



A mild, efficient, and selective procedure for transprotection of acetonides to acetates catalyzed with $\text{HClO}_4\text{--SiO}_2$

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ABSTRACT

The transformation of acetonides into acetates is frequently required in synthetic chemistry. An efficient procedure for direct conversion of acetonides into acetates in the presence of $\text{HClO}_4\text{--SiO}_2$ under mild conditions was developed. The acetonides of primary hydroxy groups are directly converted to diacetates, and the anomeric acetonides of furanosides are stereoselectively transformed into the corresponding acetyl β -D-furanosides with a 2-acetoxyisopropyl group.

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1. Introduction

Both acetonides and acetates are most frequently used as protecting groups in synthetic chemistry because they are readily introduced and removed,^{1–3} particularly, in carbohydrate,^{4–9} nucleoside,^{10–13} and polyhydroxy compound chemistries.^{14–16} In view of the fact that the transformation of acetonides into corresponding diacetates is often required in organic synthesis due to the complementary stability of these two kinds of protecting groups,^{17–24} studies in this transformation, here termed ‘transprotection’, are considerably worthy in spite of the limited methods that are currently available.²⁵

We herein wish to report an effective method for transprotection of isopropylidene acetals into acetates by making use of a cheaper, non-toxic, and reusable catalyst reported by Chakraborti and Gulhane,²⁶ namely, $\text{HClO}_4\text{--SiO}_2$ (Scheme 1), which has been used to catalyze a wide range of transformations such as the formation of acetals,^{27–29} thioacetals,³⁰ and acylal,^{31,32} acetylation of carbohydrates,^{33–35} glycosylation,^{36,37} synthesis of quinoxalines,³⁸ thio-Michael addition,³⁹ and *N*-Boc formation.⁴⁰

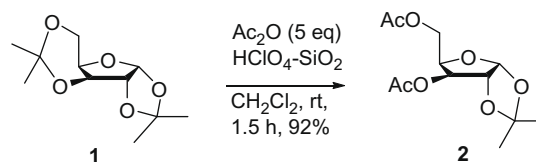
2. Results and discussion

In order to transform 1,2:3,5-di-*O*-isopropylidene- α -D-xylofuranose (**1**) straightforwardly into 3,5-di-*O*-acetyl-1,2-*O*-isopropylidene- α -D-xylofuranose (**2**),

we found that $\text{HClO}_4\text{--SiO}_2$ (0.5 mmol/g) is an extremely efficient substitute for $\text{Bi}(\text{OTf})_3\cdot x\text{H}_2\text{O}$ ²⁵ and is readily removed and recovered via filtration after the reaction is complete.³⁷

Treatment of **1** (0.15 mol) with Ac_2O (5 equiv) in CH_2Cl_2 at rt in the presence of $\text{HClO}_4\text{--SiO}_2$ (7.5 g) afforded **2** in 92% isolated yield (Table 1, entry a). Therefore, we expected to investigate the generality of the application of $\text{HClO}_4\text{--SiO}_2$ in the transprotection of acetonides into acetates. It could be seen from the results listed in Table 1 that glucose derivatives **3**, **5**, **6**, and **8** were all smoothly and immediately transformed into corresponding acetates **4**, **7**, and **9**, respectively, at rt in quite good yields (Table 1, entries b–e). Additionally, these results also demonstrated that the isopropylidene acetals of primary hydroxyl groups could be selectively transformed into the corresponding acetates in the presence of anomeric acetonides under these one-pot conditions.

As anticipated, both isopropylidene and acyl groups in **10** were simultaneously transformed, respectively, into acetates and acylal

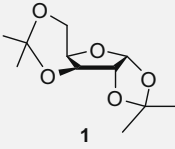
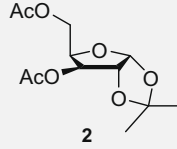
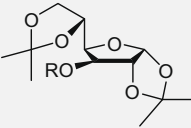
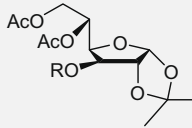
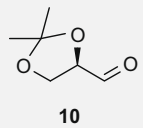
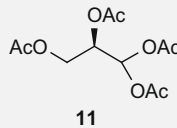
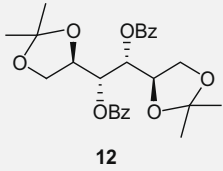
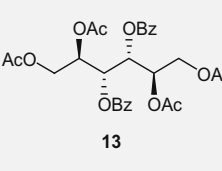
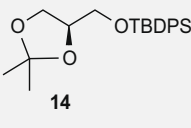
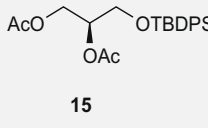
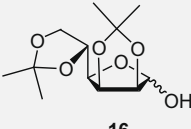
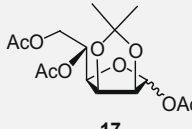
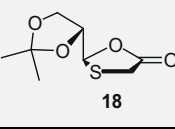
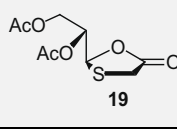


Scheme 1. Selective transprotection of terminal acetonides into diacetates.

