 74%);mp 177-178 °C (decomposes); $R_f=0.32$ (toluene/acetone 4/1); IR (ATR): $\nu=3064$, 3004, 2941, 1729, 1645, 1565, 1279, 1220, 1119, 1042, 907, 776, 697 cm⁻¹; ¹H NMR (DMSO-*d*₆, 200 MHz): δ 3.35 (s, 3H, CH₃), 3.56 (d, J=17.4 Hz, 1H, CH_aH_bS), 4.17 (d, J=17.4 Hz, 1H, CH_aH_bS), 7.09 (s, 1H, =CH), 7.54–7.73 (m, 3H, m- and p-Ph), 8.13–8.17 (m, 2H, o-Ph); ¹³C NMR (DMSO-*d*₆, 50.3 MHz): δ 29.8 (CH₃), 51.9 (CH₂S), 101.9 (=CH), 128.6 and 129.1 (o- and m-Ph), 133.6 (p-Ph), 137.6 (C1-Ph), 161.1 (C=), 170.6 (CO_{lactam}), 187.7 (CO_{ketone}); ¹H NMR (CDCl₃, 200 MHz): δ 3.31 (s, 3H, CH₃), 3.66 (d, *J*=17.7 Hz, 1H, CH_aH_bS), 3.87 (d, *J*=17.6 Hz, 1H, CH_aH_bS), 6.81 (s, 1H, =CH), 7.48-7.68 (m, 3H, m- and p-Ph), 8.00-8.06 (m, 2H, o-Ph); ¹³C NMR (CDCl₃, 50.3 MHz): δ 29.8 (CH₃), 51.4 (CH₂S), 102.7 (=CH), 128.4 and 128.6 (o- and *m*-Ph), 133.6 (*p*-Ph), 137.3 (C1-Ph), 159.2 (C=), 169.0 (CO_{lactam}), 187.6 (CO_{ketone}) HRMS: calcd for C₁₂H₁₂NO₃S [M+H]⁺ 250.0532, found 250.0542; Anal. Calcd for C₁₂H₁₁NO₃S: C, 57.82; H, 4.45; N, 5.62; S, 12.86; found: C, 57.58; H, 4.53: N. 5.54: S. 12.41.

5.6.5. (*Z*)-(3-Benzyl-1,4-dioxothiazolidin-2-ylidene)-N-phenylethanamide (**5e**). Compound **5e** was obtained from **1e** (97.3 mg; 0.30 mmol; 4 mL CH₂Cl₂) and *m*-CPBA (107.1 mg; 0.45 mmol; 5 mL CH₂Cl₂) according to the general procedure (reaction time 60 min). Column chromatography (eluent: gradient toluene/acetone 100/ 0 to 70/30) gave pure **5e** as a white solid (73.1 mg; 72%);mp 142–144 °C (decomposes); R_f =0.33 (toluene/acetone 7/3); IR (ATR): ν =3036, 1722, 1671, 1613, 1548, 1311, 1495, 1446, 1164, 1018, 758, 696 cm⁻¹; ¹H NMR (DMSO-*d*₆, 500 MHz): δ 3.61 (d, *J*=17.2 Hz, 1H, CH_aH_bS), 4.26 (d, *J*=17.2 Hz, 1H, CH_aH_bS), 4.83 (d, *J*=16.0 Hz, 1H, CH_aH_bPh), 4.91 (d, *J*=16.0 Hz, 1H, CH_aH_bPh), 6.00 (s, 1H, =CH), 7.07 (t, *J*=5.5 Hz, 1H, *p*-Ph), 7.30–7.40 (m, 7H, 2× Ph), 7.61 (d, *J*=8.0 Hz, 2H, *o*-Ph), 10.25 (s, 1H, NH_{amide}); ¹³C NMR (DMSO-*d*₆, 125.8 MHz): δ 45.4 (CH₂Ph), 51.7 (CH₂S), 102.8 (=CH), 119.2, 123.6, 126.6, 127.6, 128.7, 128.8, 134.2, 138.8, 155.7 (C=), 161.9 (CO_{amide}), 170.4 (CO_{lactam}); HRMS: calcd for $C_{18}H_{16}N_2NaO_3S$ [M+Na]⁺ 363.0774, found 363.0775.

5.6.6. (Z)-Ethyl (3-benzyl-1,4-dioxothiazolidin-2-ylidene)ethanoate (5f). Compound 5f was obtained from 1f (69.3 mg: 0.25 mmol: 3 mL CH₂Cl₂) and *m*-CPBA (76.0 mg; 0.35 mmol; 4 mL CH₂Cl₂) according to the general procedure (reaction time 30 min). Column chromatography (eluent: gradient toluene/acetone 100/0 to 80/20) gave pure **5f** as a white solid (69.6 mg; 95%);mp 128–129 °C; *R*_f=0.30 (toluene/acetone 4/1); IR (ATR): *v*=2985, 2933, 1706, 1616, 1292, 1156, 1048, 834, 733, 698 cm⁻¹; ¹H NMR (DMSO-d₆, 200 MHz): δ 1.20 (t, J=7.1 Hz, 3H, CH₃), 3.68 (d, J=17.4 Hz, 1H, CH_AH_BS), 4.14 (q, J=7.1 Hz, 2H, CH₂O), 4.29 (d, J=17.4 Hz, 1H, CH_AH_BS), 4.93 (m, 2H, CH₂Ph), 5.78 (s, 1H, =CH), 7.25–7.40 (m, 5H, Ph); ¹³C NMR (DMSO-*d*₆, 50.3 MHz): δ 14.2 (CH₃), 45.0 (CH₂Ph), 52.1 (CH₂S), 60.6 (CH₂O), 98.9 (=CH), 127.0 (o-Ph), 127.9 (p-Ph), 128.9 (*m*-Ph), 134.5 (C1–Ph), 159.4 (C=), 164.7 (CO_{ester}), 170.9 (CO_{lactam}); ¹H NMR (CDCl₃, 200 MHz): δ 1.28 (t, *J*=7.3 Hz, 3H, CH₃), 3.65 (d, J=17.5 Hz, 1H, CH_AH_BS), 3.89 (d, J=17.5 Hz, 1H, CH_AH_BS), 4.23 (q, J=7.3 Hz, 2H, CH₂O), 4.87 (m, 2H, CH₂Ph), 5.66 (s, 1H, =CH), 7.19–7.36 (m, 5H, Ph); ¹³C NMR (CDCl₃, 50.3 MHz): δ 14.0 (CH₃), 46.4 (CH₂Ph), 51.4 (CH₂S), 61.3 (CH₂O), 101.2 (=CH), 126.8 (o-Ph), 128.2 (p-Ph), 129.1 (m-Ph), 132.8 (C1-Ph), 157.3 (C=), 164.5 (CO_{es-} ter), 169.2 (CO_{lactam}); HRMS: calcd for $C_{14}H_{16}NO_4S$ [M+H]⁺ 294.0795, found 294.0786.

