

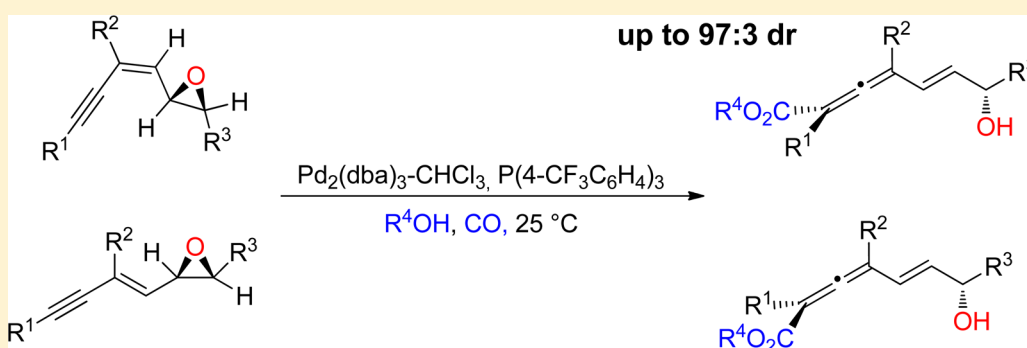
Palladium-Catalyzed Alkoxy carbonylation of Conjugated Enyne Oxiranes: A Diastereoselective Method for the Synthesis of 7-Hydroxy-2,3,5-trienoates

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S Supporting Information



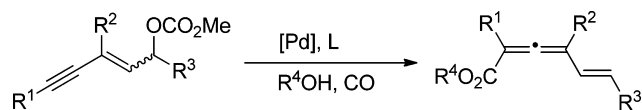
ABSTRACT: Palladium-catalyzed alkoxy carbonylation of 1,5-substituted conjugated enyne oxiranes provides a diastereoselective route to (*E*)-configured 7-hydroxy-2,3,5-trienoates. The reactions proceeded in a highly stereoselective manner, possibly through sequential formation of π -allylpalladium and σ -vinylallenyl palladium complexes. The major diastereomeric form of the product is determined by the configuration of the allenyl moiety of the substrate.

INTRODUCTION

Vinylallenes are, interestingly, reactive compounds toward various cycloaddition and cyclization reactions.¹ They exhibit particularly higher activity and selectivity with Diels–Alder reactions, because their configurational equilibrium is more on the side of *s-cis* conformer—a prerequisite for a [4 + 2] cycloaddition reaction to occur effectively—than for 1,3-dienes.² In spite of their synthetic utility in organic reactions, there are only a few methods that can generate vinylallene structures.^{1j,o,r,3}

Recently, we introduced new methods for the construction of functionalized vinylallenes, which involved palladium- or rhodium-catalyzed reactions of carbonate or acetate derivatives of 2-en-4-yne alcohols.⁴ 2,3,5-Trienoates were a form of the vinylallene structure that was accomplished via palladium-catalyzed alkoxy carbonylation of 2,4-enyne carbonates (Scheme 1).^{4a}

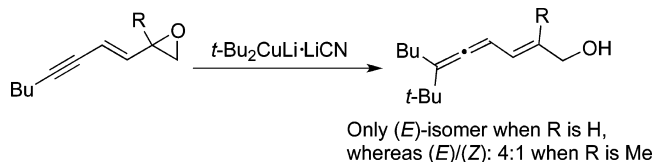
Scheme 1. Previous Study: Alkoxy carbonylation of Enyne Carbonates^{4a,c}



It was later found that in some cases, this reaction proceeds with high center-to-axial chirality transfer.^{4c}

Encouraged by these findings, we have decided to extend the methodology to enyne oxiranes in order to obtain more functionalized vinylallene structures. To the best of our knowledge, the only coupling method that involved the use of enyne oxiranes was their reactions with organocuprates, which led to alkyl-substituted vinylallenes that bear a hydroxyl group in the allylic position (Scheme 2).^{3e} However, this

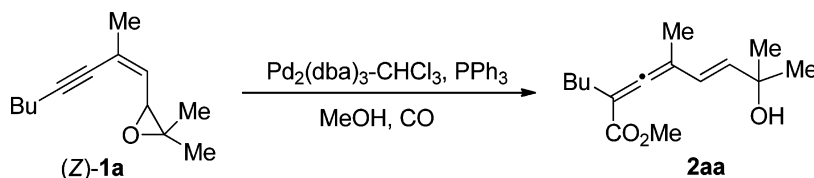
Scheme 2. Previous Study: 1,5-(S_N2'')-Substitution Reactions of Enyne Oxiranes with Organocuprates^{3e}



method provided a very limited scope for both enyne substrates and their reacting partners, and moreover, the diastereoselectivity of this method, in terms of relative configuration of allenyl moiety and allylic carbon, could not be probed.

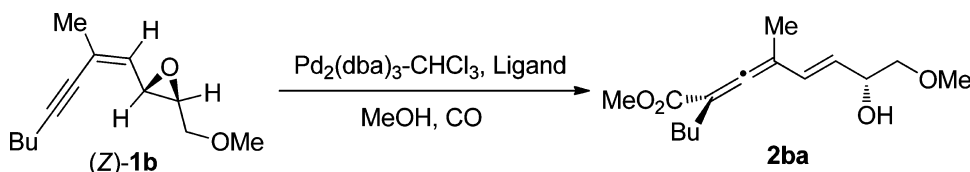
We describe here for the first time that conjugated enynes carrying an oxirane moiety as a leaving group undergo

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Table 1. Methoxycarbonylation of Enyne Oxirane (Z)-1a^a

entry	Pd %	PPh ₃ %	t (h)	yield % ^b
1	0.5	2	1	>99 ^c
2	0.25	1	2	>99
3	0.5	3	3	>99
4	0.25	0.5	5	>99
5	1	0	16	0
6	0	4	16	0

^aReaction conditions: 0.3 mmol of (Z)-1a, 2 mL of MeOH, under CO filled balloon, 25 °C. ^bDetermined by ¹H NMR analysis using benzaldehyde as the internal standard. ^cIsolated yield.

Table 2. Effect of Ligand on Methoxycarbonylation of (Z)-1b^a

entry	ligand	t (h)	yield % ^b	dr ^c
1	PPh ₃	1	96	92:8
2	PPh ₃ ^d	4	89	92:8
3	P(2-MeC ₆ H ₄) ₃	0.5	92	92:8
4	P(2-OMeC ₆ H ₄) ₃	0.5	97	87:13
5	P(4-OMeC ₆ H ₄) ₃	1	95	86:14
6	PPh ₂ Me	2.5	81	80:20
7	Tri(2-furyl)phosphine	1	94	83:17
8	P(4-CF ₃ C ₆ H ₄) ₃	0.5	94 ^e	94:6
9	dppf	1	72	90:10
10	dppe	3	96	86:14
11	dppb	1.5	89	86:14
12	Xantphos	1.5	92	86:14
13	(±)-BINAP	9.5	44 ^f	85:15
14	BIPHEP	10	0	—

^aReaction conditions: 0.1 mmol of (Z)-1b, 2 mL of MeOH, 1 mol % of Pd, P/Pd ratio is 4:1, under CO-filled balloon, 25 °C. dppf: 1,1'-bis(diphenylphosphino)ferrocene; dppe: 1,2-bis(diphenylphosphino)ethane; dppb: 1,4-bis(diphenylphosphino)butane; Xantphos: 4,5-bis(diphenylphosphino)-9,9-dimethylxanthene; BINAP: 2,2'-bis(diphenylphosphino)-1,1'-binaphthalene; BIPHEP: 2,2'-bis(diphenylphosphino)-1,1'-biphenyl. ^bDetermined by ¹H NMR analysis using benzaldehyde as the internal standard. ^cDiastereomeric ratio (dr) was determined by ¹H NMR. ^dPd(PPh₃)₄ was the catalyst of this reaction. ^eIsolated yield. ^fThe conversion of (Z)-1b was 49%.

alkoxycarbonylation in the presence of palladium under fairly mild conditions, forming hydroxy-functionalized 2,3,5-trienoates usually with high diastereoselectivities.

RESULTS AND DISCUSSION

We initiated our study using the enyne oxirane (Z)-1a whose oxirane terminus is dimethyl-substituted. We were pleased to observe that, with just our first attempt, the substrate (Z)-1a was converted to the desired 2,3,5-trienoate product 2aa, which is exclusively in (E)-configuration, and carries a hydroxyl group on the allylic position, almost quantitatively under very mild reaction conditions; the conversion was complete within just 1 h, when the reaction was performed in the presence of a Pd₂dba₃-CHCl₃ (0.5 mol % Pd) and PPh₃ (2 mol %) combination in MeOH, and under balloon pressure of CO at 25 °C (Table 1, entry 1). The catalyst loading could even be

reduced by half (0.25% Pd, 1% PPh₃) without compromising the product yield (entry 2). The optimum PPh₃/Pd ratio appears to be 4:1, because any deviation from this ratio resulted in a decrease in the reaction rate (entries 1–4). Finally, the controlled experiments proved that no conversion for the substrate would be possible in the absence of a phosphine ligand or a palladium source (entries 5 and 6).

Having determined that the enyne oxirane (Z)-1a is a perfectly amenable reagent toward palladium catalysis as a means to synthesize vinylallene structures, we next probed the diastereoselectivity—directly associated with the level of center-to-axial chirality transfer—of the method over the substrate (Z)-1b, using a variety of mono- and bidentate ligands. The corresponding vinylallene product 2ba was in 96% yield with a good diastereomeric ratio (92:8) when PPh₃ was used as the ligand (Table 2, entry 1). The relative configuration of its

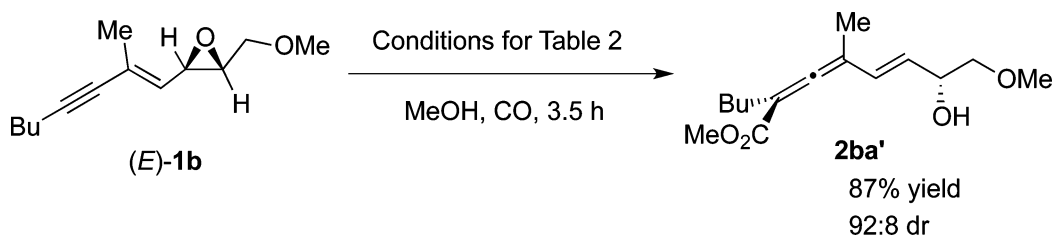
Table 3. Alkoxy carbonylation of Enyne Oxirane (Z)-1a^a

(Z)-1b $\xrightarrow[\text{ROH, CO}]{\text{Pd}_2(\text{dba})_3\text{-CHCl}_3, \text{P(4-CF}_3\text{C}_6\text{H}_4)_3}$ 2b

entry	ROH	product	<i>t</i> (h)	yield %	dr ^b
1	EtOH	2bb	1	84	80:20
2	PrOH	2bc	3	87	88:12
3	BuOH	2bd	4	83	86:14
4	<i>i</i> -PrOH	2be	2	88	86:14
5	<i>t</i> -BuOH	2bf	10	0	—
6	allyl alcohol	2bg	6	25 ^c	93:7
7 ^d			1	91	93:7

^aPerformed under the conditions of Table 2. ^bDetermined by ¹H NMR. ^cThe conversion of (Z)-1b was 75%. ^dReaction was performed with 3 mol % of Pd and 12 mol % of ligand.

Scheme 3. Methoxycarbonylation of (E)-1b



major diastereomer was determined by X-ray crystallography of its phenylcarbamate derivative (see the Supporting Information).⁵

Typically, comparable yields are obtained with a variety of monodentate phosphine ligands tested herein, with varying reaction times required for a full conversion. However, it has been found that the level of the chirality transfer is strongly dependent on the ligand type that is used. The reactions in the presence of a prepared catalyst Pd(PPh₃)₄ or P(2-MeC₆H₄)₃ ligand proceeded with similar selectivities when compared with PPh₃ (entries 2 and 3), whereas electron-rich phosphines and a heteroaryl-substituted phosphine all showed inferior stereoselectivities (entries 4–7). The best diastereoselectivity was obtained with the use of an electron-poor phosphine ligand, P(4-CF₃C₆H₄)₃, which afforded 2ba with 94% yield and 94:6 dr within just 30 min (entry 8). Finally, the use of bidentate ligands brought about dismal selectivities or low activities in some cases (entries 9–14).

Having determined P(4-CF₃C₆H₄)₃ as the optimal ligand, next we explored the suitability of a number of alcohol reagents for their reactions with (Z)-1b. Except those with *t*-BuOH (Table 3, entry 5), the vinylallene products could be obtained at yields within the range of 83–91% with all alcohol reagents tested herein. Surprisingly, however, the reactions performed with alkyl alcohols, including EtOH, PrOH, BuOH, and *i*-PrOH, all proceeded with relatively lower stereoselectivities (entries 1–5). Nevertheless, gratifyingly an additional functionality could be introduced to a 7-hydroxy-2,3,5-trienoate structure with the use of allyl alcohol, which yielded the corresponding product 2bg in 91% yield and with 93:7 dr, albeit with higher Pd loading to complete the reaction (entries 6–7).

It seems that the method is also suitable for enyne oxiranes containing an (*E*)-configured alkenyl moiety; a carbonylation reaction of (E)-1b in MeOH under the standard conditions (conditions denoted in Table 2) required remarkably longer

reaction period for complete conversion and resulted in a diastereomer of 2ba (2ba') with a high yield, but slightly decreased selectivity of 92:8 dr (Scheme 3). This result also proves that the configuration of alkenyl moiety of substrate regulates the stereomeric outcome of the product. Nonetheless, because of some difficulties encountered during the synthesis of enyne oxiranes with (*E*)-alkenyl group, such as partial isomerization of the alkenyl group, further studies were continued using only (*Z*)-configured reagents.

