



6 β ,19-Bridged androstenedione analogs as aromatase inhibitors

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ABSTRACT

Inhibition of aromatase is an efficient approach for the prevention and treatment of breast cancer. New 6 β ,19-bridged steroid analogs of androstenedione, 6 β ,19-epithio- and 6 β ,19-methano compounds **11** and **17**, were synthesized starting from 19-hydroxyandrostenedione (**6**) and 19-formylandrost-5-ene-3 β ,17 β -yl diacetate (**12**), respectively, as aromatase inhibitors. All of the compounds including known steroids 6 β ,19-epoxyandrostenedione (**4**) and 6 β ,19-cycloandrostenedione (**5**) tested were weak to poor competitive inhibitors of aromatase and, among them, 6 β ,19-epoxy steroid **4** provided only moderate inhibition (K_i : 2.2 μ M). These results show that the 6 β ,19-bridged groups of the inhibitors interfere with binding in active site of aromatase.

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1. Introduction

Aromatase is the cytochrome P-450–enzyme complex responsible for estrogen biosynthesis [1–3]. The aromatase reaction is thought to proceed through three sequential oxygenations at C-19 of the androgen androst-4-ene-3,17-dione (androstenedione) and testosterone [4–11] (Fig. 1). The first two oxygenations occur at C-19 position to produce 19-hydroxy and 19,19-dihydroxy intermediates. In the third step, C-19 and the 1 β ,2 β -hydrogens are eliminated as formic acid and water, respectively. There appear to be a major clinical role for methods of controlling estrogen-dependent breast cancer. The inhibitors of aromatase represent one route to such control [12–15].

A number of potent aromatase inhibitors, analogs of the natural substrate androstenedione, have been the subjects of clinical trials. Previously, we have reported a series of 6 β -alkylandrostenediones as aromatase inhibitors of which 6 β -ethyl compound is extremely powerful inhibitor with K_i of 1.4 nM [16–19]. The 6 β -hydroxy derivative, a polar steroid, has also high activity to aromatase [20,21]. On the other hand, 19-substituted (hydroxy, methyl and sulfur) androstenedione analogs have been reported as good to powerful competitive inhibitors of the enzyme [22–27]. Considering the facts obtained by the 6 β - and 19-substituted steroids, we aimed at compounds with 6 β ,19-bridged androstenediones as aromatase inhibitors. Previously, 6 β ,19-bridged steroids, 6 β ,19-epoxy-

[28,29], 6 β ,19-epithio- [30], 6 β ,19-methano- [31], and 6 β ,19-cyclo- [32] progesterones and its 21-hydroxy analogs, have been shown to possess new biological activities of steroid hormones (Fig. 2). However, effect of 6 β ,19-bridged structures on the aromatase activity have not been explored so far. In this paper, we describe the synthesis of 6 β ,19-bridged steroids, epoxy-, cyclo-, epithio-, and methano-androstenediones (**4**, **5**, **11**, and **17**), and their inhibitory activity of human placental aromatase.

2. Experimental

2.1. Materials and general methods

6 β ,19-Epoxyandrost-4-ene-3,17-dione (**4**) [33] and 6 β ,19-cycloandrost-4-ene-3,17-dione (**5**) [34] were synthesized by the previous reported methods. [1 β -³H] androstenedione (specific activity, 27.5 Ci/mmol; ³H-distribution 74–79%) was obtained from New England Nuclear (Boston, MA, USA). NADPH was purchased from Sigma–Aldrich (St. Louise, MO, USA).

Melting points were measured on a Yanagimoto melting point apparatus (Kyoto, Japan) and are uncorrected. Infra red (IR) spectra were recorded on a PerkinElmer FT-IR 1725X spectrophotometer in a KBr pellet or Nujol form, and ultra-violet (UV) spectra were determined in 95% ethanol on a Hitachi 150-20 spectrophotometer (Tokyo, Japan). Nuclear magnetic resonance (NMR) spectra were obtained in CDCl₃ solution with JEOL LA 400 (400 MHz for ¹H) and JEOL LA 600 (600 MHz for ¹H and 150 MHz for ¹³C) spectrometer (Tokyo, Japan) using tetramethylsilane as an internal standard, and mass (MS) spectra (electron impact mode) and high resolution

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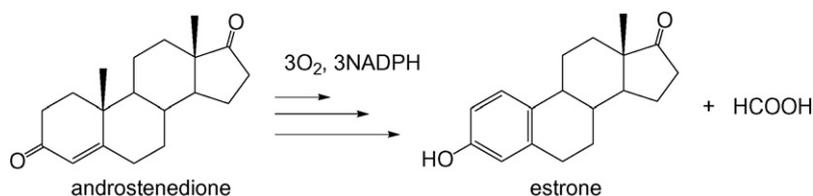


Fig. 1. Aromatase reaction of androstenedione by human placental aromatase.

(HR)-MS with a JEOL JMS-700 spectrometer. Thin-layer chromatography (TLC) was performed on E. Merck precoated plates (silica gel 60F-254, Darmstadt, Germany). Column chromatography was conducted with silica gel 60, 70–230 mesh (E. Merck).