5.6.7. (Z)-(3-Benzvl-1.4-dioxothiazolidin-2-vlidene)-1-phenvlethanone (5g). Compound 5g was obtained from 1g (77.3 mg; 0.25 mmol; 3 mL CH₂Cl₂) and *m*-CPBA (76.3 mg; 0.375 mmol; 4 mL CH₂Cl₂) according to the general procedure (reaction time 40 min). Column chromatography (eluent: gradient toluene/acetone 100/0 to 90/10) gave pure 5g as a yellow solid (55.8 mg; 69%);mp 173-175 °C (decomposes); *R*_f=0.30 (toluene/acetone 4/1); IR (KBr): *v*=3062, 3006, 2932, 1723, 1649, 1563, 1322, 1176, 1038, 1018, 892, 780, 740, 697 cm⁻¹; ¹H NMR (CDCl₃, 200 MHz): δ 3.71 (d, J=17.5 Hz, 1H, CH_aH_bS), 3.93 (d, J=17.5 Hz, 1H, CH_aH_bS), 4.94 (d, J=15.7 Hz, 1H, CH_aH_bPh), 5.06 (d, J=15.7 Hz, 1H, CH_aH_bPh), 6.75 (s, 1H, =CH), 7.28–7.77 (m, 10H, 2× Ph); ¹³C NMR (CDCl₃, 50.3 MHz): δ 46.9 (CH₂Ph), 51.4 (CH₂S), 104.3 (=CH), 127.0, 128.2, 128.5, 128.8, 129.3, 133.1, 133.5, 137.2, 157.7 (C=), 169.5 (CO_{lactam}), 187.6 (CO_{ketone}); HRMS: calcd for C₁₈H₁₆NO₃S [M+H]⁺ 326.0845, found 326.0845; Anal. Calcd for C₁₈H₁₅NO₃S: C, 66.44; H, 4.65; N, 4.30; S, 9.85; found: C, 66.12; H, 4.80; N, 4.28; S, 9.48.

5.6.8. (Z)-Ethyl (3-(3-bromopropyl)-1,4-dioxothiazolidin-2-ylidene) ethanoate (5h). Compound 5h was obtained from 1h (77.0 mg; 0.25 mmol; 3 mL CH₂Cl₂) and *m*-CPBA (89.2 mg; 0.375 mmol; 4 mL CH₂Cl₂) according to the general procedure (reaction time 60 min). Column chromatography (eluent: gradient toluene/acetone 100/ 0 to 75/25) gave pure **5h** as a colourless oil (67.7 mg; 84%); *R*_f=0.47 (toluene/acetone 7/3); IR (ATR): v=2985, 1708, 1614, 1306, 1237, 1181, 1144, 1048, 836, 735 cm⁻¹; ¹H NMR (CDCl₃, 500 MHz): δ 1.35 (t, J=7.2 Hz, 3H, CH₃), 2.17–2.24 (m, 2H, CH₂), 3.40–3.48 (m, 3H, CH₂Br), 3.61 (d, J=18.0 Hz, 1H, CH_AH_BS), 3.80 (d, J=18.0 Hz, 1H, CH_AH_BS), 3.77–3.83 (m, 1H, NCH_aH_b), 3.87–3.93 (m, 1H, NCH_aH_b), 4.32 (q, J=7.2 Hz, 2H, CH₂O), 5.84 (s, 1H, =CH); ¹³C NMR (CDCl₃, 125.8 MHz): δ 14.1 (CH₃), 28.8 (CH₂CH₂CH₂), 29.5 (CH₂Br), 41.8 (NCH₂), 51.5 (CH₂S), 61.5 (CH₂O), 100.5 (=CH), 157.5 (C=), 164.6 (CO_{ester}), 168.9 (CO_{lactam}); HRMS: calcd for $C_{10}H_{15}BrNO_4S [M+H]^+$ 323.9900, found 323.9901.

5.6.9. (*Z*)-*Ethyl*-(3-*ethoxycarbonylmethyl*-1,4-*dioxothiazolidin*-2*ylidene)ethanoate* (**5***i*). Compound **5***i* was obtained from **1***i* (82.0 mg; 0.30 mmol; 5 mL CH₂Cl₂) and *m*-CPBA (107.0 mg; 0.45 mmol; 6 mL CH₂Cl₂) according to the general procedure (reaction time 40 min). Column chromatography (eluent: gradient

toluene/acetone 100/0 to 80/20) gave pure **5i** as a white solid (73.0 mg; 84%);mp 87–89 °C; R_f =0.32 (toluene/acetone 4/1); IR (ATR): ν =3065, 2985, 2938, 1739, 1708, 1620, 1359, 1293, 1213, 1162, 1052, 971, 841 cm⁻¹; ¹H NMR (CDCl₃, 500 MHz): δ 1.30 (t, *J*=7.2 Hz, 3H, CH₃), 1.34 (t, *J*=7.2 Hz, 3H, CH₃), 3.65 (d, *J*=17.8 Hz, 1H, CH_AH_BS), 3.91 (d, *J*=17.8 Hz, 1H, CH_AH_BS), 4.25 (q, *J*=7.2 Hz, 2H, CH₂O), 4.31 (q, *J*=7.2 Hz, 2H, CH₂O), 4.12 (d, *J*=17.2 Hz, 1H, NCH_aH_b), 4.72 (d, *J*=17.2 Hz, 1H, NCH_aH_b), 5.58 (s, 1H, =CH); ¹³C NMR (CDCl₃, 125.8 MHz): δ 13.9 (CH₃), 14.0 (CH₃), 44.0 (NCH₂), 51.4 (CH₂S), 61.5 (CO_{ester}), 168.6 (CO_{lactam}); HRMS: calcd for C₁₁H₁₆NO₆S [M+H]⁺ 290.0693, found 290.0696; Anal. Calcd for C₁₁H₁₅NO₆S: C, 45.67; H, 5.23; N, 4.84; S, 11.08; found: C, 45.84; H, 5.11; N, 4.82; S, 10.77.

5.6.10. (Z)-(3,5-Dimethyl-1,4-dioxothiazolidin-2-ylidene)-N-phenylethanamide (5i). Compound 5i was obtained from 1i (98.4 g; 0.375 mmol; 8 mL CH₂Cl₂) and *m*-CPBA (178.6 mg; 0.75 mmol; 6 mL CH_2Cl_2) according to the general procedure (reaction time 60 min). Column chromatography (eluent: gradient toluene/acetone 100/ 0 to 60/40) gave pure **5j** as a pale yellow solid (74.3 mg; 71%; *syn*/ anti 85/15);mp 204–207 °C (decomposes); Rf=0.42 and 0.47 (toluene/acetone 3/2); IR (ATR): v=3330, 2914, 1722, 1673, 1606, 1538, 1311, 1039, 820, 758, 697 cm⁻¹; ¹H NMR (DMSO-*d*₆, 500 MHz): *syn*-**5j** δ 1.36 (d, *J*=7.5 Hz, 3H, CH₃CH), 3.12 (s, 3H, NCH₃), 3.89 (q, J=7.5 Hz, 1H, CHS), 6.11 (s, 1H, =CH), 7.09 (t, J=7.2 Hz, 1H, p-Ph), 7.35 (m, 2H, *m*-Ph), 7.66 (d, *J*=8.0 Hz, 2H, *o*-Ph), 10.34 (s, 1H, NH); anti-5j δ 1.40 (d, J=8.0 Hz, 3H, CH₃CH), 3.12 (s, 3H, NCH₃), 3.65 (q, *I*=8.0 Hz, 1H, CHS), 6.13 (s, 1H, =CH), 7.09 (t, *I*=7.2 Hz, 1H, *p*-Ph), 7.35 (m, 2H, *m*-Ph), 7.66 (d, *J*=8.0 Hz, 2H, o-Ph), 10.34 (s, 1H, NH); ¹³C NMR (DMSO-*d*₆, 125.8 MHz): *syn*-**5j** δ 6.6 (CH₃CH), 29.4 (NCH₃), 52.5 (CHS), 103.2 (=CH), 119.1 (o-Ph), 123.6 (p-Ph), 128.9 (m-Ph), 138.9 (C1–Ph), 154.7 (C=), 162.0 (CO_{amide}), 172.5 (CO_{lactam}); anti-5j δ 11.7 (CH₃CH), 29.4 (NCH₃), 59.5 (CHS), 103.8 (=CH), 119.1 (o-Ph), 123.6 (p-Ph), 128.9 (m-Ph), 138.9 (C1-Ph), 154.7 (C=), 162.0 (CO_{a-} mide), 172.5 (CO_{lactam}); HRMS: calcd for $C_{13}H_{14}N_2NaO_3S$ [M+Na]⁺ 301.0617, found 301.0624.

