

## Synthesis and properties of terthiophene-modified oligodeoxynucleotides

Kiyohiko Kawai,\* Akira Sugimoto, Hiroko Yoshida, Sachiko Tojo,  
Mamoru Fujitsuka and Tetsuro Majima\*

The Institute of Scientific and Industrial Research (SANKEN), Osaka University, Mihogaoka 8-1, Ibaraki, Osaka 567-0047, Japan

Received 26 April 2005; revised 24 June 2005; accepted 1 July 2005

Available online 26 August 2005

**Abstract**—Synthesis and properties of oligodeoxynucleotides (ODNs) containing terthiophene (Thp) were described. One-electron oxidation of Thp-modified ODN resulted in the formation of Thp radical cation ( $\text{Thp}^+$ ), which remained stable in the experimental time window up to 200  $\mu\text{s}$ , showing that charge may be carried along DNA by Thp as  $\text{Thp}^+$ .  
© 2005 Elsevier Ltd. All rights reserved.

DNA is a versatile molecule that can be used to construct nanometer-sized higher-ordered assemblies and architectures based on the stored information.<sup>1–4</sup> In view of the potential use of DNA as a molecular wire for electronic applications, mechanistic studies of charge transfer in DNA have attracted considerable attention.<sup>5–13</sup> Gel electrophoretic analysis<sup>14,15</sup> and our recent time-resolved transient absorption measurement<sup>16</sup> clearly demonstrated that the hole generated on DNA upon one-electron oxidation can migrate along DNA over 100 Å. However, since DNA is inherently unstable upon oxidation, DNA itself cannot be a good conductive material.<sup>17–20</sup> Another promising way is to use DNA as a scaffold for arranging organic molecules whose radical ion is stable. Previously, we have shown that a charge can be carried by pyrene and phenothiazine as their radical cations along DNA.<sup>21</sup> However, only a few compounds, e.g., pyrene,<sup>22,23</sup> phenothiazine,<sup>24–26</sup> methylindole,<sup>27,28</sup> and ferrocene,<sup>29,30</sup> were tested as carriers of positive charge along DNA. Here, we report the synthesis of terthiophene-modified oligodeoxynucleotides (ODNs), showing that a hole can be carried as a radical cation of Thp ( $\text{Thp}^+$ ) along DNA.

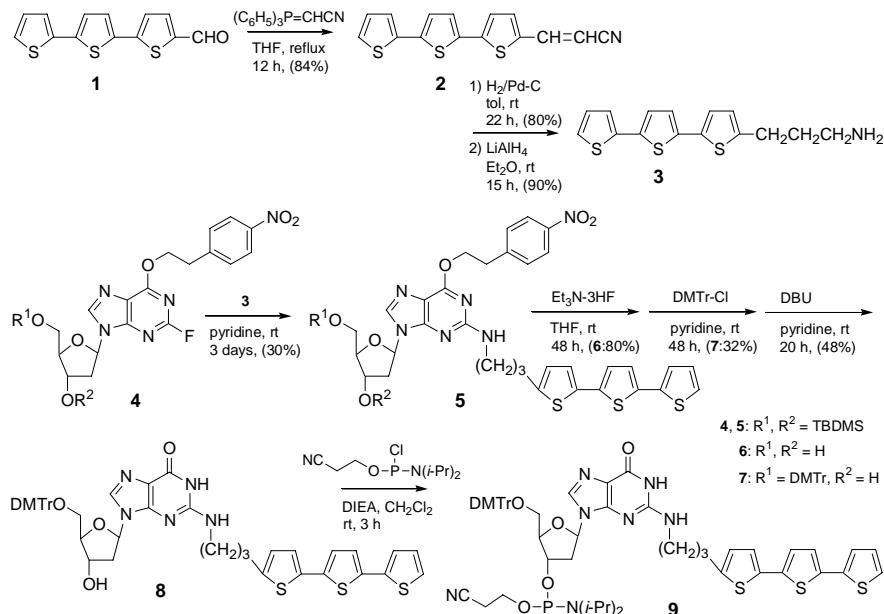
Polythiophenes and substituted polythiophenes have received much attention as among the most studied conducting polymers worldwide.<sup>31–33</sup> We selected Thp