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Table 1Direct transprotection of terminal acetonides into diacetates in the presence of $\text{HClO}_4\text{-SiO}_2$

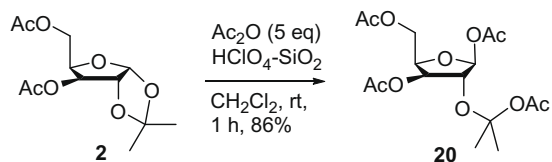
Entry	Substrate	Product ^a	Time (h)	Yield ^b (%)
a			1.5	92 ²⁵
b			7	86 ^{25,43}
c	3R H	4R Ac	7	88 ^{25,43}
d	5R Ac	4R Ac	7	90 ²⁵
e	6R Bz	7R Bz	8	90 ²⁵
	8R Ts	9R Ts	6	90 ²⁵
f			8	83 ⁴¹
g			4 (reflux)	81
h			2	78
i			8	89 ²⁵
j			0.5	82 ⁴²

^a Characterized by ¹H NMR, ¹³C NMR, and ESIMS: Calcd for.^b Isolated yields.

groups to give **11** in 83% yield (Table 1, entry f).⁴¹ Surprisingly, transprotection of the two terminal isopropylidene acetals of D-mannitol derivative **12** failed at rt, and the tetra-acetyl product **13** was obtained under reflux in CH_2Cl_2 (Table 1, entry g). Moreover, it was also observed that the *tert*-butyldiphenylsilyl (TBDPS)-protecting group remained intact as the isopropylidene group was selectively transformed (Table 1, entry h). Furthermore, the formation of both products **17** and **19** provided further illustrations of the selectivity and efficiency of this one-pot transprotection procedure: (1) A terminal hydroxyl acetonide was selectively replaced by acetates in the presence of secondary hydroxyl isopropylidene acetal, and (2) the anomeric hydroxy function was concomitantly acetylated (Table 1, entry i).

In addition, we have also tested the application of this one-pot methodology to transprotection of anomeric acetonides (Scheme 2).

Treatment of compound **2** with Ac_2O under the same conditions described as above afforded compound **20**, in which the anomeric acetonide was partially cleaved to generate a 2-acetoxyisopropyl group (Table 2, entry a). This isopropylidene chain-opened structure is similar to those that result from catalysis by $\text{Bi}(\text{OTf})_3 \cdot x\text{H}_2\text{O}$.²⁵ Following these one-pot conditions, all similar transprotections shown in Table 2 were performed smoothly as more candidates were employed as starting materials. It is noteworthy that hydroxyl group could not interrupt this type of transprotection, and these were acetylated simultaneously (Table 2, entries b and e). We also first observed that the primary *tert*-butyldimethylsilyl (TBDMS) ether group in **36** was selectively transprotected to the corresponding acetate along with the transprotection of the anomeric isopropylidene group to give **37** (Table 2, entry k). As to anomeric acetonide groups in a pyranose, a similar selective transformation was also observed



Scheme 2. Transprotection of anomeric acetanilides of furanoses into corresponding acetylated 2-O-(2'-acetoxyisopropyl)-β-D-furanosides.

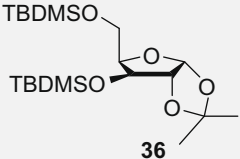
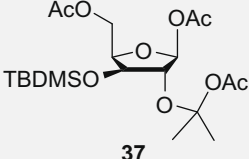
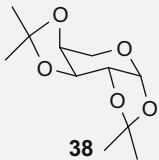
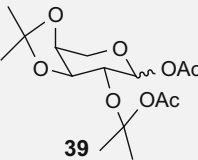
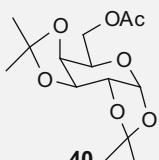
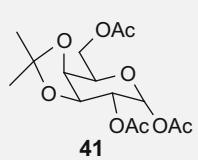
as L-arabinopyranose derivative **38** was converted into an anomeric mixture **39** in quite good yield (Table 2, entry l). The anomeric acetanilide group in 6-O-acetyl-1,2:3,4-di-O-isopropylidene-α-D-galactopyranose (**40**), however, was selectively transprotected to give a pure isomer **41** without an isopropylidene group at the 2-position (Table 2, entry m), which might arise from the participation of the 6-acetoxy group.

The assignment of the isopropylidene acetal chain-opened structures listed in Table 2 are supported by ^1H NMR, ^{13}C NMR,

Table 2
Transprotection of anomeric acetanilides of furanoses and pyranoses into corresponding 2-O-(2'-acetoxyisopropyl)-1-O-acetyl-β-D-furanosides and 2-O-(2'-acetoxyisopropyl)-1-O-acetyl-β-D-pyranosides with $\text{HClO}_4\text{-SiO}_2$

Entry	Substrate	Product ^a	Time (h)	Yield ^b (%)
a			1	86 ²⁵
b			1	87
c			1	84
d			1	90 ²⁵
e			4	90
f			1	85 ²⁵
g			1.5	89 ²⁵
h			2	89 ²⁵
i			1.5	87 ²⁵
j			2	83 ²⁵

Table 2 (continued)

Entry	Substrate	Product ^a	Time (h)	Yield ^b (%)
k	 36	 37	0.5	84
l	 38	 39	7.5	82
m	 40	 41	8	88

^a Characterized by ¹H NMR, ¹³C NMR, and ESIMS: Calcd for.

^b Isolated yields.

and ESIMS: Calcd for data, which also provide evidence that all these products are pure anomeric furanosides rather than a mixture. As to compound **20**, the molecular structure was further confirmed to be a β anomer by a NOESY spectrum. In the ¹H–¹H dimension of the NMR spectrum of compound **20**, two cross-peaks are observed between the resonance signals of the anomeric proton (at δ 6.16) and H-3, 4 (at δ 5.33), and the methyl protons in the isopropylidene group (at δ 1.45), respectively. Furthermore, it is also observed that the methyl protons of the isopropylidene group (at δ 1.45) interact with H-3, 4 (at δ 5.33) and not with H-5 (at δ 4.09–4.36). We believe that these NOESY spectroscopic data provide strong evidence for the β -anomeric configurational assignments.

