



Note

Palladium-catalyzed hydroamination of C-(tetra-O-acetyl-β-D-galactopyranosyl)allene

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ARTICLE INFO

Article history:

Received 15 June 2010

Received in revised form 4 November 2010

Accepted 8 November 2010

Available online 9 December 2010

Keywords:

Hydroamination

Amine

Palladium

Allene

Galactopyranosyl

ABSTRACT

Both palladium(0) and palladium(II) methods for catalyzed hydroamination of C-(tetra-O-acetyl-β-D-galactopyranosyl)allene with a variety of aromatic amines have been successfully developed.

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The amine functionality is one of the most ubiquitous in organic chemistry. Its importance is exemplified by the diversity of molecules that contain this functional group, which includes natural products and pharmacological agents.¹ For this reason C–N bond formation is of great interest, which is apparent from the number of methodologies that have been developed for this purpose.² Hydroamination, the formal addition of an N–H bond across a C=C unsaturated bond, is a highly desirable and an atom-economical process for the preparation of amine derivatives.^{2d,3} Allenes constitute an important class of organic compounds with unusual chemical properties due to the cumulated double bond.⁴ The catalytic hydroamination of allenes should be a straightforward method for the synthesis of allylamines. The intramolecular hydroamination of amino allenes is catalyzed by organolanthanide⁵ Group 4 early transition-metal complexes,⁶ and late transition-metal (Pd, Ag, Au) complexes.^{2b,7} Only a small number of catalytic intermolecular hydroamination reactions of allenes have been reported.⁸ We report herein on the palladium-catalyzed hydroamination of a C-(tetra-O-acetyl-β-D-galactopyranosyl)allene (**1**) with amines in the presence of a carboxylic acid. Initially the reaction of C-(tetra-O-acetyl-β-D-galactopyranosyl)allene (**1**) with aniline (**2**) using Yamamoto's conditions (5 mol % Pd₂(dba)₃·CHCl₃, 10 mol % Ph₃P, 20 mol % CH₃COOH in THF at 60 °C for 24 h) was studied.^{8f} Quite unexpectedly, a mixture of diallylated acetate **3** and diallylated amine **4** was obtained in 35% and 10% yields, respectively, along with trace amounts of **5** (Scheme 1). When the same reaction was carried out in the presence of trifluoroacetic

acid (TFA), the mixture of desired allylic amine **5** (29%) and a minor amount (18%) of dienic amine was obtained (entry 1 in Table 1).

The reaction conditions were adjusted to find a suitable catalyst–ligand–solvent–carboxylic acid system for the desired transformation. The results are summarized in Table 1. These results suggested that both Pd(0) and Pd(II) catalysts and carboxylic acid play a dramatic part on the yields of the desired allylic amine **5**. In addition, a profound solvent effect on the reaction was observed. THF was found to be the best solvent (Table 1, entry 18).

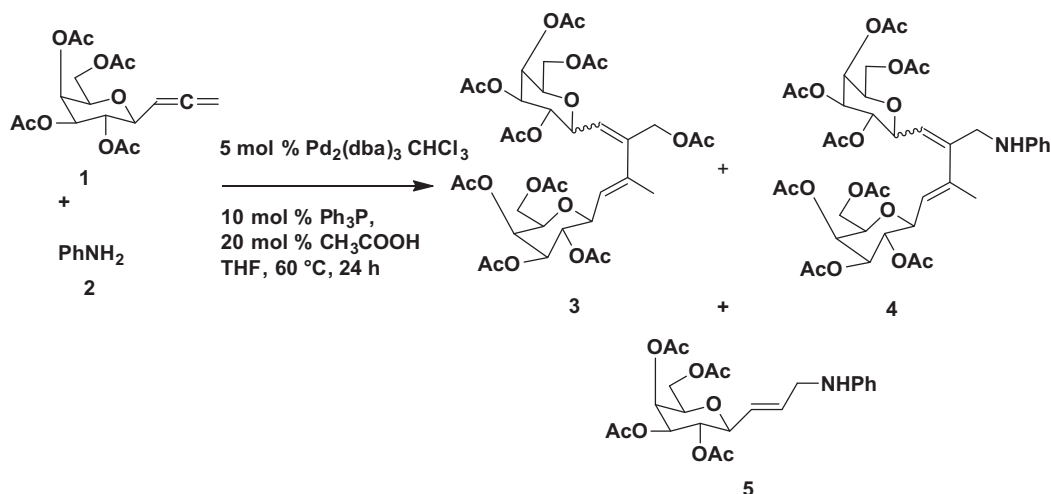
Under the optimized reaction conditions [Pd(OAc)₂ was used as a catalyst, TFA as the carboxylic acid and THF as the solvent], a series of amines were tested in the reaction, and the results are summarized in Table 2.