to assemble along DNA because  $\text{Thp}^+$  is shown to be stable in water.<sup>34–36</sup> The oxidation potential of Thp ( $E^\circ = 1.23 \text{ V}$  vs NHE in  $\text{CH}_3\text{CN}$ )<sup>37</sup> is lower than that of guanine which has the lowest oxidation potential among the four DNA bases ( $E^\circ = 1.47 \text{ V}$  vs NHE in  $\text{CH}_3\text{CN}$ ).<sup>38</sup> Therefore, similar to the previously studied pyrene<sup>22,23</sup> and phenothiazine,<sup>24–26</sup> Thp was expected to serve as a hole trap in DNA and may be useful to carry a hole along DNA. The minor groove of DNA has been demonstrated to serve as a good candidate for arranging heterocyclic compounds.<sup>39–41</sup> Thus, we synthesized a nucleoside derivative that possesses a Thp group on guanine N2 ( ${}^{\text{Th}}\text{G}$ ) to arrange Thp along the minor groove of DNA through the reaction of 3',5'-protected 2-fluorodeoxyinosine derivative (**4**) with 5-(3-aminopropyl)-2,2',5',2'-terthiophene (**3**) as shown in Scheme 1.<sup>42,43</sup>  ${}^{\text{Th}}\text{G}$  was converted to the phosphoramidite derivative (**9**) and  ${}^{\text{Th}}\text{G}$ -modified ODNs were synthesized by DNA synthesizer according to the standard procedure. Incorporation of  ${}^{\text{Th}}\text{G}$  into ODNs was confirmed by digestion with snake venom phosphodiesterase/nuclease P1/alkaline phosphatase to 2'-deoxyribonucleosides and MALDI-TOF mass spectra.<sup>44</sup>

First, to check the duplex stability of the  ${}^{\text{Th}}\text{G}$ -modified ODNs, melting temperatures were measured (Table 1).  ${}^{\text{Th}}\text{G}$ -modified ODN (**11**) showed slightly higher thermostability compared to the corresponding unmodified ODN (**10**). Incorporation of two  ${}^{\text{Th}}\text{G}$  (**12**) showed a similar  $T_m$  value with that of **10**. Thus,  ${}^{\text{Th}}\text{G}$  was incorporated into DNA without large disturbance of the duplex stability.

**Keywords:** DNA; Modified oligonucleotide; Charge transfer; Thiophene.

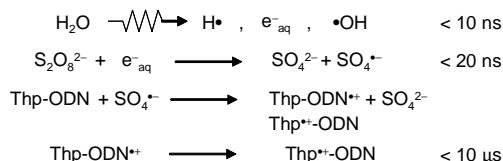
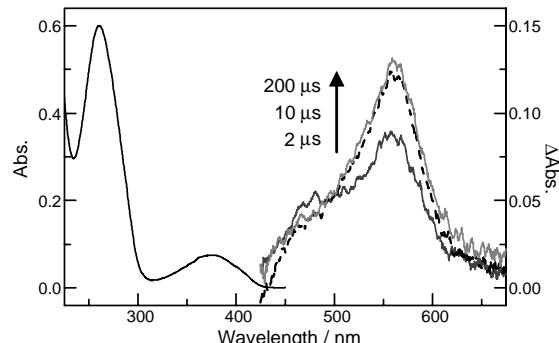
\* Corresponding authors. Tel.: +81 6 6879 8496 (K.K.); +81 6 6879 8495 (T.M.); fax: +81 6 6879 8499; e-mail addresses: kiyohiko@sanken.osaka-u.ac.jp; majima@sanken.osaka-u.ac.jp

**Scheme 1.** Synthetic route of the  $^{Th}\text{G}$ -phosphoramidite 9.**Table 1.** Melting temperatures of  $^{Th}\text{G}$ -modified ODNs

ODNs	Sequence	$T_m$ ( $^{\circ}\text{C}$ ) <sup>a</sup>
<b>10</b>	5'GCAGAACACG 3'CGTCTTGTGC	34
<b>11</b>	5' $^{Th}\text{GCA}$ GAACACG 3'CGTCTTGTGC	36
<b>12</b>	5' $^{Th}\text{GCA}$ GAACACG 3'CGTCTT $^{Th}\text{GTGC}$	34

<sup>a</sup> UV melting measurements were carried out in a pH 7.0 Na phosphate buffer 20 mM at a total strand concentration of 8  $\mu\text{M}$ .

Next, to obtain the information on the charge-carrying properties of  $^{Th}\text{G}$ , pulse radiolysis of  $^{Th}\text{G}$ -modified ODN was performed. A hole was produced in the reaction with  $\text{SO}_4^{\cdot-}$  that was generated during the pulse radiolysis (28 MeV, 8 ns) of Ar-saturated aqueous solution containing 10 mM  $\text{K}_2\text{S}_2\text{O}_8$ , 100 mM *t*-BuOH, 20 mM Na phosphate buffer (pH 7.0), and 0.1 mM ODN (strand conc) (Scheme 2). There are two possible processes for the formation of  $\text{Thp}^+$ : the direct diffusional collision between  $\text{SO}_4^{\cdot-}$  and  $^{Th}\text{G}$ , and the hole transfer from the radical cation of nucleobases generated in DNA to  $^{Th}\text{G}$  as shown in Scheme 2.<sup>22,23</sup> Interestingly, a transient absorption with a peak at 560 nm assigned to  $\text{Thp}^+$  was observed after the electron pulse during the pulse radiolysis and was stable within our experimental timescale up to 200  $\mu\text{s}$  (Fig. 1).