In conclusion, we have developed a useful procedure for the direct transprotection of terminal hydroxy acetonides into their corresponding acetates in an efficient process and under mild conditions with HClO₄–SiO₂ as catalyst. This procedure also provides excellent selectivity in transprotection of primary hydroxy acetonides into acetates in the presence of anomeric acetonides of furanoses. Anomeric acetonides of furanoses are stereoselectively transformed into the corresponding acetyl 2-*O*-(2'-acetoxyisopropyl)- β -D-furanosides. In view of economy and environmental concerns, this one-pot mild procedure might be remarkably valuable in synthetic chemistry. The application of this methodology to the synthesis of nucleosides and their transformations is underway in our laboratory, and the results will be reported in due course.

3. Experimental

3.1. General

CH₂Cl₂ was dried over P₂O₅ and distilled. Ac₂O was distilled before use. All reactions were monitored by thin-layer chromatography (TLC) on silica gel F₂₅₄ plates. ¹H NMR and ¹³C NMR spectra were recorded on a Varian Mercury 400 spectrometer or a Bruker AV300 spectrometer in CDCl₃ or DMSO-*d*₆ and using TMS as the internal standard. Exact mass measurements were performed on

a LCMS2010 spectrometer equipped with a standard electrospray-ionization (ESI) interface.

3.2. Preparation of HClO₄–SiO₂

HClO₄ (1.8 g) was added to a stirred suspension of SiO₂ (23.7 g, 300–400 mesh) in EtOH (70 mL) and stirred for 10 min. The mixture was concentrated under reduced pressure to give a free-flowing powder (HClO₄, 0.5 mmol/g) for application.

3.3. Typical synthetic procedure

HClO₄–SiO₂ (0.5 mmol/g, 70 mg) was added to a stirred solution of substrate (1.0 mmol) and Ac₂O (0.45 mL, 5.0 mmol) in CH₂Cl₂ (5 mL) at rt or in CH₂Cl₂ (5 mL) heated under reflux. After complete conversion and filtration to remove catalyst, a satd aq solution of NaHCO₃ (10 mL) was added and separated. The aqueous solution was extracted with CH₂Cl₂ (2 $\times$ 10 mL). The organic layer was combined, washed with brine (10 mL), dried (MgSO₄), and concentrated under reduced pressure. The crude product was purified through short column chromatography on silica gel, which was eluted with EtOAc–petroleum ether (bp 60–90 °C) to give the products that are described in the following.

3.3.1. 3,5-Di-*O*-acetyl-1,2-*O*-isopropylidene- α -D-xylofuranose (**2**)

Oil; ¹H NMR (400 MHz, CDCl₃): δ _H 1.32 (s, 3H), 1.53 (s, 3H), 2.09 (s, 3H), 2.10 (s, 3H), 4.19 (dd, *J* 7.2 Hz and 11.6 Hz, 1H), 4.29 (m, 1H), 4.50 (m, 2H), 5.25 (d, *J* 3.1 Hz, 1H), 5.94 (d, *J* 3.7 Hz, 1H); ¹³C NMR (CDCl₃): δ _C 20.9, 21.0, 26.4, 26.9, 61.6, 76.3, 77.3, 83.6, 105.1, 112.5, 169.8, 170.8; ESIMS: Calcd for C₁₂H₁₈O₇, *m/z* 274.11; found, 297.2 (M+Na)⁺.

3.3.2. 3,5,6-Tri-*O*-acetyl-1,2-*O*-isopropylidene- α -D-glucofuranose (**4**)

Colorless solid, mp 76–77 °C (lit.⁴³ mp 78 °C); ¹H NMR (400 MHz, CDCl₃): δ _H 1.32 (s, 3H), 1.53 (s, 3H), 2.02 (s, 3H), 2.06

(s, 6H), 4.13 (dd, *J* 5.6 Hz and 12.3 Hz, 1H), 4.42 (dd, *J* 3.0 Hz and 9.4 Hz, 1H), 4.49 (d, *J* 3.6 Hz, 1H), 4.57 (dd, *J* 2.4 and 12.3 Hz, 1H), 5.22 (m, 1H), 5.35 (d, *J* 2.8 Hz, 1H), 5.93 (d, *J* 4.0 Hz, 1H); ^{13}C NMR (CDCl_3): δ_{C} 20.9, 21.0, 26.4, 27.0, 63.6, 67.7, 74.9, 77.0, 83.4, 105.4, 112.7, 169.8, 169.9, 170.8; ESIMS: Calcd for $\text{C}_{15}\text{H}_{22}\text{O}_9$, *m/z* 346.13; found, *m/z* 369.1 ($\text{M}+\text{Na}$) $^+$.

3.3.3. 5,6-Di-*O*-acetyl-3-*O*-benzoyl-1,2-*O*-isopropylidene- α -D-glucofuranose (7)

Oil; ^1H NMR (CDCl_3): δ_{H} 1.34 (s, 3H), 1.57 (s, 3H), 1.94 (s, 3H), 2.06 (s, 3H), 4.19 (dd, *J* 5.2 Hz and 12.4 Hz, 1H), 4.59 (m, 3H), 5.32 (m, 1H), 5.54 (d, *J* 2.4 Hz, 1H), 5.99 (d, *J* 3.2 Hz, 1H), 7.45 (m, 2H), 7.58 (m, 1H), 7.99 (d, *J* 8.0 Hz, 2H); ^{13}C NMR (CDCl_3): δ_{C} 20.9, 21.0, 26.5, 27.0, 63.4, 68.1, 75.9, 77.3, 83.5, 105.4, 112.8, 128.8, 129.2, 130.0, 133.9, 165.5, 169.6, 170.8; ESIMS: Calcd for $\text{C}_{20}\text{H}_{24}\text{O}_9$, *m/z* 408.14; found, *m/z* 431.1 ($\text{M}+\text{Na}$) $^+$.