Next we explored the scope of the protocol over a variety of substrates containing (*Z*)-alkenyl and (*E*)-oxirane moieties. Except in one case, alkoxy carbonylation reactions proceeded rather smoothly and provided the expected products at yields within the range of 81–92%.

It was interesting to observe that the form of pendant oxygen functionality within R³ group is an important factor for the stereoselectivity of the process. Substituting the methoxymethyl group in R³ position with a benzyloxymethyl group ((Z)-1c) had no detectable effect on both the reactivity and stereoselectivity (Table 4, entry 1). However, when R³ was a carbinol group ((Z)-1d) or its hydroxyl group was silyl-protected ((Z)-1e), the corresponding methoxycarbonylated products 2da and 2ea were obtained with 90:10 and 89:11 dr, respectively (entries 2 and 3).

The same trend was also obvious for the reactions of (Z)-1f and (Z)-1g, where R³ is dimethylcarbinol and dimethylmethoxy methyl, respectively. Though comparably good yields could be achieved with both reagents, the products 2fa and 2ga were obtained with significantly different level of stereoselectivities, 85:15 and 95:5 dr, respectively (entries 4 and 5).

It seems that for an effective stereoselectivity, the alkenyl carbon that is proximal to the alkynyl moiety is required to be fully substituted, because the carbonylative reaction of the enyne oxirane containing a disubstituted alkenyl moiety ((Z)-1h) with

Table 4. Alkoxyacylation of Enyne Oxiranes^a

Reaction scheme: (Z)-1 $\xrightarrow[\text{R}^4\text{OH, CO}]{\text{Pd}_2(\text{dba})_3\text{-CHCl}_3, \text{P}(4\text{-CF}_3\text{C}_6\text{H}_4)_3}$ 2

entry	product	time (min)	yield ^b (%)	dr ^c	entry	product	time (min)	yield ^b (%)	dr ^c
1		30	90	94:6	11		300	81	88:12
2		100	84	90:10	12		90	92	94:6
3		50	82	89:11	13		90	89	97:3
4		60	90	85:15	14		45	86	96:4
5		45	89	95:5	15		30	87	94:6
6		45	87	85:15	16		45	84	96:4
7		90	82	93:7	17		120	91	95:5
8		70	87	82:18	18		360	88	94:6
9		50	89	94:6	19 ^c		60	88	95:5
10		110	68	95:5	20		45	87	88:12
			14	ND					

^aPerformed under the conditions of Table 2. ^bDetermined by ¹H NMR. ^cReaction was performed with 3 mol % of Pd and 12 mol % of ligand.

MeOH proceeded with an inferior stereoselectivity (entry 6). As a model for substrates with all alkyl substituents, the enyne oxirane (Z)-1i, where all R¹, R², and R³ positions were occupied by butyl groups, also gave rise to the expected product 2ia with

a high selectivity of 93:7 dr (entry 7). However, (Z)-1j possessing a methyl group in the R³ interestingly afforded the lowest dr (82:18) as compared to other enyne oxiranes tested herein (entry 8).⁶ While the method tolerated a larger cyclohexyl

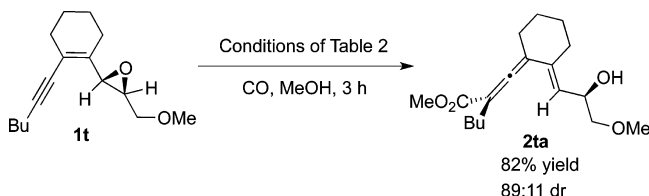
group in the R² (entry 9) under the reaction conditions, the presence of the *tert*-butyl group at this position ((*Z*)-**1l**) brought about the formation of an allylic esterified byproduct (**3la**) along with the desired product **2la** with a high dr (entry 10).⁷

It was also intriguing to find that there appears to be a direct relation between the diastereoselectivity and the size of R¹ (entries 11–13).⁸ For instance, when (*Z*)-**1m**, which contains a terminal alkynyl moiety, was subjected to the methoxycarbonylation, the product **2ma** was obtained with only 88:12 dr. Nevertheless, the substrate (*Z*)-**1o** (R¹ = *t*-Bu) afforded the expected product **2oa** in an excellent level of selectivity (97:3 dr).

The remarkable improvement on the stereoselectivity of the process that was gained with the presence of the *tert*-butyl group on the alkynyl moiety appears enduring; in contrast to the results obtained with (*Z*)-**1b**, when R¹ = *tert*-butyl, the replacement of the methoxy group with benzyloxy- ((*Z*)-**1p**), hydroxyl- ((*Z*)-**1q**), or silyl-protected ((*Z*)-**1r**) groups within R³ only slightly impacted the dr of the corresponding products (entries 14–16). Moreover, the carbonylative reaction of (*Z*)-**1o** with other alcohols also provided the corresponding vinylallenes with high dr values (entries 17–19).

The enyne oxirane (*Z*)-**1s** with a phenyl substituent at alkynyl terminus yielded the product **2sa** with 88:12 dr (entry 20), and the substrate **1t** having an endocyclic double bond also successfully underwent methoxycarbonylation to generate an exocyclic conjugated vinylallene structure **2ta** with a relatively lower but synthetically still useful dr (Scheme 4).

Scheme 4. Methoxycarbonylation of **1t**



The mechanisms illustrated in Scheme 5 are proposed to account for the stereoselective formation of **2** and **2'**.⁹ The reaction cycle should begin with ring-opening by the attack of a palladium complex to both (*Z*)-**1** and (*E*)-**1** in anti-mode,¹⁰ leading to the formation of π -allylpalladium complexes **A** and **A'**, respectively.^{10d,11} Given the formation of vinylallenes exclusively in (*E*)-configuration for both intermediates (**A** and **A'**), the alkanolate group (CHR³O[−]) on the planar allyl ligand should be oriented syn with respect to the middle allylic C–H. These intermediates additionally undergo a palladium shift to the far alkynyl carbon proceeding mainly with retention to form vinylallenyl palladium complexes **B** and **B'**, respectively. The relative position of the R² group and alkynyl moiety on **A** and **A'** may also be inverted via the π – σ – π interconversion into **C** and **C'**,^{4c,10c} respectively, before undergoing such a Pd shift. The favored configuration of this side of the complex should be regulated by the inherent structure of R². For instance, when R² is hydrogen the thermodynamically more stable π -allyl complexes should bear alkynyl and CHR³O[−] moieties in syn,syn positions (**A'** and **C**).^{10c,d,11}

However, such configurational transitions cannot alter the outcome of the reaction, because both isomeric intermediates **C** and **C'** would also lead to **B** and **B'**, respectively. These σ -allenylpalladium complexes undergo successive CO insertion and reductive elimination, as verified previously,^{4,12} to yield **2** and **2'** stereoselectively.

The loss of stereochemical integrity of the resulting products should take place during the course of the reaction cycle because subjecting the purified **2ba** again to the standard conditions for 4 h had no influence on its original diastereomeric ratio.¹³ Though we cannot completely disregard the occurrence of the π -allylpalladium isomerization or its formation through syn-attack of the palladium, the back side migration of the palladium over the alkynyl side (by inversion of configuration) could be the primary basis for the formation of the minor diastereomeric form in each case, and particularly increased sizes of R¹ and R² might have limited the occurrence of such transitions in an inverted manner. However, the presence of a highly encumbered group, such as *tert*-butyl, at R² also retards the effective migration of the π -allyl coordinated palladium, and as a result (*Z*)-**1l** partly underwent allylic alkoxycarbonylation to form the byproduct **3la**.

Finally, the enyne oxirane (*Z*)-**1i'** containing a (*Z*)-configured oxirane moiety was also subjected to the standard reaction conditions. The reaction proceeded smoothly in accordance to the proposed mechanism, and thus yielded the vinylallene product **2ia'**, which is the diastereomer of **2ia** (see entry 7 of Table 4), with a high level of dr (Scheme 6).

CONCLUSION

In summary, we have established an unprecedented, mild, and efficient palladium-catalyzed diastereoselective 1,5-type alkoxycarbonylative substitution over conjugated enyne oxiranes, which provides a highly selective route to anti-substituted (*E*)-configured vinylallenes bearing a hydroxyl group on the allylic position. The reactions proceeded through successive formation of zwitterionic π -allyl- and σ -allenylpalladium complexes.

EXPERIMENTAL SECTION

General Methods. THF, DMF, and DCM solvents were all purified by a solvent purification system. Et₂O was distilled from benzophenone-ketyl under argon prior to use. Methanol and ethanol were dried over Mg turnings in the presence of iodine and stored on molecular sieve 3 Å under Ar. 1-Propanol and 2-propanol were dried first by stirring over anhydrous CaO followed by refluxing over Mg turnings in the presence of iodine. 1-Butanol was dried first by stirring over anhydrous MgSO₄ and then refluxed over Mg turnings in the presence of iodine. *tert*-BuOH was stirred over CaH₂ (5% w/v), distilled, and stored on molecular sieve 3 Å under Ar.

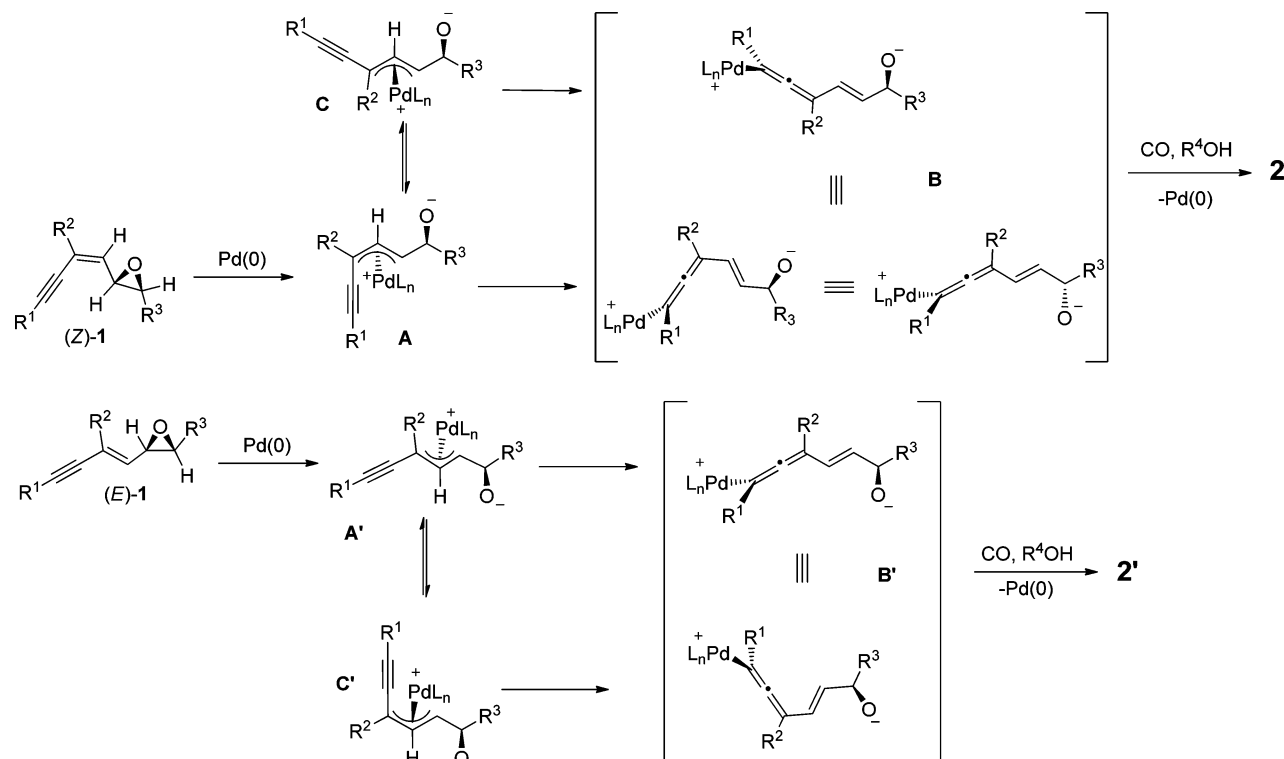
Syntheses of all enyne oxirane starting materials (**1** and **1'**) were performed under Ar gas, and purification of all synthesized molecules was performed by column chromatography on silica gel. Silica gel material used for the purification of enyne oxirane substrates had a particle size range of 60–200 mesh and was treated by NEt₃ before use. It must be noted that the column chromatography of the substrate **1** and **1'** on an untreated silica gel always resulted in decomposition. All other column purifications were performed on silica gel 60 (35–70 μ m). All substrates and vinylallene products appeared either colorless or as pale yellow oils.

The synthesized reactants and carbonylation products were analyzed by GC and GC-MS. NMR spectra were recorded on a 400 MHz spectrometer. Chemical shifts are reported in ppm downfield from Me₄Si. Infrared spectra were obtained by ATR method with neat samples. High-resolution mass spectral analyses of new compounds were performed using EI-High Resolution Double Focusing Magnetic Sector (ionization mode: 70 eV, emission current: 1 mA, source temperature: 160 °C, resolution: 10 000 (10% valley definition)) and ESI-LTQ Orbitrap (source voltage: +3.8 kV, capillary voltage: 41 V, capillary temperature: 275 °C, tube lens voltage: 140 V, system resolution: 60 000 (10% valley definition)).