**Scheme 2.** Mechanistic scheme for generation of oxidizing reagent  $\text{SO}_4^{\cdot-}$  during pulse radiolysis, hole generation, and transfer in Thp-modified ODN.**Figure 1.** Ground-state absorption (left axis) and transient absorption spectra (right axis) obtained at 2, 10, and 200  $\mu\text{s}$  during the pulse radiolysis of ODN 11. Ground-state absorption spectrum was measured in a 20 mM Na phosphate buffer (pH 7.0) at a total strand concentration of 8  $\mu\text{M}$ . Pulse radiolysis was carried out in Ar-saturated aqueous solution in the presence of 10 mM  $\text{K}_2\text{S}_2\text{O}_8$ , 100 mM *t*-BuOH, and 20 mM Na phosphate buffer (pH 7.0), at a total strand concentration of 200  $\mu\text{M}$ .

In conclusion, a phosphoramidite derivative of  $^{Th}\text{G}$  was synthesized to assemble Thp along the DNA minor groove as a hole carrier. It has been demonstrated that  $^{Th}\text{G}$  can be incorporated into DNA without large alternation of the duplex stability. Pulse radiolysis of  $^{Th}\text{G}$ -modified ODN clearly demonstrated that  $\text{Thp}^+$  can be stably generated in DNA, showing that Thp serves as a hole trap. Hence, by arranging several  $^{Th}\text{G}$  along DNA, it may be useful to carry a hole along DNA.

### Acknowledgments

This work has been partly supported by a Grant-in-Aid for Scientific Research on Priority Area (417), 21st Century COE Research, and others from the Ministry

of Education, Culture, Sports, Science and Technology (MEXT) of Japanese Government.

### Supplementary data

Supplementary data associated with this article can be found in the online version at doi:10.1016/j.bmcl.2005.07.004.