A possible reaction pathway for Pd(0)-catalyzed hydroamination is showed in Scheme 2. The two principal mechanistic pathways to be considered for hydroamination involve either activation of the amine (path a) or palladacycle formation (path b). The oxidative addition of carboxylic acid to Pd(0) produced hydridopalladium(II) intermediate species (**I**), which on reaction with amine would give intermediate (**III**) and carboxylic acid. Species **III** would form the π-allylpalladium intermediate (**VIII**) with C-(tetra-O-acetyl-β-D-galactopyranosyl)allene (**1**) which, after reductive elimination, would give the hydroamination product (**IX**). The formation of diallylated acetate (**VII**) and diallylated amine (**XII**) could arise by a combination of either path a or path b, intermediate (**V**, **X**) (Scheme 2, path a) and intermediate (**II**) (Scheme 2, path b).

On the other hand, hydroamination of C-(tetra-O-acetyl-β-D-galactopyranosyl)allene (**1**) gave a yield of 42% allylic amine **5** using Pd(OAc)₂ and TFA in THF (Table 1, entry 18). The two possible mechanistic pathways to be considered for hydroamination

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Scheme 1. Palladium-catalyzed hydroamination of C-(tetra-O-acetyl-β-D-galactopyranosyl)allene.

Table 1

Effect of reaction parameters on hydroamination of C-(tetra-O-acetyl-β-D-galactopyranosyl)allene (**1**)^a with aniline^b

Entry	Palladium salt and ligand (mol %)	Carboxylic acid and additive (mol %)	Solvent	Temperature (°C)	% Yield ^c	
					5	4
1	Pd ₂ (dba) ₃ ·CHCl ₃ (5), Ph ₃ P (10)	TFA (20)	THF	60	29	18
2	Pd ₂ (dba) ₃ ·CHCl ₃ (5), Ph ₃ P (10)	TFA (20)	THF	rt	10	46
3	Pd ₂ (dba) ₃ (5), Ph ₃ P (10)	TFA (20)	THF	60	15	42
4	Pd ₂ (dba) ₃ (5), Ph ₃ P (10)	TFA (20)	THF	rt	13	42
5	Pd ₂ (dba) ₃ (5), TFP (10)	TFA (20)	THF	rt	6	47
6	Pd ₂ (dba) ₃ (5), Ph ₃ P (10)	Formic acid (20)	THF	rt	ND	14
7	Pd ₂ (dba) ₃ (5), Ph ₃ P (10)	Benzoic acid (20)	THF	rt	ND	17
8	Pd(OAc) ₂ (5), Ph ₃ P (20)	TFA (20)	THF	rt	6	48
9	Pd ₂ (dba) ₃ (5)	TFA (20)	THF	rt	21	12
10	Pd ₂ (dba) ₃ (5)	TFA (20)	THF	60	23	12
11	Pd ₂ (dba) ₃ (10)	TFA (20)	THF	60	14	26
12	Pd ₂ (dba) ₃ (5)	TFA (50)	THF	rt	25	13
13	Pd ₂ (dba) ₃ (5)	TFA (100)	THF	rt	22	10
14	Pd ₂ (dba) ₃ (5)	TFA (10)	THF	rt	13	7
15	Pd ₂ (dba) ₃ (5)	TFA (5)	THF	rt	15	13
16	Pd ₂ (dba) ₃ (5)	TFA (5)	DMF	rt	ND	17
17	Pd ₂ (dba) ₃ (5)	TFA (5)	CH ₃ CN	rt	8	14
18	Pd(OAc) ₂ (5)	TFA (20)	THF	rt	42	12
19	Pd(OAc) ₂ (5)	TFA (20)	THF	60	32	9
20	Pd(OAc) ₂ (5)	Acetic acid (20)	THF	rt	25	9
21	Pd(OAc) ₂ (5)	—	THF	rt	17	3
22	Pd(OAc) ₂ (5)	—	THF	60	12	5
23	Pd(OAc) ₂ (10)	TFA (20)	THF	rt	21	13
24	Pd(OAc) ₂ (5)	TFA (20), K ₂ S ₂ O ₈ (250)	THF	rt	25	19
25	Pd(OAc) ₂ (5)	TFA (20), K ₂ S ₂ O ₈ (100)	THF	rt	17	12
26	Pd(OAc) ₂ (5)	TFA (20), LiCl (200)	THF	rt	31	ND
27	Pd(OAc) ₂ (5)	LiCl (200)	THF	rt	32	ND
28	Pd(OAc) ₂ (5)	TFA (50)	THF	rt	31	16
29	Pd(OAc) ₂ (5)	TFA (20)	DCM	rt	30	14
30	Pd(OAc) ₂ (5)	TFA (20)	CH ₃ CN	rt	33	14
31	Pd(OAc) ₂ (5)	TFA (20)	Toluene	rt	24	15
32	Pd(OAc) ₂ (5)	TFA (20)	Dioxane	rt	14	9

ND = Not detected.