3.3.4. 5,6-Di-*O*-acetyl-1,2-*O*-isopropylidene-3-*O*-*p*-toluenesulfonyl- α -D-glucofuranose (9)

Oil; ^1H NMR (CDCl_3): δ_{H} 1.28 (s, 3H), 1.48 (s, 3H), 1.95 (s, 3H), 2.03 (s, 3H), 2.44 (s, 3H), 4.06 (dd, *J* 4.0 Hz and 12.4 Hz, 1H), 4.45 (t, *J* 6.8 Hz, 1H), 4.57 (d, *J* 12.4 Hz, 1H), 4.69 (d, *J* 3.2 Hz, 1H), 5.03 (m, 2H), 5.89 (d, *J* 3.2 Hz, 1H), 7.34 (d, *J* 8.0 Hz, 2H), 7.77 (d, *J* 8.0 Hz, 2H); ^{13}C NMR (CDCl_3): δ_{C} 21.0, 21.9, 26.5, 26.9, 29.9, 62.8, 67.8, 76.4, 80.2, 83.0, 105.1, 113.0, 128.3, 130.2, 132.8, 145.8, 169.8, 170.7; ESIMS: Calcd for $\text{C}_{20}\text{H}_{26}\text{O}_{10}\text{S}$, *m/z* 458.12; found, *m/z* 481.2 ($\text{M}+\text{Na}$) $^+$.

3.3.5. (*R*)-Propane-1,1,2,3-tetraol tetraacetate (1,1,2,3-tetra-*O*-acetyl-D-glyceraldehyde hydrate) (11)

Oil; ^1H NMR ($\text{DMSO}-d_6$): δ_{H} 2.01 (s, 3H), 2.04 (m, 6H), 2.09 (m, 3H), 4.16 (dd, *J* 7.2 Hz and 12.4 Hz, 1H), 4.29 (dd, *J* 3.2 Hz and 12 Hz, 1H), 5.20 (m, 1H), 6.84 (dd, *J* 1.6 Hz and 4.4 Hz, 1H); ^{13}C NMR ($\text{DMSO}-d_6$): δ_{C} 20.9, 21.0, 21.1, 61.4, 69.9, 86.5, 169.0, 170.2, 170.7. The NMR spectra matched that reported⁴¹ in the literature, allowing for the fact that the latter spectra were measured in CDCl_3 rather than in $\text{DMSO}-d_6$ solution.

3.3.6. 1,2,5,6-Tetra-*O*-acetyl-3,4-di-*O*-benzoyl-D-mannitol (13)

Oil; ^1H NMR ($\text{DMSO}-d_6$): δ_{H} 1.93 (s, 12H), 4.14 (dd, *J* 5.2 Hz and 12.0 Hz, 2H), 4.28 (dd, *J* 3.2 Hz and 12.4 Hz, 2H), 5.24 (m, 2H), 5.72 (d, *J* 7.2 Hz, 2H), 7.56 (m, 4H), 7.70 (m, 2H), 8.01 (m, 4H); ^{13}C NMR ($\text{DMSO}-d_6$): δ_{C} 20.5, 20.6, 61.6, 68.4, 68.5, 128.8, 129.1, 129.7, 134.1, 164.8, 169.4, 170.0; ESIMS: Calcd for $\text{C}_{28}\text{H}_{30}\text{O}_{12}$, *m/z* 558.17; found, *m/z* 581.3 ($\text{M}+\text{Na}$) $^+$.

3.3.7. 1,2-Di-*O*-acetyl-3-*O*-*tert*-butyldiphenylsilyl-D-glycerol (15)

Oil; ^1H NMR (CDCl_3): δ_{H} 1.06 (s, 9H), 2.04 (s, 6H), 3.78 (m, 2H), 4.23 (dd, *J* 6.0 Hz and 12.0 Hz, 1H), 4.41 (m, 1H), 5.17 (m, 1H), 7.42 (m, 6H), 7.66 (m, 4H); ^{13}C NMR (CDCl_3): δ_{C} 19.5, 21.0, 21.2, 26.9, 62.4, 62.9, 71.9, 128.0, 130.1, 133.2, 133.3, 135.8, 170.5, 170.9; ESIMS: Calcd for $\text{C}_{23}\text{H}_{30}\text{O}_5\text{Si}$, *m/z* 414.19; found, *m/z* 437.2 ($\text{M}+\text{Na}$) $^+$.

3.3.8. 1,5,6-Tri-*O*-acetyl-2,3-*O*-isopropylidene- α , β -D-mannofuranoside (17)

Oil; ^1H NMR (CDCl_3): δ_{H} 1.30 (s, 3H), 1.46 (s, 3H), 2.08 (s, 9H), 4.16 (dd, *J* 5.6 Hz and 12.4 Hz, 1H), 4.23 (dd, *J* 3.6 Hz and 7.6 Hz, 1H), 4.60 (dd, *J* 2.4 Hz and 12.4 Hz, 1H), 4.67 (d, *J* 6.0 Hz, 1H), 4.80 (dd, *J* 4.0 Hz and 6.0 Hz, 1H), 5.29 (m, 1H), 6.16 (s, 1H); ^{13}C NMR (CDCl_3): δ_{C} 21.0, 21.1, 21.2, 25.1, 26.2, 63.2, 69.2, 79.4, 80.2, 84.9, 100.8, 113.7, 169.4, 169.8 and 170.8; ESIMS: Calcd for $\text{C}_{15}\text{H}_{22}\text{O}_9$, *m/z* 346.33; found, 369.3 ($\text{M}+\text{Na}$) $^+$.