### References and notes

- Liao, S.; Seeman, N. C. *Science* **2004**, *306*, 2072.
- Seeman, N. C. *Nature* **2003**, *421*, 427.
- Endo, M.; Majima, T. *Angew. Chem., Int. Ed.* **2003**, *42*, 5744.
- Endo, M.; Majima, T. *J. Am. Chem. Soc.* **2003**, *125*, 13654.
- Lewis, F. D.; Letsinger, R. L.; Wasielewski, M. R. *Acc. Chem. Res.* **2001**, *34*, 159.
- Boon, E. M.; Ceres, D. M.; Drummond, T. G.; Hill, M. G.; Barton, J. K. *Nat. Biotech.* **2000**, *18*, 1096.
- Giese, B. *Acc. Chem. Res.* **2000**, *33*, 631.
- Schuster, G. B. *Acc. Chem. Res.* **2000**, *33*, 253.
- Okamoto, A.; Kamei, T.; Tanaka, K.; Saito, I. *J. Am. Chem. Soc.* **2004**, *126*, 14732.
- Yamana, K.; Kumamoto, S.; Hasegawa, T.; Nakano, H.; Sugie, Y. *Chem. Lett.* **2002**, *506*.
- Yamana, K.; Kumamoto, S.; Nakano, H.; Matsuo, Y.; Sugie, Y. *Chem. Lett.* **2001**, *1132*.
- Takada, T.; Kawai, K.; Cai, X.; Sugimoto, A.; Fujitsuka, M.; Majima, T. *J. Am. Chem. Soc.* **2004**, *126*, 1125.
- Kawai, K.; Osakada, Y.; Takada, T.; Fujitsuka, M.; Majima, T. *J. Am. Chem. Soc.* **2004**, *126*, 12843.
- Nunez, M. E.; Noyes, K. T.; Barton, J. K. *Chem. Biol.* **2002**, *9*, 403.
- Henderson, P. T.; Jones, D.; Hampikian, G.; Kan, Y. Z.; Schuster, G. B. *Proc. Natl. Acad. Sci. U.S.A.* **1999**, *96*, 8353.
- Takada, T.; Kawai, K.; Fujitsuka, M.; Majima, T. *Proc. Natl. Acad. Sci. U.S.A.* **2004**, *101*, 14002.
- Burrows, C. J.; Muller, J. G. *Chem. Rev.* **1998**, *98*, 1109.
- Armitage, B. *Chem. Rev.* **1998**, *98*, 1171.
- Kawai, K.; Cai, X.; Sugimoto, A.; Tojo, S.; Fujitsuka, M.; Majima, T. *Angew. Chem., Int. Ed.* **2004**, *43*, 2406.
- Kawai, K.; Takada, T.; Nagai, T.; Cai, X.; Sugimoto, A.; Fujitsuka, M.; Majima, T. *J. Am. Chem. Soc.* **2003**, *125*, 16198.
- Takada, T.; Kawai, K.; Tojo, S.; Majima, T. *Tetrahedron Lett.* **2003**, *44*, 3851.
- Kawai, K.; Takada, T.; Tojo, S.; Ichinose, N.; Majima, T. *J. Am. Chem. Soc.* **2001**, *123*, 12688.
- Takada, T.; Kawai, K.; Tojo, S.; Majima, T. *J. Phys. Chem. B* **2003**, *107*, 14052.
- Tierney, M. T.; Sykora, M.; Khan, S. I.; Grinstaff, M. W. *J. Phys. Chem. B* **2000**, *104*, 7574.
- Kawai, K.; Takada, T.; Tojo, S.; Majima, T. *J. Am. Chem. Soc.* **2003**, *125*, 6842.
- Kawai, K.; Takada, T.; Tojo, S.; Majima, T. *Tetrahedron Lett.* **2002**, *43*, 89.
- Delaney, S.; Yoo, J.; Stemp, E. D. A.; Barton, J. K. *Proc. Natl. Acad. Sci. U.S.A.* **2004**, *101*, 10511.
- Pascaly, M.; Yoo, J.; Barton, J. K. *J. Am. Chem. Soc.* **2002**, *124*, 9083.
- Pike, A. R.; Ryder, L. C.; Horrocks, B. R.; Clegg, W.; Connolly, B. A.; Houlton, A. *Chem. Eur. J.* **2005**, *11*, 344.
- Ihara, T.; Maruo, Y.; Takenaka, S.; Takagi, M. *Nucleic Acids Res.* **1996**, *24*, 4273.
- Otsubo, T.; Aso, Y.; Takimiya, K. *Bull. Chem. Soc. Jpn.* **2001**, *74*, 1789.
- Sumi, N.; Nakanishi, H.; Ueno, S.; Takimiya, K.; Aso, Y.; Otsubo, T. *Bull. Chem. Soc. Jpn.* **2001**, *74*, 979.
- Izumi, T.; Kobashi, S.; Takimiya, K.; Aso, Y.; Otsubo, T. *J. Am. Chem. Soc.* **2003**, *125*, 5286.
- Hapiot, P.; Lagrost, C.; Aeyiach, S.; Jouini, M.; Lacroix, J. C. *J. Phys. Chem. B* **2002**, *106*, 3622.
- Emmi, S. S.; D'Angelantonio, M.; Beggiato, G.; Poggi, G.; Geri, A.; Pietropaolo, D.; Zotti, G. *Radiat. Phys. Chem.* **1999**, *54*, 263.
- Guyard, L.; Hapiot, P.; Jouini, M.; Lacroix, J. C.; Lagrost, C.; Neta, P. *J. Phys. Chem. A* **1999**, *103*, 4009.
- Hill, M. G.; Penneau, J. F.; Zinger, B.; Mann, K. R.; Miller, L. L. *Chem. Mater.* **1992**, *4*, 1106.
- Seidel, C. A. M.; Schulz, A.; Sauer, M. H. M. *J. Phys. Chem.* **1996**, *100*, 5541.
- Dervan, P. B.; Fechter, E. J.; Edelson, B. S.; Gottesfeld, J. M. Regulation of gene expression with pyrrole-imidazole polyamides. In *Pseudo-Peptides in Drug Discovery*; 2004; pp 121–152.
- Wang, M.; Silva, G. L.; Armitage, B. A. *J. Am. Chem. Soc.* **2000**, *122*, 9977.
- Sugiyama, H.; Lian, C. Y.; Isomura, M.; Saito, I.; Wang, A. H. *J. Proc. Natl. Acad. Sci. U.S.A.* **1996**, *93*, 14405.
- Nakatani, K.; Dohno, C.; Saito, I. *J. Am. Chem. Soc.* **2001**, *123*, 9681.
- Adib, A.; Potier, P. F.; Doronina, S.; Huc, I.; Behr, J.-P. *Tetrahedron Lett.* **1997**, *38*, 2989.
- Calcd for 5'-GCA<sup>Th</sup>GAACACG-3': 3334.5, found 3334.2. Calcd for 5'-CGT<sup>Th</sup>GTTCTGC-3': 3298.4, found 3299.0.