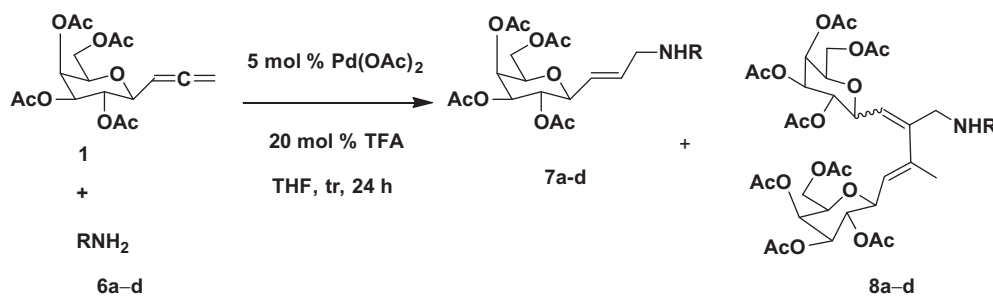
^a C-(Tetra-O-acetyl-β-D-galactopyranosyl)allene (**1**) was prepared according to the literature method.⁹

^b All reactions were carried out with conditions: C-(tetra-O-acetyl-β-D-galactopyranosyl)allene (**1**) (0.27 mmol) and aniline (0.81 mmol) in 0.5 mL of solvent for 24 h.

^c Isolated yields.

involve either activation of the amine or activation of the allene. In the allene activation (Scheme 3, path c) the CH₂=C double bond of allene is activated by coordination to the palladium, and the C–N bond is formed by nucleophilic attack of amine on the coordinative allene. To liberate the product, the palladium–carbon bond in the resulting ammonioallyl complex has to be cleaved. This can be

brought about either by direct protonolysis or by protonation at the palladium with subsequent C–H reductive elimination. Amine activation (Scheme 3, path d) proceeds via oxidative addition of the amine N–H bond to the coordinatively unsaturated palladium center, forming the amido hydrido complex, followed by allene coordination, insertion of the allene into the palladium–nitrogen

Table 2Hydroamination of C-(tetra-O-acetyl-β-D-galactopyranosyl)allene (**1**) with aromatic amines^a

Entry	Amine	Product ^b (%)	
1	 6a	7a (16)	8a (ND)
2	 6b	7b (22)	8b (ND)
3	 6c	7c (25)	8c (3)
4	 6d	7d (17)	8d (6)

ND = Not detected.

^a Reaction conditions: C-(tetra-O-acetyl-β-D-galactopyranosyl)allene (**1**) (0.27 mmol); amine (0.81 mmol); Pd(OAc)₂ (5 mol %); TFA (20 mol %); THF (0.5 mL); room temperature for 24 h.^b Isolated yields.

bond, and finally C–H reductive elimination, liberating the product and closing the catalytic cycle.

In conclusion, we have successfully developed both Pd(0)- and Pd(II)-catalyzed methods for hydroamination of C-(tetra-O-acetyl-β-D-galactopyranosyl)allene (**1**) with a variety of aromatic amines.

1. Experimental

1.1. General methods

All reagents were commercially available and used without purification. Flash column chromatography was performed using Silica Gel 60 (230–400 mesh). Accurate molecular masses were recorded on a microTOF machine. Infrared spectra were recorded using a Perkin–Elmer FTIR spectrometer. The IR samples were prepared as thin films by evaporation of a solution of the compound in CH₂Cl₂ onto a germanium plate. Nuclear magnetic resonance spectra were determined at 400 MHz (¹H) and at 100 MHz (¹³C) on Bruker spectrometers. Chemical shift values are quoted in parts per million (ppm) downfield from TMS, and coupling constants are in Hertz. Chemical shift multiplicities are reported as s = singlet,