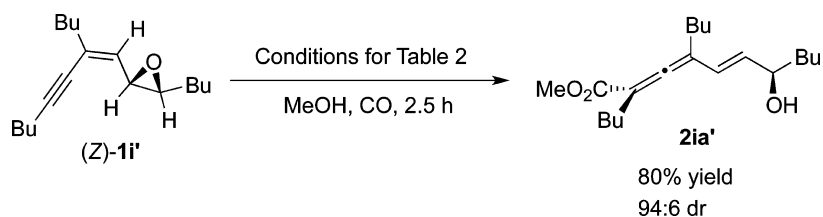
The Pd₂(dba)₃–CHCl₃ complex was synthesized in the laboratory.¹⁴

Synthesis of (*Z*)-1i** and (*Z*)-**1i'**.** 1-Hexyne (2.3 mL, 20 mmol) was added dropwise to a solution of BBr₃ (22 mL, 22 mmol, 1 M) in dry

Scheme 5. Mechanism of Alkoxy carbonylation of Conjugated Enyne Oxiranes



Scheme 6. Methoxycarbonylation of (Z)-1i'



DCM at -78°C . The mixture was allowed to warm to 0°C and stirred for 1 h. The reaction mixture was recooled to -78°C , and a solution of 2,3-dimethylbutan-2,3-diol (2.84 g, 24 mmol) in dry DCM (20 mL) was added dropwise. Following this addition, the mixture was warmed to ambient temperature and stirred for 1 h. Then, the mixture was quenched by brine and extracted with DCM. The combined organic layers were dried over MgSO_4 , filtered, and evaporated. The residue was purified on silica gel column to afford **S1** as a colorless oil (hexane–EtAc, yield: 3.57 g, 62%).¹⁵

To a stirred solution of **S1** (4.64 g, 16 mmol) and $\text{PdCl}_2(\text{PPh}_3)_2$ (1 mol %, 112 mg, 0.16 mmol) in dry THF (20 mL) was added a solution of 1-hexynylzinc chloride, which was generated by treatment of 1-hexyne (1.6 g, 19.2 mmol) with BuLi (8.5 mL, 20.7 mmol, 2.5 M in hexane) in dry THF (20 mL) for 30 min at -78°C and following treatment with ZnCl_2 (2.72 g, 19.2 mmol) for 30 min at 0°C . The resultant reaction mixture was stirred at room temperature (rt) for 1 h before quenching by 0.5 M HCl(aq). The content of the reaction medium was extracted with ether, washed with saturated NaHCO_3 and brine, dried over MgSO_4 , filtered, and concentrated under reduced pressure. The residue was purified on silica gel column to afford **S2** as a colorless oil (hexane–EtAc, yield: 3.9 g, 84%).¹⁵

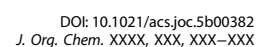
A mixture of 1.5 mol % of $\text{Pd}_2(\text{dba})_3$ (137 mg, 0.15 mmol), 6 mol % of AsPh_3 (184 mg, 0.6 mmol), and 15 mL of 1 M KOH(aq) in 50 mL of THF was cooled to 0°C before the addition of the compound **S2** (2.9 g, 10 mmol). Then, a THF solution (10 mL) of *trans*-¹⁶ or *cis*-1-iodo-1-hexene¹⁷ (2.52 g, 12 mmol), which were synthesized according to the literature procedures, was added dropwise, and the reaction

stirred for 30 min at room temperature. After the completion of the reaction, water was added to the resulting mixture before extracting with ether. The combined extracts were dried over MgSO_4 , filtered, and concentrated under reduced pressure. The residue was purified on silica gel column to afford the (*SE*,*7Z*)-8-butyltetradeca-5,7-dien-9-yne (**S3i**) or (*SZ*,*7Z*)-8-butyltetradeca-5,7-dien-9-yne (**S3i'**) as yellowish oil products (hexane–EtAc, yield of **S3i**: 1.89 g, 77%; yield of **S3i'**: 1.25 g, 51%).¹⁸

To a solution of **S3i** or **S3i'** (492 mg, 2 mmol) in DCM (30 mL) was added 12 mL solution of Na_2CO_3 (25%) followed by 3.4 mmol (587 mg) of *m*-CPBA dropwise at 0°C . The mixture was stirred at the same temperature and monitored with TLC until the reactant was consumed completely. At the end of the epoxidation process, the mixture was extracted with DCM, dried over anhydrous MgSO_4 , filtered, and concentrated under reduced pressure. The crude mixture was chromatographed on NEt_3 -pretreated short silica gel column which afforded the enyne oxirane (**Z**)-**1i** or (**Z**)-**1i'** in about 85% purities as pale yellow oil products (hexane–EtAc, yield of (**Z**)-**1i**: 300 mg, 57%; yield of (**Z**)-**1i'**: 158 mg, 30%).

(**Z**)-**1i**: ^1H NMR (400 MHz, C_6D_6) δ : 5.32 (d, $J = 8.7$ Hz, 1H), 3.90 (dd, $J = 8.7, 2.3$ Hz, 1H), 2.71 (td, $J = 5.5, 2.3$ Hz, 1H), 2.11 (t, $J = 6.8$ Hz, 2H), 2.10 (t, $J = 6.4$ Hz, 2H), 1.54–1.15 (m, 14H), 0.84–0.70 (m, 9H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, C_6D_6) δ : 137.5, 133.3, 95.7, 78.6, 59.6, 56.4, 37.4, 31.8, 30.7, 30.3, 28.0, 22.4, 22.0, 21.8, 19.0, 13.7, 13.6, 13.3.

(**Z**)-**1i'**: ^1H NMR (400 MHz, CDCl_3) δ : 5.38 (d, $J = 9.0$ Hz, 1H), 3.90 (dd, $J = 9.0, 4.8$ Hz, 1H), 3.11 (q, $J = 4.8$ Hz, 1H), 2.37 (t, $J = 6.8$ Hz,



1H), 2.06 (t, $J = 6.8$ Hz, 2H), 1.71 (d, $J = 1.6$ Hz, 3H), 1.29–1.16 (m, 4H), 0.7 (t, $J = 7.2$ Hz, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, C_6D_6) δ : 138.5, 132.8, 128.2, 127.4, 127.3, 124.8, 95.5, 79.1, 72.8, 70.0, 58.2, 53.8, 30.6, 23.5, 21.8, 18.9, 13.3.

(Z)-1d: ^1H NMR (400 MHz, CDCl_3) δ : 5.25 (dd, $J = 9.0, 1.4$ Hz, 1H), 3.96 (dd, $J = 12.6, 2.3$ Hz, 1H), 3.87 (dd, $J = 9.0, 2.3$ Hz, 1H), 3.68 (dd, $J = 12.6, 4.1$ Hz, 1H), 3.11–3.09 (m, 1H), 2.36 (t, $J = 7.0$ Hz, 2H), 1.88 (d, $J = 1.4$ Hz, 3H), 1.56–1.39 (m, 4H), 0.92 (t, $J = 7.2$ Hz, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, C_6D_6) δ : 132.5, 124.8, 95.7, 79.1, 61.1, 59.5, 53.7, 30.6, 23.5, 21.8, 19.0, 13.3.

(Z)-1e: ^1H NMR (400 MHz, CDCl_3) δ : 5.22 (dd, $J = 9.1, 1.5$ Hz, 1H), 3.86 (dd, $J = 12.1, 3.2$ Hz, 1H), 3.76 (dd, $J = 9.1, 2.4$ Hz, 1H), 3.71 (dd, $J = 12.1, 4.8$ Hz, 1H), 3.00 (ddd, $J = 4.8, 3.2, 2.4$ Hz, 1H), 2.33 (t, $J = 7.2$ Hz, 2H), 1.86 (d, $J = 1.5$ Hz, 3H), 1.56–1.37 (m, 4H), 0.91 (t, $J = 6.8$ Hz, 3H), 0.89 (s, 9H), 0.07 (s, 3H), 0.06 (s, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ : 131.9, 125.3, 95.7, 78.7, 63.0, 60.1, 54.3, 30.8, 25.8, 23.8, 22.0, 19.1, 18.3, 13.6, –5.3, –5.4.

(Z)-1h: ^1H NMR (400 MHz, CDCl_3) δ : 5.75 (dtd, $J = 10.8, 2.4, 0.8$ Hz, 1H), 5.44 (dd, $J = 10.8, 8.8$ Hz, 1H), 3.80 (dd, $J = 8.8, 2.2$ Hz, 1H), 3.74 (dd, $J = 11.2, 2.8$ Hz, 1H), 3.44–3.38 (m, 1H), 3.41 (s, 3H), 3.11 (dt, $J = 5.4, 2.6$ Hz, 1H), 2.36 (td, $J = 6.8, 2.3$ Hz, 2H), 1.58–1.41 (m, 4H), 0.92 (t, $J = 7.2$ Hz, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ : 137.2, 115.4, 97.1, 72.3, 59.2, 58.2, 53.3, 50.4, 30.6, 21.9, 19.2, 13.5.

(Z)-1k: ^1H NMR (400 MHz, CDCl_3) δ : 5.21 (d, $J = 8.7$ Hz, 1H), 3.77 (dd, $J = 8.7, 2.4$ Hz, 1H), 3.72 (dd, $J = 11.5, 3.1$ Hz, 1H), 3.38 (dd, $J = 11.5, 5.7$ Hz, 1H), 3.40 (s, 3H), 3.07 (dt, $J = 5.7, 2.4$ Hz, 1H), 2.37 (t, $J = 7.2$ Hz, 2H), 2.02–1.97 (m, 1H), 1.76–1.20 (m, 14H), 0.92 (t, $J = 7.2$ Hz, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ : 136.3, 129.1, 97.0, 76.7, 72.6, 59.2, 58.5, 54.3, 45.4, 31.7, 31.6, 30.8, 29.7, 26.2, 26.0, 22.0, 19.2, 13.6.

(Z)-1l: ^1H NMR (400 MHz, CDCl_3) δ : 5.24 (d, $J = 8.9$ Hz, 1H), 3.81 (dd, $J = 8.9, 2.4$ Hz, 1H), 3.74 (dd, $J = 11.2, 3.1$ Hz, 1H), 3.42–3.38 (m, 1H), 3.41 (s, 3H), 3.10 (ddd, $J = 5.6, 3.1, 2.4$ Hz, 1H), 2.39 (t, $J = 7.2$ Hz, 2H), 1.59–1.41 (m, 4H), 1.11 (s, 9H), 0.93 (t, $J = 7.6$ Hz, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ : 140.2, 127.4, 97.4, 72.7, 59.2, 58.5, 54.7, 36.1, 30.8, 28.9, 21.9, 19.2, 13.6.

(Z)-1m: ^1H NMR (400 MHz, C_6D_6) δ : 5.10 (dq, $J = 8.9, 1.0$ Hz, 1H), 3.84 (dd, $J = 8.9, 2.0$ Hz, 1H), 3.25 (dd, $J = 11.5, 3.0$ Hz, 1H), 3.06–2.99 (m, 1H), 3.05 (s, 3H), 2.83 (ddd, $J = 5.5, 3.0, 2.0$ Hz, 1H), 2.74 (s, 1H), 1.57 (d, $J = 1.0$ Hz, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, C_6D_6) δ : 136.0, 122.9, 82.4, 81.5, 72.2, 58.4, 58.0, 53.2, 22.7.

(Z)-1n: ^1H NMR (400 MHz, C_6D_6) δ : 5.14 (dq, $J = 9.0, 1.4$ Hz, 1H), 4.01 (dd, $J = 9.0, 2.0$ Hz, 1H), 3.35 (dd, $J = 11.3, 3.2$ Hz, 1H), 3.16 (dd, $J = 11.3, 5.5$ Hz, 1H), 3.09 (s, 3H), 2.91 (ddd, $J = 5.5, 3.2, 2.0$ Hz, 1H), 2.34 (m, 1H), 1.71 (d, $J = 1.4$ Hz, 3H), 1.66–1.03 (m, 10H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, C_6D_6) δ : 132.7, 124.7, 99.5, 79.2, 72.4, 58.5, 58.1, 53.7, 32.5, 32.4, 29.6, 25.7, 24.6, 23.5.

(Z)-1o: ^1H NMR (400 MHz, C_6D_6) δ : 5.12 (dd, $J = 8.5, 1.3$ Hz, 1H), 3.95 (dd, $J = 8.5, 2.3$ Hz, 1H), 3.34 (dd, $J = 11.5, 3.1$ Hz, 1H), 3.16 (dd, $J = 11.5, 5.5$ Hz, 1H), 3.09 (s, 3H), 2.90 (ddd, $J = 5.5, 3.1, 2.3$ Hz, 1H), 1.68 (d, $J = 1.3$ Hz, 3H), 1.12 (s, 9H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, C_6D_6) δ : 132.8, 124.5, 103.4, 77.7, 72.4, 58.5, 58.0, 53.7, 30.6, 27.9, 23.4.

(Z)-1p: ^1H NMR (400 MHz, CDCl_3) δ : 7.35–7.28 (m, 5H), 5.22 (dq, $J = 8.9, 1.4$ Hz, 1H), 4.61 (d, A of AB, $J_{AB} = 12.0$ Hz, 1H), 4.57 (d, B of AB, $J_{AB} = 12.0$ Hz, 1H), 3.81 (dd, $J = 11.3, 3.0$ Hz, 1H), 3.76 (dd, $J = 8.9, 2.2$ Hz, 1H), 3.53 (dd, $J = 11.3, 5.5$ Hz, 1H), 3.13 (ddd, $J = 5.5, 3.0, 2.2$ Hz, 1H), 1.86 (d, $J = 1.4$ Hz, 3H), 1.24 (s, 9H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ : 140.0, 137.9, 131.3, 128.4, 127.7, 125.7, 104.0, 77.1, 73.3, 70.1, 58.6, 54.3, 31.0, 28.1, 23.8.

(Z)-1q: ^1H NMR (400 MHz, C_6D_6) δ : 5.17 (dd, $J = 8.9, 1.1$ Hz, 1H), 4.1 (dd, $J = 8.9, 2.3$ Hz, 1H), 3.54–3.48 (m, 1H), 3.34–3.30 (m, 1H), 2.75–2.73 (m, 1H), 1.73 (d, $J = 1.1$ Hz, 3H), 1.17 (s, 9H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, C_6D_6) δ : 132.5, 124.6, 103.7, 77.6, 61.0, 59.4, 53.6, 30.6, 27.9, 23.4.