3.3.9. 2-(*R*)-[1'(*R*), 2'-Di-*O*-acetyethyl]-[1,3]oxathiolan-5-one (19)

Colorless solid, mp 75–77 °C; ^1H NMR ($\text{DMSO}-d_6$): δ_{H} 2.00 (s, 3H), 2.09 (s, 3H), 3.78 (d, *J* 16.8 Hz, 1H), 3.85 (d, *J* 16.8 Hz, 1H), 4.16 (dd, *J* 6.8 Hz and 12.0 Hz, 1H), 4.26 (dd, *J* 4.4 Hz and 12.0 Hz, 1H), 5.21 (m, 1H), 5.84 (d, *J* 4.0 Hz, 1H); ^{13}C NMR ($\text{DMSO}-d_6$): δ_{C} 20.6, 20.7, 30.0, 61.9, 72.5, 78.8, 169.6, 170.2, 173.2; HRESIMS: Calcd for $\text{C}_9\text{H}_{12}\text{O}_6\text{S}$, *m/z* 248.0355; found, *m/z* 248.0357 ($\text{M}+\text{Na}$) $^+$.

3.3.10. 1,3,5-Tri-*O*-acetyl-2-*O*-(2-acetoxyisopropyl)- β -D-xylofuranoside (20)

Oil; ^1H NMR (CDCl_3): δ_{H} 1.45 (s, 3H), 1.46 (s, 3H), 2.05 (s, 3H), 2.06 (s, 3H), 2.07 (s, 3H), 2.10 (s, 3H), 4.10 (dd, *J* 3.6 Hz and 8.0 Hz, 1H), 4.35 (m, 2H), 5.33 (m, 2H), 6.16 (d, *J* 1.6 Hz, 1H); ^{13}C NMR (CDCl_3): δ_{C} 13.6, 13.7, 13.8, 14.1, 19.6, 20.0, 54.9, 62.4, 62.5, 73.3, 89.4, 106.8, 162.8, 162.9, 163.0, 163.5; ESIMS: Calcd for $\text{C}_{16}\text{H}_{24}\text{O}_{10}$, *m/z* 376.14; found, *m/z* 399.1 ($\text{M}+\text{Na}$) $^+$.

3.3.11. 1,3-Di-*O*-acetyl-2-*O*-(2-acetoxyisopropyl)-5-deoxy- β -D-xylofuranoside (22)

Oil; ^1H NMR ($\text{DMSO}-d_6$): δ_{H} 1.14 (d, *J* 6.4 Hz, 3H), 1.39 (s, 3H), 1.41 (s, 3H), 1.99 (s, 3H), 2.03 (s, 3H), 2.08 (s, 3H), 4.35 (dd, *J* 2.4 Hz and 6.4 Hz, 1H), 5.05 (m, 1H), 5.12 (t, *J* 4.2 Hz, 1H), 6.13 (d, *J* 2.3 Hz, 1H); ^{13}C NMR ($\text{DMSO}-d_6$): δ_{C} 16.2, 20.6, 20.8, 21.0, 26.7, 26.9, 68.2, 72.6, 80.2, 95.6, 112.7, 169.7, 170.0; ESIMS: Calcd for $\text{C}_{14}\text{H}_{22}\text{O}_8$, *m/z* 318.13; found, *m/z* 341.1 ($\text{M}+\text{Na}$) $^+$.

3.3.12. 2-*O*-(2-Acetoxyisopropyl)-1,3-di-*O*-acetyl-5-*O*-*p*-toluenesulfonyl- β -D-xylofuranoside (24)

Oil; ^1H NMR (CDCl_3): δ_{H} 1.44 (s, 6H), 2.00 (s, 3H), 2.07 (s, 3H), 2.09 (s, 3H), 2.45 (s, 3H), 4.23 (dd, *J* 2.0 Hz and 4.0 Hz, 2H), 4.38 (dd, *J* 2.4 Hz and 4.0 Hz, 1H), 5.23 (d, *J* 5.2 Hz, 1H), 5.3 (dd, *J* 4.0 Hz and 5.6 Hz, 1H), 6.13 (d, *J* 1.0 Hz, 1H), 7.36 (d, *J* 4.0 Hz, 2H), 7.78 (d, *J* 4.0 Hz, 2H); ^{13}C NMR (CDCl_3): δ_{C} 20.8, 20.9, 21.4, 21.9, 26.9, 27.2, 67.3, 69.3, 69.5, 80.3, 96.3, 114.1, 128.3, 130.2, 132.7, 145.4, 169.9, 170.2; ESIMS: Calcd for $\text{C}_{21}\text{H}_{28}\text{O}_{11}\text{S}$, *m/z* 488.14; found, *m/z* 511.2 ($\text{M}+\text{Na}$) $^+$.

3.3.13. 2-*O*-(2-Acetoxyisopropyl)-1-*O*-acetyl-3,5-di-*O*-benzoyl- β -D-xylofuranoside (26)

Oil; ^1H NMR (CDCl_3): δ_{H} 1.50 (s, 3H), 1.54 (s, 3H), 2.02 (s, 6H), 4.52 (m, 1H), 4.62 (m, 2H), 5.74 (m, 2H), 6.33 (d, *J* 2.4 Hz, 1H), 7.44 (m, 4H), 7.57 (m, 2H), 8.03 (m, 4H); ^{13}C NMR (CDCl_3): δ_{C} 20.8, 21.1, 26.7, 26.8, 62.9, 69.6, 70.1, 80.5, 96.5, 113.8, 128.4, 128.6, 128.9, 129.4, 129.7, 129.9, 133.3, 133.6, 165.5, 166.1, 170.0, 171.0; ESIMS: Calcd for $\text{C}_{26}\text{H}_{28}\text{O}_{10}$, *m/z* 500.49; found, *m/z* 523.1 ($\text{M}+\text{Na}$) $^+$.