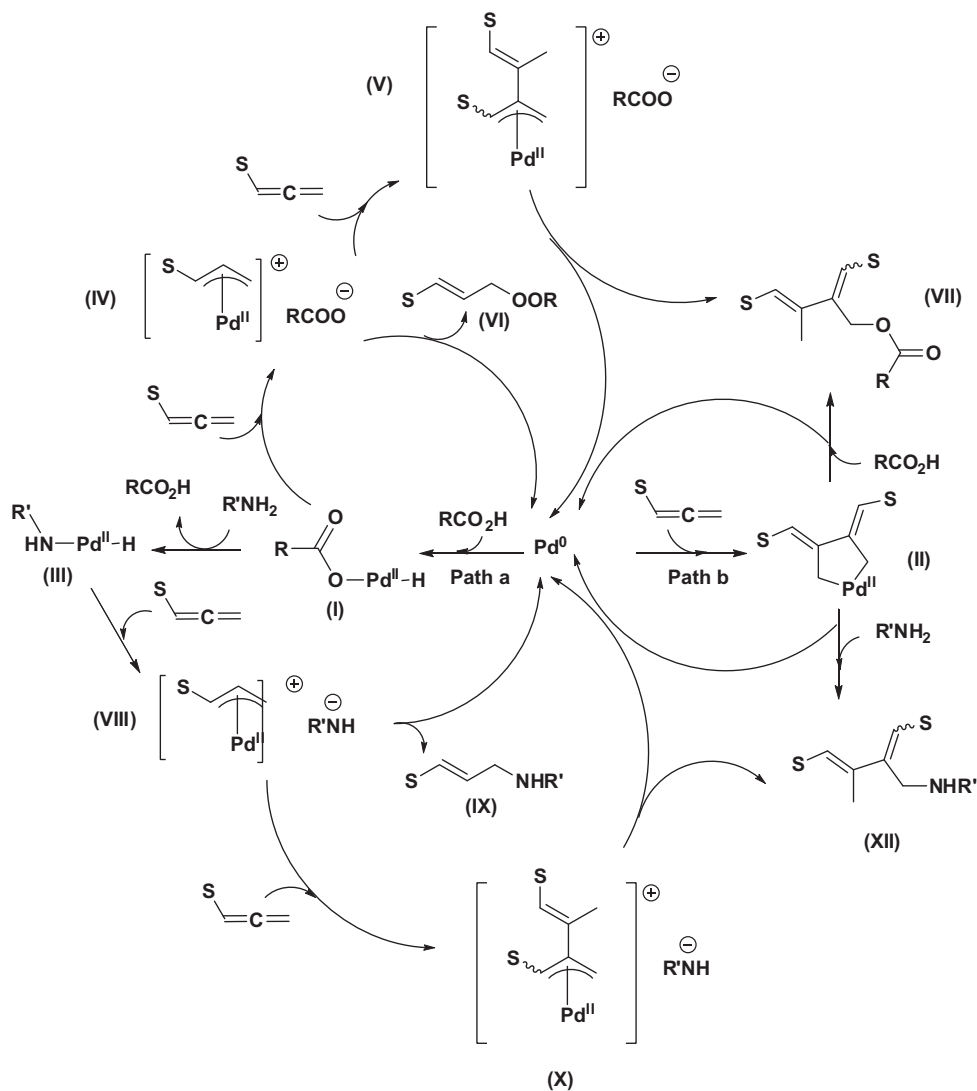
d = doublet, dd = doublet of doublets, t = triplet, m = multiplet, br s = broad singlet, app. t = apparent triplet. C-(Tetra-O-acetyl-β-D-galactopyranosyl)allene [(1S)-2,3,4,6-(tetra-O-acetyl-1,5-anhydro-1-C-propadienyl-D-galactitol, **1**] was prepared according to the literature method.⁹