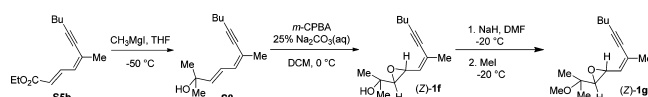
(Z)-1r: ^1H NMR (400 MHz, CDCl_3) δ : 5.21 (d, $J = 9.0$ Hz, 1H), 3.87 (dd, $J = 12.0, 3.2$ Hz, 1H), 3.74 (m, 2H), 3.01 (m, 1H), 1.86 (s, 3H), 1.26 (s, 9H), 0.90 (s, 9H), 0.08 (s, 6H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ : 131.6, 125.3, 103.8, 77.1, 62.9, 60.2, 54.3, 31.0, 28.1, 25.9, 23.9, 18.3, –5.3.

(Z)-1s: ^1H NMR (400 MHz, CDCl_3) δ : 7.46–7.44 (m, 2H), 7.34–7.32 (m, 3H), 5.38 (dq, $J = 9.0, 1.5$ Hz, 1H), 3.84 (dd, $J = 9.0, 2.4$ Hz, 1H), 3.74 (dd, $J = 11.7, 3.2$ Hz, 1H), 3.43 (dd, $J = 11.7, 6.0$ Hz, 1H), 3.42 (s, 3H), 3.14 (ddd, $J = 6.0, 3.2, 2.4$ Hz, 1H), 2.00 (d, $J = 1.5$ Hz, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ : 133.1, 131.5, 128.5, 128.3, 124.9, 122.9, 94.5, 87.3, 72.5, 59.2, 58.5, 54.1, 23.4.

Synthesis of (Z)-1f and (Z)-1g. To the dry Et_2O (15 mL) solution of **S5b** (1.1 g, 5 mmol) was added an ethereal (15 mL) solution of 2.1 eq. MeMgI (5.25 mL, 10.5 mmol, 2 M) dropwise at -50°C , and then the mixture stirred for 6 h at the same temperature. The mixture was allowed to warm to 0°C and quenched by the addition of 30 mL of saturated $\text{NH}_4\text{Cl(aq)}$ solution before extracting with Et_2O . The combined extracts were dried over MgSO_4 , filtered, and evaporated. The resulting residue was purified on a silica gel column to afford **S8** (hexane– EtAc , yield: 0.49 g, 80%).

The epoxidation of **S8** (412 mg, 2 mmol) and isolation of the product (Z)-1f were performed as specified for (Z)-1i (hexane– EtOAc , yield: 0.27 g, 60%).

The hydroxyl group of (Z)-1f (222 mg, 1.0 mmol) was methylated as described above furnishing the enyne oxirane (Z)-1g in 90% yield (0.21 g).



(Z)-1f: ^1H NMR (400 MHz, CDCl_3) δ : 5.26 (dd, $J = 8.8, 1.6$ Hz, 1H), 3.89 (dd, $J = 8.8, 2.3$ Hz, 1H), 2.88 (d, $J = 2.3$ Hz, 1H), 2.36 (t, $J = 7.0$ Hz, 2H), 1.87 (d, $J = 1.6$ Hz, 3H), 1.78 (d, $J = 0.8$ Hz, 1H), 1.57–1.40 (m, 4H), 1.31 (s, 3H), 1.25 (s, 3H), 0.92 (t, $J = 6.8$ Hz, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ : 132.0, 125.1, 78.8, 73.8, 64.7, 53.3, 50.7, 30.7, 23.7, 22.2, 21.9, 21.0, 19.1, 13.5.

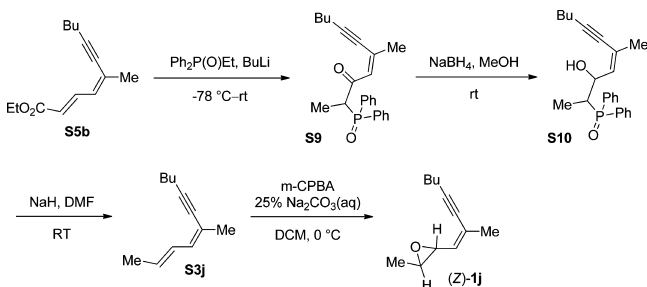
(Z)-1g: ^1H NMR (400 MHz, CDCl_3) δ : 5.24 (dd, $J = 8.9, 1.5$ Hz, 1H), 3.69 (dd, $J = 8.9, 2.3$ Hz, 1H), 3.29 (s, 3H), 2.87 (d, $J = 2.3$ Hz, 1H), 2.36 (t, $J = 6.7$ Hz, 2H), 1.87 (d, $J = 1.5$ Hz, 3H), 1.56–1.39 (m, 4H), 1.18 (s, 3H), 1.15 (s, 3H), 0.92 (t, $J = 7.2$ Hz, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ : 132.2, 125.3, 95.8, 79.0, 74.0, 64.9, 53.4, 50.9, 30.9, 23.9, 22.4, 22.0, 21.1, 19.2, 13.7.

Synthesis of (Z)-1j. To a stirred solution of diphenylethylphosphine oxide (4.6 g, 20 mmol) in dry THF (70 mL) was added BuLi (2.5 M in hexane, 8.8 mL) dropwise at 0°C and stirred for a further 30 min. The solution was cooled to -78°C , and then the diene **S5b** (3.88 g, 20 mol) was added dropwise. The solution was allowed to warm to ambient temperature and subsequently stirred overnight. Saturated NH_4Cl solution was added, and subsequently its THF content was removed under reduced pressure. The aqueous residue was diluted with brine (20 mL) and extracted with DCM. The combined organic phases were dried over MgSO_4 , filtered, and concentrated under reduced pressure. The product **S9** was purified by column chromatography on silica gel (hexane– EtAc , yield: 1.9 g, 25%).²⁴

To a stirred solution of **S9** (1.9 g, 5 mmol) in ethanol (50 mL) was added NaBH_4 (189 mg, 5 mmol) in one portion and stirred for a further 8 h at ambient temperature. The reaction afforded **S10** enriched in threo form. Saturated NH_4Cl (15 mL) was added, and subsequently its ethanol content was removed under reduced pressure. The aqueous mixture was diluted with brine (20 mL) and extracted with DCM. The combined organic layers were dried over MgSO_4 , filtered, and concentrated under reduced pressure. The product **S10** was isolated in pure threo form by column chromatography on silica gel (hexane/ EtAc , yield: 1.31 g, 69%).²⁴

To a stirred solution of **S10** (1.31 g, 3.45 mmol) in DMF (50 mL) was added NaH (60% dispersion in oil; 138 mg, 3.45 mmol) in one portion at ambient temperature and stirred for a further 3 h. The reaction was quenched by the addition of 25 mL of water and 15 mL of brine and subsequently extracted with Et_2O . The combined extracts were washed with water, dried over MgSO_4 , filtered, and concentrated under reduced pressure. The product **S3j** was purified by column chromatography on silica gel (hexane, yield: 330 mg, 59%).²⁴

The epoxidation of **S3j** (162 mg, 1 mmol) and isolation of the product (**Z**)-**1j** were performed as specified for (**Z**)-**1i** (hexane–EtAc, yield: 35.6 mg, 20%).



S9: ^1H NMR (400 MHz, CDCl_3) δ : 7.86–7.74 (m, 4H), 7.55–7.43 (m, 6H), 6.34 (d, J = 1.2 Hz, 1H), 3.99 (dq, J = 13.5, 7.0 Hz, 1H), 2.42 (t, J = 7.0 Hz, 2H), 1.89 (d, J = 1.2 Hz, 3H), 1.60–1.51 (m, 2H), 1.47–1.38 (m, 5H), 0.92 (t, J = 7.0 Hz, 3H).

S10: ^1H NMR (400 MHz, CDCl_3) δ : 7.83–7.71 (m, 4H), 7.56–7.43 (m, 6H), 5.53 (dd, J = 8.3, 1.2 Hz, 1H), 4.80 (dt, J = 13.8, 8.3 Hz, 1H), 2.82–2.71 (m, 1H), 2.15 (t, J = 7.2 Hz, 2H), 1.65 (d, J = 1.2 Hz, 3H), 1.40–1.28 (m, 4H), 1.04 (dd, J = 17.2, 7.4 Hz, 3H), 0.78 (t, J = 7.2 Hz, 3H).

S3j: ^1H NMR (400 MHz, CDCl_3) δ : 6.48 (ddq, J = 15.4, 10.8, 1.6 Hz, 1H), 6.11 (d, J = 10.8 Hz, 1H), 4.80 (dq, J = 15.4, 7.0 Hz, 1H), 2.39 (t, J = 6.8 Hz, 2H), 1.86 (s, 3H), 1.79 (d, J = 7.0 Hz, 3H), 1.60–1.52 (m, 2H), 1.51–1.44 (m, 2H), 0.94 (t, J = 6.8 Hz, 3H).

(**Z**)-**1j**: ^1H NMR (400 MHz, C_6D_6) δ : 5.16 (dd, J = 8.9, 1.4 Hz, 1H), 3.75 (dd, J = 8.9, 2.1 Hz, 1H), 2.64 (qd, J = 5.4, 2.1 Hz, 1H), 2.08 (t, J = 6.8 Hz, 2H), 1.73 (d, J = 1.4 Hz, 3H), 1.31–1.20 (m, 4H), 1.04 (d, J = 5.2 Hz, 3H), 0.72 (t, J = 7.2 Hz, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, C_6D_6) δ : 133.7, 123.9, 95.2, 79.3, 57.4, 55.2, 30.6, 23.4, 21.8, 19.0, 17.3, 13.2.

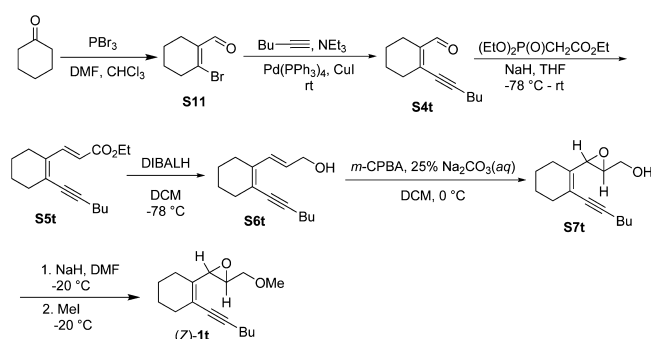
Synthesis of (Z**)-1t**. PBr_3 (1.4 mL, 13.8 mmol) was added dropwise to a mixture of DMF (1.2 mL, 15.3 mmol) and chloroform (10 mL) at 0 °C, and then the resulting mixture stirred for 1 h. Subsequently, 0.5 g of cyclohexanone (6 mmol) was added dropwise and stirred for 8 h at room temperature. The reaction was terminated with water, neutralized with the addition of solid NaHCO_3 , and extracted with DCM. The extract was washed with brine, dried over MgSO_4 , filtered, and concentrated under reduced pressure. The purification on short silica gel column provided the compound **S11** (hexane/EtAc, 0.92 g, 81%).²⁵

A mixture of **S11** (945 mg, 5 mmol), $\text{Pd}(\text{PPh}_3)_4$ (123 mg, 0.1 mmol, 2 mol % of Pd), and CuI (21 mg, 0.1 mmol, 2 mmol % of Cu) in 10 mL of Et_3N was stirred for 10 min at room temperature followed by the addition of 1-hexyne (0.5 g, 6 mmol). After being stirred for 3 h at room temperature, water was added and extracted with Et_2O . The combined organic phases were dried over MgSO_4 , filtered, and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel to obtain endocyclic enyne aldehyde **S4t** (hexane/EtAc, 0.84 g, 90%).²⁵

The conversion of **S4t** (840 mg, 4.42 mmol) to dienyne ester **S5t** was performed by HWE reaction as described above (hexane/EtAc, 0.96 g, 84%). Further successive synthetic procedures, which involved the reduction of the ester **S5t** (960 mg, 3.7 mmol) to the enyne alcohol **S6t** (730 mg, 91% yield), the epoxidation of **S6t** (436 mg, 2 mmol) to **S7t** (260 mg, 55% yield), and finally methyl derivatization of hydroxyl group of **S7t** (260 mg, 1.1 mmol) to obtain (**Z**)-**1t** (0.22 g, 90%) were all conducted as described above.

(**Z**)-**1t**: ^1H NMR (400 MHz, CDCl_3) δ : 4.00–3.97 (m, 1H), 3.73 (dt, J = 11.2, 2.8 Hz, 1H), 3.45–3.30 (m, 4H), 3.20 (sext, J = 2.8 Hz, 1H), 2.34 (t, J = 6.8 Hz, 2H), 2.21–2.10 (m, 2H), 2.02–1.91 (m, 2H), 1.79–1.32 (m, 6H), 0.98–0.79 (m, 5H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ : 137.3, 121.7, 94.2, 79.5, 73.1, 59.2, 56.2, 55.2, 30.9, 30.8, 22.4, 22.2, 21.9, 21.7, 19.2, 13.6.

Carbonylation Reactions. A palladium complex, phosphine ligand, and 1 mL of alcohol were added successively to a Schlenk apparatus that is attached to an Ar line and stirred 1 h at 25 °C before the addition of the substrate. A CO full balloon was then fixed to the



reaction vessel, and 1 mL of alcohol solution of the substrate **1** or **1'** was added. When the reaction was complete as judged by TLC analysis, the solvent content was evaporated and the residue was purified by column chromatography on silica gel to afford **2** or **2'** as a pale yellow oil. The coupling constants of olefinic protons and NOE studies confirmed (*E*)-configured structures. ^1H NMR analyses of vinylallene products, except that of **2ia'**, were performed in C_6D_6 . With this solvent, the ^1H NMR signals of diastereomers were resolved adequately, allowing to determine diastereomeric ratios smoothly. In contrast, when using CDCl_3 solvent, diastereomeric signals were all overlapped.