3.3.14. 2-*O*-(2-Acetoxyisopropyl)-1,3-di-*O*-acetyl-5-*O*-benzoyl- β -D-xylofuranoside (28)

Oil; ^1H NMR ($\text{DMSO}-d_6$): δ_{H} 1.42 (s, 3H), 1.43 (s, 3H), 2.05 (s, 3H), 2.07 (s, 3H), 2.09 (s, 3H), 4.34 (dd, *J* 6.8 Hz and 12.0 Hz, 1H), 4.48 (m, 2H), 5.42 (m, 2H), 6.18 (d, *J* 2.0 Hz, 1H), 7.54 (m, 2H), 7.67 (m, 1H), 7.92 (m, 2H); ^{13}C NMR ($\text{DMSO}-d_6$): δ_{C} 20.6, 20.7, 21.0, 26.72, 27.0, 62.8, 69.1, 69.7, 80.2, 95.7, 113.0, 129.0, 129.4, 133.7, 165.4, 169.8, 169.9; ESIMS: Calcd for $\text{C}_{21}\text{H}_{26}\text{O}_{10}$, *m/z* 438.15; found, *m/z* 461.2 ($\text{M}+\text{Na}$) $^+$.

3.3.15. 2-*O*-(2-Acetoxyisopropyl)-1-*O*-acetyl-5-*O*-benzoyl-3-*O*-*p*-toluenesulfonyl- β -D-xylofuranoside (30)

Oil; ^1H NMR (CDCl_3): δ_{H} 1.36 (s, 3H), 1.45 (s, 3H), 2.06 (s, 3H), 2.08 (s, 3H), 2.39 (s, 3H), 4.39 (m, 2H), 4.53 (m, 1H), 5.14 (dd, *J* 3.6 Hz and 5.2 Hz, 1H), 5.52 (dd, *J* 5.2 Hz and 10.4 Hz, 1H), 6.33 (d, *J* 2.2 Hz, 1H), 7.28 (t, *J* 8.0 Hz, 2H), 7.46 (m, 2H), 7.59 (m, 1H), 7.81 (d, *J* 8.3 Hz, 2H), 8.01 (d, *J* 0.7 Hz, 2H); ^{13}C NMR (CDCl_3): δ_{C}

20.8, 21.1, 26.4, 26.8, 29.7, 62.2, 68.9, 77.0, 80.4, 96.5, 114.2, 127.8, 128.5, 129.3, 129.7, 129.9, 133.3, 145.4, 165.7, 169.9, 170.0; ESIMS: Calcd for $C_{26}H_{30}O_{11}S$, m/z 550; found, m/z 550 (M^+).

3.3.16. 2-O-(2-Acetoxyisopropyl)-1-O-acetyl-3,5-di-O-benzyl- β -D-xylofuranoside (32)

Oil; 1H NMR ($CDCl_3$): δ_H 1.45 (s, 3H), 1.48 (s, 3H), 2.07 (s, 3H), 2.08 (s, 3H), 3.68 (m, 2H), 3.94 (t, J 4.8 Hz, 1H), 4.34 (dd, J 2.4 Hz and 3.6 Hz, 1H), 4.51 (dd, J 12.0 Hz and 22.4 Hz, 2H), 4.72 (s, 2H), 5.30 (m, 1H), 6.26 (d, J 2.8 Hz, 1H), 7.32 (m, 10H); ^{13}C NMR ($CDCl_3$): δ_C 21.3, 21.4, 26.7, 27.0, 68.1, 72.0, 73.4, 74.6, 76.2, 81.6, 96.8, 113.2, 128.0, 128.1, 128.6, 138.0, 138.1, 170.4, 170.5; ESIMS: Calcd for $C_{26}H_{32}O_8$, m/z 472.53; found, m/z 495.3 ($M+Na$) $^+$.

3.3.17. 2-O-(2-Acetoxyisopropyl)-1,5,6-tri-O-acetyl-3-O-benzoyl- β -D-glucufuranoside (33)

Oil; 1H NMR ($DMSO-d_6$): δ_H 1.39 (s, 6H), 1.92 (s, 3H), 1.99 (s, 3H), 2.02 (s, 3H), 2.05 (s, 3H), 4.12 (m, 1H), 4.28 (dd, J 2.8 Hz and 12.3 Hz, 1H), 4.51 (dd, J 2.0 Hz and 4.6 Hz, 1H), 5.15 (m, 1H), 5.47 (dd, J 3.7 Hz and 6.8 Hz, 1H), 5.8 (t, J 4.2 Hz, 1H), 6.22 (d, J 2.0 Hz, 1H), 7.57 (m, 2H), 7.70 (m, 1H), 7.96 (m, 2H); ^{13}C NMR ($DMSO-d_6$): δ_C 20.4, 20.5, 20.8, 26.6, 26.9, 61.4, 68.3, 68.8, 69.2, 80.7, 95.5, 112.9, 128.8, 128.9, 129.3, 133.9, 164.9, 169.2, 169.5, 169.6, 170.0; ESIMS: Calcd for $C_{24}H_{30}O_{12}$, m/z 510.17; found, m/z 533.2 ($M+Na$) $^+$.

3.3.18. 2-O-(2-Acetoxyisopropyl)-1,3,5,6-tetra-O-acetyl- β -D-glucufuranoside (34)

Oil; 1H NMR ($CDCl_3$): δ_H 1.44 (s, 3H), 1.45 (s, 3H), 2.04 (s, 3H), 2.06 (s, 3H), 2.07 (s, 3H), 2.13 (s, 3H), 2.10 (s, 3H), 4.12 (dd, J 3.2 Hz and 8.4 Hz, 1H), 4.22 (d, J 8.4 Hz, 1H), 4.32 (s, 1H), 5.13 (m, 1H), 5.34 (m, 1H), 5.44 (d, J 4.2 Hz, 1H), 6.20 (s, 1H); ^{13}C NMR ($CDCl_3$): δ_C 20.9, 21.0, 21.1, 21.4, 26.4, 27.1, 61.9, 68.2, 68.6, 68.9, 81.2, 96.4, 113.9, 170.0, 170.1, 170.3, 170.8; ESIMS: Calcd for $C_{19}H_{28}O_{12}$, m/z 448.16; found, m/z 487.2 ($M+K$) $^+$.