1.2. Typical general procedure

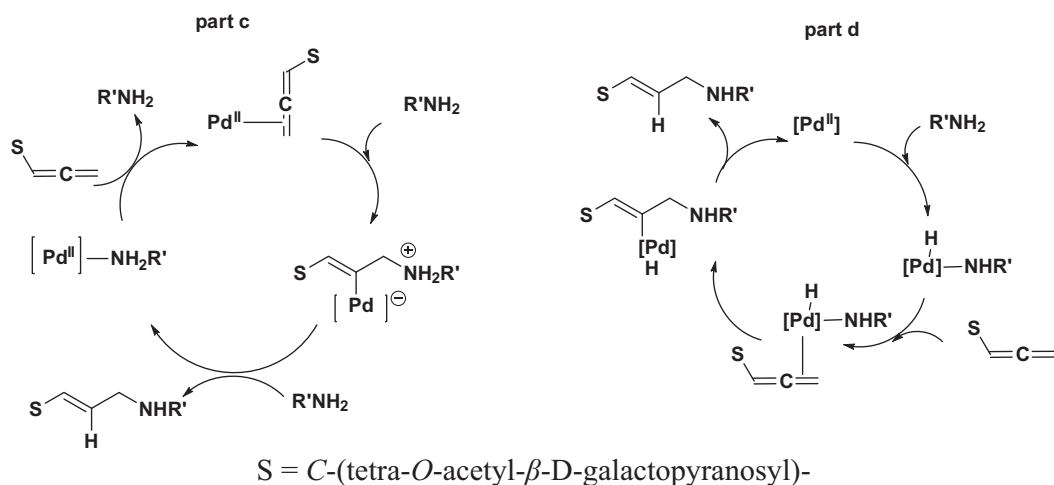
All reactions were carried out in an atmosphere of air. A reaction tube was charged with the C-(tetra-O-acetyl-β-D-galactopyranosyl)allene (**1**) (0.5 mmol), amines (1.5 mmol), Pd salts (catalyst, 5–10 mol %), Ph₃P (ligand, 5 mol %), carboxylic acid (10–100 mol %), LiCl or LiBr (additive, 5 mol %), and solvent (1 mL/mmol). The mixture was either stirred at room temperature or heated at 60 °C for 24 h, then the solvent was evaporated under reduced pressure, and the residue was purified by flash column chromatography on silica gel to give the product.

1.2.1. Data for **3**

IR (film) cm^{−1}: 3477, 2964, 1747, 1436, 1372, 1229, 1119, 1049; ¹H NMR (CDCl₃): δ 5.92 (1H, d, *J* 6.4), 5.80 (1H, d, *J* 6.0), 5.41–5.45 (2H, m), 5.31–5.36 (2H, m), 5.24 (1H, t, *J* 2.4), 5.21 (1H, t, *J* 3.6), 5.15



Scheme 2. Proposed mechanistic pathways for palladium(0)-catalyzed hydroamination.



Scheme 3. Proposed mechanistic pathways for palladium(II)-catalyzed hydroamination.

(1H, t, *J* 6.4), 5.03–5.06 (2H, m), 4.82 (1H, d, *J* 13.2), 4.02–4.20 (6H, m), 1.90–2.20 (30H, 10 × *s*); ¹³C NMR (CDCl₃): δ 170.42, 170.32, 170.29, 170.15, 170.11, 170.01, 169.96, 169.82, 169.57, 145.01, 143.28, 123.86, 120.77, 69.62, 68.88, 68.76, 68.40, 68.24, 68.20, 67.98, 67.85, 67.82, 67.44, 61.67, 61.32, 59.27, 20.70, 20.66, 20.64, 20.60, 15.61; HRMS: calcd for C₃₆H₄₈O₂₀Na (M+Na): *m/z* 823.2637; found: *m/z* 823.2566.

1.2.2. Data for 4

IR (film) cm⁻¹: 3440, 2361, 1747, 1637, 1372, 1226, 1023; ¹H NMR (CDCl₃): δ 7.16 (2H, app. t, *J* 8 and 7.6), 6.68–6.72 (1H, m), 6.58 (2H, d, *J* 8.4), 5.83 (1H, d, *J* 6.8), 5.75 (1H, d, *J* 6.4), 5.41–5.45 (2H, m), 5.28–5.34 (2H, m), 5.21–5.26 (2H, m), 5.10–5.14 (3H, m), 5.00 (1H, app. t, *J* 6.4 and 5.6), 4.00–4.20 (6H, m), 1.80–2.10 (27H, 9 × *s*); ¹³C NMR (CDCl₃): δ 170.54, 170.41, 170.13, 170.09, 169.98, 169.90, 169.83, 169.81, 147.84, 144.58, 129.34, 129.24, 121.82, 120.62, 117.82, 112.75, 69.48, 69.06, 68.77, 68.36, 68.26, 68.10, 67.99, 67.91, 67.52, 61.82, 61.51, 42.14, 20.78, 20.75, 20.73, 20.67, 20.65, 15.87; HRMS: calcd for C₄₀H₅₁NO₁₈Na (M+Na): *m/z* 856.3004; found: *m/z* 856.3036.