Product Data. 2aa: Yield: 24.9 mg (99%); ^1H NMR (400 MHz, CDCl_3) δ : 6.13 (d, J = 16.0 Hz, 1H), 5.76 (d, J = 16.0 Hz, 1H), 3.67 (s, 3H), 2.22 (t, J = 7.6 Hz, 2H), 1.85 (s, 3H), 1.37–1.27 (m, 10H), 0.86 (t, J = 7.0 Hz, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ : 213.4, 167.7, 137.8, 122.9, 103.1, 99.7, 70.9, 52.0, 30.1, 29.8, 28.6, 22.1, 14.8, 13.8; FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3409, 2957, 2930, 2871, 1940, 1714, 1436, 1378, 1264, 1133, 1083, 1019, 966, 912, 723, 696; MS (EI, m/z): 252 (1, M^+), 234 (2), 205 (7), 194 (7), 179 (5), 151 (23), 135 (35), 121 (15), 107 (32), 91 (43), 77 (25), 65 (15), 59 (46), 43 (100); HRMS (ESI) $\text{C}_{15}\text{H}_{25}\text{O}_3$ (MH^+), 253.1798 (calculated); 253.1799 (found).

2ba: Yield: 25.2 mg (94%); ^1H NMR (400 MHz, C_6D_6) δ : 6.43 (dd, J = 15.7, 1.6 Hz, 1H, major), 6.41 (dd, J = 15.7, 1.6 Hz, 1H, minor), 5.51 (dd, J = 15.7, 5.5 Hz, 1H, major), 5.51 (dd, J = 15.7, 5.5 Hz, 1H, minor), 4.23–4.19 (m, 1H), 3.36 (s, 3H, major), 3.35 (s, 3H, minor), 3.03 (dd, A of ABX, J_{AB} = 9.2 Hz, J_{AX} = 4.0 Hz, 1H), 2.98 (dd, B of ABX, J_{AB} = 9.2 Hz, J_{BX} = 7.2 Hz, 1H), 2.95 (s, 3H), 2.39 (td, J = 7.5, 2.2 Hz, 2H), 2.23 (bs, 1H), 1.69 (s, 3H), 1.47–1.39 (m, 2H), 1.23 (sext, J = 7.6 Hz, 2H), 0.78 (t, J = 7.2 Hz, 3H, major), 0.78 (t, J = 7.2 Hz, 3H, minor); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, C_6D_6) δ : 213.5, 166.9, 129.2, 127.3, 103.1, 76.5, 70.6, 58.2, 51.3, 30.3, 28.9, 22.2, 14.5, 13.7; FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3419, 2955, 2928, 2870, 1943, 1715, 1456, 1436, 1379, 1266, 1241, 1193, 1129, 1016, 966; MS (EI, m/z): 268 (1, M^+), 236 (1), 223 (4), 205 (1), 191 (6), 151 (2), 135 (7), 121 (6), 93 (8), 91 (14), 77 (10), 59 (6), 45 (100); HRMS (ESI) $\text{C}_{15}\text{H}_{25}\text{O}_4$ (MH^+), 269.1747 (calculated); 269.1747 (found).

2ba': Yield: 23.3 mg (87%); ^1H NMR (400 MHz, C_6D_6) δ : 6.42 (dd, J = 15.7, 1.6 Hz, 1H, major), 6.40 (dd, J = 15.7, 1.6 Hz, 1H, minor), 5.52 (dd, J = 15.7, 5.7 Hz, 1H), 4.22–4.20 (m, 1H), 3.37 (s, 3H, major), 3.35 (s, 3H, minor), 3.03 (dd, A of ABX, J_{AB} = 9.6 Hz, J_{AX} = 4.0 Hz, 1H), 2.99 (dd, B of ABX, J_{AB} = 9.6 Hz, J_{BX} = 8.0 Hz, 1H), 2.96 (s, 3H), 2.38 (td, J = 7.6 Hz, 1.8 Hz, 2H), 2.29 (bs, 1H), 1.70 (s, 3H), 1.48–1.40 (m, 2H), 1.25 (sext, J = 7.6 Hz, 2H), 0.79 (t, J = 7.6 Hz, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, C_6D_6) δ : 213.5, 166.9, 129.2, 103.1, 100.0, 76.5, 70.6, 58.2, 51.3, 30.3, 28.9, 22.2, 14.5, 13.7; FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3419, 2955, 2928, 2870, 1943, 1715, 1456, 1436, 1379, 1266, 1241, 1193, 1129, 1016, 966; MS (EI, m/z): 268 (1, M^+), 236 (1), 223 (4), 205 (1), 191 (6), 151 (2), 135 (7), 121 (6), 93 (8), 91 (14), 77 (10), 59 (6), 45 (100); HRMS (ESI) $\text{C}_{15}\text{H}_{25}\text{O}_4$ (MH^+), 269.1747 (calculated); 269.1747 (found).

2bb: Yield: 23.7 mg (84%); ^1H NMR (400 MHz, C_6D_6) δ : 6.44 (dd, J = 15.4, 1.6 Hz, 1H, major), 6.43 (dd, J = 15.4, 1.6 Hz, 1H, minor), 5.54 (dd, J = 15.4, 5.6 Hz, 1H), 4.24–4.18 (m, 1H), 3.99 (q, J = 7.1 Hz, 2H), 3.03 (dd, A of ABX, J_{AB} = 9.6 Hz, J_{AX} = 4.0 Hz, 1H), 2.98 (dd, B of ABX, J_{AB} = 9.6 Hz, J_{BX} = 7.6 Hz, 1H), 2.95 (s, 3H), 2.41 (td, J = 7.2, 2.4 Hz, 2H), 2.28 (bs, 1H), 1.71 (s, 3H), 1.49–1.41

(m, 2H), 1.25 (sext, $J = 7.2$ Hz, 2H), 0.92 (t, $J = 7.1$ Hz, 3H, major), 0.91 (t, $J = 7.1$ Hz, 3H, minor), 0.79 (t, $J = 7.2$ Hz, 3H, major), 0.79 (t, $J = 7.2$ Hz, 3H, minor); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ : 213.5, 167.1, 128.7, 127.8, 103.1, 100.3, 76.4, 71.1, 60.8, 59.1, 30.2, 28.6, 22.2, 14.7, 14.3, 13.9; FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3418, 2956, 2926, 2858, 1940, 1708, 1458, 1367, 1322, 1261, 1240, 1172, 1118, 1062, 1023, 965, 864, 713. MS (EI, m/z): 282 (1, M^+), 238 (1), 237 (4), 209 (2), 191 (5), 135 (3), 121 (5), 107 (6), 93 (8), 91 (12), 77 (7), 65 (5), 55 (7), 45 (100); HRMS (ESI): $\text{C}_{16}\text{H}_{26}\text{O}_4$ (MH) $^+$, 283.1904 (calculated), 283.1909 (found).

2bc: Yield: 25.8 mg (87%); ^1H NMR (400 MHz, C_6D_6) δ : 6.44 (dd, $J = 15.8, 1.6$ Hz, 1H, major), 6.43 (dd, $J = 15.8, 1.6$ Hz, 1H, minor), 5.53 (dd, $J = 15.8, 5.6$ Hz, 1H), 4.23–4.19 (m, 1H), 4.02–3.92 (m, 2H), 3.03 (dd, A of ABX, $J_{\text{AB}} = 9.6$ Hz, $J_{\text{AX}} = 4.4$ Hz, 1H), 2.98 (dd, B of ABX, $J_{\text{AB}} = 9.6$ Hz, $J_{\text{BX}} = 7.2$ Hz, 1H), 2.94 (s, 3H), 2.41 (td, $J = 7.3, 1.6$ Hz, 2H), 2.22 (bs, 1H), 1.72 (s, 3H), 1.49–1.33 (m, 4H), 1.25 (sext, $J = 7.4$ Hz, 2H), 0.79 (t, $J = 7.4$ Hz, 3H, major), 0.80 (t, $J = 7.4$ Hz, 3H, minor), 0.66 (t, $J = 7.3$ Hz, 3H, major), 0.65 (t, $J = 7.3$ Hz, 3H, minor); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ : 213.6, 167.2, 128.6, 127.9, 103.1, 100.3, 76.5, 71.1, 66.3, 59.1, 30.2, 28.5, 22.2, 22.0, 14.7, 13.9, 10.3; FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3406, 2958, 2928, 1941, 1709, 1458, 1397, 1322, 1263, 1172, 1132, 1062, 1015, 965, 713; MS (EI, m/z): 296 (1, M^+), 278 (1), 251 (3), 236 (1), 209 (3), 191 (5), 135 (4), 121 (5), 91 (10), 79 (6), 65 (3), 45 (100); HRMS (ESI): $\text{C}_{17}\text{H}_{28}\text{O}_4$ (MH) $^+$, 297.2060 (calculated), 297.2065 (found).

2bd: Yield: 25.7 mg (83%); ^1H NMR (400 MHz, C_6D_6) δ : 6.45 (dd, $J = 16.0, 1.2$ Hz, 1H, major), 6.43 (dd, $J = 16.0, 1.2$ Hz, 1H, minor), 5.53 (dd, $J = 16.0, 5.6$ Hz, 1H, major), 5.53 (dd, $J = 16, 5.6$ Hz, 1H, minor), 4.24–4.19 (m, 1H), 4.09–3.99 (m, 2H), 3.04 (dd, A of ABX, $J_{\text{AB}} = 9.6$ Hz, $J_{\text{AX}} = 4.0$ Hz, 1H), 2.99 (dd, B of ABX, $J_{\text{AB}} = 9.6$ Hz, $J_{\text{BX}} = 7.6$ Hz, 1H), 2.95 (s, 3H), 2.41 (td, $J = 7.4, 2.4$ Hz, 2H), 2.26 (bs, 1H), 1.75 (s, 3H, minor), 1.73 (s, 3H, major), 1.46 (quint, $J = 7.2$ Hz, 2H), 1.36 (quint, $J = 7.4$ Hz, 2H), 1.25 (sext, $J = 7.2$ Hz, 2H), 1.12 (sext, $J = 7.4$ Hz, 2H), 0.80 (t, $J = 7.2$ Hz, 3H, minor), 0.79 (t, $J = 7.2$ Hz, 3H, major), 0.68 (t, $J = 7.4$ Hz, 3H, major), 0.67 (t, $J = 7.4$ Hz, 3H, minor); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ : 213.6, 167.2, 128.6, 127.8, 103.1, 100.3, 76.4, 71.1, 64.7, 59.1, 30.6, 30.2, 28.5, 22.2, 19.2, 14.7, 13.9, 13.7; FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3428, 2958, 2929, 2873, 1940, 1710, 1458, 1379, 1322, 1261, 1131, 1062, 1016, 965, 713; MS (EI, m/z): 310 (1, M^+), 293 (1), 265 (1), 235 (2), 207 (4), 191 (4), 161 (3), 121 (3), 105 (5), 91 (10), 79 (5), 57 (15), 45 (100); HRMS (ESI): $\text{C}_{18}\text{H}_{31}\text{O}_4$ (MH) $^+$, 311.2217 (calculated), 311.2219 (found).

2be: Yield: 26.1 mg (88%); ^1H NMR (400 MHz, C_6D_6) δ : 6.43 (dd, $J = 16.0, 1.6$ Hz, 1H), 5.51 (dd, $J = 16.0, 5.6$ Hz, 1H), 5.05 (sept, $J = 6.4$ Hz, 1H, major), 5.05 (sept, $J = 6.4$ Hz, 1H, minor), 4.23–4.16 (m, 1H), 3.02 (dd, A of ABX, $J_{\text{AB}} = 9.2$ Hz, $J_{\text{AX}} = 4.0$ Hz, 1H), 2.97 (dd, B of ABX, $J_{\text{AB}} = 9.2$ Hz, $J_{\text{BX}} = 7.6$ Hz, 1H), 2.94 (s, 3H), 2.42 (td, $J = 7.6, 2.0$ Hz, 2H), 2.21 (bs, 1H), 1.71 (s, 3H), 1.45 (quint, $J = 7.6$ Hz, 2H), 1.25 (sext, $J = 7.6$ Hz, 2H), 1.02 (d, $J = 6.4$ Hz, 3H), 1.01 (d, $J = 6.4$ Hz, 3H), 0.79 (t, $J = 7.6$ Hz, 3H, major), 0.79 (t, $J = 7.6$ Hz, 3H, minor); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ : 213.5, 166.7, 128.9, 127.6, 103.0, 100.7, 76.5, 71.1, 68.1, 59.1, 30.2, 28.6, 22.2, 21.9, 21.8, 14.7, 13.9; FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3442, 2957, 2927, 1942, 1706, 1457, 1373, 1322, 1264, 1107, 1062, 1014, 966, 831, 713; MS (EI, m/z): 296 (1, M^+), 282 (1), 251 (2), 209 (3), 191 (4), 180 (1), 121 (6), 107 (4), 91 (7), 77 (6), 55 (4), 45 (100); HRMS (ESI): $\text{C}_{17}\text{H}_{29}\text{O}_4$ (MH) $^+$, 297.2060 (calculated), 297.2060 (found).