3.3.19. 2-O-(2-Acetoxyisopropyl)-1,5,6-tri-O-acetyl-3-O-p-toluenesulfonyl- β -D-glucufuranoside (35)

Oil; 1H NMR ($CDCl_3$): δ_H 1.32 (s, 3H), 1.42 (s, 3H), 2.06 (s, 3H), 2.07 (s, 3H), 2.09 (s, 3H), 2.13 (s, 3H), 2.45 (s, 3H), 4.14 (dd, J 4.8 Hz and 12.4 Hz, 1H), 4.37 (dd, J 2.0 Hz and 12.4 Hz, 1H), 4.46 (s, 1H), 5.18 (dd, J 3.6 Hz and 8.0 Hz, 2H), 5.51 (dd, J 4.4 Hz and 6.8 Hz, 1H), 6.30 (s, 1H), 7.35 (d, J 8.4 Hz, 2H), 7.81 (d, J 8.0 Hz, 2H); ^{13}C NMR ($CDCl_3$): δ_C 20.9, 21.2, 21.3, 21.9, 26.5, 26.8, 29.9, 61.3, 68.7, 69.4, 76.0, 81.0, 96.5, 114.2, 128.2, 130.0, 133.5, 145.5, 169.8, 170.0, 170.3, 170.9; ESIMS: Calcd for $C_{24}H_{32}O_{13}S$, m/z 560.16; found, m/z 583.2 ($M+Na$) $^+$.

3.3.20. 2-O-(2-Acetoxyisopropyl)-1,5-di-O-acetyl-3-O-tert-butyltrimethylsilyl- β -D-xylofuranoside (37)

Oil; 1H NMR ($CDCl_3$): δ_H 0.13 (s, 6H), 0.9 (s, 9H), 1.48 (s, 6H), 2.05 (s, 3H), 2.07 (s, 3H), 2.10 (s, 3H), 4.05 (t, J 4.0 Hz, 1H), 4.22 (dd, J 3.6 Hz and 12.0 Hz, 1H), 4.28 (m, 1H), 4.35 (dd, J 3.6 Hz and 12.0 Hz, 1H), 5.19 (t, J 4.0 Hz, 1H), 6.30 (d, J 2.4 Hz, 1H); ^{13}C NMR ($CDCl_3$): δ_C -4.5, 18.2, 21.0, 21.2, 21.4, 25.7, 25.8, 26.7, 62.76, 69.7, 72.0, 81.9, 96.8, 113.4, 170.5, 170.8; ESIMS: Calcd for $C_{20}H_{36}O_9Si$, m/z 448.21; found, m/z 471.2 ($M+Na$) $^+$.

3.3.21. 2-O-(2-Acetoxyisopropyl)-1-O-acetyl-3,4-O-isopropylidene- α,β -L-arabinopyranoside (39)

Oil; 1H NMR ($DMSO-d_6$): δ_H 1.22–1.45 (m, 12H), 1.94–2.08 (m, 6H), 3.95–4.44 (m, 5H), 6.06–6.27 (m, 1H); ^{13}C NMR ($DMSO-d_6$): δ_C 25.2, 25.6, 26.0, 26.6, 26.7, 27.0, 27.7, 27.9, 28.1, 62.4, 63.4, 73.8, 74.4, 74.7, 75.6, 77.0, 77.3, 80.1, 83.9, 93.2, 96.7, 108.4,

108.6, 111.3, 112.5, 169.9, 170.0, 170.1, 170.3; ESIMS: Calcd for $C_{15}H_{24}O_8$, m/z 332.15; found, 355.2 ($M+Na$) $^+$.

3.3.22. 1,2,6-Tri-O-acetyl-3,4-O-isopropylidene- α -D-galactopyranoside (41)

Oil; 1H NMR ($DMSO-d_6$): δ_H 1.29 (s, 3H), 1.42 (s, 3H), 2.03 (s, 3H), 2.05 (s, 3H), 2.10 (s, 3H), 4.18 (m, 2H), 4.32 (m, 1H), 4.38 (m, 2H), 4.88 (dd, J 3.6 Hz and 7.2 Hz, 1H), 6.04 (d, J 3.6 Hz, 1H); ^{13}C NMR ($DMSO-d_6$): δ_C 20.5, 20.6, 25.9, 27.4, 62.8, 67.7, 69.2, 71.8, 72.3, 88.4, 109.3, 169.0, 169.8, 170.2; ESIMS: Calcd for $C_{15}H_{22}O_9$, m/z 346.13; found, 369.2 ($M+Na$) $^+$.

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Supplementary data

Supplementary data associated with this article can be found, in the online version, at [doi:10.1016/j.carres.2009.08.035](https://doi.org/10.1016/j.carres.2009.08.035).