1.2.3. Data for 5

IR (film) cm⁻¹: 3418, 2361, 1746, 1633, 1604, 1507, 1371, 1226, 1051; ¹H NMR (CDCl₃): δ 7.17 (2H, app. t *J* 8.4 and 7.2), 6.71 (1H, app. t *J* 8.4 and 7.2), 6.60 (2H, d, *J* 7.6), 5.98 (1H, dt, *J* 15.6 and 5.2), 5.82 (1H, q, *J* 15.6 and 5.2), 5.37 (1H, d, *J* 3.2), 5.28 (1H, dd, *J* 10.4 and 5.6), 5.10 (1H, dd, *J* 10.4 and 3.2), 4.80 (1H, t, *J* 3.2), 4.00–4.14 (3H, m), 3.84 (2H, d, *J* 5.2) 1.90–2.10 (12H, 4 × *s*); ¹³C NMR (CDCl₃): δ 170.52, 170.19, 170.06, 169.87, 147.64, 134.47, 129.27, 123.49, 117.87, 113.14, 72.53, 68.31, 68.29, 68.01, 61.83, 45.62, 20.73, 20.69, 20.66; HRMS: calcd for C₂₃H₃₀NO₉ (M+H): *m/z* 464.1921; found: *m/z* 464.1968.

1.2.4. Data for 7a

IR (film) cm⁻¹: 3442, 2362, 1747, 1638, 1515, 1372, 1231, 1023; ¹H NMR (CDCl₃): δ 6.79 (2H, d, *J* 8.8), 6.60 (2H, d, *J* 8.8), 5.97 (1H, dt, *J* 15.6 and 5.2), 5.85 (1H, q, *J* 15.6 and 5.2), 5.42 (1H, d, *J* 3.2), 5.29 (1H, dd, *J* 10.4 and 5.6), 5.12 (1H, dd, *J* 10.4 and 3.2), 4.79 (1H, t, *J* 3.2), 4.00–4.10 (3H, m), 3.80 (2H, d, *J* 5.2), 1.90–2.10 (15H, 5 × *s*); ¹³C NMR (CDCl₃): δ 170.53, 170.23, 170.13, 170.08, 152.44, 141.82, 134.79, 123.41, 114.92, 114.57, 72.50, 68.40, 68.33, 68.04, 68.01, 61.87, 45.94, 20.72, 20.70, 20.66; HRMS: calcd for C₂₄H₃₁NO₁₀Na (M+Na): *m/z* 516.1846; found: *m/z* 516.2006.

1.2.5. Data for 7b

IR (film) cm⁻¹: 3439, 2362, 1747, 1636, 1604, 1474, 1372, 1312, 1228, 1113, 1048; ¹H NMR (CDCl₃): δ 8.09 (2H, d, *J* 8.8), 6.58 (2H, d, *J* 9.2), 5.96 (1H, dt, *J* 15.6 and 5.2), 5.88 (1H, q, *J* 15.6 and 5.2), 5.40 (1H, d, *J* 3.2), 5.30 (1H, dd, *J* 10.4 and 5.6), 5.10 (1H, dd, *J* 10.4 and 3.2), 4.82 (1H, t, *J* 3.2), 4.01–4.18 (3H, m), 3.95 (2H, d, *J* 5.2), 1.93–2.14 (12H, 4 × *s*); ¹³C NMR (CDCl₃): δ 170.64, 170.17, 170.08, 169.77, 152.98, 138.32, 132.15, 126.36, 124.85, 111.39, 72.24, 68.45, 68.23, 67.95, 67.82, 61.73, 44.89, 20.73, 20.69, 20.64; HRMS: calcd for C₂₃H₂₈N₂O₁₁Na (M+Na): *m/z* 531.1591; found: *m/z* 531.1670.

1.2.6. Data for 7c

IR (film) cm⁻¹: 3398, 2926, 1748, 1622, 1532, 1351, 1228, 1050, 738; ¹H NMR (CDCl₃): δ 7.55 (1H, d, *J* 7.6), 7.40 (1H, s), 7.32 (1H, t, *J* 8.4), 6.90 (1H, d, *J* 1.2), 5.99 (1H, dt, *J* 15.6 and 5.2), 5.92 (1H, q, *J* 15.6 and 5.2), 5.39 (1H, d, *J* 3.2), 5.33 (1H, dd, *J* 10.4 and 5.6), 5.11 (1H, dd, *J* 10.4 and 3.2), 4.82 (1H, t, *J* 3.2), 4.02–4.15 (3H, m), 3.93 (2H, d, *J* 5.2), 1.97–2.14 (12H, 4 × *s*); ¹³C NMR (CDCl₃): δ 170.61, 170.18, 170.07, 169.85, 149.44, 148.44, 132.92, 129.81, 124.40, 119.04, 112.38, 106.52, 72.39, 68.35, 68.25, 67.92, 61.73,

45.31, 20.72, 20.69, 20.66; HRMS: calcd for C₂₃H₂₈N₂O₁₁Na (M+Na): *m/z* 531.1591; found: *m/z* 531.1668.