2.2. 6 β ,19-Cycloandrost-4-ene-3,17-dione (5)

p-Toluenesulfonyl chloride (1.13 g, 5.93 mmol) was added to a solution of 19-hydroxyandrostenedione (**6**) (556 mg, 1.84 mmol) in pyridine (27.8 mL) at 0 °C [34]. The reaction mixture was stand for 30 min and allowed to warm to room temperature. After 116 h, the mixture was poured to ice water, acidified by 10% HCl, extracted with EtOAc. The organic layer was washed with 5% NaHCO₃ solution and water and dried with Na₂SO₄. The evaporation of the organic solvent afforded 19-tosylate (802 mg, 96%). 28% NaOCH₃ in CH₃OH solution (0.48 mL) was added to the 19-tosylate (402 mg, 0.88 mmol) in CH₃OH (12.4 mL) under nitrogen. The mixture was heated under reflux for 5.5 h, diluted with EtOAc, washed with sat. NaHCO₃ solution and water, and dried with Na₂SO₄. Evaporation of the solvent gave an oil which was purified by column chromatography (hexane–EtOAc, 5:1, v/v) followed by preparative TLC (hexane–EtOAc, 1:1, v/v). A solid material was recrystallized from acetone to give 6 β ,19-cycloandrostenedione (**5**) (110 mg, 44%), mp 128–132 °C (reported, 132 °C [34]). ¹H NMR δ : 0.97 (3H, s, 18-CH₃), 3.24 (1H, t, *J* = 5.6 Hz, 6-H), 5.61 (1H, s, 4-H). ¹³C NMR δ : 14.43 (C-18), 21.28 (C-11), 24.39 (C-15), 30.67 (C-12), 31.69 (C-2), 32.35 (C-1), 33.72 (C-8), 33.95 (C-19), 35.73 (C-16), 43.24 (C-6), 49.02 (C-13), 49.48 (C-9), 49.79 (C-14), 49.99 (C-10), 111.06 (C-4), 180.19 (C-5), 198.88 (C-3), 219.46 (C-17). IR (KBr) cm⁻¹: 1732 and 1655 (C=O). UV λ_{max} nm (ϵ): 242 (13,900). MS *m/z*: 284 (M⁺, 100%). Analysis calculated for C₁₉H₂₄O₂: C, 80.24; H, 8.51. Found: C, 80.15; H, 8.56.

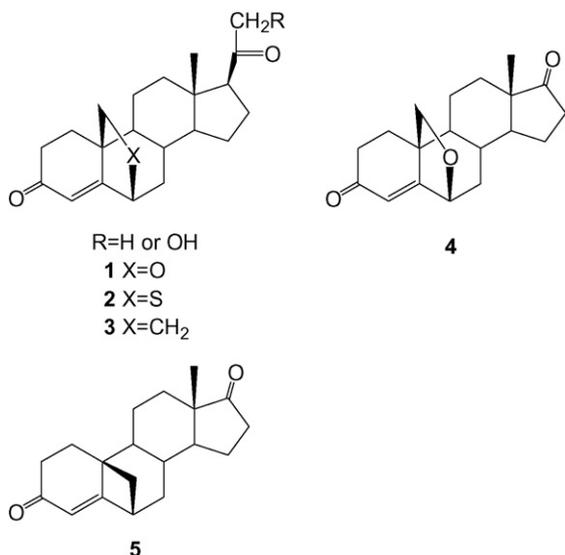


Fig. 2. Structures of 6,19-bridged progesterone analogs and androstenediones.

2.3. 19-Acetylthioandrost-4-ene-3,17-dione (8)

A solution of 19-hydroxyandrostenedione (**6**) (1.08 g, 3.57 mmol) in cold pyridine (10.8 mL) was added dropwise to a stirred solution of trifluoromethanesulfonic anhydride (1.34 mL, 7.96 mmol) in cold pyridine (7.4 mL) under nitrogen [30]. The mixture was allowed to warm to room temperature for 1.5 h and diluted with EtOAc. After acidification with 5% HCl, the organic layer was washed with sat. NaHCO₃ solution and water and dried with Na₂SO₄. A yellow solid, a crude 19-triflate (1.42 g), obtained by evaporation of the solvent, was used without further purification in the following step.

A solution of the 19-triflate **7** (1.42 g) and potassium thioacetate (1.42 g, 12.45 mmol) in acetone (73 mL) was stirred at room temperature for 24 h under nitrogen. The reaction mixture was then diluted with EtOAc, washed with sat. NaHCO₃ solution and water, and dried with Na₂SO₄. Evaporation of the solvent yielded an oil which was purified by column chromatography (hexane–EtOAc, 3:1, v/v) to afford 19-thioacetate **8** as pale yellow oil (558 mg, 43%). ¹H NMR δ : 0.95 (3H, s, 18-CH₃), 2.34 (3H, s, SCOCH₃), 3.20 and 3.52 (1H each, *d*, *J* = 14.0 Hz, 19-H₂), 5.90 (1H, s, 4-H). IR (Nujor) cm⁻¹: 3586 (OH), 1733, 1683, and 1652 (C=O). UV λ_{max} nm (ϵ): 238 (16,600). MS *m/z*: 360 (M⁺, 31%), 317 (58), 284 (18), 272 (100). HR-MS calculated for C₂₁H₂₈O₃S: 360.1759. Found: 360.1766.