2bg: Yield: 26.8 mg (91%); ^1H NMR (400 MHz, C_6D_6) δ : 6.43 (dd, $J = 15.6, 0.8$ Hz, 1H), 5.72–5.62 (m, 1H), 5.52 (dd, $J = 15.6, 5.6$ Hz, 1H), 5.08 (dq, $J = 18.8, 1.6$ Hz, 1H), 4.89 (dq, $J = 6.4, 1.6$ Hz, 1H), 4.53–4.41 (m, 2H), 4.23–4.19 (m, 1H), 3.04 (dd, A of ABX, $J_{\text{AB}} = 9.6$ Hz, $J_{\text{AX}} = 4.0$ Hz, 1H), 2.99 (dd, B of ABX, $J_{\text{AB}} = 9.6$ Hz, $J_{\text{BX}} = 7.2$ Hz, 1H), 2.95 (s, 3H), 2.39 (td, $J = 7.2, 2.0$ Hz, 2H), 2.31 (bs, 1H), 1.70 (s, 3H), 1.44 (quint, $J = 7.2$ Hz, 2H), 1.24 (sext, $J = 7.2$ Hz, 2H), 0.79 (t, $J = 7.2$ Hz, 3H, minor), 0.78 (t, $J = 7.2$ Hz, 3H, major); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ : 213.7, 166.8, 132.4, 128.5, 128.1, 117.4, 103.2, 100.0, 76.4, 71.1, 65.2, 59.1, 30.1, 28.5, 22.2, 14.7, 13.9; FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3417, 2956, 2928, 2873, 1939, 1710, 1647, 1456, 1379, 1322, 1259, 1236, 1172, 1129, 1062, 1015, 965, 928, 837, 713. MS (EI, m/z): 294 (1, M^+), 261 (1), 249 (2), 221 (1), 191 (5),

165 (3), 121 (6), 91 (11), 77 (9), 65 (4), 55 (7), 45 (100); HRMS (ESI): $\text{C}_{17}\text{H}_{27}\text{O}_4$ (MH) $^+$, 295.1904 (calculated), 295.1910 (found).

2ca: Yield: 31.0 mg (90%); ^1H NMR (400 MHz, C_6D_6) δ : 7.24–7.01 (m, 5H), 6.44 (dd, $J = 15.8, 1.6$ Hz, 1H, major), 6.42 (dd, $J = 15.8, 1.6$ Hz, 1H, minor), 5.49 (dd, $J = 15.8, 5.7$ Hz, 1H), 4.29–4.23 (m, 1H), 4.21 (d, A of AB, $J_{\text{AB}} = 12.0$ Hz, 1H), 4.19 (d, B of AB, $J_{\text{AB}} = 12.0$ Hz, 1H), 3.36 (s, 3H, major), 3.34 (s, 3H, minor), 3.18 (dd, A of ABX, $J_{\text{AB}} = 9.2$ Hz, $J_{\text{AX}} = 3.6$ Hz, 1H), 3.11 (dd, B of ABX, $J_{\text{AB}} = 9.2$ Hz, $J_{\text{BX}} = 7.6$ Hz, 1H), 2.38 (td, $J = 7.6, 2.8$ Hz, 2H), 1.69 (s, 3H), 1.42 (quint, $J = 7.6$ Hz, 2H), 1.23 (sext, $J = 7.6$ Hz, 2H), 0.78 (t, $J = 7.6$ Hz, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ : 213.6, 167.6, 137.7, 128.5, 128.4, 128.0, 127.9, 127.8, 103.2, 99.9, 74.0, 73.4, 71.2, 52.1, 30.1, 28.6, 22.2, 14.7, 13.9; FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3416, 2954, 2928, 2869, 1943, 1714, 1454, 1435, 1379, 1322, 1266, 1134, 1016, 965, 738, 698. MS (EI, m/z): 344 (1, M^+), 312 (1), 279 (3), 253 (2), 191 (2), 167 (20), 149 (77), 113 (10), 104 (12), 91 (65), 70 (35), 57 (42), 57 (54), 43 (80), 41 (100); HRMS (ESI): $\text{C}_{21}\text{H}_{29}\text{O}_4$ (MH) $^+$, 345.2060 (calculated), 345.2065 (found).

2da: Yield: 21.3 mg (84%); ^1H NMR (400 MHz, C_6D_6) δ : 6.35 (d, $J = 16.0$ Hz, 1H, major), 6.34 (d, $J = 16.0$ Hz, 1H, minor), 5.48 (dd, $J = 16.0, 5.9$ Hz, 1H), 4.10–4.04 (m, 1H), 3.44–3.24 (m, 2H), 3.37 (s, 3H), 2.39 (td, $J = 7.6, 3.2$ Hz, 2H), 1.70 (s, 3H), 1.44 (quint, $J = 7.6$ Hz, 2H), 1.25 (sext, $J = 7.6$ Hz, 2H), 0.80 (t, $J = 7.6$ Hz, 3H, major), 0.80 (t, $J = 7.2$ Hz, 3H, minor); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, C_6D_6) δ : 213.6, 167.0, 129.2, 128.0, 103.2, 100.1, 72.8, 66.3, 51.4, 30.3, 28.9, 22.2, 14.5, 13.7; FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3384, 2954, 2928, 2871, 1940, 1715, 1436, 1379, 1265, 1240, 1131, 1076, 1017, 965, 875; LC-MS/MS: 255 (100, MH^+), 237 (20), 227 (8), 225 (20), 223 (42), 207 (12), 205 (50), 195 (12), 193 (15), 177 (18), 175 (18), 163 (15), 149 (8); HRMS (ESI): $\text{C}_{14}\text{H}_{23}\text{O}_4$ (MH) $^+$, 255.1596 (calculated), 255.1591 (found).

2ea: Yield: 30.2 mg (82%); ^1H NMR (400 MHz, C_6D_6) δ : 6.45 (dd, $J = 15.8, 1.6$ Hz, 1H, major), 6.34 (dd, $J = 15.8, 1.6$ Hz, 1H, minor), 5.54 (dd, $J = 15.8, 5.6$ Hz, 1H, major), 5.50 (dd, $J = 15.8, 5.6$ Hz, 1H, minor), 4.19–4.10 (m, 1H), 3.42 (dd, A of ABX, $J_{\text{AB}} = 9.6$ Hz, $J_{\text{AX}} = 4.4$ Hz, 1H), 3.37 (s, 3H), 3.31 (dd, B of ABX, $J_{\text{AB}} = 9.6$ Hz, $J_{\text{BX}} = 7.2$ Hz, 1H), 2.39 (td, $J = 7.2, 1.6$ Hz, 2H), 2.24 (d, $J = 3.6$ Hz, 1H), 1.72 (s, 3H, major), 1.69 (s, 3H, minor), 1.44 (quint, $J = 7.2$ Hz, 2H), 1.24 (sext, $J = 7.2$ Hz, 2H), 0.89 (s, 9H, minor), 0.85 (s, 9H, major), 0.79 (t, $J = 7.2$ Hz, 3H), –0.06 (s, 6H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, C_6D_6) δ : 213.5, 166.9, 129.2, 127.5, 103.1, 100.0, 72.4, 67.2, 51.3, 30.3, 28.9, 25.6, 22.2, 18.1, 14.5, 13.7, –5.6, –5.7; FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3465, 2954, 2928, 2858, 1940, 1716, 1463, 1435, 1361, 1258, 1118, 963, 837, 777; MS (EI, m/z): 368 (1, M^+), 339 (1), 311 (2), 279 (1), 249 (3), 219 (2), 191 (4), 175 (5), 159 (6), 145 (3), 121 (5), 117 (15), 105 (30), 91 (20), 77 (20), 75 (100), 73 (50), 57 (85), 41 (60); HRMS (ESI): $\text{C}_{20}\text{H}_{37}\text{O}_4\text{Si}$ (MH) $^+$, 369.2456 (calculated), 369.2470 (found).

2fa: Yield: 25.4 mg (90%); ^1H NMR (400 MHz, C_6D_6) δ : 6.29 (dd, $J = 15.7, 1.2$ Hz, 1H, minor), 6.28 (dd, $J = 15.7, 1.2$ Hz, 1H, major), 5.63 (dd, $J = 15.7, 6.6$ Hz, 1H, minor), 5.62 (dd, $J = 15.7, 6.6$ Hz, 1H, major), 3.73 (d, $J = 6.6$ Hz, 1H), 3.36 (s, 3H, major), 3.34 (s, 3H, minor), 2.38 (td, $J = 7.2, 1.6$ Hz, 2H), 2.15 (bs, 1H), 2.00 (bs, 1H), 1.71 (s, 3H, minor), 1.71 (s, 3H, major), 1.43 (quint, $J = 7.2$ Hz, 2H), 1.23 (sext, $J = 7.2$ Hz, 2H), 1.06 (s, 3H, major), 1.05 (s, 3H, minor), 1.03 (s, 3H, minor), 1.02 (s, 3H, major), 0.78 (t, $J = 7.2$ Hz, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, C_6D_6) δ : 213.5, 166.9, 129.3, 128.4, 103.2, 100.0, 79.2, 72.3, 51.3, 30.3, 28.8, 26.1, 23.8, 22.1, 14.5, 13.6; FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3407, 2956, 2929, 2860, 1940, 1714, 1436, 1377, 1265, 1242, 1133, 1015, 965, 898; MS (EI, m/z): 282 (2, M^+), 250 (2), 222 (1), 191 (3), 164 (4), 134 (4), 121 (20), 91 (7), 77 (16), 65 (5), 59 (100), 43 (37); HRMS (ESI): $\text{C}_{16}\text{H}_{27}\text{O}_4$ (MH) $^+$: 283.1904 (calculated), 283.1908 (found).

2ga: Yield: 26.3 mg (89%); ^1H NMR (400 MHz, C_6D_6) δ : 6.45 (dd, $J = 15.8, 1.6$ Hz, 1H), 5.68 (dd, $J = 15.8, 6.2$ Hz, 1H), 3.96 (d, $J = 6.2$ Hz, 1H), 3.36 (s, 3H, major), 3.33 (s, 3H, minor), 2.86 (s, 3H), 2.44 (bs, 1H), 2.38 (td, $J = 7.6, 1.6$ Hz, 2H), 1.72 (s, 3H), 1.43 (quint, $J = 7.6$ Hz, 2H), 1.23 (sext, $J = 7.6$ Hz, 2H), 0.93 (s, 3H), 0.88 (s, 3H), 0.78 (t, $J = 7.6$ Hz, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, C_6D_6) δ : 213.4, 166.9, 129.1, 103.2, 99.9, 77.6, 77.1, 51.3, 48.6, 30.3, 28.9, 22.1, 20.2, 19.2, 14.6, 13.7; FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3446, 2953, 2928, 2859, 1939,

1712, 1459, 1435, 1380, 1321, 1264, 1241, 1118, 1062, 1015, 965, 861, 833, 740, 713; MS (EI, m/z): 296 (1, M^+), 265 (1), 233 (1), 221 (1), 191 (1), 121 (3), 91 (5), 74 (4), 73 (100), 59 (5), 43 (20); HRMS (ESI): $C_{17}H_{29}O_4$ (MH) $^+$: 297.2060 (calculated), 297.2063 (found).

2ha: Yield: 22.1 mg (87%); 1H NMR (400 MHz, C_6D_6) δ : 6.24 (ddd, J = 15.2, 10.6, 1.6 Hz, 1H), 5.98 (dt, J = 10.6, 2.7 Hz, 1H), 5.46 (ddd, J = 15.2, 4.8, 0.8 Hz, 1H), 4.16–4.08 (m, 1H), 3.36 (s, 3H, major), 3.35 (s, 3H, minor), 2.99 (dd, A of ABX, J_{AB} = 9.6 Hz, J_{AX} = 4.0 Hz, 1H), 2.93 (dd, B of ABX, J_{AB} = 9.6 Hz, J_{BX} = 7.6 Hz, 1H), 2.93 (s, 3H), 2.41–2.29 (m, 2H), 2.20 (bs, 1H), 1.42 (quint, J = 7.2 Hz, 2H), 1.23 (sext, J = 7.2 Hz, 2H), 0.79 (t, J = 7.2 Hz, 3H, minor), 0.78 (t, J = 7.2 Hz, 3H, major); $^{13}C\{^1H\}$ NMR (100 MHz, C_6D_6) δ : 213.8, 166.6, 132.7, 124.1, 101.9, 97.0, 76.3, 70.3, 58.2, 51.3, 30.2, 28.6, 22.1, 13.6. FTIR (ν_{max}/cm^{-1}): 3423, 2955, 2930, 2872, 1941, 1716, 1436, 1322, 1268, 1133, 1062, 967, 853, 713; MS (EI, m/z): 254 (1, M^+), 223 (1), 209 (2), 177 (4), 149 (3), 135 (4), 125 (6), 107 (8), 91 (7), 77 (10), 65 (5), 59 (10), 45 (100); HRMS (ESI): $C_{14}H_{23}O_4$ (M) $^+$: 255.1591 (calculated), 255.1593 (found).

2ia: Yield: 26.4 mg (82%); 1H NMR (400 MHz, $CDCl_3$) δ : 6.08 (d, J = 15.8 Hz, 1H), 5.72 (dd, J = 15.8, 6.8 Hz, 1H), 4.16 (q, J = 6.8 Hz, 1H), 3.70 (s, 3H), 2.31–2.15 (m, 4H), 1.62–1.28 (m, 16H), 0.93 (m, 9H); $^{13}C\{^1H\}$ NMR (100 MHz, $CDCl_3$) δ : 213.1, 167.8, 132.8, 126.2, 108.2, 101.4, 73.1, 52.0, 37.0, 30.3, 29.6, 28.7, 28.3, 27.6, 22.6, 22.4, 22.3, 14.0, 13.9, 13.8; FTIR (ν_{max}/cm^{-1}): 3418, 2956, 2929, 2860, 1937, 1716, 1465, 1435, 1378, 1265, 1133, 1072, 964; MS (EI, m/z): 322 (1, M^+), 236 (3), 194 (5), 163 (5), 119 (5), 105 (6), 91 (17), 85 (12), 69 (30), 57 (50), 41 (100); HRMS (ESI): $C_{20}H_{35}O_3$ (MH) $^+$: 323.2581 (calculated), 323.2584 (found).