5.6.11. (Z)-(3,5-Dimethyl-1,4-dioxothiazolidin-2-ylidene)-N-(2phenylethyl)ethanamide (5k). Compound 5k was obtained from 1k (147.5 mg; 0.51 mmol; 8 mL CH₂Cl₂) and *m*-CPBA (241.8 mg; 1.01 mmol; 7 mL CH₂Cl₂) according to the general procedure (reaction time 60 min). Column chromatography (eluent: gradient toluene/acetone 100/0 to 60/40) gave pure 5k as a colourless oil (100.8 mg; 65%; *syn/anti* 86/14); *R*_f=0.28 (toluene/acetone 3/2); IR (ATR): *v*=3299, 3066, 2937, 1721, 1660, 1610, 1288, 1046, 848, 735, 700 cm⁻¹; ¹H NMR (DMSO- d_6 , 500 MHz): *syn*-**5k** δ 1.33 (d, J=7.5 Hz, 3H, CH₃), 2.79 (t, J=7.3 Hz, 2H, CH₂Ph), 3.04 (s, 1H, NCH₃), 3.40–3.46 (m, 2H, NCH₂), 3.82 (q, J=7.5 Hz, 1H, CHS), 5.93 (s, 1H, =CH), 7.20–7.32 (m, 5H, Ph), 8.33 (t, J=5.7 Hz, 1H, NH); anti-**5k** δ 1.35 (d, *J*=7.5 Hz, 3H, CH₃), 2.79 (t, *J*=7.3 Hz, 2H, CH₂Ph), 3.04 (s, 1H, NCH₃), 3.40–3.46 (m, 2H, NCH₂), 3.56 (q, *J*=7.5 Hz, 1H, CHS), 5.96 (s, 1H, =CH), 7.20-7.32 (m, 5H, Ph), 8.33 (t, J=5.7 Hz, 1H, NH); ¹³C NMR (DMSO- d_6 , 125.8 MHz): syn-**5k** δ 6.5 (CH₃CH), 29.3 (NCH₃), 35.0 (CH₂Ph), 40.3 (NCH₂), 52.5 (CHS), 103.4 (=CH), 126.2 (p-Ph), 128.4 and 128.6 (o- and m-Ph), 139.3 (C1-Ph), 153.2 (C=), 163.3 (CO_{amide}), 172.4 (CO_{lactam}); anti-**5k** δ 11.7 (CH₃CH), 29.3 (NCH₃), 40.0 (CH₂Ph), 40.3 (NCH₂), 59.3 (CHS), 104.1 (=CH), 126.5 (p-Ph), 128.6 and 128.7 (o- and m-Ph), 138.6 (C1-Ph), 153.2 (C=), 163.6 (CO_{amide}), 172.4 (CO_{lactam}); ¹H NMR (CDCl₃, 500 MHz): *syn*-**5k** δ 1.57 (d, *J*=7.5 Hz, 3H, CH₃), 2.85 (t, *J*=7.2 Hz, 2H, CH₂Ph), 3.11 (s, 1H, NCH₃), 3.31 (q, J=7.5 Hz, 1H, CHS), 3.50-3.68 (m, 2H, NCH₂), 5.76 (s, 1H, ==CH), 6.54 (t, J=5.7 Hz, 1H, NH), 7.18-7.30 (m, 5H, Ph); anti-**5k** δ 1.49 (d, *J*=7.5 Hz, 3H, CH₃), 2.85 (t, *J*=7.2 Hz, 2H, CH₂Ph), 3.11 (s, 1H, NCH₃), 3.50 (q, J=7.5 Hz, 1H, CHS), 3.50-3.68 (m, 2H, NCH₂), 5.78 (s, 1H, =CH), 6.54 (t, J=5.7 Hz, 1H, NH), 7.18–7.30 (m, 5H, Ph); ¹³C NMR (CDCl₃, 125.8 MHz): syn-**5k** δ 6.8 (CH₃CH), 29.7 (NCH₃), 35.4 (CH₂Ph), 40.9 (NCH₂), 53.2 (CHS), 103.8 (=CH), 126.5 (*p*-Ph), 128.6 and 128.7 (*o*- and *m*-Ph), 138.6 (C1–Ph), 153.5 (C=), 163.6 (CO_{amide}), 171.7 (CO_{lactam}); *anti*-**5k** δ 12.4 (CH₃CH), 29.6 (NCH₃), 35.4 (CH₂Ph), 40.9 (NCH₂), 59.6 (CHS), 104.1 (=CH), 126.5 (*p*-Ph), 128.6 and 128.7 (*o*- and *m*-Ph), 138.6 (C1–Ph), 153.5 (C=), 163.6 (CO_{amide}), 171.7 (CO_{lactam}); HRMS: calcd for C₁₅H₁₈N₂NaO₃S [M+Na]⁺ 329.0930, found 329.09304.

5.6.12. (Z)-Ethyl (3,5-dimethyl-1,4-dioxothiazolidin-2-ylidene)ethanoate (51). Compound 51 was obtained from 11 (150.7 mg; 0.70 mmol; 12 mL CH₂Cl₂) and *m*-CPBA (333.3 mg; 1.40 mmol; 10 mL CH₂Cl₂) according to the general procedure (reaction time 40 min). Column chromatography (eluent: gradient toluene/acetone 100/0 to 80/20) gave pure **51** as a white solid (150.6 mg; 93%; syn/anti 86/14);mp 97–99 °C; R_f=0.37 (toluene/acetone 4/1); IR (KBr): *v*=2983, 2937, 1705, 1614, 1262, 1183, 1131, 1056, 839, 732, 688 cm⁻¹; ¹H NMR (DMSO-*d*₆, 500 MHz): *syn*-**5l** δ 1.26 (t, *J*=7.0 Hz, 3H, CH₃CH₂), 1.36 (d, J=7.5 Hz, 3H, CH₃CH), 3.10 (s, 3H, NCH₃), 3.95 (q, J=7.5 Hz, 1H, CHS), 4.22 (q, J=7.0 Hz, 2H, CH₂O), 5.93 (s, 1H, =CH); anti-**5l** δ 1.26 (t, J=7.0 Hz, 3H, CH₃CH₂), 1.44 (d, *J*=7.5 Hz, 3H, CH₃CH), 3.10 (s, 3H, NCH₃), 3.74 (q, *J*=7.5 Hz, 1H, CHS), 4.20 (q, *J*=7.0 Hz, 2H, CH₂O), 5.94 (s, 1H, =CH); ¹³C NMR (DMSO-*d*₆, 125.8 MHz): syn-51 & 6.6 (CH₃CH), 14.1 (CH₃CH₂), 29.5 (NCH₃), 52.5 (CHS), 60.4 (CH₂O), 98.9 (=CH), 158.4 (C=), 164.6 (CO_{ester}), 172.9 (CO_{lactam}); anti-5l δ 11.5 (CH₃CH), 14.1 (CH₃CH₂), 29.5 (NCH₃), 59.9 (CHS), 60.4 (CH₂O), 99.3 (=CH), 158.6 (C=), 164.5 (CO_{ester}), 172.1 (CO_{lactam}); ¹H NMR (CDCl₃, 500 MHz): *syn*-**5l** δ 1.35 (t, *J*=7.1 Hz, 3H, CH₃CH₂), 1.63 (d, *J*=7.4 Hz, 3H, CH₃CH), 3.18 (s, 3H, NCH₃), 3.42 (q, J=7.4 Hz, 1H, CHS), 4.32 (q, J=7.1 Hz, 2H, CH₂O), 5.73 (s, 1H, =CH); anti-51 & 1.35 (t, J=7.1 Hz, 3H, CH₃CH₂), 1.56 (d, J=7.9 Hz, 3H, CH₃CH), 3.18 (s, 3H, NCH₃), 3.63 (q, *I*=7.9 Hz, 1H, CHS), 4.32 (q, J=7.1 Hz, 2H, CH₂O), 5.76 (s, 1H, =CH); ¹³C NMR (CDCl₃, 125.8 MHz): syn-**5l** δ 6.7 (CH₃CH), 14.1 (CH₃CH₂), 29.7 (NCH₃), 53.3 (CHS), 61.3 (CH₂O), 100.6 (=CH), 157.1 (C=), 164.6 (CO_{ester}), 172.0 (CO_{lactam}); anti-51 & 12.3 (CH₃CH), 14.1 (CH₃CH₂), 29.8 (NCH₃), 59.9 (CHS), 61.3 (CH₂O), 100.9 (=CH), 157.1 (C=), 164.6 (CO_{ester}), 172.0 (CO_{lactam}); HRMS: calcd for C₉H₁₄NO₄S [M+H]⁺ 232.0638, found 232.0635.