2.4. 3,17-Diethylenedioxy-19-acetylthioandrost-5-ene (9)

A mixture of the 19-thioacetate **8** (502 mg, 1.39 mmol) and ethylene glycol (3.5 mL) in benzene (16 mL) was distilled for 15 min to remove traces water. *p*-Toluenesulfonic acid monohydrate (15 mg) was added to the reaction mixture and the mixture was heated under reflux for 4 h, diluted with EtOAc, and added sat. NaHCO₃ solution. The organic layer was washed with water, dried with Na₂SO₄, and evaporated to give a semi-solid which was purified by column chromatography (hexane–EtOAc, 20:1, v/v) to afford ketal **9** (432 mg, 69%) as an oil. ¹H NMR δ : 0.87 (3H, s, 18-CH₃), 2.31 (3H, s, SCOCH₃), 3.05 and 3.43 (1H each, *d*, *J* = 14.2 Hz, 19-H₂), 3.85 (8H, m, 3,17-OCH₂CH₂O-), 5.46 (1H, m, 6-H). IR (Nujor) cm⁻¹: 1689 (C=O). UV λ_{max} nm (ϵ): 235 (5200). MS *m/z*: 448 (M⁺, 21%), 405 (17), 359 (32), 297 (69), 99 (100). HR-MS calculated for C₂₅H₃₆O₅S: 448.2284. Found: 448.2292.

2.5. 6 β ,19-Epithioandrost-4-ene-3,17-dione (11)

A solution of the ketal **9** (287 mg, 0.64 mmol) in dry CH₃OH (8 mL) was deoxygenated by bubbling dry nitrogen for 15 min. Then a solution of potassium hydroxide (58 mg, 1.04 mmol) in CH₃OH (0.4 mL) was added and the mixture was stirred at room temperature for 30 min. The reaction mixture was neutralized with 5% HCl and diluted EtOAc. The organic layer was washed with sat. NaHCO₃ solution and water, and dried with Na₂SO₄, and evaporated to afford the crude 19-thiol **10** (176 mg). Iodine (234 mg, 0.93 mmol) and triethylamine (61 mL, 0.43 mmol) in dry CH₂Cl₂ (115 mL) were added to a solution of the crude 19-thiol **10** (176 mg) in CH₂Cl₂ (72 mL) at 0 °C, then the mixture was stirred and allowed to warm to room temperature. After 2 h, the reaction mixture was diluted with EtOAc and washed with sat. Na₂S₂O₃ solution and water, and dried with

Na₂SO₄. Evaporation of the solvent gave a solid which was purified by column chromatography (hexane–EtOAc, 3:1, v/v) to yield a crude 6,19-epithioandrostenedione **11** (33 mg, 16%). Recrystallization of the crude compound from acetone afforded compound **11**, mp 251–256 °C. ¹H NMR δ: 0.99 (3H, s, 18-CH₃), 2.61 and 3.07 (1H each, d, *J* = 10.3 Hz, 19-H₂), 3.95 (1H, m, 6-H), 5.81 (1H, s, 4-H). IR (KBr) cm⁻¹: 1738 and 1667 (C=O). UV λ_{max} nm (ε): 241 (14,000). MS *m/z*: 316 (M⁺, 100%), 288 (10), 260 (18), 242 (15), 152 (34). Analysis calculated for C₁₉H₂₄O₂S: C, 72.11; H, 7.64; S, 10.13. Found: C, 72.35; H, 7.30; S, 10.20.

2.6. 5α-Chloro-19a-hydroxy-6β,19-methanoandrostane-3β,17β-yl diacetate (**13**)

A solution of 19-formylandrost-5-ene-3β,17β-yl diacetate (**12**) (608 mg, 1.51 mmol) in dry CH₂Cl₂ (6 mL) was added to a solution of titanium chloride (3 mL) in CH₂Cl₂ (10 mL) at –70 °C under a nitrogen atmosphere [31]. The reaction mixture was stirred at –30 °C for 2 h, and C₂H₅OH–water (1:3, 12 mL) was added. The resulting solution was further stirred at 0 °C for 15 min. The mixture was diluted with EtOAc, washed with sat. NaHCO₃ solution and water, and dried with Na₂SO₄. The solvent was evaporated to give a solid which was purified by column chromatography (hexane–EtOAc, 8:1, v/v) and recrystallization from acetone to afford 5α-chloro-19a-hydroxy-6β,19-methanoandrostane-3β,17β-yl diacetate (**13**) (266 mg, 40%), mp 158–162 °C. ¹H NMR δ: 0.79 (3H, s, 18-CH₃), 2.72 (1H, t, *J* = 12.8 Hz, 6α-H), 4.27 (1H, d, *J* = 7.7 Hz, 19a-CH), 4.60 (1H, t, *J* = 8.4 Hz, 17-H), 5.15 (1H, m, 3-H). ¹³C NMR δ: 12.49 (C-18), 21.14 and 21.36 (3β- or 17β-OCOCH₃), 21.67 (C-11), 22.96 (C-15), 25.89 (C-1), 26.71 (C-2), 27.58 (C-16), 29.86 (C-7), 33.87 (C-8), 36.54 (C-19), 36.70 (C-12), 41.45 (C-4), 43.31 (C-10), 47.50 (C-14), 48.03 (C-13), 49.15 (C-9), 54.35 (C-6), 70.30 (C-3), 73.31 (C-19a), 82.08 (C-5), 82.41 (C-17), 170.45 and 171.10 (3β- or 17β-OCOCH₃). IR (KBr) cm⁻¹: 3465 (OH) and 1723 (C=O). MS *m/z*: 438 (M⁺, 0.5%), 378 (11), 334 (100), 298 (85). Analysis calculated for C₂₄H₃₅ClO₅: C, 65.66; H, 8.04. Found: C, 65.87; H, 8.25.