1.2.7. Data for 8c

IR (film) cm⁻¹: 3736, 2362, 1744, 1540, 1370, 1225, 1023; ¹H NMR (CDCl₃): δ 7.53 (1H, d, *J* 6.4), 7.41 (1H, app. t, *J* 2.4 and 2.0), 7.27 (1H, s), 6.89 (1H, d, *J* 2.0), 5.91 (1H, d, *J* 7.2), 5.40–5.46 (3H, m), 5.27–5.31 (2H, m), 5.22–5.26 (2H, m) 5.12–5.18 (4H, m), 4.01–4.15 (6H, m), 2.00–2.13 (27H, 9 × *s*); ¹³C NMR (CDCl₃): δ 170.61, 170.52, 170.44, 170.19, 170.05, 170.01, 169.96, 169.91, 149.44, 148.82, 144.33, 142.93, 129.74, 123.26, 118.69, 117.18, 112.09, 106.16, 70.07, 69.06, 69.02, 68.16, 67.96, 67.73, 67.59, 61.85, 61.63, 42.34, 20.80, 20.75, 20.70, 20.65, 14.12; HRMS: calcd for C₄₀H₅₀N₂O₂₀Na (M+Na): *m/z* 901.2855; found: *m/z* 901.2904.

1.2.8. Data for 7d

IR (film) cm⁻¹: 3440, 2361, 1748, 1584, 1533, 1372, 1228, 1050, 773; ¹H NMR (CDCl₃): δ 7.83–7.86 (1H, m), 7.77–7.82 (1H, m), 7.45–7.53 (2H, m), 7.36 (1H, d, *J* 7.6), 7.30 (1H, d, *J* 6.8), 6.61 (1H, d, *J* 7.2), 6.12 (1H, dt, *J* 15.6 and 5.2), 5.93 (1H, q, *J* 15.6 and 5.2), 5.38 (1H, d, *J* 3.2), 5.33 (1H, dd, *J* 10.4 and 5.6), 5.14 (1H, dd, *J* 10.4 and 3.2), 4.83 (1H, t, *J* 3.2), 4.07–4.15 (3H, m), 4.05 (2H, d, *J* 5.2), 1.90–2.13 (12H, 4 × *s*); ¹³C NMR (CDCl₃): δ 170.58, 170.20, 170.05, 169.88, 142.73, 134.34, 133.99, 128.71, 126.45, 125.79, 124.86, 123.99, 123.53, 119.84, 117.95, 105.02, 72.50, 68.39, 68.33, 68.04, 68.01, 61.87, 45.94, 20.72, 20.70, 20.66; HRMS: calcd for C₂₇H₃₁NO₉Na (M+Na): *m/z* 536.1897; found: *m/z* 536.1963.

1.2.9. Data for 8d

IR (film) cm⁻¹: 3442, 2362, 2066, 1746, 1638, 1372, 1227, 1048; ¹H NMR (CDCl₃): δ 7.83 (1H, d, *J* 8.8), 7.77 (1H, d, *J* 7.6), 7.35–7.46 (2H, m), 7.33 (1H, d, *J* 7.6), 7.24 (1H, d, *J* 8.4), 6.59 (1H, d, *J* 7.6), 5.94 (1H, d, *J* 7.2), 5.40–5.45 (3H, m), 5.31–5.36 (3H, m), 5.22–5.30 (2H, m), 5.16–5.21 (3H, m), 4.02–4.25 (6H, m), 1.90–2.10 (27H, 9 × *s*); ¹³C NMR (CDCl₃): δ 170.64, 170.56, 170.51, 170.47, 170.23, 170.20, 170.12, 170.03, 145.22, 143.30, 134.36, 134.26, 129.29, 128.99, 128.67, 128.48, 126.54, 125.84, 122.82, 120.39, 117.79, 104.13, 69.60, 69.14, 69.07, 68.56, 68.40, 68.28, 67.90, 67.86, 67.70, 61.68, 61.64, 61.14, 43.52, 20.69, 20.67, 20.63, 14.12; HRMS: calcd for C₄₄H₅₃NO₁₈Na (M+Na): *m/z* 906.3160; found: *m/z* 906.3220.

Acknowledgments

This work was financially supported by PERCH-CIC and Mahasarakham University.