5.6.13. (Z)-(3,5-Dimethyl-1,4-dioxothiazolidin-2-ylidene)-1-phenyle thanone (5m). Compound 5m was obtained from 1m (74.2 mg; 0.30 mmol; 4 mL CH₂Cl₂) and *m*-CPBA (142.8 mg; 0.60 mmol; 7 mL CH₂Cl₂) according to the general procedure (reaction time 45 min). Column chromatography (eluent: gradient toluene/acetone 100/ 0 to 75/25) gave pure 5m as a white solid (57.8 mg; 73%; syn/anti 85/15); mp 132–134 °C (decomposes); *R*_f=0.30 and 0.40 (toluene/ acetone 4/1); IR (ATR): v=3065, 2915, 1729, 1644, 1558, 1348, 1224, 1038, 780, 702 cm⁻¹; ¹H NMR (DMSO- d_6 , 500 MHz): syn-**5m** δ 1.38 (d, *I*=7.5 Hz, 3H, CH₃), 3.27 (s, 1H, NCH₃), 3.98 (q, *I*=7.5 Hz, 1H, CHS), 7.15 (s, 1H, =CH), 7.59 (m, 2H, m-Ph), 7.69 (t, J=7.5 Hz, 1H, p-Ph), 8.16 (d, J=8.2 Hz, 2H, o-Ph); anti-**5m** δ 1.48 (d, J=7.7 Hz, 3H, CH₃), 3.25 (s, 1H, NCH₃), 3.78 (q, J=7.7 Hz, 1H, CHS), 7.14 (s, 1H, =CH), 7.59 (m, 2H, m-Ph), 7.69 (t, J=7.5 Hz, 1H, p-Ph), 8.16 (d, J=8.2 Hz, 2H, o-Ph); ¹³C NMR (DMSO- d_6 , 125.8 MHz): *syn*-**5m** δ 6.8 (CH₃CH), 29.9 (NCH₃), 52.1 (CHS), 102.6 (=CH), 128.3 (o-Ph), 128.8 (m-Ph), 133.4 (*p*-Ph), 137.3 (C1–Ph), 158.8 (C=), 173.3 (CO_{lactam}), 187.5 (CO_{ketone}); *anti*-**5m** δ 11.5 (CH₃CH), 29.9 (NCH₃), 59.7 (CHS), 102.9 (=CH), 128.3 (o-Ph), 128.8 (m-Ph), 133.4 (p-Ph), 137.4 (C1-Ph), 159.4 (C=), 172.2 (CO_{lactam}), 187.5 (CO_{ketone}); ¹H NMR (CDCl₃, 500 MHz): syn-5m δ 1.65 (d, J=7.5 Hz, 3H, CH₃), 3.28 (s, 1H, NCH₃), 3.44 (q, J=7.5 Hz, 1H, CHS), 6.80 (s, 1H, =CH), 7.51 (m, 2H, m-Ph), 7.61 (t, J=7.2 Hz, 1H, p-Ph), 8.01 (d, J=8.5 Hz, 2H, o-Ph); anti-5m δ 1.59 (d, J=7.5 Hz, 3H, CH₃), 3.30 (s, 1H, NCH₃), 3.67 (q, J=7.5 Hz, 1H, CHS), 6.84 (s, 1H, =CH), 7.51 (m, 2H, m-Ph), 7.61 (t, J=7.2 Hz, 1H, p-Ph), 8.01 (d, J=8.5 Hz, 2H, o-Ph); 13 C NMR (CDCl₃, 125.8 MHz): syn-**5m** δ 7.0

(CH₃CH), 30.0 (NCH₃), 53.1 (CHS), 103.1 (=CH), 128.3 and 128.8 (*o*-and *m*-Ph), 133.5 (*p*-Ph), 137.5 (C1–Ph), 157.8 (C=), 172.5 (CO_{lactam}), 187.6 (CO_{ketone}); *anti*-**5m** δ 12.4 (CH₃CH), 30.0 (NCH₃), 59.7 (CHS), 103.3 (=CH), 128.3 and 128.8 (*o*- and *m*-Ph), 133.5 (*p*-Ph), 137.5 (C1–Ph), 158.0 (C=), 172.3 (CO_{lactam}), 187.6 (CO_{ketone}); HRMS: calcd for C₁₃H₁₃NNaO₃S [M+Na]⁺ 286.0508, found 286.0514.

5.6.14. (Z)-(3-Benzvl-5-methyl-1.4-dioxothiazolidin-2-vlidene)-Nphenylethanamide (5n). Compound 5n was obtained from 1n (67.7 mg; 0.20 mmol; 6 mL CH₂Cl₂) and *m*-CPBA (95.2 mg; 0.40 mmol; 5 mL CH₂Cl₂) according to the general procedure (reaction time 30 min). Column chromatography (eluent: gradient toluene/acetone 100/0 to 75/25) gave pure 5n as a colourless oil (53.9 mg; 76%; *syn/anti* 87/13); *R*_f=0.36 and 0.42 (toluene/acetone 7/3); IR (ATR): v=3300, 3198, 3173, 3060, 2931, 1726, 1671, 1613, 1549, 1467, 1445, 1322, 1163, 1040, 1010, 735, 697 cm⁻¹; ¹H NMR (DMSO-*d*₆, 500 MHz): *syn*-**5n** δ 1.42 (d, *J*=7.5 Hz, 3H, CH₃), 4.11 (q, J=7.5 Hz, 1H, CHS), 4.84 (d, J=16.5 Hz, 1H, CH_AH_BPh), 4.90 (d, J=16.5 Hz, 1H, CH_AH_BPh), 6.07 (s, 1H, =CH), 7.06–7.62 (m, 10H, 2× Ph), 10.28 (s, 1H, NH); anti-**5n** δ 1.46 (d, J=8.0 Hz, 3H, CH₃), 3.80 (q, J=8.0 Hz, 1H, CHS), 4.76 (d, J=16.3 Hz, 1H, CH_AH_BPh), 4.96 (d, J=16.3 Hz, 1H, CH_AH_BPh), 6.12 (s, 1H, =CH), 7.06–7.62 (m, 10H, 2× Ph), 10.28 (s, 1H, NH); 13 C NMR (DMSO- d_6 , 125.8 MHz): syn-**5n** δ 6.6 (CH₃), 45.7 (CH₂Ph), 52.6 (CHS), 103.8 (=CH), 119.3, 123.7, 126.6, 127.6, 128.8, 128.9, 134.2, 138.8, 153.8 (C=), 161.9 (CO_{amide}), 173.1 (CO_{lactam}); anti-**5n** δ 11.7 (CH₃), 45.7 (CH₂Ph), 59.2 (CHS), 104.6 (=CH), 119.2, 125.3, 126.7, 128.2, 134.2, 137.4, 153.8 (C=), 161.9 (CO_{amide}) , 173.1 (CO_{lactam}) ; HRMS: calcd for $C_{19}H_{19}N_2O_2S [M+H]^+$ 355.1116, found 355.1120.