2.7. 5α-Chloro-6β,19-methanoandrostane-3β,17β-yl diacetate (**14**)

A mixture of DBU (110 μL) and CS₂ (348 μL) was added to a solution of the 19a-hydroxy-6β,19-methano steroid **13** (91 mg, 0.21 mmol) in dimethylformamide (1.25 mL) under nitrogen atmosphere. The reaction mixture was stirred for 1 h, then CH₃I (2.5 mL) was added [35]. After 2 h, the mixture was diluted with EtOAc which was washed successively with sat. KHSO₄ solution, sat. NaHCO₃ solution, and water. The organic layer was dried with Na₂SO₄. Evaporation of the solvent gave the residue which was dissolved in dry toluene (2 mL). Bu₃SnH (73 μL) was added to the solution under nitrogen and the mixture was heated under reflux for 17 h and another Bu₃SnH (73 μL) was added to the mixture which was heated under reflux for 6.5 h. Then the reaction mixture was diluted with EtOAc and insoluble materials were filtered off. Evaporation of the solvent yielded a solid which was purified by column chromatography (hexane–EtOAc, 10:1, v/v) followed by recrystallization from CH₃OH to give 5α-chloro-6β,19-methanoandrostane-3β,17β-yl diacetate **14** (47 mg, 54%), mp 110–114 °C. ¹H NMR δ: 0.82 (3H, s, 18-CH₃), 4.60 (1H, t, *J* = 8.3 Hz, 17-H), 5.15 (1H, m, 3-H). IR (KBr) cm⁻¹: 1733 (C=O). MS *m/z*: 422 (M⁺, 6%), 362 (45), 326 (76), 119 (100). Analysis calculated for C₂₄H₃₅ClO₄: C, 68.15; H, 8.34. Found: C, 68.33; H, 8.51.

2.8. 6β,19-Methanoandrost-4-ene-3,17-dione (**17**)

Potassium hydroxide (23.5 mg, 0.42 mmol) in CH₃OH (0.71 mL) was added to a solution of the diacetate **14** (50 mg, 0.12 mmol)

in CH₃OH (1.5 mL) under nitrogen and warmed at 50 °C. After 1.5 h, the reaction mixture was diluted with EtOAc and neutralized with 5% HCl. The organic layer was washed with sat. NaHCO₃ solution and water, and dried with Na₂SO₄. Evaporation of the solvent gave a crude 5α-chloro-6β,19-methanoandrostane-3β,17β-diol (**15**) (43 mg, 99%).

Pyridinium chlorochromate (113 mg, 0.53 mmol) and BaCO₃ (66 mg, 0.34 mmol) were added to a solution of the crude diol **15** (43 mg, 0.13 mmol) in CH₂Cl₂ (1.5 mL) under nitrogen atmosphere. The reaction mixture was stirred for 1.5 h and passed through silica gel column (5 g) to remove a dark brown solid. Elution with CH₂Cl₂ and evaporation of the solvent gave a crude 5α-chloro-6β,19-methanoandrostane-3,17-dione (**16**) (31 mg, 72%).

A solution of the crude 5α-androstane steroid **16** (29 mg, 0.09 mmol) in CH₂Cl₂ (1 mL) was stirred for 6 h in the presence of Al₂O₃ (192 mg, 1.88 mmol). Al₂O₃ was removed by filtration and the solvent was evaporated to yield a solid. The solid was recrystallized from acetone to afford 6β,19-methanoandrost-4-ene-3,17-dione (**17**) (10 mg, 42%), mp 196–199 °C. ¹H NMR δ: 0.95 (3H, s, 18-CH₃), 2.85 (1H, br. s, 6-H), 5.77 (1H, s, 4-H). IR (KBr) cm⁻¹: 2927 (SR), 1738 and 1670 (C=O). UV λ_{max} nm (ε): 243 (14,400). MS *m/z*: 298 (M⁺, 100%), 256 (84), 148 (44), 135 (82), 91 (39). Analysis calculated for C₂₀H₂₆O₂: C, 80.50; H, 8.78. Found: C, 80.28; H, 8.90.

2.9. Enzyme preparation

Human placental microsomes (sedimented after 60 min at 105,000 × *g*) were obtained as described by Ryan [36]. They were washed once with 0.05 mM dithiothreitol, lyophilized and stored at –80 °C. No significant loss of activity occurred during the period (6 months) of this study. The preparation of human placental microsomes was conducted under the approval of the ethical review committee of Tohoku Pharmaceutical University in accordance with the standard of the Helsinki Declaration.