References

- (a) Gibson, M. S.; Patai, S. *The Chemistry of the Amino Group*; Interscience: New York, 1968; p 61; (b) Staunton, J.; Wilkinson, B. *Top. Curr. Chem.* **1998**, *195*, 49–92.
- (a) Hultzsich, K. C. *Org. Biomol. Chem.* **2005**, *3*, 1819–1824; (b) Hartwig, J. F. *Pure Appl. Chem.* **2004**, *76*, 507–516; (c) Pohlki, F.; Doye, S. *Chem. Soc. Rev.* **2003**, *32*, 104–114; (d) Müller, T. E.; Beller, M. *Chem. Rev.* **1998**, *98*, 675–704; (e) Beller, M.; Seayad, J.; Tillack, A.; Jiao, H. *Angew. Chem., Int. Ed.* **2004**, *43*, 3368–3398.
- (a) Roesky, P. W.; Müller, T. E. *Angew. Chem., Int. Ed.* **2003**, *42*, 2708–2710; (b) Alonso, F.; Beletskaya, I. P.; Yus, M. *Chem. Rev.* **2004**, *104*, 3079–3159; (c) Beller, M.; Eichberger, M.; Trauthwein, H. *Angew. Chem., Int. Ed.* **1997**, *36*, 2225–2227; (d) Kawatsura, M.; Hartwig, J. F. *J. Am. Chem. Soc.* **2000**, *122*, 9546–9547.
- (a) Brummond, K. M.; DeForrest, J. *Synthesis* **2007**, 795–818; (b) Krause, N.; Hashmi, A. S. K. *Modern Allene Chemistry*; Wiley: Weinheim, 2004.
- (a) Gagné, M. R.; Stern, C. L.; Marks, T. J. *J. Am. Chem. Soc.* **1992**, *114*, 275–294; (b) Ryu, J. S.; Li, G. Y.; Marks, T. J. *J. Am. Chem. Soc.* **2003**, *125*, 12584–12605.
- (a) Bytschkov, I.; Doye, S. *Eur. J. Org. Chem.* **2003**, 935–946; (b) Senn, H. M.; Blöchl, P. E.; Togni, A. *J. Am. Chem. Soc.* **2000**, *122*, 4098–4107; (c) Müller, T. E.; Grosche, M.; Herdtweck, E.; Pleier, E. K.; Walter, E.; Yan, K. Y. *Organometallics* **2000**, *19*, 170–183.
- (a) Shimada, T.; Bajracharya, G. B.; Yamamoto, Y. *Eur. J. Org. Chem.* **2005**, 59–62; (b) Takaya, J.; Hartwig, J. F. *J. Am. Chem. Soc.* **2005**, *127*, 5756–5757; (c) Li, K.; Phua, P. H.; Hii, K. K. *Tetrahedron* **2005**, *61*, 6237–6242; (d) Li, X.; Chianese, A. R.;

- Vogel, T.; Crabtree, R. H. *Org. Lett.* **2005**, 7, 5437–5440; (e) Meguro, M.; Yamamoto, Y. *Tetrahedron Lett.* **1998**, 39, 5421–5424; (f) Nedolya, N. A.; Schlyakhtina, N. I.; Zinov'eva, V. P.; Albanov, A. I.; Brandsma, L. *Tetrahedron Lett.* **2002**, 43, 1569–1571; (g) Billet, M.; Schoenfelder, A.; Klotz, P.; Mann, A. *Tetrahedron Lett.* **2002**, 43, 1453–1456; (h) Mizushima, E.; Hayashi, T.; Tanaka, M. *Org. Lett.* **2003**, 5, 3349–3352; (i) Hill, A. W.; Elsegood, M. R. J.; Kimber, M. C. *J. Org. Chem.* **2010**, 75, 5406–5409.
8. (a) Walsh, P. J.; Baranger, A. M.; Bergman, R. G. *J. Am. Chem. Soc.* **1992**, 114, 1708; (b) Johnson, J. S.; Bergman, R. G. *J. Am. Chem. Soc.* **2001**, 123, 2923–2924; (c) Ayinla, R. O.; Schafer, L. L. *Inorg. Chim. Acta* **2006**, 359, 3097–3102; (d) De Renzi, A.; Ganis, P.; Panunzi, A.; Vitagliano, A. *J. Am. Chem. Soc.* **1980**, 105, 1716–1722; (e) Besson, L.; Goré, J.; Cazes, B. *Tetrahedron Lett.* **1995**, 36, 3857–3860; (f) Al-Masum, M.; Meguro, M.; Yamamoto, Y. *Tetrahedron Lett.* **1997**, 38, 6071–6074; (g) Zhang, J.; Yang, C. G.; He, C. *J. Am. Chem. Soc.* **2006**, 128, 1798–1799.
9. (a) Zhu, Y.-H.; Vogel, P. *Synlett* **2001**, 79–81; (b) Kröger, L.; Henkensmeier, D.; Schäfer, A.; Thiem, J. *Bioorg. Med. Chem. Lett.* **2004**, 14, 73–75; (c) Schäfer, A.; Thiem, J. *J. Org. Chem.* **2000**, 65, 24–29.