2ia': Yield: 25.5 mg (80%); 1H NMR (400 MHz, $CDCl_3$) δ : 6.08 (d, J = 15.7 Hz, 1H), 5.72 (dd, J = 15.7, 6.7 Hz, 1H), 4.17 (q, J = 6.7 Hz, 1H), 3.71 (s, 3H), 2.30–2.14 (m, 4H), 1.71–1.18 (m, 15H), 0.97–0.82 (m, 9H); $^{13}C\{^1H\}$ NMR (100 MHz, $CDCl_3$) δ : 213.1, 167.8, 132.8, 126.1, 108.2, 101.4, 73.0, 52.0, 37.1, 30.4, 29.6, 28.7, 28.4, 27.6, 22.6, 22.4, 22.3, 14.0, 13.9; FTIR (ν_{max}/cm^{-1}): 3422, 2956, 2928, 2859, 1937, 1716, 1465, 1435, 1379, 1264, 1132, 1074, 963; MS (EI, m/z): 322 (1, M^+), 236 (20), 231 (21), 205 (20), 195 (36), 194 (90), 179 (35), 177 (42), 165 (39), 163 (50), 152 (38), 135 (55), 121 (61), 107 (68), 105 (60), 93 (72), 91 (74), 85 (69), 79 (62), 57 (100), 55 (41); HRMS (ESI): $C_{20}H_{35}O_3$ (MH) $^+$: 323.2581 (calculated), 323.2584 (found).

2ja: Yield: 20.7 mg (87%); 1H NMR (400 MHz, C_6D_6) δ : 6.16 (dd, J = 15.6, 1.2 Hz, 1H), 5.50 (dd, J = 15.6, 6.2 Hz, 1H, minor), 5.49 (dd, J = 15.6, 6.2 Hz, 1H, major), 4.03 (quint, J = 6.2 Hz, 1H), 3.38 (s, 3H, major), 3.37 (s, 3H, minor), 2.39 (td, J = 7.4, 1.2 Hz, 2H), 1.70 (s, 3H), 1.49–1.41 (m, 2H), 1.25 (sext, J = 7.4 Hz, 2H), 1.04 (d, J = 6.2 Hz, 3H), 0.80 (t, J = 7.4 Hz, 3H, minor), 0.80 (t, J = 7.4 Hz, 3H, major); $^{13}C\{^1H\}$ NMR (100 MHz, C_6D_6) δ : 213.3, 166.9, 134.7, 125.2, 103.1, 99.9, 68.0, 51.2, 30.3, 28.9, 23.1, 22.1, 14.5, 13.6; FTIR (ν_{max}/cm^{-1}): 3410, 2956, 2928, 1941, 1715, 1436, 1322, 1266, 1135, 1062, 1016, 965, 714; MS (EI, m/z): 238 (3, M^+), 205 (5), 194 (15), 181 (15), 164 (13), 151 (55), 135 (45), 121 (100), 107 (30), 93 (90), 77 (45), 65 (10), 55 (14), 43 (92); HRMS (ESI): $C_{14}H_{23}O_3$ (MH) $^+$: 239.1648 (calculated), 239.1652 (found).

2ka: Yield: 29.9 mg (89%); 1H NMR (400 MHz, C_6D_6) δ : 6.27 (dd, J = 15.8, 1.4 Hz, 1H, major), 6.26 (dd, J = 15.8, 1.4 Hz, 1H, minor), 5.78 (dd, J = 15.8, 5.5 Hz, 1H), 4.27–4.23 (m, 1H), 3.38 (s, 3H, major), 3.37 (s, 3H, minor), 3.05 (dd, A of ABX, J_{AB} = 9.6 Hz, J_{AX} = 4.0 Hz, 1H), 3.02 (dd, B of ABX, J_{AB} = 9.6 Hz, J_{BX} = 7.2 Hz, 1H), 2.94 (s, 3H), 2.46–2.41 (m, 2H), 2.34 (bs, 1H), 2.16–2.09 (m, 1H), 2.03–1.99 (m, 1H), 1.92–1.87 (m, 1H), 1.66–1.60 (m, 2H), 1.54–1.45 (m, 3H), 1.33–1.01 (m, 7H), 0.81 (t, J = 7.2 Hz, 3H, major), 0.81 (t, J = 7.2 Hz, 3H, minor); ^{13}C NMR (100 MHz, C_6D_6) δ : 212.3, 167.1, 129.0, 125.9, 113.8, 102.7, 76.5, 70.8, 58.2, 51.3, 38.0, 32.8, 32.4, 30.6, 29.0, 26.4, 26.4, 26.2, 22.4, 13.7; FTIR (ν_{max}/cm^{-1}): 3426, 2925, 2853, 1924, 1713, 1435, 1322, 1263, 1240, 1195, 1173, 1119, 1062, 1016, 964, 890, 841, 713; MS (EI, m/z): 336 (2, M^+), 291 (60), 273 (8), 259 (100), 241 (20), 231 (30), 213 (27), 203 (30), 189 (19), 175 (21), 161 (22), 145 (21), 133 (21), 119 (19), 105 (25), 91 (47), 79 (33), 67 (21), 55 (45), 45 (65); HRMS (ESI): $C_{20}H_{33}O_4$ (MH) $^+$: 337.2380 (calculated), 337.2384 (found).

2la: Yield: 21.1 mg (68%); 1H NMR (400 MHz, C_6D_6) δ : 6.22 (dd, J = 15.6, 1.6 Hz, 1H, major), 6.21 (dd, J = 15.6, 1.6 Hz, 1H, minor), 5.92 (dd, J = 15.6, 5.2 Hz, 1H), 4.23–4.15 (m, 1H), 3.38 (s, 3H), 3.01 (dd, A of ABX, J_{AB} = 9.2 Hz, J_{AX} = 3.6 Hz, 1H), 2.95 (dd, B of ABX, J_{AB} = 9.2 Hz, J_{BX} = 7.6 Hz, 1H), 2.92 (s, 3H), 2.44–2.37 (m, 2H), 2.29 (bs, 1H), 1.51–1.46 (m, 2H), 1.26 (sext, J = 7.2 Hz, 2H), 1.07 (s, 9H), 0.81 (t, J = 7.2 Hz, 3H); $^{13}C\{^1H\}$ NMR (100 MHz, C_6D_6) δ : 208.6, 167.4, 132.0, 123.6, 117.1, 102.7, 76.4, 70.6, 58.2, 51.3, 33.8, 30.5, 30.1, 28.9, 22.4, 13.7; FTIR (ν_{max}/cm^{-1}): 3455, 2957, 2929, 2871, 1940, 1713, 1459, 1435, 1395, 1363, 1322, 1264, 1239, 1195, 1172, 1132, 1062, 1017, 963, 713; LC-MS/MS: 311 (67, M^+), 292 (19), 279 (80), 261 (27), 255 (10), 251 (5), 247 (10), 237 (9), 229 (60), 223 (100), 219 (8), 205 (8), 201 (9), 191 (8), 179 (26), 177 (8), 169 (10); HRMS (ESI): $C_{18}H_{31}O_4$ (MH) $^+$: 311.2223 (calculated), 311.2220 (found).

2ma: Yield: 17.2 mg (81%); 1H NMR (400 MHz, C_6D_6) δ : 6.29 (d, J = 16.0 Hz, 1H, major), 6.26 (d, J = 15.9 Hz, 1H, minor), 5.69–5.66 (m, 1H), 5.47 (ddd, J = 15.9, 5.2, 1.2 Hz, 1H), 4.17–4.12 (m, 1H), 3.32 (s, 3H, major), 3.31 (s, 3H, minor), 3.00 (dd, A of ABX, J_{AB} = 9.6 Hz, J_{AX} = 4.0 Hz, 1H), 2.95 (dd, B of ABX, J_{AB} = 9.6 Hz, J_{BX} = 7.2 Hz, 1H), 2.95 (s, 3H), 2.08 (bs, 1H), 1.55 (d, J = 2.7 Hz, 3H); $^{13}C\{^1H\}$ NMR (100 MHz, C_6D_6) δ : 216.1, 165.0, 130.2, 126.0, 103.5, 87.5, 76.3, 70.5, 58.2, 51.1, 14.0; FTIR (ν_{max}/cm^{-1}): 3429, 2925, 1944, 1721, 1438, 1395, 1260, 1196, 1153, 1126, 1028, 966, 840; MS (EI, m/z): 212 (2, M^+), 193 (1), 167 (3), 135 (6), 110 (6), 107 (11), 81 (5), 77 (20), 67 (6), 59 (11), 45 (100); HRMS (ESI): $C_{11}H_{17}O_4$ (MH) $^+$: 213.1121 (calculated), 213.1120 (found).

2na: Yield: 27.0 mg (92%); 1H NMR (400 MHz, C_6D_6) δ : 6.46 (dd, J = 15.7, 1.6 Hz, 1H), 5.53 (dd, J = 15.7, 5.5 Hz, 1H), 4.27–4.20 (m, 1H), 3.36 (s, 3H, major), 3.35 (s, 3H, minor), 3.05 (dd, A of ABX, J_{AB} = 9.6 Hz, J_{AX} = 4.0 Hz, 1H), 3.00 (dd, B of ABX, J_{AB} = 9.6 Hz, J_{BX} = 7.2 Hz, 1H), 2.96 (s, 3H), 2.64 (tt, J = 11.4, 3.4 Hz, 1H), 2.33 (bs, 1H), 1.95 (d, J = 11.4 Hz, 2H), 1.71 (s, 3H), 1.64–1.55 (m, 2H), 1.53–1.45 (m, 1H), 1.30–0.94 (m, 5H); ^{13}C NMR (100 MHz, C_6D_6) δ : 213.2, 166.6, 129.1, 105.9, 104.3, 76.5, 70.6, 58.2, 51.2, 37.6, 32.8, 26.4, 26.3, 26.0, 14.6; FTIR (ν_{max}/cm^{-1}): 3420, 2924, 2851, 1936, 1712, 1435, 1369, 1322, 1254, 1228, 1192, 1128, 1093, 1071, 1026, 997, 965, 891, 872, 780; MS (EI, m/z): 294 (2, M^+), 262 (1), 249 (2), 217 (3), 189 (1), 171 (4), 135 (2), 119 (3), 105 (5), 91 (6), 77 (6), 67 (4), 55 (8), 45 (100); HRMS (ESI): $C_{17}H_{27}O_4$ (MH) $^+$: 295.1904 (calculated), 295.1908 (found).

2oa: Yield: 23.9 mg (89%); 1H NMR (400 MHz, C_6D_6) δ : 6.43 (dd, J = 15.8, 1.6 Hz, 1H), 5.51 (dd, J = 15.8, 5.6 Hz, 1H), 4.25–4.20 (m, 1H), 3.32 (s, 3H, major), 3.30 (s, 3H, minor), 3.04 (dd, A of ABX, J_{AB} = 9.6 Hz, J_{AX} = 4.4 Hz, 1H), 2.99 (dd, B of ABX, J_{AB} = 9.6 Hz, J_{BX} = 7.2 Hz, 1H), 2.96 (s, 3H), 2.34 (bs, 1H), 1.68 (s, 3H), 1.29 (s, 9H); $^{13}C\{^1H\}$ NMR (100 MHz, C_6D_6) δ : 212.4, 165.9, 128.9, 108.8, 103.2, 76.5, 70.7, 58.2, 50.9, 34.2, 29.3, 14.6; FTIR (ν_{max}/cm^{-1}): 3419, 2955, 2870, 1934, 1717, 1435, 1362, 1322, 1258, 1226, 1132, 1062, 965; MS (EI, m/z): 268 (1, M^+), 236 (1), 223 (2), 191 (3), 163 (5), 145 (5), 135 (15), 119 (5), 105 (6), 91 (12), 79 (15), 65 (5), 59 (7), 57 (25), 45 (100); HRMS (ESI): $C_{15}H_{25}O_4$ (MH) $^+$: 269.1747 (calculated), 269.1748 (found).

2ob: Yield: 25.7 mg (91%); 1H NMR (400 MHz, C_6D_6) δ : 6.44 (dd, J = 15.5, 1.6 Hz, 1H, major), 6.42 (dd, J = 15.5, 1.6 Hz, 1H, minor), 5.52 (dd, J = 15.5, 6.0 Hz, 1H), 4.23–4.19 (m, 1H), 3.95 (qd, J = 7.2 Hz, 1.2 Hz, 2H), 3.03 (dd, A of ABX, J_{AB} = 9.6 Hz, J_{AX} = 4.0 Hz, 1H), 2.99 (dd, B of ABX, J_{AB} = 9.6 Hz, J_{BX} = 7.2 Hz, 1H), 2.95 (s, 3H), 2.32 (bs, 1H), 1.70 (s, 3H), 1.31 (s, 9H), 0.89 (t, J = 7.2 Hz, 3H); $^{13}C\{^1H\}$ NMR (100 MHz, $CDCl_3$) δ : 212.1, 166.2, 129.0, 127.4, 109.2, 103.1, 76.5, 71.1, 60.3, 59.1, 34.2, 29.5, 14.8, 14.2; FTIR (ν_{max}/cm^{-1}): 3420, 2959, 1939, 1713, 1459, 1366, 1322, 1255, 1223, 1112, 1060, 965; MS (EI, m/z): 282 (2, M^+), 249 (1), 237 (3), 208 (2), 191 (4), 163 (6), 135 (15), 105 (7), 91 (11), 77 (16), 65 (4), 57 (18), 45 (100); HRMS (ESI): $C_{16}H_{27}O_4$ (MH) $^+$: 283.1904 (calculated), 283.1908 (found).

2oe: Yield: 26.0 mg (88%); 1H NMR (400 MHz, C_6D_6) δ : 6.43 (dd, J = 15.6, 1.6 Hz, 1H), 5.53 (dd, J = 15.6, 5.6 Hz, 1H), 5.01 (sept, J = 6.4 Hz, 1H), 4.22–4.19 (m, 1H), 3.03 (dd, A of ABX, J_{AB} = 9.6 Hz, J_{AX} = 4.4 Hz, 1H), 2.99 (dd, B of ABX, J_{AB} = 9.6 Hz, J_{BX} = 7.6 Hz, 1H),

2.95 (s, 3H, major), 2.94 (s, 3H, minor), 2.26 (bs, 1H), 1.71 (s, 3H), 1.31 (s, 9H), 1.00 (d, $J = 6.4$ Hz, 3H), 0.99 (d, $J = 6.4$ Hz, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ : 211.8, 165.9, 129.1, 127.3, 109.7, 103.1, 76.5, 71.1, 67.8, 59.0, 34.1, 29.5, 21.8, 21.8, 14.7; FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3410, 2960, 2926, 2896, 2825, 1935, 1708, 1458, 1373, 1322, 1255, 1226, 1049, 1021, 964, 833, 783, 713; MS (EI, m/z): 296 (1, M^+), 278 (1), 251 (2), 236 (2), 191 (6), 163 (7), 135 (15), 105 (14), 91 (17), 77 (16), 65 (5), 57 (40), 45 (100); HRMS (ESI): $\text{C}_{17}\text{H}_{29}\text{O}_4$ (MH) $^+$: 297.2060 (calculated); 297.2064 (found).