2.10. Aromatase assay procedure

Aromatase activity was measured essentially according to the original procedure of Siiteri and Thompson [37]. The screening assay for determination of IC₅₀ value and the kinetic assay were carried out essentially according to the assay methods described in our previous work [21]. Briefly, 20 μg of protein from the lyophilized microsomes and 20-min incubation period were used for the screening assay, and 20 μg of protein from the microsomes and a 5-min period were used for the kinetic assay. The assays were carried out at 37 °C in 67 mM phosphate buffer, pH 7.4, in the presence of NADPH under air.

Apparent *K_i* values were calculated using non-linear regression analysis with GraFit software [38].

3. Results and discussion

3.1. Chemistry

6β,19-Epoxyandrostenedione (**4**) [33] and 6β,19-cycloandrostenedione (**5**) [34] were prepared according to the reported methods. The structure of the 6β,19-cyclo compound **5**, obtained by reaction of 19-tosylate of 19-hydroxyandrostenedione (**6**) with sodium methoxide, was reconfirmed by two-dimensional NMR spectroscopy.

Burton et al. [30,31] have reported synthesis of series of 6β,19-epithio- and 6β,19-methano-progesterone analogs to expect new biological activities as steroid hormones. In the course of our studies, we were of interest in 6β,19-bridged androstenedione analogs as the aromatase inhibitors. We initially tried

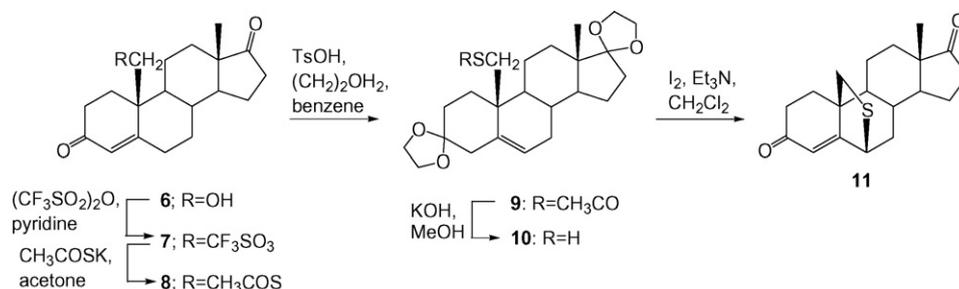


Fig. 3. Synthesis of 6,19-epithioandrostenedione (11).

synthesis of 6 β ,19-epithioandrostenedione (11) starting from 19-hydroxyandrostenedione (6) (Fig. 3). The 19-hydroxy group of compound 6 was converted to triflate 7 by treatment with trifluoromethanesulfonic anhydride in pyridine and displaced with fluoromethanesulfonic anhydride in pyridine and displaced with potassium thioacetate in acetone to give 19-thioacetyl derivative 8. The ^1H NMR spectrum of this compound showed a singlet peak of an acetylthio group (3H, 2.34 ppm) and two doublet peaks of 19- CH_2 (3.20 and 3.52 ppm, $J = 14.0$ Hz). Compound 8 was converted to ethyleneketal 9 in which the double bond migrated to the 5,6-position (^1H NMR: multiplet signal at 5.46 ppm of C-6) as required for the 6 β ,19-cyclization [30], and the 19-thioacetate was hydrolyzed with potassium hydroxide in methanol to give the free thiol 10. Treatment of the thiol 10 by triethylamine and iodine resulted in the one-pot conversion of compound 10 into 6 β ,19-epithio compound 11 based upon the ^1H NMR spectrum which showed 2.61 and 3.07 ppm (1H each, d, $J = 10.3$ Hz) for 19-H, and 3.95 ppm (1H, m) for 6-H.

Next, 6 β ,19-methanoandrostenedione (17) was synthesized from 19-formylandrost-5-ene-3 β ,17 β -yl diacetate (12) through six steps (Fig. 4). The key step in the synthesis was the cyclization of an additional carbon atom at C-19a [31]. Titanium tetrachloride has been the Lewis acid of choice for the Prins reaction to give bridged chloroalcohol 13. Treatment of compound 12 with the titanium salt in CH_2Cl_2 at -50°C gave the Prins reaction product, 5 α -chloro-6 β ,19-methano steroid 13, with a hydroxyl group at the C-19a position. The ^1H NMR spectrum showed a methine proton at 4.27 ppm ($J = 7.7$ Hz) for the 19a-position and 2.72 ppm ($J = 12.8$ Hz) for the 6 α -position. Thus the formation of the 6 β ,19-methano ring proceeded through a stereo-controlled way; the stereochemistry of the 19a-hydroxy group was not established by the NMR spectrometry. This hydroxyl group was eliminated by Barton deoxygenation

procedure [40]. The 19a-alcohol 13 was converted into the 19a-xanthate derivative by reaction with CS_2 and methyl iodide and this was treated with tributyltin hydride in toluene to give the deoxygenated compound 14. The ^1H NMR spectrum of this compound showed an absence of a hydroxymethine resonance at 3.5–4.5 ppm. The 3 β ,17 β -acetoxy moieties of compound 14 was eliminated by the alkaline hydrolysis to yield 3 β ,17 β -dihydroxy compound 15. Pyridinium chlorochromate oxidation of compound 15 followed by treatment with Al_2O_3 afforded the other desired compound, 6 β ,19-methano compound 17.