References

- Kocienski, P. J. *Protecting Groups*; Georg Thieme: Stuttgart, 2004.
- Sartori, G.; Ballini, R.; Bigi, F.; Bosica, G.; Maggi, R.; Righi, P. *Chem. Rev.* **2004**, *104*, 199–250.
- Wu, Q.; Chen, W.; Wang, Y.; Qu, Y.; Zhang, Q. *Lett. Org. Chem.* **2006**, *3*, 271–274.
- Ichikawa, E.; Kato, K. *Curr. Med. Chem.* **2001**, *8*, 385–402.
- Gruner, S. A. W.; Locardi, E.; Lohof, E.; Kessler, H. *Chem. Rev.* **2002**, *102*, 491–514.
- Nicolaou, K. C.; Mitchell, H. J. *Angew. Chem., Int. Ed.* **2001**, *40*, 1576–1624.
- Kunz, H. *Pure Appl. Chem.* **1995**, *67*, 1627–1633.
- Hultin, P. G.; Earle, M. A.; Sudharshan, M. *Tetrahedron* **1997**, *53*, 14823–14870.
- Schulze, O.; Voss, J.; Adiwidjaja, G. *Carbohydr. Res.* **2005**, *340*, 561–587.
- Wu, Q.; Simons, C. *Synthesis* **2004**, 1533–1553.
- Simons, C. In *Nucleoside Mimetics, Their Chemistry and Biological Properties*; Gordon and Breach Science: Australia, Canada, 2001; 6.
- Gunaga, P.; Moon, H. R.; Choi, W. J.; Shin, D. H.; Park, J. G.; Jeong, L. S. *Curr. Med. Chem.* **2004**, *11*, 2585–2590.
- Kodama, T.; Matsuda, A.; Shuto, S. *Tetrahedron* **2006**, *62*, 10011–10018.
- Koo, B.; McDonald, F. E. *Org. Lett.* **2005**, *7*, 3621–3624.
- Kelly, T. R.; Cai, X.; Tu, B.; Elliott, E. L.; Grossmann, G.; Laurent, P. *Org. Lett.* **2004**, *6*, 4953–4956.
- Miyashita, K.; Ikejiri, M.; Kawasaki, H.; Maemura, S.; Imanishi, T. *J. Am. Chem. Soc.* **2003**, *125*, 8238–8243.
- Chen, S.-H.; Lin, S.; King, I.; Spinka, T.; Dutschman, G. E.; Gullen, E. A.; Cheng, Y.-C.; Doyle, T. W. *Bioorg. Med. Chem. Lett.* **1998**, *8*, 3245–3250.
- Gilbert, C. L. K.; Lisek, C. R.; White, R. L.; Gumina, G. *Tetrahedron* **2005**, *61*, 8339–8344.
- Sairam, P.; Puranik, R.; Rao, B. S.; Swamy, P. V.; Chandra, S. *Carbohydr. Res.* **2003**, *338*, 303–307.
- Ozols, A. M.; Azharyev, A. V.; Dyatkina, N. B.; Krayevsky, A. A. *Synthesis* **1980**, 557–560.
- Roy, A.; Achari, B.; Mandal, S. B. *Tetrahedron Lett.* **2006**, *47*, 3875–3878.
- Sabitha, G.; Bhaskar, V.; Reddy, S. S.; Yadav, J. S. *Tetrahedron* **2008**, *64*, 10207–10213.
- Chakraborty, T. K.; Purkait, S. *Tetrahedron Lett.* **2008**, *49*, 5502–5504.
- Sengupta, J.; Bhattacharjya, A. *J. Org. Chem.* **2008**, *73*, 6860–6863.
- Wu, Q.-P.; Zhou, M.-X.; Xi, X.-D.; Song, D.; Wang, Y.; Liu, H.-X.; Li, Y.-Zh.; Zhang, Q.-Sh. *Tetrahedron Lett.* **2008**, *49*, 2714–2718.
- Chakraborti, A. K.; Gulhane, R. *Chem. Commun. (Cambridge)* **2003**, 1896–1897.
- Chakraborti, A. K.; Gulhane, R. Indian Patent, 266/DEL/2003, March 10, 2003.
- Khan, A. T.; Parvin, T.; Choudhury, L. H. *Synthesis* **2006**, 2450–2497.
- Rudrawar, S.; Besra, R. C.; Chakraborti, K. A. *Synthesis* **2006**, 2767–2770.
- Kumar, R.; Kumar, D.; Chakraborti, A. K. *Synthesis* **2007**, 299–303.
- Khan, A. T.; Choudhury, L. H.; Ghosh, S. J. *Mol. Catal. A: Chem.* **2006**, *255*, 230–233.
- Kumar, R.; Tiwari, P.; Maulik, P. R.; Misra, A. K. *J. Mol. Catal. A: Chem.* **2006**, *247*, 27–31.
- Misra, A. K.; Tiwari, P.; Madhusudan, S. K. *Carbohydr. Res.* **2005**, *340*, 325–329.
- Mukhopadhyay, B.; Russell, D. A.; Field, R. A. *Carbohydr. Res.* **2005**, *340*, 1075–1080.
- Agnihotri, G.; Tiwari, P.; Misra, A. K. *Carbohydr. Res.* **2005**, *340*, 1393–1396.
- Du, Y.; Wei, G.; Cheng, S.; Hua, Y.; Linhardt, R. J. *Tetrahedron Lett.* **2006**, *47*, 307–310.

37. Mukhopadhyay, B.; Collet, B.; Field, R. A. *Tetrahedron Lett.* **2005**, 46, 5923–5925.
38. Das, B.; Venkateswarlu, K.; Suneel, K.; Majhi, A. *Tetrahedron Lett.* **2007**, 48, 5371–5374.
39. Khatik, L. G.; Sharma, G.; Kumar, R.; Chakraborti, A. K. *Tetrahedron* **2007**, 63, 1200–1210.
40. Chakraborti, A. K.; Chankeshwara, S. V. *Org. Biomol. Chem.* **2006**, 4, 2769–2771.
41. Angyal, S. J.; Wheen, R. G. *Aust. J. Chem.* **1980**, 33, 1001–1011.
42. Wu, Q.-P.; Shi, D.-X.; Wang, H.; Zhang, Q.-Sh. *Acta Crystallogr., Sect. E* **2009**, 65, o1483.
43. Tiwari, P.; Misra, A. K. *Carbohydr. Res.* **2006**, 341, 339–350.