2og: Yield: 25.9 mg (88%); ^1H NMR (400 MHz, C_6D_6) δ : 6.42 (dd, $J = 15.6$, 1.6 Hz, 1H, major), 6.40 (dd, $J = 15.6$, 1.6 Hz, 1H, minor), 5.64 (ddt, $J = 17.2$, 10.8, 4.8 Hz, 1H), 5.53 (dd, $J = 15.6$, 5.6 Hz, 1H), 5.08 (dq, $J = 17.2$, 1.6 Hz, 1H), 4.87 (dq, $J = 10.8$, 1.6 Hz, 1H), 4.48–4.37 (m, 2H), 4.24–4.19 (m, 1H), 3.03 (dd, A of ABX, $J_{\text{AB}} = 9.6$ Hz, $J_{\text{AX}} = 4.0$ Hz, 1H), 2.99 (dd, B of ABX, $J_{\text{AB}} = 9.6$ Hz, $J_{\text{BX}} = 7.2$ Hz, 1H), 2.95 (s, 3H, major), 2.94 (s, 3H, minor), 2.28 (bs, 1H), 1.69 (s, 3H), 1.30 (s, 9H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ : 212.4, 165.7, 132.5, 128.7, 127.7, 117.1, 108.9, 103.2, 76.5, 71.1, 64.7, 59.1, 34.2, 29.4, 14.8; FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3410, 2957, 2927, 1935, 1713, 1647, 1457, 1398, 1362, 1321, 1252, 1217, 1172, 1131, 1061, 1027, 1016, 964, 928, 836, 781, 713; MS (EI, m/z): 294 (1, M^+), 249 (3), 236 (2), 191 (4), 163 (5), 135 (6), 105 (5), 91 (10), 77 (9), 65 (3), 57 (16), 45 (100); HRMS (EI): $\text{C}_{17}\text{H}_{27}\text{O}_4$ (MH) $^+$: 295.1904 (calculated), 295.1907 (found).

2pa: Yield: 29.6 mg (86%); ^1H NMR (400 MHz, C_6D_6) δ : 7.17–7.05 (m, 5H), 6.43 (dd, $J = 15.8$, 1.6 Hz, 1H), 5.49 (dd, $J = 15.8$, 5.6 Hz, 1H), 4.28–4.24 (m, 1H), 4.20 (s, 2H, major), 4.17 (s, 2H, minor), 3.31 (s, 3H), 3.19 (dd, A of ABX, $J_{\text{AB}} = 9.2$ Hz, $J_{\text{AX}} = 3.6$ Hz, 1H), 3.12 (dd, B of ABX, $J_{\text{AB}} = 9.2$ Hz, $J_{\text{BX}} = 7.2$ Hz, 1H), 2.32 (bs, 1H), 1.67 (s, 3H), 1.34 (s, 9H, minor), 1.29 (s, 9H, major); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ : 212.4, 166.5, 137.8, 128.8, 128.5, 127.9, 127.8, 127.7, 108.8, 103.1, 74.0, 73.4, 71.2, 51.6, 34.2, 29.4, 14.8; FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3422, 2954, 2865, 1934, 1715, 1496, 1479, 1454, 1434, 1391, 1362, 1257, 1225, 1104, 1060, 1029, 965, 781, 737, 698; MS (EI, m/z): 344 (2, M^+), 311 (2), 296 (2), 223 (2), 191 (3), 168 (4), 135 (11), 107 (7), 91 (100), 57 (50), 41 (25); HRMS (ESI): $\text{C}_{21}\text{H}_{29}\text{O}_4$ (MH) $^+$: 345.2060 (calculated), 345.2068 (found).

2qa: Yield: 22.1 mg (87%); ^1H NMR (400 MHz, C_6D_6) δ : 6.32 (dd, $J = 15.8$, 1.6 Hz, 1H, major), 6.09 (dd, $J = 15.8$, 6.0 Hz, 1H, minor), 5.74 (dd, $J = 15.8$, 1.6 Hz, 1H, minor), 5.43 (dd, $J = 15.8$, 6.0 Hz, 1H, major), 4.03–3.99 (m, 1H), 3.35 (dd, A of ABX, $J_{\text{AB}} = 11.2$ Hz, $J_{\text{AX}} = 3.6$ Hz, 1H), 3.32 (s, 3H), 3.23 (dd, B of ABX, $J_{\text{AB}} = 11.2$ Hz, $J_{\text{BX}} = 7.2$ Hz, 1H), 2.44 (bs, 1H), 2.22 (bs, 1H), 1.67 (s, 3H, major), 1.57 (s, 3H, minor), 1.34 (s, 9H, minor), 1.29 (s, 9H, major); ^{13}C NMR (100 MHz, C_6D_6) δ : 212.4, 165.9, 128.9, 108.9, 103.2, 72.8, 66.3, 50.9, 34.1, 29.3, 14.6; NMR (100 MHz, CDCl_3) δ : 212.4, 166.5, 129.2, 127.9, 108.9, 103.1, 73.0, 66.4, 51.7, 34.2, 29.4, 14.8; FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3368, 2955, 2926, 2865, 1934, 1717, 1641, 1458, 1434, 1363, 1257, 1225, 1060, 1029, 965, 925, 862, 781; MS (EI, m/z): 254 (3, M^+), 207 (7), 194 (11), 179 (10), 148 (13), 119 (15), 105 (20), 91 (35), 73 (20), 59 (20), 57 (100), 41 (50); HRMS (EI): $\text{C}_{14}\text{H}_{23}\text{O}_4$ (MH) $^+$: 255.1591 (calculated), 255.1590 (found).

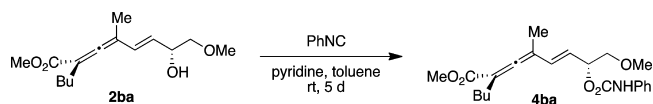
2ra: Yield: 30.9 mg (84%); ^1H NMR (400 MHz, C_6D_6) δ : 6.43 (dd, $J = 15.6$, 1.6 Hz, 1H, major), 6.41 (dd, $J = 15.6$, 1.6 Hz, 1H, minor), 5.53 (dd, $J = 15.6$, 5.6 Hz, 1H), 4.17–4.12 (m, 1H), 3.42 (dd, A of ABX, $J_{\text{AB}} = 10.0$ Hz, $J_{\text{AX}} = 4.0$ Hz, 1H), 3.32 (dd, B of ABX, $J_{\text{AB}} = 10.0$ Hz, $J_{\text{BX}} = 7.6$ Hz, 1H), 3.32 (s, 3H), 2.23 (bs, 1H), 1.71 (s, 3H), 1.29 (s, 9H), 0.85 (s, 9H), –0.06 (s, 6H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, C_6D_6) δ : 212.4, 165.9, 129.1, 108.8, 103.2, 72.5, 67.2, 50.9, 34.1, 29.3, 25.6, 18.1, 14.6, –5.6, –5.7; FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3467, 2954, 2927, 2857, 1935, 1718, 1462, 1434, 1362, 1256, 1225, 1111, 1062, 1005, 963, 837, 777; MS (EI, m/z): 368 (1, M^+), 351 (1), 311 (3), 279 (2), 237 (10), 223 (8), 191 (10), 177 (15), 163 (20), 159 (43), 145 (20), 135 (25), 105 (30), 89 (45), 75 (85), 73 (100), 57 (55), 41 (38); HRMS (ESI): $\text{C}_{20}\text{H}_{37}\text{O}_4\text{Si}$ (MH) $^+$: 369.2456 (calculated), 369.2463 (found).

2sa: Yield: 25.1 mg (87%); ^1H NMR (400 MHz, C_6D_6) δ : 7.71–7.67 (m, 2H), 7.14–7.11 (m, 2H), 7.03–6.99 (m, 1H), 6.41 (dd, $J = 15.8$, 1.2 Hz, 1H, major), 6.38 (dd, $J = 15.8$, 1.2 Hz, 1H, minor), 5.56 (dd, $J = 15.8$, 5.6 Hz, 1H), 4.21–4.16 (m, 1H), 3.38 (s, 3H, major), 3.36 (s, 3H, minor), 3.03–2.96 (m, 2H), 2.96 (s, 3H), 2.30 (bs, 1H),

1.68 (s, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, C_6D_6) δ : 215.9, 165.8, 133.2, 130.5, 128.7, 128.2, 127.5, 126.2, 104.4, 102.7, 76.4, 70.6, 58.2, 51.5, 14.2; FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3419, 2922, 2851, 1926, 1716, 1492, 1434, 1369, 1321, 1273, 1195, 1171, 1123, 1062, 1039, 964, 918, 898, 781, 694; MS (EI, m/z): 288 (2, M^+), 256 (3), 211 (5), 183 (5), 155 (17), 115 (8), 89 (4), 77 (9), 51 (5), 45 (100); HRMS (ESI): $\text{C}_{17}\text{H}_{21}\text{O}_4$ (MH) $^+$: 289.1434 (calculated), 289.1439 (found).

2ta: Yield: 25.4 mg (82%); ^1H NMR (400 MHz, C_6D_6) δ : 5.80 (d, $J = 8.2$, 1.6 Hz, 1H, minor), 5.75 (dt, $J = 8.2$, 1.6 Hz, 1H, major), 4.55–4.50 (m, 1H), 3.36 (s, 3H), 3.08–3.03 (m, 2H), 2.96 (s, 3H), 2.40 (td, $J = 7.6$, 2.8 Hz, 2H), 2.25–2.11 (m, 4H), 1.60–1.20 (m, 4H), 0.86–0.78 (m, 5H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, C_6D_6) δ : 207.1, 167.4, 137.2, 125.6, 109.3, 99.6, 76.3, 66.9, 58.3, 51.4, 30.7, 30.5, 28.9, 28.5, 25.6, 25.5, 22.2, 13.9; FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3456, 2928, 2857, 2859, 1946, 1713, 1436, 1436, 1266, 1241, 1128, 1063, 957; MS (EI, m/z): 308 (1, M^+), 276 (25), 231 (65), 201 (35), 175 (40), 161 (50), 117 (55), 91 (95), 77 (40), 55 (30), 45 (100); HRMS (ESI): $\text{C}_{18}\text{H}_{29}\text{O}_4$ (MH) $^+$: 310.2067 (calculated), 310.2067 (found).

Carbamate Derivatization of 2ba and Crystal Growth for X-ray Analysis. The mixture of vinylallene ester (45.2 mg, 0.2 mmol), phenyl isocyanate (31 mg, 0.26 mmol), and dry pyridine (15.8 mg, 0.2 mmol) in 2 mL of toluene was stirred at room temperature for 5 days. The mixture was taken into Et_2O and washed with brine, dried over MgSO_4 , and filtered. The solvent was removed in vacuum, and the crude residue was purified by column chromatography on silica gel (hexane– EtAc ; 5:1, yield: 0.62 g, 80%).²⁶ The pure white solid material was dissolved in a boiled hexane and gradually cooled down to 30 ° overnight.



4ba: M.P.: 67–69 °C; ^1H NMR (400 MHz, CDCl_3) δ : 7.38 (d, $J = 7.6$ Hz, 2H), 7.32–7.28 (m, 2H), 7.06 (tt, $J = 7.6$, 1.2 Hz, 1H), 6.72 (bs, 1H), 6.33 (dd, $J = 15.8$, 1.2 Hz, 1H), 5.70 (dd, $J = 15.8$ Hz, 6.4 Hz, 1H), 5.53 (qd, $J = 5.6$, 1.2 Hz, 1H), 3.70 (s, 3H), 3.60 (d, $J = 2.4$ Hz, 1H), 3.59 (d, $J = 0.8$ Hz, 1H), 3.42 (s, 3H), 2.25 (t, $J = 7.6$ Hz, 2H), 1.89 (s, 3H), 1.40–1.28 (m, 4H), 0.88 (t, $J = 7.6$ Hz, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ : 213.8, 167.5, 152.6, 137.7, 130.0, 129.1, 124.8, 123.5, 118.6, 103.0, 100.0, 74.1, 73.5, 59.3, 52.1, 30.1, 28.6, 22.1, 14.7, 13.9.

■ ASSOCIATED CONTENT

■ Supporting Information

X-ray crystallographic data of **4ba** and copies of ^1H NMR and ^{13}C NMR spectra for the substrates **1** and **1'**, the products **2** and **2'**, and **4ba**. The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.joc.5b00382.

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Notes

The authors declare no competing financial interest.

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■ DEDICATION

Dedicated to the memory of Prof. Dr. Ayhan S. Demir.

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- (5) CCDC 1019595 contains the supplementary crystallographic data of phenylcarbamate derivative of **2ba**. This data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.
- (6) Major diastereomeric products could usually be enriched by vigorous column purification. For instance, it has been possible to improve the dr of **2ja** to 99:1 by column chromatography.
- (7) The preparation of enyne oxiranes containing a phenyl group in the R² or R³ positions failed owing to their decomposition during epoxidation process.
- (8) Such a trend was also observed for the alkoxycarbonylation reactions of conjugated 2,4-enyne carbonates (ref 4c).
- (9) The mechanism of this method is in keeping with those of analogue reactions of 2,4-enyne carbonates (ref 4c).
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