The structures of the compounds synthesized were confirmed by the spectrometric analysis, HR-MS, and elemental analysis.

3.2. Biochemical properties

The inhibitory activities of human placental aromatase by the 6 β ,19-bridged steroids 4, 5, 11 and 17 were tested. Aromatization activity in placental microsomes was determined using a radiometric assay in which tritiated water released from [1β - ^3H] androstenedione into the incubation medium during aromatization was measured [37]. To characterize the nature of inhibitor binding to the active site of aromatase, aromatization was measured at several concentrations and substrate concentrations. The results of these studies were plotted on a typical Lineweaver–Burk plot in all cases. The results are given in Table 1. In these studies, apparent K_m and V_{max} values for androstenedione were 35 nM and 110 pmol/min/mg protein, respectively. All of the steroids showed clear-cut competitive inhibition (Fig. 5). The apparent K_i values were obtained by Dixon plots. The bridged steroids 4, 5, 11 and 17 were weak to poor competitive inhibitors to aromatase in human placental microsomes in which the 6 β ,19-epoxy steroid 4 was the

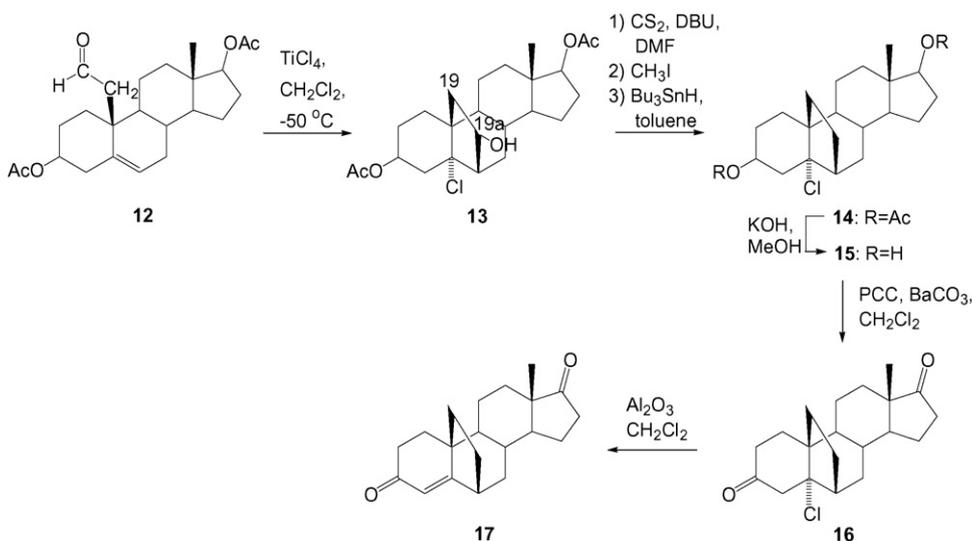


Fig. 4. Synthesis of 6,19-methanoandrostenedione (17).

Table 1
In vitro aromatase inhibition by 6 β ,19-bridged ADs (**4**, **5**, **11** and **17**).

Compound	IC ₅₀ (μ M ^a)	Apparent K _i (μ M ^b)
6,19-epoxy, 4	44 \pm 2.0	2.2 \pm 0.17
6,19-cyclo, 5	>100	18 \pm 1.0
6,19-epithio, 11	>100	18 \pm 1.4
6,19-methano, 17	>100	16 \pm 1.0

^a 300 nM of [1β -³H]AD 20 μ g protein from human placental microsomes and 20-min incubation time were used.

^b All of the inhibitors examined showed a competitive type inhibition on the basis of the Lineweaver–Burk plot and apparent inhibition constant (K_i) was obtained by Dixon plot. Twenty micrograms protein from the placental microsomes and 5-min incubation time were used.

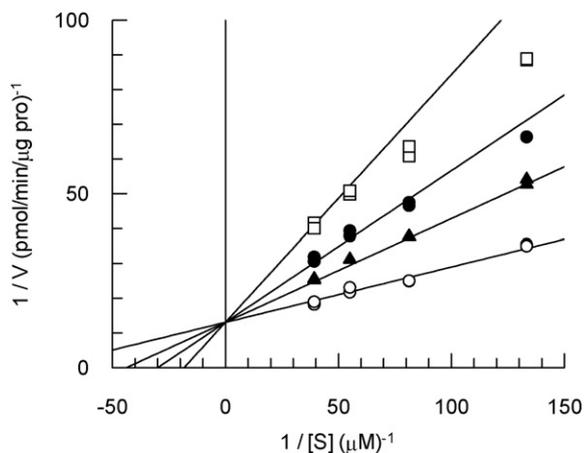


Fig. 5. Lineweaver–Burk plots of aromatase inhibition by 6 β ,19-oxidoandrostenedione (**4**). Concentrations of the inhibitor: control (0 μ M) (○); 1.92 μ M (▲); 3.83 μ M (●); 7.66 μ M (□).

most potent one among the steroids examined (K_i value: 2.2 μ M of compound **4** and 16–18 μ M of compound **5**, **11**, and **17**, respectively).