5.6.15. (Z)-Ethyl (3-benzyl-5-methyl-1,4-dioxothiazolidin-2-ylidene) ethanoate (50). Compound 50 was obtained from 10 (58.3 mg; 0.20 mmol; 3 mL CH₂Cl₂) and *m*-CPBA (81.2 mg; 0.40 mmol; 4 mL CH_2Cl_2) according to the general procedure (reaction time 30 min). Column chromatography (eluent: gradient toluene/acetone 100/ 0 to 85/15) gave pure **50** as a colourless oil (52.3 mg; 85%; syn/anti 86/14); $R_{f}=0.30$ (toluene/acetone 9/1); IR (ATR): $\nu=3063$, 2983, 2937, 1727, 1707, 1616, 1293, 1154, 1056, 836, 734, 698 cm⁻¹; ¹H NMR (DMSO-*d*₆, 500 MHz): *syn*-**50** δ 1.21 (t, *J*=7.3 Hz, 3H, CH₃CH₂), 1.42 (d, J=7.0 Hz. 3H, CH₃CH), 4.12-4.19 (m, 3H, CHS and CH₂O), 4.92 (d, *J*=16.0 Hz, 1H, CH_AH_BPh), 4.94 (d, *J*=16.0 Hz, 1H, CH_AH_BPh), 5.85 (s, 1H, =CH), 7.25 (d, J=7.0 Hz, 2H, o-Ph), 7.29 (t, J=7.3 Hz, 1H, *p*-Ph), 7.35 (m, 2H, *m*-Ph); *anti*-**50** δ 1.20 (t, *J*=7.3 Hz, 3H, CH₃CH₂), 1.48 (d, J=7.5 Hz, 3H, CH₃CH), 3.90 (q, J=7.5 Hz, 1H, CHS), CH₂O is covered by syn-50, 4.82 (d, J=16.0 Hz, 1H, CH_AH_BPh), 4.99 (d, *J*=16.0 Hz, 1H, CH_A*H*_BPh), 5.88 (s, 1H, =CH), 7.24–7.37 (m, 5H, Ph); ¹³C NMR (DMSO- d_6 , 125.8 MHz): syn-**50** δ 6.6 (CH₃CH), 14.0 (CH₃CH₂), 45.1 (CH₂Ph), 52.7 (CHS), 60.5 (CH₂O), 99.6 (=CH), 126.7 (o-Ph), 127.6 (p-Ph), 128.7 (m-Ph), 134.3 (C1-Ph), 157.2 (C=), 164.5 (CO_{ester}), 173.4 (CO_{lactam}); anti-**50** δ 11.5 (CH₃CH), 14.0 (CH₃CH₂), 45.2 (CH₂Ph), 59.6 (CHS), 60.5 (CH₂O), 100.3 (=CH), 126.8 (o-Ph), 127.7 (p-Ph), 128.7 (m-Ph), 134.3 (C1-Ph), 157.3 (C=), 164.3 (CO_{es-} ter), 173.0 (CO_{lactam}); ¹H NMR (CDCl₃, 500 MHz): syn-**50** δ 1.29 (t, J=7.3 Hz, 3H, CH₃CH₂), 1.67 (d, J=7.5 Hz, 3H, CH₃CH), 3.48 (q, J=7.5 Hz, 1H, CHS), 4.24 (q, J=7.3 Hz, 2H, CH₂O), 4.79 (d, J=15.5 Hz, 1H, CH_AH_BPh), 4.94 (d, *J*=15.5 Hz, 1H, CH_AH_BPh), 5.67 (s, 1H, =CH), 7.18–7.39 (m, 5H, Ph); anti-**50** δ 1.29 (t, J=7.3 Hz, 3H, CH₃CH₂), 1.58 (d, J=7.5 Hz, 3H, CH₃CH), 3.71 (q, J=7.5 Hz, 1H, CHS), 4.24 (q, J=7.3 Hz, 2H, OCH₂), 4.81 (d, J=15.5 Hz, 1H, CH_AH_BPh), 4.91 (d, *J*=15.5 Hz, 1H, CH_AH_BPh), 5.70 (s, 1H, =CH), 7.18–7.39 (m, 5H, Ph); ¹³C NMR (CDCl₃, 125.8 MHz): *syn*-**50** δ 6.7 (CH₃CH), 14.0 (CH₃CH₂), 46.8 (CH₂Ph), 53.5 (CHS), 61.3 (CH₂O), 101.7 (=CH), 126.7 (o-Ph), 128.2 (p-Ph), 129.1 (m-Ph), 133.1 (C1-Ph), 155.9 (C=), 164.5 (CO_{ester}), 172.4 (CO_{lactam}); anti-**50** δ 12.4 (CH₃CH), 14.0 (CH₃CH₂), 46.6 (CH₂Ph), 59.6 (CHS), 61.3 (CH₂O), 102.1 (=CH), assignation for Ph is not certain, 133.0 (C1-Ph), 156.0 (C=), 164.5 (CO_{ester}), 172.8 (CO_{lactam}); HRMS: calcd for $C_{15}H_{18}NO_4S \ [M+H]^+$ 308.0951, found 308.0963.

5.6.16. (Z)-Ethyl (3-(3-bromopropyl)-5-methyl-1,4-dioxothiazolidin-2-ylidene)ethanoate (5p). Compound 5p was obtained from 1p (64.4 mg; 0.20 mmol; 4 mL CH₂Cl₂) and *m*-CPBA (95.1 mg; 0.40 mmol; 4 mL CH₂Cl₂) according to the general procedure (reaction time 35 min). Column chromatography (eluent: gradient toluene/acetone 100/0 to 85/15) gave pure 5p as a colourless oil (57.5 mg; 85%; syn/anti 87/13); Rf=0.39 (toluene/acetone 4/1); IR (ATR): v=2983, 2939, 1709, 1615, 1306, 1236, 1182, 1145, 1059, 838 cm⁻¹; ¹H NMR (CDCl₃, 500 MHz): syn-**5p** δ 1.35 (t, *J*=7.0 Hz, 3H, CH₃CH₂), 1.62 (d, J=7.0 Hz, 3H, CH₃CH), 2.16-2.21 (m, 2H, CH₂CH₂CH₂), 3.39 (q, *J*=7.0 Hz, 1H, CHS), 3.37–3.44 (m, 2H, CH₂Br), 3.76–3.82 (m, 1H, NCH_aH_b), 3.85–3.91 (m, 1H, NCH_aH_b), 4.32 (q, J=7.0 Hz, 2H, CH₂O), 5.85 (s, 1H, =CH); anti-**5p** δ 1.35 (t, J=7.0 Hz, 3H, CH₃CH₂), 1.54 (d, J=8.0 Hz, 3H, CH₃CH), 2.16-2.21 (m, 2H, CH₂CH₂CH₂), 3.37-3.44 (m, 2H, CH₂Br), 3.63 (q, J=8.0 Hz, 1H, CHS), 3.76-3.82 (m, 1H, NCH_aH_b), 3.85-3.91 (m, 1H, NCH_aH_b), 4.32 (q, J=7.0 Hz, 2H, CH₂O), 5.88 (s, 1H, =CH); ¹³C NMR (CDCl₃, 125.8 MHz): *syn*-**5p** δ 6.6 (CH₃CH), 14.2 (CH₃CH₂), 28.9 (CH₂CH₂CH₂), 29.5 (CH₂Br), 41.9 (NCH₂), 53.5 (CHS), 61.4 (CH₂O), 100.8 (=CH), 156.1 (C=), 164.6 (CO_{ester}), 172.3 (CO_{lactam}); anti-**5p** δ 12.2 (CH₃CH), 14.2 (CH₃CH₂), 28.9 (CH₂CH₂CH₂), 29.5 (CH₂Br), 41.9 (NCH₂), 59.6 (CHS), 61.4 (CH₂O), 101.1 (=CH), 156.1 (C=), 164.6 (CO_{ester}), 172.3 (CO_{lactam}); HRMS: calcd for C₁₁H₁₇BrNO₄S [M+Na]⁺ 338.0056, found 338.0041.