The affinity of the polar steroid, 19-hydroxyandrostenedione (**6**), to aromatase is known to be about one-third of androstenedione [27] and that of the other polar steroid, 6 β -hydroxyandrostenedione, is relatively high to be 130 nM of the K_i value [20,21]. The previous reports have shown that the affinity of 19-methyl-substituted androstenedione for aromatase is comparable to that of androstenedione [22] and the 6 β -methyl compound [16] has very high affinity for the active site (K_i: 11 nM). It has shown that 19-thio- and 19-methylthio-androstenediones act as very strong inhibitors of aromatase [23,24]. However, it was demonstrated that the affinities of the 6 β ,19-bridged compounds **4**, **5**, **11** and **17** were extremely low, compared to the above reported ones, and the K_i values were more than 2.2 μ M. In the bridged compounds, the 19-substituents cannot rotate freely around the C₁₀–C₁₉ bond. This may cause the low affinities. On the other hand, in the 4-en-3-one steroids, the 6 β ,19-bridged structure bends the steroid skeleton at the A/B ring junction stabilizing the quasi-cis conformation with an inverted ring A 1 β -half chair [28]. The inverted ring A conformation caused by the 6 β ,19-bridged moiety may also prevent the access of the active site of aromatase through the steric reason.

4. Conclusion

6 β ,19-Bridged androstenediones (**4**, **5**, **11**, and **17**) were synthesized and tested the inhibitory activities of the placental aromatase. The synthesis of the compounds **11** and **17** were achieved starting from the 19-hydroxy steroid **6** and the 19-formyl steroid **12**, respec-

tively. The aromatase inhibitory activities of the 6 β ,19-bridged compounds were disappointingly low, however, their new biological properties such as androgenic/antiandrogenic activities are of interest and studies of them will be carried in our laboratory.

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References

- [1] Thompson Jr EA, Siiteri PK. The involvement of human placental microsomal cytochrome P-450 in aromatization. *J Biol Chem* 1974;249:5373–8.
- [2] Kellis Jr J, Vickery LE. Purification and characterization of human placental aromatase cytochrome P-450. *J Biol Chem* 1987;262:4413–20.
- [3] Yoshida N, Osawa Y. Purification of human placental aromatase cytochrome P-450 with monoclonal antibody and its characterization. *Biochemistry* 1991;30:3003–10.
- [4] Meyer AS. Conversion of 19-hydroxy-4-androstene-3,17-dione to estrone by endocrine tissue. *Biochim Biophys Acta* 1995;17:441–2.
- [5] Arigoni D, Battaglia R, Akhtar M, Smith T. Stereospecificity of oxidation at C-19 in oestrogen biosynthesis. *J Chem Soc Chem Commun* 1975:185–7.
- [6] Caspi E, Arunachalam Y, Nelson PA. Biosynthesis of estrogens: aromatization of (19R)-, (19S)-, and (19RS)-[19-³H,²H,¹H]-3 β -hydroxy-androst-5-en-17-one by human placental aromatase. *J Am Chem Soc* 1986;108:1847–52.
- [7] Akhtar M, Calder MR, Corina DL, Wright JN. Mechanistic studies on C-19 demethylation in oestrogen biosynthesis. *Biochem J* 1982;201:569–80.
- [8] Akhtar M, Corina D, Pratt J, Smith T. Studies on the removal of C-19 in oestrogen biosynthesis using ¹⁸O₂. *J Chem Soc Chem Commun* 1976:854–6.
- [9] Bednarski PJ, Nelson SD. Dissociation of 19-hydroxy-, 19-oxo-, and aromatizing activities in human placental microsomes through the use of suicide substrate to aromatase. *J Steroid Biochem* 1989;32:309–16.
- [10] Townsley JD, Brodie HJ. Studies on the mechanism of estrogen biosynthesis. III. The stereochemistry of aromatization of C₁₉ and C₁₈ steroids. *Biochemistry* 1968;7:33–40.
- [11] Cole PA, Robinson CH. Conversion of 19-oxo-[2 β -²H]-androgens into oestrogens by human placental aromatase. *Biochem J* 1990;268:553–61.
- [12] Brodie AMH. Aromatase inhibitors in the treatment of breast cancer. *J Steroid Biochem Mol Biol* 1994;49:281–7.
- [13] Harper-Wynne C, Dowsett M. Recent advances in the clinical application of aromatase inhibitors. *J Steroid Biochem Mol Biol* 2001;76:179–86.
- [14] Howell A, Howell SJ, Clarke R, Anderson E. Where do selective estrogen receptor modulators (SERMs) and aromatase inhibitors (AIs) now fit into breast cancer treatment algorithms? *J Steroid Biochem Mol Biol* 2001;79:227–37.
- [15] Lombardi P. Exemestane, a new steroidal aromatase inhibitor of clinical relevance. *Biochim Biophys Acta* 2002;1587:326–37.
- [16] Numazawa M, Oshibe M. 6-Alkyl- and 6-aryl-androst-4-ene-3,17-diones as aromatase inhibitors. Synthesis and structure-activity relationships. *J Med Chem* 1994;37:1312–9.
- [17] Numazawa M, Oshibe M. Further studies on 6-alkyl-androst-4-ene-3,17-diones as aromatase inhibitors: elongation of the 6-alkyl chain. *Steroids* 1995;60:506–11.
- [18] Numazawa M, Oshibe M, Yamaguchi S, Tachibana M. Time-dependent inactivation of aromatase by 6-alkyl-androsta-1,4-diene-3,17-diones. Effects of length and configuration of the 6-alkyl group. *J Med Chem* 1996;39:1033–8.
- [19] Numazawa M, Oshibe M, Yamaguchi S. 6-Alkyl-androsta-4,6-diene-3,17-diones and their 1,4,6-triene analogs as aromatase inhibitors. *Steroids* 1997;62:595–602.
- [20] Tan L, Hrycaj EG, Matsumoto K. Synthesis and properties of the epimeric 6-hydroperoxyandrostenediones, new substrates/inhibitors of human placental aromatase. *J Steroid Biochem* 1983;19:1329–38.
- [21] Numazawa M, Shelangouski M, Nagaoka M. Probing the binding pocket of the active site of aromatase with 6-ether or 6-ester substituted androst-4-ene-3,17-diones and their diene and triene analogs. *Steroids* 2000;65:871–82.
- [22] Marcotte PA, Robinson CH. Synthesis and evaluation of 10 β -substituted 4-estrene-3,17-diones as inhibitors of human placental microsomal aromatase. *Steroids* 1982;39:325–44.
- [23] Wright NJ, Calder MR, Akhtar M. Steroidal C19 sulphur and nitrogen derivatives designed as aromatase inhibitors. *J Chem Soc Chem Commun* 1985:1733.
- [24] Bednarski JJ, Porubek DJ, Nelson SD. Thiol-containing androgens as suicide substrates of aromatase. *J Med Chem* 1985;28:775–9.
- [25] Banting L, Nicholls PJ, Shaw MA, Smith HJ. Recent developments in aromatase inhibition as a potential treatment for oestrogen-dependent breast cancer. *Prog Med Chem* 1989;26:253–98.
- [26] Johnston JO. Aromatase inhibitors. *Crit Rev Biochem Mol Biol* 1998;33:375–405.
- [27] Covey DF. Aromatase inhibitors: specific inhibitors of oestrogen biosynthesis. In: Berg D, Plempel M, editors. *Steroid biosynthesis inhibitors*. Chichester (England): Ellis Horwood Ltd.; 1988. p. 534–71.