5.6.17. (Z)-(3-Benzvl-5-ethoxvcarbonvlmethvl-1.4-dioxothiazolidin-2-vlidene)-N-phenvlethanamide (5a). Compound 5a was obtained from 1q (102.6 mg; 0.25 mmol; 5 mL CH₂Cl₂) and m-CPBA (119.0 mg; 0.50 mmol; 8 mL CH₂Cl₂) according to the general procedure (reaction time 45 min). Column chromatography (eluent: gradient toluene/acetone 100/0 to 70/30) gave pure 5q as a colourless oil (88.4 mg; 83%; syn/anti 79/21); Rf=0.42 and 0.61 (toluene/acetone 7/3); IR (KBr): v=3259, 3196, 3061, 2981, 2930, 1723, 1673, 1602, 1549, 1323, 1165, 1018, 839, 756, 695 cm⁻¹; ¹H NMR (DMSO-*d*₆, 500 MHz): *syn*-**5q** δ 1.23 (t, *J*=7.0 Hz, 3H, CH₃), 2.94 (dd, J_{AB}=17.8 Hz, J_{AX}=10.5 Hz, 1H, CH_AH_BCO₂Et), 3.09 (dd, JAB=17.8 Hz, JBX=4.0 Hz, 1H, CHAHBCO2Et), 4.16 (q, J=7.0 Hz, 2H, CH₂O), 4.47 (dd, J_{AX}=10.5 Hz, J_{BX}=4.0 Hz, 1H, CH_XS), 4.86 (d, J=16.2 Hz, 1H, CH_AH_BPh), 4.90 (d, J=16.2 Hz, 1H, CH_AH_BPh), 6.08 (s, 1H, =CH), 7.06-7.64 (m, 10H, 2× Ph), 10.29 (s, 1H, NH); anti-5q δ 1.15 (t, J=7.2 Hz, 3H, CH₃), 3.23 (dd, J_{AB}=18.4 Hz, J_{AX}=5.5 Hz, 1H, CH_AH_BCO₂Et), 3.54 (dd, J_{AB}=18.4 Hz, J_{BX}=4.2 Hz, 1H, CH_AH_BCO₂Et), 3.93 (m, J_{AX}=5.5 Hz, J_{BX}=4.2 Hz, 1H, CH_XS), 4.03-4.11 (q, J=7.2 Hz, 2H, CH₂O), CH_AH_BPh are covered by syn-5q, 6.08 (s, 1H, =CH), 7.06–7.64 (m, 10H, $2 \times$ Ph), 10.26 (s, 1H, NH); ¹³C NMR (DMSO- d_6 , 125.8 MHz): syn-5q δ 14.0 (CH₃), 27.4 (CH₂CO₂Et), 45.7 (CH₂Ph), 53.8 (CHS), 60.8 (CH₂O), 103.9 (=CH), 119.2, 123.7, 126.6, 127.6, 128.7, 128.8, 134.0, 138.8, 154.2 (C=), 161.9 (CO_{amide}), 170.2 and 171.6 (CO_{lactam} and CO_{ester}); anti-**5q** δ 13.8 (CH₃), 31.8 (CH₂CO₂Et), 45.7 (CH₂Ph), 61.2 (CH₂O), 61.6 (CHS), 102.8 (=CH), 119.2, 123.6, 125.3, 126.8, 127.7, 128.2, 134.1, 138.8, 154.8 (C=), 161.8 (CO_{amide}), 170.7 and 171.3 (CO_{lactam} and CO_{ester}); HRMS: calcd for C₂₂H₂₃N₂O₅S [M+H]⁺ 427.1322, found 427.1313.

5.6.18. (*Z*)-(3-Benzyl-5-ethoxycarbonylmethyl-1,4-dioxothiazolidin-2-ylidene)-1-phenylethanone (**5***r*). Compound **5***r* was obtained from **1***r* (79.1 mg; 0.20 mmol; 3 mL CH₂Cl₂) and *m*-CPBA (81.2 mg; 0.40 mmol; 4 mL CH₂Cl₂) according to the general procedure (reaction time 90 min). Column chromatography (eluent: gradient toluene/ acetone 100/0 to 60/40) gave pure **5***r* as a colourless oil (58.7 mg; 71%; syn/anti 77/23); *R*_f=0.30 (toluene/acetone 7/3); IR (KBr): *v*=3063, 3032, 2980, 2930, 1720, 1650, 1575, 1320, 1292, 1175, 1042, 1019, 778, 740, 699 cm⁻¹; ¹H NMR (CDCl₃, 500 MHz): syn-**5***r* δ 1.30 (t, *J*=7.0 Hz, 3H, CH₃), 3.23 (dd, *J*_{AB}=18.5 Hz, *J*_{AX}=9.5 Hz, 1H, CH_AH_BCO₂Et), 3.27

12

Z. Džambaski et al. / Tetrahedron xxx (2013) 1–12

(dd, J_{AB}=18.5 Hz, J_{BX}=4.3 Hz, 1H, CH_AH_BCO₂Et), 4.01 (dd, J_{AX}=9.5 Hz, J_{BX}=4.3 Hz, 1H, CH_XS), 4.23 (q, J=7.0 Hz, 2H, CH₂O), 4.86 (d, J=15.5 Hz, 1H, CH_AH_BPh), 5.11 (d, J=15.5 Hz, 1H, CH_AH_BPh), 6.78 (s, 1H, =CH), 7.26–7.74 (m, 10H, 2× Ph); anti-**5r** δ 1.21 (t, *J*=7.0 Hz, 3H, CH₃), 3.41 (dd, J_{AB}=18.5 Hz, J_{AX}=4.3 Hz, 1H, CH_AH_BCO₂Et), 3.44 (dd, J_{AB}=18.5 Hz, J_{BX}=4.3 Hz, 1H, CH_AH_BCO₂Et), 3.75 (t, J_{AX}=J_{BX}=4.3 Hz, 1H, CH_XS), 4.07–4.14 (m, 2H, CH₂O), 4.90 (d, *J*=16.0 Hz, 1H, CH_AH_BPh), 5.16 (d, I=16.0 Hz, 1H, CH_AH_BPh), 6.75 (s, 1H, =CH), 7.26-7.74 (m, 10H, 2× Ph); ¹³C NMR (CDCl₃, 125.8 MHz): syn-**5r** δ 14.1 (CH₃), 27.3 (CH₂CO₂Et), 47.1 (CH₂Ph), 54.6 (CHS), 61.6 (CH₂O), 105.4 (=CH), 126.9, 128.2, 128.7, 129.2, 133.2, 133.4, 137.2, 156.4 (C=), 170.3 and 171.5 (CO_{lactam} and CO_{ester}), 187.4 (CO_{ketone}); anti-5r δ 13.9 (CH₃), 32.3 (CH₂CO₂Et), 47.4 (CH₂Ph), 62.1 and 62.2 (CH₂O and CHS), 103.3 (=CH), 127.0, 128.4, 128.6, 129.1, 133.1, 133.2, 137.4, 157.6 (C=), 170.4 and 171.3 (CO_{lactam} and CO_{ester}), 187.4 (CO_{ketone}); ¹H NMR (DMSO-d₆, 500 MHz): syn-5r δ 1.22 (t, J=7.0 Hz, 3H, CH₃), 2.95 (dd, J_{AB}=17.8 Hz, J_{AX}=10.5 Hz, 1H, CH_AH_BCO₂Et), 3.10 (dd, J_{AB}=17.8 Hz, J_{BX}=4.0 Hz, 1H, CH_AH_BCO₂Et), 4.14 (q, J=7.0 Hz, 2H, CH₂O), 4.48 (dd, J_{AX}=10.5 Hz, J_{BX}=4.0 Hz, 1H, CH_XS), 5.10 (s, 2H, CH₂Ph), 7.10 (s, 1H, =CH), 7.27–7.97 (m, 10H, 2× Ph); ¹³C NMR (DMSO-d₆, 125.8 MHz): syn-5r δ 14.1 (CH₃), 27.7 (CH₂CO₂Et), 44.6 (CH₂Ph), 53.7 (CHS), 61.0 (CH₂O), 103.6 (=CH), 127.2, 128.2, 128.8, 129.0, 133.7, 134.5, 137.3, 158.1 (C=), 170.3 and 172.4 (CO_{lactam} and CO_{ester}), 187.6 (CO_{ketone}); anti-5r δ 13.5 (CH₃), 31.8 (CH₂CO₂Et), 45.4 (CH₂Ph), 61.3 and 61.5 (CH₂O and CHS), 102.6 (=CH), assignation for Ph is not certain, 158.7 (C=), 170.9 and 171.7 (CO_{lactam} and CO_{ester}), 187.2 (CO_{ketone}); HRMS: calcd for C₂₂H₂₂NO₅S [M+H]⁺ 412.1213, found 412.1211.