- [28] Vicent GP, Monteserin MC, Veleiro AS, Burton G, Lantos CP, Galigniana MD. 21-Hydroxy-6,19-oxidoprogesterone: a novel synthetic steroid with specific antiglucocorticoid properties in the rat. *Mol Pharmacol* 1997;52:749–53.
- [29] Veleiro AS, Rosenstein R, Grilli ML, Jaliffa C, Speroni F, Burton G. Synthesis and GABA_A receptor activity of a 6,19-oxido analogue of pregnanolone. *Bioorg Med Chem Lett* 2003;13:343–6.
- [30] Veleiro AS, Pecci A, Monteserin MC, Baggio R, Garland MT, Lantos CP, et al. 6,19-Sulfur-bridged progesterone analogues with antiimmunosuppressive activity. *J Med Chem* 2005;48:5675–83.
- [31] Joselevich M, Ghini AA, Burton G. 6,19-Carbon-bridged steroids. Synthesis of 6,19-methanoprogesterone. *Org Biomol Chem* 2003;1:939–43.
- [32] Chenna PH, Veleiro AS, Sonogo JM, Ceballos NR, Garland MT, Baggio RF, et al. Synthesis of 6,19-cyclopregnanes. Constrained analogues of steroid hormones. *Org Biomol Chem* 2007;5:2453–7.
- [33] Kalvoda J, Heusler K, Ueberwasser H, Anner G, Wettstein A. Steroids. CXCVIII. 19-Norsteroids. 4. Reductive ether cleavage of 5 α -halo-6 β ,19-epoxysteroids. *Helv Chim Acta* 1963;46:1361–9.
- [34] Bonet JJ, Wehrli H, Schaffner K. Zu Solvolysversuchen mit Δ^4 - und Δ^5 -ungesättigten 19-mesyloxy-steroiden. *Helv Chim Acta* 1962;155:2615–8.
- [35] Barton DHR, McCombie SW. A new method for the deoxygenation of secondary alcohols. *J Chem Soc, Perkin Trans I* 1975:1574–85.
- [36] Ryan KJ. Biological aromatization of steroids. *J Biol Chem* 1959;234:268–72.
- [37] Siiteri PK, Thompson Jr EA. Human placental aromatase. *J Steroid Biochem* 1975;6:317–22.
- [38] Grafit version 5.0.3. Surrey, UK: Erithacus Software Limited; 2001.
- [39] Andersen NH, Hadley SW, Kelly JD, Bacon ER. Intramolecular olefinic aldehyde Prins reactions for the construction of five-membered rings. *J Org Chem* 1985;50:4144–51.
- [40] Barton DHR, Jang DO, Jaszberenyi JC. The invention of radical reactions. Part XXXI. Diphenylsilane: a reagent for deoxygenation of alcohols via their thiocarbonyl derivatives, deamination via isonitriles, and dehalogenation of bromo and iodo compounds by radical chain chemistry. *Tetrahedron* 1993;49:7193–214.