Acknowledgements

This work was supported by the Ministry of Education, Science and Technological Development of the Republic of Serbia, Grant No. 172020 and Deutscher Akademischer Austauschdienst (DAAD) project ID: 504 252 70.

Supplementary data

¹H NMR spectra for *syn*- and *anti*-**51**, absolute energies and *x*, *y*, *z* coordinates of the optimized structures. Supplementary data related to this article can be found at http://dx.doi.org/10.1016/j.tet.2013.05.087.

References and notes

- (a) Jain, A. K.; Vaidya, A.; Ravichandran, V.; Kashaw, S. K.; Agrawal, R. K. Bioorg. Med. Chem. 2012, 20, 3378; (b) Prabhakar, Y. S.; Solomon, V. R.; Gupta, M. K.; Katti, S. B. Top. Heterocycl. Chem. 2006, 4, 161.
- (a) Vintonyak, V. V.; Warburg, K.; Kruse, H.; Grimme, S.; Hübel, K.; Rauh, D.; Waldmann, H. *Angew. Chem., Int. Ed.* **2010**, 49, 5902; (b) Küçükgüzel, Ş. G.; Oruç, E. E.; Rollas, S.; Şahin, F.; Özbek, A. *Eur. J. Med. Chem.* **2002**, *37*, 197.
- (a) Rawal, R. K.; Katti, S. B.; Kaushik-Basu, N.; Arora, P.; Pan, Z. *Bioorg. Med. Chem. Lett.* **2008**, *18*, 6110; (b) Hoye, T. R.; Ayyad, S.-E. N.; Eklov, B. M.; Hashish, N. E.; Shier, W. T.; El Sayed, K. A.; Hamann, M. T. *J. Am. Chem. Soc.* **2002**, *124*, 7405.
- Allen, S.; Newhouse, B.; Anderson, A. S.; Fauber, B.; Allen, A.; Chantry, D.; Eberhardt, C.; Odingo, J.; Burgess, L. E. Bioorg. Med. Chem. Lett. 2004, 14, 1619
- Ahmed, S. A.; Odde, S.; Daga, P. R.; Bowling, J. J.; Mesbah, M. K.; Youssef, D. T.; Khalifa, S. I.; Doerksen, R. J.; Hamann, M. T. Org. Lett. 2007, 9, 4773.
- Knutsen, L. J. S.; Hobbs, C. J.; Earnshaw, C. G.; Fiumana, A.; Gilbert, J.; Mellor, S. L.; Radford, F.; Smith, N. J.; Birch, P. J.; Burley, J. R.; Ward, S. D. C.; James, I. F. *Bioorg. Med. Chem. Lett.* **2007**, *17*, 662.
- Zhou, H.; Wu, S.; Zhai, S.; Liu, A.; Sun, Y.; Li, R.; Zhang, Y.; Ekins, S.; Swaan, P. W.; Fang, B.; Zhang, B.; Yan, B. J. Med. Chem. 2008, 51, 1242.
- Rock, D. M.; McLean, M. J.; Macdonald, R. L.; Catterall, W. A.; Taylor, C. P. Epilepsy Res. 1991, 8, 197.
- Gududuru, V.; Hurh, E.; Dalton, J. T.; Miller, D. D. Bioorg. Med. Chem. Lett. 2004, 14, 5289.
- (a) Marković, R.; Baranac, M.; Jovanović, V.; Džambaski, Z. J. Chem. Educ. 2004, 81, 1026; (b) Marković, R.; Baranac, M.; Džambaski, Z.; Stojanović, M.; Steel, P. J. Tetrahedron 2003, 59, 7803.
- 11. In all oxidation reactions, sulfones were formed in less than 3%.
- 12. For the determination of configuration around the CdbndC double bond in these and similar compounds, and isomerization studies, see: (a) Marković, R.;

Baranac, M.; Juranić, N.; Macura, S.; Cekić, I.; Minić, D. J. Mol. Struct. 2006, 800, 85; (b) Marković, R.; Shirazi, A.; Džambaski, Z.; Baranac, M.; Minić, D. J. Phys. Org. Chem. 2004, 17, 118; (c) Marković, R.; Džambaski, Z.; Baranac, M. Tetrahedron 2001, 57, 5833; (d) Marković, R.; Baranac, M. Heterocycles 1998, 48, 893 and Ref 10 in the present paper.

- (a) Buchanan, G. W.; Durst, T. Tetrahedron Lett. 1975, 1683; (b) Harrison, C. R.; Hodge, P. J. Chem. Soc., Perkin Trans. 1 1976, 1772; (c) Cook, M. J.; Tonge, A. P. J. Chem. Soc., Perkin Trans. 2 1974, 767.
- (a) Green, C. H.; Hellier, D. G. J. Chem. Soc., Perkin Trans. 2 1972, 458; (b) Buck, K. W.; Foster, A. B.; Pardoe, W. D.; Qadir, M. H.; Webber, J. M. J. Chem. Soc., Chem. Commun. 1966, 759.
- 15. Abraham, R. J.; Byrne, J. J.; Griffiths, L. Magn. Reson. Chem. 2008, 46, 667.
- 16. For the application of theoretical methods for stereochemical determinations of sulfoxide-containing compounds, see: (a) Dračínský, M.; Pohl, R.; Slavěťínská, L.; Janků, J.; Buděšínský, M. *Tetrahedron: Asymmetry* 2011, 22, 356; (b) Dračínský, M.; Pohl, R.; Slavěťínská, L.; Hřebabecký, H.; Buděšínský, M. *Tetrahedron: Asymmetry* 2011, 22, 1797; (c) Pihlaja, K.; Sinkkonen, J.; Stájer, G.; Koch, A.; Kleinpeter, E. *Magn. Reson. Chem.* 2011, 49, 443; (d) Kovács, J.; Tóth, G.; Simon, A.; Lévai, A.; Koch, A.; Kleinpeter, E. *Magn. Reson. Chem.* 2003, 41, 193; (e) Li, W.; Hwang, D. J.; Cremer, D.; Joo, H.; Kraka, E.; Kim, J.; Ross, C. R., II; Nguyen, V. Q.; Dalton, J. T.; Miller, D. D. *Chirality* 2009, 21, 578.
- For the calculation of ¹H NMR chemical shifts, see: Jain, R.; Bally, T.; Rablen, P. R. J. Org. Chem. 2009, 74, 4017.
- (a) Becke, A. D. J. Chem. Phys. 1993, 98, 5648; (b) Lee, C.; Yang, W.; Parr, R. G. Phys. Rev. B 1988, 37, 785.
- (a) Ditchfield, R. Mol. Phys. 1974, 27, 789; (b) Wolinski, K.; Hinton, J. F.; Pulay, P. J. Am. Chem. Soc. 1990, 112, 8251.
- 20. Inclusion of solvent in the chemical shift calculations gave similar correlation with R^2 =0.997.
- (a) Sandström, J. Top. Stereochem. 1983, 14, 83; (b) Fischer, G.; Rudorf, W.-D.; Kleinpeter, E. Magn. Reson. Chem. 1991, 29, 212; (c) Benassi, R.; Bertarini, C.; Kleinpeter, E.; Taddei, F. THEOCHEM 2000, 498, 217; (d) Kleinpeter, E.; Klod, S.; Rudorf, W.-D. J. Org. Chem. 2004, 69, 4317; (e) Rattananakin, P.; Pittman, C. R., Jr.; Collier, W. E.; Saebø, S. Struct. Chem. 2007, 18, 399.
- Ye, G.; Chatterjee, S.; Li, M.; Zhou, A.; Song, Y.; Barker, B. L; Chen, C.; Beard, D. J.; Henry, W. P.; Pittman, C. U. *Tetrahedron* **2010**, *66*, 2919 and references therein.
- (a) Kleinpeter, E. J. Serb. Chem. Soc. 2006, 71, 1; (b) Kleinpeter, E.; Schulenburg, A. Tetrahedron Lett. 2005, 46, 5995; (c) Baranac-Stojanović, M.; Klaumünzer, U.; Marković, R.; Kleinpeter, E. Tetrahedron 2010, 66, 8958.
- (a) Kleinpeter, E.; Thomas, St.; Uhlig, G.; Rudorf, W.-D. Magn. Reson. Chem. 1993, 31, 714; (b) Kleinpeter, E.; Heydenreich, M.; Chatterjee, S. K.; Rudorf, W.-D. Magn. Reson. Chem. 1994, 32, 473; (c) Kleinpeter, E.; Koch, A.; Heydenreich, M.; Chatterjee, S. K.; Rudorf, W.-D. J. Mol. Struct. 1995, 356, 25; (d) Mueller, J. L.; Gibson, M. S.; Hartman, J. S. Can. J. Chem. 1996, 74, 1329; (e) Chiara, J. L.; Gómez-Sánchez, A.; Bellanato, J. J. Chem. Soc., Perkin Trans. 2 1998, 1797; (f) Kleinpeter, E.; Heydenreich, M.; Woller, J.; Wolf, G.; Koch, A.; Kempter, G. J. Chem. Soc., Perkin Trans. 2 1998, 1877; (g) Meier, H.; Mühling, B.; Gerold, J.; Jacob, D.; Oehlhof, A. Eur, J. Org. Chem. 2007, 625.
- (a) Glendening, E. D.; Landis, C. R.; Weinhold, F. WIREs Comput. Mol. Sci. 2012, 2, 1; (b) Weinhold, F.; Landis, C. R. Discovering Chemistry with Natural Bond Orbitals; John Wiley & Sons: Hoboken, NJ, 2012.
- 26. Rosini, C.; Donnoli, M. I.; Superchi, S. Chem.-Eur. J. 2001, 7, 72.
- (a) Rozwadowska, M. D.; Sulima, A. *Tetrahedron* 2003, 59, 1173; (b) Clay, M. P.; Hanssen, B. R.; Surapaneni, S. S.; Lindstrom, T. D. *Chirality* 1999, *11*, 233.
- 28. For the literature about the CH…O hydrogen bonds, see: (a) Koch, U.; Popelier, P. L. A. J. Phys. Chem. 1995, 99, 9747; (b) Yoshida, H.; Harada, T.; Ohno, K.; Matsuura, H. Chem. Commun. 1997, 2213; (c) Tsuzuki, S.; Houjou, H.; Nagawa, Y.; Hiratani, K. J. Chem. Soc., Perkin Trans. 2 2001, 1951; (d) Scheiner, S. Int. J. Quantum Chem. 2010, 110, 2775 For the literature about the hydrogen bonds, see; (e) Desiraju, G. R. Acc. Chem. Res. 2002, 35, 565.
- (a) Yates, J. R.; Pham, T. N.; Pickard, C. J.; Mauri, F.; Amado, A. M.; Gil, A. M.; Brown, S. P. J. Am. Chem. Soc. 2005, 127, 10216; (b) Wang, B.; Hinton, J. F.; Pulay, P. J. Phys. Chem. A 2003, 107, 4683; (c) Scheiner, S.; Gu, Y.; Kar, T. J. Mol. Struct. (Theochem) 2000, 500, 441.
- 30. Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Montgomery, J. A., Jr.; Vreven, T.; Kudin, K. N.; Burant, J. C.; Millam, J. M.; Iyengar, S. S.; Tomasi, J.; Barone, V.; Mennuci, B.; Cossi, M.; Scalmani, G.; Rega, N.; Petersson, G. A.; Nakatsuji, H.; Hada, M.; Ehara, M.; Toyota, K.; Fukuda, R.; Hasegawa, J.; Ishida, M.; Nakajima, T.; Honda, Y.; Kitao, O.; Nakai, H.; Klene, M.; Li, X.; Knox, J. E.; Hratchian, H. P.; Cross, J. B.; Adamo, C.; Jaramillo, J.; Gomperts, R.; Stratmann, R. E.; Yazyev, O.; Austin, A. J.; Cammi, R.; Pomelli, C.; Ochterski, J. W.; Ayala, P. Y.; Morokuma, K.; Voth, G. A.; Salvador, P.; Dannenberg, J. J.; Zakrzewski, V. G.; Dapprich, S.; Daniels, A. D.; Strain, M. C.; Farkas, O.; Malick, D. K.; Rabuck, A. D.; Raghavachari, K.; Foresman, J. B.; Ortiz, J. V.; Cui, Q.; Babuol, A. G.; Clifford, S.; Cioslowski, J.; Stefanov, B. B.; Liu, G.; Liashenko, A.; Piskorz, P.; Komaromi, I.; Martin, R. L.; Fox, D. J.; Keith, T.; Al-Laham, M. A.; Peng, C. Y.; Nanayakkara, A.; Challacombe, M.; Gill, P. M. W.; Johnson, B.; Chen, W.; Wong, M. W.; Gonzalez, C.; Pople, J. A. *Gaussian 03* (*Revision C.02*); Gaussian: Wallingford CT, 2004.
- (a) Cancés, E.; Mennucci, B.; Tomasi, J. J. Chem. Phys. 1997, 107, 3032; (b) Mennucci, B.; Cancés, E.; Tomasi, J. J. Phys. Chem. B 1997, 101, 10506.
- Stojanović, M.; Marković, R.; Kleinpeter, E.; Baranac-Stojanović, M. Org. Biomol. Chem. 2012, 10, 575.
- Stojanović, M.; Marković, R.; Kleinpeter, E.; Baranac-Stojanović, M. Tetrahedron 2011, 67, 9541.